# MEETING OF THE OIE

**TERRESTRIAL ANIMAL HEALTH STANDARDS COMMISSION**

Paris, 6–17 September 2010

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## List of participants

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*OIE Terrestrial Animal Health Standards Commission / September 2010*
MEETING OF THE OIE
TERRESTRIAL ANIMAL HEALTH STANDARDS COMMISSION

Paris, 6–17 September 2010

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Adopted agenda

A. MEETING OF THE DIRECTOR GENERAL WITH THE CODE COMMISSION AND THE SCIENTIFIC COMMISSION

Welcome - Director General

B. JOINT MEETING OF THE CODE COMMISSION AND THE SCIENTIFIC COMMISSION

Item 1 Glossary: new definition for wildlife; review of definition for infected zone

Item 2 OIE criteria for listing diseases: report of the ad hoc Group on Notification of Animal Diseases and Pathogenic Agents

Item 3 Foot and mouth disease
   a) Proposal for OIE to approve national FMD control strategies: next steps
   b) Compartmentalisation – development of a checklist or other guidance
   c) Trade in animal products (“commodities”) – next steps
   d) Others: update on beef casings; articles on fresh meat from free countries/zones

Item 4 Brucellosis: clarification on approach – Terrestrial Code chapters to be based on animal species (bovidae, small ruminants, suidae and camelidae) or on pathogen (B.abortus, B.melitensis, B.suis).

Item 5 Rabies: report of the ad hoc Group, revised chapter and next steps

Item 6 Scrapie: policy on the issue of animal genotype; and on notification of atypical scrapie

Item 7 Two documents: (1) Labelling of veterinary products and (2) Good Practices for the use of Veterinary Products
C. EXAMINATION OF MEMBER COMMENTS AND WORK OF RELEVANT EXPERT GROUPS

1. Update on reports of other commissions; harmonisation with the OIE *Aquatic Animal Health Code*; other relevant activities of the OIE (Conflict of Interest criteria) - President of the Commission

2. *Terrestrial Code* revision

Item 1 General comments

Item 2 Glossary

Item 3 Notification of diseases and epidemiological information and criteria for listing diseases (Chapters 1.1. and 1.2.)

Item 4 Animal health surveillance (Chapter 1.4.)

Item 5 Status for OIE listed diseases (Chapter 1.6.)

Item 6 OIE Import Risk Analysis Handbook

Item 7 Evaluation of Veterinary Services

   a) Revisions to Chapters 3.1. and 3.2.

   b) Global veterinary legislation initiative

Item 8 Design and implementation of identification systems to achieve animal traceability (Chapter 4.2.)

Item 9 Zoning and compartmentalisation

   a) Zoning and compartmentalisation (Chapter 4.3.)

   b) Application of compartmentalisation (Chapter 4.4.)

   c) Update on compartmentalisation projects supported by the OIE

Item 10 Semen and embryos

   a) Collection and processing of bovine, small ruminant and porcine semen (Chapter 4.6.)

   b) Collection and processing of *in vitro* produced embryos / oocytes from livestock and horses (Chapter 4.8.)
Item 11  Disposal of dead animals (Chapter 4.12.)

Item 12  Veterinary certificate
   a)  General obligations related to certification (Chapter 5.1.)
   b)  Certification procedures (Chapter 5.2.)

Item 13  Control of hazards of animal health and public health importance in animal feed (Chapter 6.3.)

Item 14  Control of OIE listed diseases in heat treated, shelf stable pet food (new draft Chapter)

Item 15  Salmonellosis
   a)  Prevention, detection and control of Salmonella in poultry (Chapter 6.5.)
   b)  Biosecurity procedures in poultry production (revised Chapter 6.4.)

Item 16  Introduction to the recommendations for controlling antimicrobial resistance (Chapter 6.7.)

Item 17  Animal welfare
   a)  Chapters on transport of animals (Chapters 7.3 and 7.4.)
   b)  Slaughter of animals (Chapter 7.5.)
   c)  Killing of animals for disease control purposes (Chapter 7.6.)
   d)  Stray dog population control (Chapter 7.7.)
   e)  Use of animals in research and education (Chapter 7.8.)
   f)  Report of the OIE Animal Welfare Working Group (June 2010 meeting)
   g)  Report of the ad hoc Group on Animal Welfare and Broiler Chicken Production Systems
   h)  Animal welfare and beef production systems
   i)  Proposal to use risk analysis principles in developing animal welfare standards
   j)  Guidelines on the establishment of OIE Regional Animal Welfare Strategies

Item 18  Anthrax (Chapter 8.1.)
Annex II (contd)

Item 19  Aujeszky's disease (Chapter 8.2.)
Item 20  Bluetongue (Chapter 8.3.)
Item 21  Foot and mouth disease
  a)  Chapter 8.5.
  b)  Revised FMD questionnaire
  c)  OIE recognition of a national FMD control programme
Item 22  Rabies
  a)  Chapter 8.10.
  b)  Model international veterinary certificate for dogs and cats originating from rabies infected countries (Chapter 5.11.)
Item 23  Vesicular stomatitis (Chapter 8.15.)
Item 24  Diseases of bees (Chapters 4.14., 9.1. – 6.)
Item 25  Avian influenza (Chapter 10.4.)
Item 26  Newcastle disease (Chapter 10.13.)
Item 27  Bovine brucellosis (Chapter 11.3.)
Item 28  Bovine spongiform encephalopathy (Chapter 11.5.)
Item 29  Bovine tuberculosis (Chapter 11.6.)
Item 30  Contagious bovine pleuropneumonia
Item 31  Lumpy skin disease (Chapter 11.12.)
Item 32  Equine diseases
  a)  African horse sickness status (Chapter 12.1.)
  b)  Equine influenza (Chapter 12.6.)
  c)  Equine viral arteritis (Chapter 12.9.)
Item 33  Enzootic abortion of ewes (Ovine chlamydiosis) (Chapter 14.5.)
Item 34  Scrapie (Chapter 14.9.)
Item 35  Classical swine fever (Chapter 15.2.)
Item 36  Swine vesicular disease (Chapter 15.4.)

Item 37  Report of the ad hoc Group on Communication

Item 38  Report of the ad hoc Group on Veterinary Education

3. Other issues

Item 39  OIE work programme on standard setting for foodborne pathogens

Item 40  Update on the OIE’s work on private standards

Item 41  Porcine reproductive and respiratory syndrome (PRRS)

Item 42  Future work programme of the Code Commission

Item 43  Other issues:

  Request for approval as an OIE Collaborating Centre on Animal Welfare (Sweden)
GLOSSARY

For the purposes of the Terrestrial Code:

**Antimicrobial agent**

means a naturally occurring, semi-synthetic or synthetic substance that at in vivo concentrations exhibits antimicrobial activity (kill or inhibit the growth of micro-organisms) at concentrations attainable in vivo. Anthelmintics and substances classed as disinfectants or antiseptics are excluded from this definition.

**Captive wild animal**

means an animal that has a phenotype not significantly affected by human selection but that are captive or otherwise live under supervision or control by humans.

**Euthanasia**

means the act of inducing death using a method that causes a rapid and irreversible loss of consciousness with minimum pain and distress to the animal.

**Feral animal**

means a previously domestic animal that now live without supervision, control by or dependence on humans.

**Veterinary legislation**

means laws, regulations and all associated legal instruments that pertain to the veterinary domain.

**Wild animal**

means an animal that has a phenotype unaffected by human selection and live independent of direct human supervision or control.

**Wildlife**

means any combination of feral animals, captive wild animals and wild animals.

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Annex IV

CHAPTER 1.1.

NOTIFICATION OF DISEASES AND EPIDEMIOLOGICAL INFORMATION

Article 1.1.1.
For the purposes of the Terrestrial Code and in terms of Articles 5, 9 and 10 of the OIE Organic Statutes, every OIE Member of the organization shall recognize the right of the Headquarters to communicate directly with the Veterinary Authority of its territory or territories.

All notifications and all information sent by the OIE to the Veterinary Authority shall be regarded as having been sent to the country concerned and all notifications and all information sent to the OIE by the Veterinary Authority shall be regarded as having been sent by the country concerned.

Article 1.1.2.
1. Members shall make available to other Members, through the OIE, whatever information is necessary to minimize the spread of important animal diseases and to assist in achieving better worldwide control of these diseases.

2. To achieve this, Members shall comply with the notification requirements specified in Article 1.1.3.

3. To assist in the clear and concise exchange of information, reports shall conform as closely as possible to the official OIE disease reporting format.

4. Recognizing that scientific knowledge concerning the relationship between disease agents and diseases is constantly developing and that the presence of an infectious agent does not necessarily imply the presence of a disease, Members shall ensure through their reports that they comply with the spirit and intention of point 1 above.

5. In addition to notifying new findings in accordance with Article 1.1.3., Members shall also provide information on the measures taken to prevent the spread of diseases; including quarantine measures and restrictions on the movement of animals, animal products and biological products and other miscellaneous objects which could by their nature be responsible for transmission of disease. In the case of diseases transmitted by vectors, the measures taken against such vectors shall also be specified.

Article 1.1.3.

Veterinary Authorities shall, under the responsibility of the national Delegate, send to the Headquarters:

1. in accordance with relevant provisions in the disease specific chapters, notification from the national Delegate to the OIE by telegram, fax or e-mail, within 24 hours, of any of the following events:

   a. first occurrence of a listed disease and/or infection in a country, a zone or a compartment;

   b. re-occurrence of a listed disease and/or infection in a country, a zone or a compartment following a report declared the outbreak ended;

   c. first occurrence of a new strain of a pathogen of an OIE listed disease in a country, a zone or a compartment;

   d. a sudden and unexpected increase in the distribution, incidence, morbidity or mortality of a listed disease prevalent within a country, a zone or a compartment;

   e. an emerging disease with significant morbidity or mortality, or zoonotic potential;
Annex IV (contd)

f. evidence of change in the epidemiology of a listed disease (including host range, pathogenicity, strain) in particular if there is a zoonotic impact;

2. weekly reports by telegram, fax or e-mail subsequent to a notification under point 1 above, to provide further information on the evolution of an incident which justified urgent notification; these reports should continue until the situation has been resolved through either the disease being eradicated or it becoming endemic so that six-monthly reporting under point 3 will satisfy the obligation of the Member to the OIE; in any case, a final report on the incident should be submitted;

3. a six-monthly report on the absence or presence, and evolution of listed diseases listed by the OIE and information of epidemiological significance to other Members;

4. an annual report concerning any other information of significance to other Members.

Article 1.1.4.

1. The Veterinary Authority of a territory in which an infected zone was located shall inform the Headquarters when this zone is free from the disease.

2. An infected zone for a particular disease shall be considered as such until a period exceeding the infective period specified in the Terrestrial Code has elapsed after the last reported case, and when full prophylactic and appropriate animal health measures have been applied to prevent possible reappearance or spread of the disease. These measures will be found in detail in the various chapters of Volume 2 of the Terrestrial Code.

3. A Member may be considered to regain freedom from a specific disease when all conditions given in the relevant chapters of the Terrestrial Code have been fulfilled.

4. The Veterinary Authority of a Member which sets up one or several free zones shall inform the OIE giving necessary details, including the criteria on which the free status is based, the requirements for maintaining the status and indicating clearly the location of the zones on a map of the territory of the Member.

Article 1.1.5.

1. The Headquarters shall send by telegram, fax, e-mail or Disease Information to the Veterinary Authorities concerned, all notifications received as provided in Articles 1.1.2. to 1.1.4.

2. The Headquarters shall dispatch to the Delegates information on new outbreaks of listed diseases.

3. The Headquarters, on the basis of information received and of any official communication, shall prepare an annual report concerning the application of the Terrestrial Code and its effects on international trade.

Article 1.1.6.

All telegrams or faxes sent by Veterinary Authorities in pursuance of Articles 1.1.3. and 1.1.5. shall receive priority in accordance with the circumstances. Communications by telephone, telegram or fax, sent in the case of exceptional urgency when there is danger of spread of a notifiable epizootic disease, shall be given the highest priority accorded to these communications by the International Arrangements of Telecommunications.

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**CHAPTER 1.2.**

**CRITERIA FOR LISTING DISEASES**

Article 1.2.1.

The criteria for the inclusion of a *disease* in the OIE List are as follows:

<table>
<thead>
<tr>
<th>Basic criteria</th>
<th>Parameters (at least one ‘yes’ answer means that the criterion has been met)</th>
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<tbody>
<tr>
<td><strong>International Spread</strong></td>
<td>Has international spread been proven on three or more occasions?  <strong>OR</strong> Are more than three countries with populations of susceptible animals free of the <em>disease</em> or facing impending freedom (based on the relevant provisions of the <em>Terrestrial Code</em>, and in particular those contained in Chapter 1.4.)?  <strong>OR</strong> Do OIE annual reports indicate that a significant number of countries with susceptible populations have reported absence of the <em>disease</em> for several consecutive years?</td>
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<tr>
<td><strong>Zoonotic Potential</strong></td>
<td>Has transmission to humans been proven? (with the exception of artificial circumstances)  <strong>AND</strong>  Is human infection associated with severe consequences? (death or prolonged illness)</td>
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<td><strong>Significant Spread within Naïve Populations</strong></td>
<td>Does the <em>disease</em> exhibit significant mortality at the level of a country or a zone?  <strong>OR</strong>  Does the <em>disease</em> exhibit significant morbidity at the level of a country or a zone?</td>
</tr>
<tr>
<td><strong>Emerging Diseases</strong></td>
<td>Are there apparent zoonotic properties or is there a rapid spread?</td>
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Article 1.2.2.

The criteria in Article 1.2.1. above are applied according to the decision-making model shown below:
The following diseases are included in the OIE List.

In case of modifications of this list of animal diseases adopted by the General Assembly, the new list comes into force on 1 January of the following year.

1. The following diseases are included within the category of multiple species diseases:

   - Anthrax
   - Aujeszky's disease
   - Bluetongue
   - Brucellosis (*Brucella abortus*)
   - Brucellosis (*Brucella melitensis*)
   - Brucellosis (*Brucella suis*)
   - Crimean Congo haemorrhagic fever
   - Echinococcosis/hydatidosis
   - Epizootic haemorrhagic disease
   - Equine encephalomyelitis (Eastern)
   - Foot and mouth disease
- Heartwater
- Japanese encephalitis
- Leptospirosis
- New world screwworm (*Cochliomyia hominivorax*)
- Old world screwworm (*Chrysomya bezziana*)
- Paratuberculosis
- Q fever
- Rabies
- Rift Valley fever
- Rinderpest
- Surra (*Trypanosoma evansi*)
- Trichinelllosis
- Tularemia
- Vesicular stomatitis
- West Nile fever.

2. The following *diseases* are included within the category of cattle *diseases*:
- Bovine anaplasmosis
- Bovine babesiosis
- Bovine genital campylobacteriosis
- Bovine spongiform encephalopathy
- Bovine tuberculosis
- Bovine viral diarrhoea
- Contagious bovine pleuropneumonia
- Enzootic bovine leukosis
- Haemorrhagic septicaemia
- Infectious bovine rhinotracheitis/infectious pustular vulvovaginitis
- Lumpy skin disease
- Theileriosis
- Trichomonosiosis
- Trypanosomosis (tsetse-transmitted).

3. The following *diseases* are included within the category of sheep and goat *diseases*:
- Caprine arthritis/encephalitis
- Contagious agalactia
- Contagious caprine pleuropneumonia
- Enzootic abortion of ewes (ovine chlamydiosis)
- Maedi–visna
Annex IV (contd)

- Nairobi sheep disease
- Ovine epididymitis (*Brucella ovis*)
- Peste des petits ruminants
- Salmonellosis (*S. abortusovis*)
- Scrapie
- Sheep pox and goat pox.

4. The following *diseases* are included within the category of equine *diseases*:
   - African horse sickness
   - Contagious equine metritis
   - Dourine
   - Equine encephalomyelitis (Western)
   - Equine infecti-us anaemia
   - Equine influenza
   - Equine piroplasmosis
   - Equine rhinopneumonitis
   - Equine viral arteritis
   - Glanders
   - Venezuelan equine encephalomyelitis.

5. The following *diseases* are included within the category of swine *diseases*:
   - African swine fever
   - Classical swine fever
   - Nipah virus encephalitis
   - Porcine cysticercosis
   - Porcine reproductive and respiratory syndrome
   - Swine vesicular disease
   - Teschovirus encephalomyelitis (under study)
   - Transmissible gastroenteritis.

6. The following *diseases* are included within the category of avian *diseases*:
   - Avian chlamydiosis
   - Avian infectious bronchitis
   - Avian infectious laryngotracheitis
   - Avian mycoplasmosis (*Mycoplasma gallisepticum*)
   - Avian mycoplasmosis (*Mycoplasma synoviae*)
   - Duck virus hepatitis
   - Fowl cholera
   - Fowl typhoid
- Highly pathogenic avian influenza in birds and low pathogenicity notifiable avian influenza in poultry as defined in Chapter 10.4.

- Infectious bursal disease (Gumboro disease)

- Marek’s disease

- Newcastle disease

- Pullorum disease

- Turkey rhinotracheitis.

7. The following diseases are included within the category of lagomorph diseases:

- Myxomatosis

- Rabbit haemorrhagic disease.

8. The following diseases are included within the category of bee diseases:

- Acarapisosis of honey bees

- American foulbrood of honey bees

- European foulbrood of honey bees

- Small hive beetle infestation (Aethina tumida)

- Tropilaelaps infestation of honey bees

- Varroosis of honey bees.

9. The following diseases are included within the category of other diseases:

- Camelpox

- Chronic wasting disease

- Leishmaniosis.
Annex IV (contd)

CHAPTER 10.6.

AVIAN TUBERCULOSIS

Article 10.6.1.

General provisions

Standards for diagnostic tests are described in the Terrestrial Manual.

Article 10.6.2.

Recommendations for the importation of birds for breeding or rearing

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the birds:

1. showed no clinical sign of avian tuberculosis on the day of shipment;

2. come from establishments which are regularly inspected by the Veterinary Authority and which are recognised as being free from avian tuberculosis.

Article 10.6.3.

Recommendations for the importation of birds for slaughter

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the birds:

1. showed no clinical sign of avian tuberculosis on the day of shipment;

2. come from establishments which are regularly inspected by the Veterinary Authority and are recognised as being free from avian tuberculosis; or

3. come from establishments in which no case of avian tuberculosis has been reported;

4. are not being eliminated as part of an eradication programme against avian tuberculosis.

Article 10.6.4.

Recommendations for the importation of wild avian species destined for zoological gardens
Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that prior to shipment, the birds showed no clinical sign of avian tuberculosis and, as far as can be determined, had not been exposed to avian tuberculosis.

Article 10.6.5.

Recommendations for the importation of hatching eggs

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the hatching eggs:

1. come from establishments and/or hatcheries which are regularly inspected by the Veterinary Authority;
2. come from establishments and/or hatcheries which are recognised as being free from avian tuberculosis;
3. were shipped in clean and unused packages.

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Annex IV (contd)

CHAPTER 10.7.

DUCK VIRUS ENTERITIS

Article 10.7.1.

General provisions

For the purposes of the Terrestrial Code, the incubation period for duck virus enteritis (DVE) shall be 7 days (chronic carriers occur).

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 10.7.2.

Recommendations for the importation of ducks

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the birds:

1. showed no clinical sign of DVE on the day of shipment;
2. come from establishments which are regularly inspected by the Veterinary Authority;
3. come from establishments which are recognised as being free from DVE;
4. have not been vaccinated against DVE; or
5. were vaccinated against DVE (the nature of the vaccine used and the date of vaccination should also be stated in the certificate).

Article 10.7.3.

Recommendations for the importation of day-old ducks

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the day-old birds:

1. come from establishments and/or hatcheries which are regularly inspected by the Veterinary Authority;
2. have not been vaccinated against DVE; or
3. were vaccinated against DVE (the nature of the vaccine used and the date of vaccination should also be stated in the certificate);
4. are the progeny of parent flocks which:
   a. come from establishments and/or hatcheries which are recognised as being free from DVE;
   b. come from establishments and/or hatcheries in which vaccination against DVE is not practised on the parent stock; or
   c. come from establishments and/or hatcheries in which vaccination against DVE is practised on the parent stock;
5. were shipped in clean and unused packages.
Annex IV (contd)

Recommendations for the importation of hatching eggs of ducks

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the hatching eggs:

1. have been disinfected in conformity with the standards referred to in Chapter 6.4;
2. come from establishments and/or hatcheries which are regularly inspected by the Veterinary Authority;
3. were shipped in clean and unused packages.

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Annex IV (contd)

CHAPTER 10.9.

FOWL CHOLERA

Article 10.9.1.

General provisions

For the purposes of the Terrestrial Code, the incubation period for fowl cholera (FC) shall be 14 days (chronic carriers occur).

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 10.9.2.

Recommendations for the importation of domestic birds

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the birds:

1. showed no clinical sign of FC on the day of shipment;
2. come from establishments which are regularly inspected by the Veterinary Authority;
3. come from establishments which are recognised as being free from FC;
4. have not been vaccinated against FC; or
5. were vaccinated against FC (the nature of the vaccine used and the date of vaccination should also be stated in the certificate).

Article 10.9.3.

Recommendations for the importation of day-old birds

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the day-old birds:

1. come from establishments and/or hatcheries which are regularly inspected by the Veterinary Authority;
2. have not been vaccinated against FC; or
3. were vaccinated against FC (the nature of the vaccine used and the date of vaccination shall also be stated in the certificate);
4. are the progeny of parent flocks which:
   a. come from establishments and/or hatcheries which are recognised as being free from FC;
   b. come from establishments and/or hatcheries in which vaccination against FC is not practised on the parent stock; or
   c. come from establishments and/or hatcheries in which vaccination against FC is practised on the parent stock;
5. were shipped in clean and unused packages.
Annex IV (contd)

Article 10.9.4.

Recommendations for the importation of hatching eggs of domestic birds

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the hatching eggs:

1. have been disinfected in conformity with the standards referred to in Chapter 6.4;
2. come from establishments and/or hatcheries which are regularly inspected by the Veterinary Authority;
3. were shipped in clean and unused packages.

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Annex IV (contd)

CHAPTER 10.12.

MAREK’S DISEASE

Article 10.12.1.

General provisions

For the purposes of the Terrestrial Code, the incubation period for Marek’s disease (MD) shall be 4 months.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 10.12.2.

Recommendations for the importation of chickens

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the birds:

1. showed no clinical sign of Marek’s disease on the day of shipment;
2. come from an establishment which is regularly inspected by the Veterinary Authority;
3. have not been vaccinated against MD and come from an establishment which has been free from MD for at least the past 2 years; or
4. were vaccinated against MD (the nature of the vaccine used and the date of vaccination should also be stated in the certificate).

Article 10.12.3.

Recommendations for the importation of day-old birds

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the day-old birds:

1. come from establishments which are regularly inspected by the Veterinary Authority and from hatcheries which comply with the standards referred to in Chapter 6.4;
2. were vaccinated against MD (the nature of the vaccine used and the date of vaccination should also be stated in the certificate);
3. were shipped in clean and unused packages.

Article 10.12.4.

Recommendations for the importation of hatching eggs of chickens

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the hatching eggs:

1. have been disinfected in conformity with the standards referred to in Chapter 6.4;
2. come from establishments which are regularly inspected by the Veterinary Authority and from hatcheries which comply with the standards referred to in Chapter 6.4.
3.come from establishments in which vaccination against MD is practised (the nature of the vaccine used and the date of vaccination should also be stated in the certificate);

4.were shipped in clean and unused packages.

Article 10.12.5.

Recommendations for the importation of meat-meals and feather-meals

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that these products have been processed using heat treatment to ensure the destruction of the MD virus.

Article 10.12.6.

Recommendations for the importation of feathers and down

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that these products have been processed to ensure the destruction of the MD virus.

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Annex IV (contd)

CHAPTER 15.5.

TESCHOVIRUS ENCEPHALOMYELITIS
(PREVIOUSLY ENTEROVIRUS ENCEPHALOMYELITIS, TESCHEN DISEASE, TALFAN DISEASE) (UNDER STUDY)

Article 15.5.1.

General provisions

For the purposes of the Terrestrial Code, the incubation period for teschovirus encephalomyelitis shall be 40 days.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 15.5.2.

Teschovirus encephalomyelitis free country

A country may be considered free from teschovirus encephalomyelitis when it has been shown that teschovirus encephalomyelitis has not been present for at least the past 3 years.

This period shall be 6 months after the slaughter of the last affected animal for countries in which a stamping-out policy is practised with or without vaccination against teschovirus encephalomyelitis.

Article 15.5.3.

Teschovirus encephalomyelitis infected zone

A zone shall be considered as infected with teschovirus encephalomyelitis until:

1. at least 40 days have elapsed after the confirmation of the last case and the completion of a stamping-out policy and disinfection procedures, or
2. 6 months have elapsed after the clinical recovery or death of the last affected animal if a stamping-out policy was not practised.

Article 15.5.4.

Recommendations for importation from teschovirus encephalomyelitis free countries

for domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of teschovirus encephalomyelitis on the day of shipment;
2. were kept in a country free from teschovirus encephalomyelitis since birth or for at least the past 40 days.

Article 15.5.5.

Recommendations for importation from teschovirus encephalomyelitis free countries
Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of teschovirus encephalomyelitis on the day of shipment;
2. come from a country free from teschovirus encephalomyelitis;
3. were kept in a quarantine station for the 40 days prior to shipment.

Recommendations for importation from countries considered infected with teschovirus encephalomyelitis

for domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of teschovirus encephalomyelitis on the day of shipment;
2. were kept since birth, or for the past 40 days, in an establishment where no case of teschovirus encephalomyelitis was officially reported during that period, and that the establishment of origin was not situated in a teschovirus encephalomyelitis infected zone, or
3. were kept in a quarantine station for the 40 days prior to shipment;
4. have not been vaccinated against teschovirus encephalomyelitis; or
5. were vaccinated against teschovirus encephalomyelitis, not less than 30 days and not more than one year prior to shipment (the nature of the vaccine used, whether inactivated or modified live virus, and the virus types and strains included shall also be stated in the certificate).

Recommendations for importation from countries considered infected with teschovirus encephalomyelitis

for wild pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of teschovirus encephalomyelitis on the day of shipment;
2. were kept in a quarantine station for the 40 days prior to shipment;
3. have not been vaccinated against teschovirus encephalomyelitis; or
4. were vaccinated against teschovirus encephalomyelitis, not less than 30 days and not more than one year prior to shipment (the nature of the vaccine used, whether inactivated or modified live virus, and the virus types and strains included shall also be stated in the certificate).
Annex IV (contd)

Article 15.5.8.

Recommendations for importation from teschovirus encephalomyelitis free countries
for semen of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the donor animals:

1. showed no clinical sign of teschovirus encephalomyelitis on the day of collection of the semen;
2. were kept in a country free from teschovirus encephalomyelitis for not less than 40 days prior to collection.

Article 15.5.9.

Recommendations for importation from countries considered infected with teschovirus encephalomyelitis
for semen of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the donor animals:

1. showed no clinical sign of teschovirus encephalomyelitis on the day of collection of the semen;
2. were kept in the exporting country, for the 40 days prior to collection, in an establishment or artificial insemination centre where no case of teschovirus encephalomyelitis was officially reported during that period, and that the establishment or artificial insemination centre was not situated in a teschovirus encephalomyelitis infected zone.

Article 15.5.10.

Recommendations for importation from teschovirus encephalomyelitis free countries
for fresh meat of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from animals:

1. which have been kept in a country free from teschovirus encephalomyelitis since birth or for at least the past 40 days;
2. which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections for teschovirus encephalomyelitis with favourable results.

Article 15.5.11.

Recommendations for importation from countries considered infected with teschovirus encephalomyelitis
for fresh meat of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from animals:

1. which have not been kept in a teschovirus encephalomyelitis infected zone;
2. which have been slaughtered in an approved abattoir not situated in a teschovirus encephalomyelitis infected zone and have been subjected to ante-mortem and post-mortem inspections for teschovirus encephalomyelitis with favourable results.
Annex IV (contd)

Article 15.5.12.

Recommendations for importation from countries considered infected with teschovirus encephalomyelitis

for meat products of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the entire consignment of meat products comes from animals which have been slaughtered in an approved abattoir and have been subjected to ante mortem and post mortem inspections for teschovirus encephalomyelitis with favourable results;
2. the meat products have been processed to ensure the destruction of the teschovirus encephalomyelitis virus;
3. the necessary precautions were taken after processing to avoid contact of the meat with any source of teschovirus encephalomyelitis virus.

Article 15.5.13.

Recommendations for importation from teschovirus encephalomyelitis free countries

for products of animal origin (from pigs) intended for use in animal feeding or for agricultural or industrial use

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that these products come from animals which have been kept in a country free from teschovirus encephalomyelitis since birth or for at least the past 40 days.

Article 15.5.14.

Recommendations for importation from countries considered infected with teschovirus encephalomyelitis

for meal and flour from blood, meat, defatted bones, hooves and claws (from pigs)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that these products have been processed using heat treatment to ensure the destruction of teschovirus encephalomyelitis virus.

Article 15.5.15.

Recommendations for importation from countries considered infected with teschovirus encephalomyelitis

for bristles

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that these products have been processed to ensure the destruction of teschovirus encephalomyelitis virus, in premises controlled and approved by the Veterinary Authority of the exporting country.

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OIE Terrestrial Animal Health Standards Commission / September 2010
CHAPTER 3.1.

VETERINARY SERVICES

Article 3.1.1.

The quality of the Veterinary Services depends on a set of factors, which include fundamental principles of an ethical, organisational, legislative, regulatory and technical nature. The Veterinary Services shall conform to these fundamental principles, regardless of the political, economic or social situation of their country.

Compliance with these fundamental principles by the Veterinary Services of a Member is important to the establishment and maintenance of confidence in its international veterinary certificates by the Veterinary Services of other Members.

The same fundamental principles should apply in countries where the responsibility for establishing or applying certain animal health or welfare measures, or issuing some international veterinary certificates is exercised by an organisation other than the Veterinary Services, or by an authority or agency on behalf of the Veterinary Services. In all cases, the Veterinary Services retain ultimate responsibility for the application of these principles.

These fundamental principles are presented in Article 3.1.2. Other factors affecting quality are described in Volume 1 of the Terrestrial Code (notification, principles of certification, etc.).

The quality of Veterinary Services, including veterinary legislation and regulations, can be measured through an evaluation, whose general principles are described in Article 3.1.3. and in Article 3.1.4.

Recommendations on the evaluation of Veterinary Services, including veterinary legislation, are described in Chapter 3.2.

A procedure for evaluating Veterinary Services by OIE experts, on a voluntary basis, is described in Article 3.1.5.

Article 3.1.2.

Fundamental principles of quality

The Veterinary Services shall comply with the following principles to ensure the quality of their activities:

1. Professional judgement

   The personnel of Veterinary Services should have the relevant qualifications, scientific expertise and experience to give them the competence to make sound professional judgements.

2. Independence

   Care should be taken to ensure that Veterinary Services' personnel are free from any commercial, financial, hierarchical, political or other pressures which might affect their judgement or decisions.

3. Impartiality

   The Veterinary Services should be impartial. In particular, all the parties affected by their activities have a right to expect their services to be delivered under reasonable and non-discriminatory conditions.
Annex V (contd)

4. **Integrity**

The *Veterinary Services* should guarantee that the work of each of their personnel is of a consistently high level of integrity. Any fraud, corruption or falsification should be identified and corrected.

5. **Objectivity**

The *Veterinary Services* should at all times act in an objective, transparent and non-discriminatory manner.

6. **Veterinary legislation**

Veterinary legislation is prerequisite to support good governance and provide the legal framework for all key activities of the *Veterinary Services*.

Legislation should be suitably flexible to allow for judgements of equivalence and efficient responses to changing situations. In particular, it should define and document the responsibilities and structure of the organisations in charge of the *animal identification system*, control of animal movements, animal disease control and reporting systems, epidemiological *surveillance* and communication of epidemiological information.

A similar demonstration should be made by *Veterinary Services* when they are in charge of veterinary public health activities.

7. **General organisation**

The *Veterinary Services* should be able to demonstrate by means of appropriate legislation, sufficient financial resources and effective organisation that they are in a position to have control of the establishment and application of animal health and *animal welfare* measures, and of international veterinary certification activities.

The *Veterinary Services* should have at their disposal effective systems for animal disease *surveillance* and for *notification* of disease problems wherever they occur, in accordance with the provisions of the *Terrestrial Code*. Adequate coverage of animal populations should also be demonstrated. They should at all times endeavour to improve their performance in terms of animal health information systems and animal disease control.

The *Veterinary Services* should define and document the responsibilities and structure of the organisation (in particular the chain of command) in charge of issuing *international veterinary certificates*.

Each position within the *Veterinary Services* which has an impact on their quality should be described. These job descriptions should include the requirements for education, training, technical knowledge and experience.

8. **Quality policy**

The *Veterinary Services* should define and document their policy and objectives for, and commitment to, quality, and should ensure that this policy is understood, implemented and maintained at all levels in the organisation. Where conditions allow, they may implement a quality system corresponding to their areas of activity and appropriate for the type, range and volume of work that they have to perform. The recommendations for the quality and evaluation of *Veterinary Services* propose a suitable reference system, which should be used if a Member choose to adopt a quality system.

9. **Procedures and standards**

The *Veterinary Services* should develop and document appropriate procedures and standards for all providers of relevant activities and associated facilities. These procedures and standards may for example relate to:
a) programming and management of activities, including international veterinary certification activities;

b) prevention, control and notification of disease outbreaks;

c) risk analysis, epidemiological surveillance and zoning;

d) inspection and sampling techniques;

e) diagnostic tests for animal diseases;

f) preparation, production, registration and control of biological products for use in the diagnosis or prevention of diseases;

g) border controls and import regulations;

h) disinfection and disinfestation;

i) treatments intended to destroy, if appropriate, pathogens in animal products.

Inasmuch as the OIE has adopted standards on these matters, the Veterinary Services should comply with these standards when applying animal health measures and when issuing international veterinary certificates.

10. Information, complaints and appeals

The Veterinary Authority should undertake to reply to legitimate requests from Veterinary Authorities of other Members or any other authority, in particular ensuring that any requests for information, complaints or appeals that they may present are dealt with in a timely manner.

A record should be maintained of all complaints and appeals and of the relevant action taken by the Veterinary Services.

11. Documentation

The Veterinary Services should have at their disposal a reliable and up-to-date documentation system suited to their activities.

12. Self-evaluation

The Veterinary Services should undertake periodical self-evaluation especially by documenting achievements against goals, and demonstrating the efficiency of their organisational components and resource adequacy.

A procedure for evaluating Veterinary Services by OIE experts, on a voluntary basis, is described in Article 3.1.5.

13. Communication

Veterinary Services should have effective internal and external systems of communication covering administrative and technical staff and parties affected by their activities.

14. Human and financial resources

Responsible authorities should ensure that adequate resources are made available to implement effectively the above activities.
Annex V (contd)

Article 3.1.3.

For the purposes of the Terrestrial Code, every Member should recognise the right of another Member to undertake, or request it to undertake, an evaluation of its Veterinary Services where the initiating Member is an actual or a prospective importer or exporter of commodities and where the evaluation is to be a component of a risk analysis process which is to be used to determine or review sanitary measures which apply to such trade.

Any evaluation of Veterinary Services should be conducted having regard to the OIE recommendations on the evaluation of Veterinary Services presented in Chapter 3.2.

A Member has the right to expect that the evaluation of its Veterinary Services will be conducted in an objective manner. A Member undertaking evaluation should be able to justify any measure taken as a consequence of its evaluation.

Article 3.1.4.

A Member which intends to conduct an evaluation of another Member's Veterinary Services should give them notice in writing. This notice should define the purpose of the evaluation and details of the information required.

On receipt of a formal request for information to enable an evaluation of its Veterinary Services by another Member, and following bilateral agreement of the evaluation process and criteria, a Member should expeditiously provide the other country with meaningful and accurate information of the type requested.

The evaluation process should take into account the fundamental principles and other factors of quality laid down in Article 3.1.1. and in Article 3.1.2. It should also take into consideration the specific circumstances regarding quality, as described in Article 3.1.1., prevailing in the countries concerned.

The outcome of the evaluation conducted by a Member should be provided in writing as soon as possible, and in any case within 4 months of receipt of the relevant information, to the Member which has undergone the evaluation. The evaluation report should detail any findings which affect trade prospects. The Member which conducts the evaluation should clarify in detail any points of the evaluation on request.

In the event of a dispute between two Members over the conduct or the conclusions of the evaluation of the Veterinary Services, the matter should be dealt with having regard to the procedures set out in Article 5.3.8.

Article 3.1.5.

Evaluation facilitated by OIE experts under the auspices of the OIE

The OIE has established procedures for the evaluation of the Veterinary Services of a Member, upon request by the Member.

The World Assembly of OIE Delegates endorses a list of approved experts to facilitate the evaluation process.

Under these procedures, the Director General of the OIE recommends an expert(s) from that list.
The expert(s) facilitate(s) the evaluation of the *Veterinary Services* of the Member based on the provisions in Chapter 3.2., using the OIE *Tool for the Evaluation of Performance of Veterinary Services* (OIE PVS Tool).

The expert(s) produce(s) a report in consultation with the *Veterinary Services* of the Member.

The report is submitted to the Director General of the OIE and, with the consent of the Member, published by the OIE.

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CHAPTER 3.2.

EVALUATION OF VETERINARY SERVICES

Article 3.2.1.

General considerations

1. Evaluation of Veterinary Services is an important element in the risk analysis process which countries may legitimately use in their policy formulations directly applying to animal health and sanitary controls of international trade in animals, animal-derived products, animal genetic material and animal feedstuffs.

Any evaluation should be carried out with due regard for Chapter 3.1.

2. In order to ensure that objectivity is maximised in the evaluation process, it is essential for some standards of discipline to be applied. The OIE has developed these recommendations which can be practically applied to the evaluation of Veterinary Services. These are relevant for evaluation of the Veterinary Services of one country by those of another country for the purposes of risk analysis in international trade. The recommendations are also applicable for evaluation by a country of its own Veterinary Services – the process known as self-evaluation – and for periodic re-evaluation. These recommendations should be used by OIE experts when facilitating an evaluation under the auspices of the OIE, following a request of a Member. In applying these recommendations on the evaluation, the OIE Tool for the Evaluation of Performance of Veterinary Services (OIE PVS Tool) should be used.

In carrying out a risk analysis prior to deciding the sanitary/zoosanitary conditions for the importation of a commodity, an importing country is justified in regarding its evaluation of the Veterinary Services of the exporting country as critical.

3. The purpose of evaluation may be either to assist a national authority in the decision-making process regarding priorities to be given to its own Veterinary Services (self-evaluation) or to assist the process of risk analysis in international trade in animals and animal-derived products to which official sanitary and/or zoosanitary controls apply.

4. In both situations, the evaluation should demonstrate that the Veterinary Services have the capability for effective control of the sanitary and zoosanitary status of animals and animal products. Key elements to be covered in this process include adequacy of resources, management capability, legislative and administrative infrastructures, independence in the exercise of official functions and history of performance, including disease reporting.

5. Good governance is the key to competence, integrity and confidence in organisations. Mutual confidence between relevant official Veterinary Services of trading partner countries contributes fundamentally to stability in international trade in animals and animal-related products. In this situation, scrutiny is directed more at the exporting country than at the importing country.

6. Although quantitative data can be provided on Veterinary Services, the ultimate evaluation will be essentially qualitative. While it is appropriate to evaluate resources and infrastructure (organisational, administrative and legislative), it is also appropriate to place emphasis on the evaluation of the quality of outputs and performance of Veterinary Services. Evaluation should take into consideration any quality systems used by Veterinary Services.
Annex V (contd)

7. An importing country has a right of assurance that information on sanitary/zoosanitary situations provided by the Veterinary Services of an exporting country is objective, meaningful and correct. Furthermore, the Veterinary Services of the importing country are entitled to expect validity in the veterinary certification of export.

8. An exporting country is entitled to expect that its animals and animal products will receive reasonable and valid treatment when they are subjected to import inspection in the country of destination. The country should also be able to expect that any evaluation of its standards and performance will be conducted on a non-discriminatory basis. The importing country should be prepared and able to defend any position which it takes as a consequence of the evaluation.

9. As the veterinary statutory body is not a part of the Veterinary Services, an evaluation of that body should be carried out to ensure that the registration/licensing of veterinarians and authorisation of veterinary para-professionals is included.

Article 3.2.2.

Scope

1. In the evaluation of Veterinary Services, the following items may be considered, depending on the purpose of the evaluation:

   - organisation, structure and authority of the Veterinary Services;
   - human resources;
   - material (including financial) resources;
   - veterinary legislation, regulatory frameworks and functional capabilities;
   - animal health, animal welfare and veterinary public health controls;
   - formal quality systems including quality policy;
   - performance assessment and audit programmes;
   - participation in OIE activities and compliance with OIE Members’ obligations.

2. To complement the evaluation of Veterinary Services, the legislative and regulatory framework, the organisational structure and functioning of the veterinary statutory body should also be considered.

3. Article 3.2.14. outlines appropriate information requirements for:

   - self-evaluation by the Veterinary Authority which perceives a need to prepare information for national or international purposes;
   - evaluation by a prospective or actual importing country of the Veterinary Services of a prospective or actual exporting country;
   - verification or re-verification of an evaluation in the course of a visit to the exporting country by the importing country;
   - evaluation by third parties such as OIE PVS experts or regional organisations.
Evaluation criteria for the organisational structure of the Veterinary Services

1. A key element in the evaluation is the study of the organisation and structure of the official Veterinary Services. The Veterinary Services should define and set out their policy, objectives and commitment to quality systems and standards. These organisational and policy statements should be described in detail. Organisational charts and details of functional responsibilities of staff should be available for evaluation. The role and responsibility of the Chief Veterinary Officer/Veterinary Director should be clearly defined. Lines of command should also be described.

2. The organisational structure should also clearly set out the interface relationships of government Ministers and departmental Authorities with the Chief Veterinary Officer/Veterinary Director and the Veterinary Services. Formal relationships with statutory authorities and with industry organisations and associations should also be described. It is recognised that Services may be subject to changes in structure from time to time. Major changes should be notified to trading partners so that the effects of re-structuring may be assessed.

3. Organisational components of Veterinary Services which have responsibility for key functional capabilities should be identified. These capabilities include epidemiological surveillance, disease control, import controls, animal disease reporting systems, animal identification systems, traceability systems, animal movement control systems, communication of epidemiological information, training, inspection and certification. Laboratory and field systems and their organisational relationships should be described.

4. To reinforce the reliability and credibility of their services, the Veterinary Services may have set up quality systems that correspond with their fields of activity and to the nature and scale of activities that they carry out. Evaluation of such systems should be as objective as possible.

5. The Veterinary Authority alone speaks for the country as far as official international dialogue is concerned. This is also particularly important to cases where zoning and compartmentalisation are being applied. The responsibilities of the Veterinary Authority should be made clear in the process of evaluation of Veterinary Services.

6. The Veterinary Authority is defined in the Glossary of the Terrestrial Code. As some countries have some relevant roles of the Veterinary Authority vested in autonomous sub-national (state/provincial, municipal) government bodies, there is an important need to assess the role and function of these Services. Details of their roles, relationship (legal and administrative) to each other and to the Veterinary Authority should be available for evaluation. Annual reports, review findings and access to other information pertinent to the animal health activities of such bodies should also be available.

7. Similarly, where the Veterinary Authority has arrangements with other providers of relevant services such as universities, laboratories, information services, etc., these arrangements should also be described. For the purposes of evaluation, it is appropriate to expect that the organisational and functional standards that apply to the Veterinary Authority should also apply to the service providers.
Evaluation criteria for quality systems

1. The *Veterinary Services* should demonstrate a commitment to the quality of the processes and outputs of their services. Where services or components of services are delivered under a formal quality systems programme which is based on OIE recommended standards or, especially in the case of laboratory components of *Veterinary Services* other internationally recognised quality standards, the *Veterinary Services* undergoing evaluation should make available evidence of accreditation, details of the documented quality processes and documented outcomes of all relevant audits undertaken.

2. Where the *Veterinary Services* undergoing evaluation make large use of formal quality systems in the delivery of their services, it is appropriate that greater emphasis be placed on the outcomes of evaluation of these quality systems than on the resource and infrastructural components of the services.

Evaluation criteria for human resources

1. The *Veterinary Services* should demonstrate that their human resource component includes an integral core of full-time civil service employees. This core should always include *veterinarians*. It should also include administrative officials and *veterinary para-professionals*. The human resources may also include part-time and private sector *veterinarians* and *veterinary para-professionals*. It is essential that all the above categories of personnel be subject to legal disciplinary provisions. Data relating to the resource base of the *Veterinary Services* undergoing evaluation should be available.

2. In addition to raw quantitative data on this resource base, the functions of the various categories of personnel in the *Veterinary Services* should be described in detail. This is necessary for analysis and estimation of the appropriateness of the application of qualified skills to the tasks undertaken by the *Veterinary Services* and may be relevant, for example, to the roles of *veterinarians* and *veterinary para-professionals* in field services. In this case, the evaluation should provide assurances that *disease* monitoring is being conducted by a sufficient number of qualified, experienced field *veterinarians* who are directly involved in farm visits; there should not be an over-reliance on *veterinary para-professionals* for this task.

3. Analysis of these data can be used to estimate the potential of the *Veterinary Services* to have reliable knowledge of the state of animal health in the country and to support an optimal level of animal disease control programmes. A large population of private *veterinarians* would not provide the *Veterinary Services* with an effective epizootiological information base without legislative (e.g. compulsory reporting of *notifiable diseases*) and administrative (e.g. official animal health surveillance and reporting systems) mechanisms in place.

4. These data should be assessed in close conjunction with the other information described in this chapter. For example, a large field staff (*veterinarians* and *veterinary para-professionals*) need fixed, mobile and budgetary resources for animal health activities in the livestock farming territory of the country. If deficiencies are evident, there would be reason to challenge the validity of epizootiological information.
Evaluation criteria for material resources

1. **Financial**

   Actual yearly budgetary information regarding the *Veterinary Services* should be available and should include the details set out in the model questionnaire outlined in Article 3.2.14. Information is required on conditions of service for veterinary staff (including salaries and incentives), and should provide a comparison with the private sector and perhaps with other professionals. Information should also be available on non-government sources of revenue available to *veterinarians* in their official responsibilities.

2. **Administrative**

   a. **Accommodation**

      The *Veterinary Services* should be accommodated in premises suitable for efficient performance of their functions. The component parts of the *Veterinary Services* should be located as closely as possible to each other at the central level, and in the regions where they are represented, in order to facilitate efficient internal communication and function.

   b. **Communications**

      The *Veterinary Services* should be able to demonstrate that they have reliable access to effective communications systems, especially for animal health surveillance and control programmes. Inadequate communications systems within the field services components of these programmes or between outlying offices and headquarters, or between the *Veterinary Services* and other relevant administrative and professional services, signify an inherent weakness in these programmes. Adequate communications systems between laboratories and between field and laboratory components of the *Veterinary Services* should also be demonstrated.

      Examples of types of communications which should be routinely available on an adequate country-wide basis are national postal, freight and telephone networks. Rapid courier services, facsimile and electronic data interchange systems (e.g. e-mail and Internet services) are examples of useful communication services which, if available, can supplement or replace the others. A means for rapid international communication should be available to the *Veterinary Authority*, to permit reporting of changes in national disease status consistent with OIE recommendations and to allow bilateral contact on urgent matters with counterpart *Veterinary Authorities* in trading-partner countries.

   c. **Transport systems**

      The availability of sufficient reliable transport facilities is essential for the performance of many functions of *Veterinary Services*. This applies particularly to the field services components of animal health activities (e.g. emergency response visits). Otherwise, the *Veterinary Services* cannot assure counterpart services in other countries that they are in control of the animal health situation within the country.

      Appropriate means of transport are also vital for the satisfactory receipt of samples to be tested at veterinary laboratories, for inspection of imports and exports, and for the performance of *animals* and animal product inspection in outlying production or processing establishments.
3. **Technical**

Details available on laboratories should include resources data, programmes under way as well as those recently completed and review reports on the role or functions of the laboratory. Information as described in the model questionnaire should be used in the evaluation of laboratory services.

a. **Cold chain for laboratory samples and veterinary medicines**

Adequate refrigeration and freezing systems should be available and should be used throughout the country to provide suitable low temperature protection for laboratory samples in transit or awaiting analysis, as well as veterinary medical products (e.g. vaccines) when these are required for use in animal disease control programmes. If these assurances cannot be given, it may be valid to discount many types of test results, as well as the effectiveness of certain disease control programmes and the export inspection system in the country undergoing evaluation.

b. **Diagnostic laboratories**

Analysis of the laboratory service component of *Veterinary Services*, which would include official governmental laboratories and other laboratories accredited by the *Veterinary Services* for specified purposes, is an essential element of the evaluation process. The quality of the veterinary diagnostic laboratories of a country underpins the whole control and certification processes of the zoosanitary/sanitary status of exported animals and animal products, and therefore these laboratories should be subject to rigid quality assurance procedures and should use international quality assurance programmes (wherever available) for standardising test methodologies and testing proficiency. An example is the use of International Standard Sera for standardising reagents.

This emphasis is valid whether one relates it to the actual testing performed on individual export consignments or to the more broad and ongoing testing regimes which are used to determine the animal health and veterinary public health profiles of the country and to support its disease control programmes. For the purposes of evaluation, veterinary diagnostic laboratories include those which are concerned with either animal health or veterinary public health activities. The *Veterinary Services* should approve and designate these laboratories for such purposes and have them audited regularly.

c. **Research**

The scope of animal disease and veterinary public health problems in the country concerned, the stages reached in the controls which address those problems and their relative importance can be measured to some degree by analysis of information on government priorities and programmes for research in animal health. This information should be accessible for evaluation purposes.

**Article 3.2.7.**

**Legislation and functional capabilities**

1. **Animal health, animal welfare and veterinary public health**

The *Veterinary Authority* should be able to demonstrate that it has the capacity, supported by appropriate legislation, to exercise control over all animal health matters. These controls should include, where appropriate, compulsory notification of prescribed animal diseases, inspection, movement controls through systems which provide adequate traceability, registration of facilities,
quarantine of infected premises/areas, testing, treatment, destruction of infected animals or contaminated materials, controls over the use of veterinary medicines, etc. The scope of the legislative controls should include domestic animals and their reproductive material, animal products, wildlife as it relates to the transmission of disease to humans and domestic animals, and other products subject to veterinary inspection. Arrangements should exist for co-operation with the Veterinary Authorities of the neighbouring countries for the control of animal diseases in border areas and for establishing linkages to recognise and regulate transboundary activities. Within the structure of Veterinary Services, there should be appropriately qualified personnel whose responsibilities include animal welfare. Information on the veterinary public health legislation covering the production of products of animal origin for national consumption may be also considered in the evaluation.

2. **Export/import inspection**

The Veterinary Authority should have appropriate legislation and adequate capabilities to prescribe the methods for control and to exercise systematic control over the import and export processes of animals and animal products in so far as this control relates to sanitary and zoosanitary matters. The evaluation should also involve the consideration of administrative instructions to ensure the enforcement of importing country requirements during the pre-export period.

In the context of production for export of foodstuffs of animal origin, the Veterinary Authority should demonstrate that comprehensive legislative provisions are available for the oversight by the relevant authorities of the hygienic process and to support official inspection systems of these commodities which function to standards consistent with or equivalent to relevant Codex Alimentarius and OIE standards.

Control systems should be in place which permit the exporting Veterinary Authority to approve export premises. The Veterinary Services should also be able to conduct testing and treatment as well as to exercise controls over the movement, handling and storage of exports and to make inspections at any stage of the export process. The product scope of this export legislation should include, *inter alia*, animals and animal products (including animal semen, ova and embryos), and animal feedstuffs.

The Veterinary Authority should be able to demonstrate that they have adequate capabilities and legislative support for zoosanitary control of imports and transit of animals, animal products and other materials which may introduce animal diseases. This could be necessary to support claims by the Veterinary Services that the animal health status of the country is suitably stable, and that cross-contamination of exports from imports of unknown or less favourable zoosanitary status is unlikely. The same considerations should apply in respect of veterinary control of public health. The Veterinary Services should be able to demonstrate that there is no conflict of interest when certifying veterinarians are performing official duties.

Legislation should also provide the right to deny and/or withdraw official certification. Penalty provisions applying to malpractice on the part of certifying officials should be included.

The Veterinary Services should demonstrate that they are capable of providing accurate and valid certification for exports of animals and animal products, based on Chapters 5.1. and 5.2. of the Terrestrial Code. They should have appropriately organised procedures which ensure that sanitary/animal health certificates are issued by efficient and secure methods. The documentation control system should be able to correlate reliably the certification details with the relevant export consignments and with any inspections to which the consignments were subjected.
Annex V (contd)

Security in the export certification process, including electronic documentation transfer, is important. A system of independent compliance review is desirable, to safeguard against fraud in certification by officials and by private individuals or corporations. The certifying veterinarian should have no conflict of interest in the commercial aspects of the animals or animal product being certified and be independent from the commercial parties.

Article 3.2.8.

Animal health controls

1. Animal health status

An updated assessment of the present animal disease status of a country is an important and necessary procedure. For this undertaking, studies of the OIE publications such as World Animal Health, the Bulletin and Disease Information should be fundamental reference points. The evaluation should consider the recent history of the compliance of the country with its obligations regarding international notification of animal diseases. In the case of an OIE Member, failure to provide the necessary animal health reports consistent with OIE requirements will detract from the overall outcome of the evaluation of the country.

An exporting country should be able to provide further, detailed elaboration of any elements of its animal disease status as reported to the OIE. This additional information will have particular importance in the case of animal diseases which are foreign to or strictly controlled in the importing country or region. The ability of the Veterinary Services to substantiate elements of their animal disease status reports with surveillance data, results of monitoring programmes and details of disease history is highly relevant to the evaluation. In the case of evaluation of the Veterinary Services of an exporting country for international trade purposes, an importing country should be able to demonstrate the reasonableness of its request and expectations in this process.

2. Animal health control

Details of current animal disease control programmes should be considered in the evaluation. These programmes would include epidemiological surveillance, official government-administered or officially-endorsed, industry-administered control or eradication programmes for specific diseases or disease complexes, and animal disease emergency preparedness. Details should include enabling legislation, programme plans for epidemiological surveillance and animal disease emergency responses, quarantine arrangements for infected and exposed animals or herds, compensation provisions for animal owners affected by disease control measures, training programmes, physical and other barriers between the free country or zone and those infected, incidence and prevalence data, resource commitments, interim results and programme review reports.

3. National animal disease reporting systems

The presence of a functional animal disease reporting system which covers all agricultural regions of the country and all veterinary administrative control areas should be demonstrated.

An acceptable variation would be the application of this principle to specific zones of the country. In this case also, the animal disease reporting system should cover each of these zones. Other factors should come to bear on this situation, e.g. the ability to satisfy trading partners that sound animal health controls exist to prevent the introduction of disease or export products from regions of lesser veterinary control.
Veterinary public health controls

1. **Food hygiene**

   The *Veterinary Authority* should be able to demonstrate effective responsibility for the veterinary public health programmes relating to the production and processing of animal products. If the *Veterinary Authority* does not exercise responsibility over these programmes, the evaluation should include a comprehensive review of the role and relationship of the organisations (national, state/provincial, and municipal) which are involved. In such a case, the evaluation should consider whether the *Veterinary Authority* can provide guarantees of responsibility for an effective control of the sanitary status of animal products throughout the slaughter, processing, transport and storage periods.

2. **Zoonoses**

   Within the structure of *Veterinary Services*, there should be appropriately qualified personnel whose responsibilities include the monitoring and control of zoonotic diseases and, where appropriate, liaison with medical authorities.

3. **Chemical residue testing programmes**

   Adequacy of controls over chemical residues in exported *animals*, animal products and feedstuffs should be demonstrated. Statistically-based *surveillance* and monitoring programmes for environmental and other chemical contaminants in *animals*, in animal-derived foods and in animal feedstuffs should be favourably noted. These programmes should be coordinated nationwide. Correlated results should be freely available on request to existing and prospective trading partner countries. Analytical methods and result reporting should be consistent with internationally recognised standards. If official responsibility for these programmes does not rest with the *Veterinary Services*, there should be appropriate provision to ensure that the results of such programmes are made available to the *Veterinary Services* for assessment. This process should be consistent with the standards set by the Codex Alimentarius Commission or with alternative requirements set by the importing country where the latter are scientifically justified.

4. **Veterinary medicines**

   It should be acknowledged that primary control over veterinary medicinal products may not rest with the *Veterinary Authority* in some countries, owing to differences between governments in the division of legislative responsibilities. However, for the purpose of evaluation, the *Veterinary Authority* should be able to demonstrate the existence of effective controls (including nationwide consistency of application) over the manufacture, importation, export, registration, supply, sale and use of veterinary medicines, biologicals and diagnostic reagents, whatever their origin. The control of veterinary medicines has direct relevance to the areas of animal health and public health.

   In the animal health sphere, this has particular application to biological products. Inadequate controls on the registration and use of biological products leave the *Veterinary Services* open to challenge over the quality of animal disease control programmes and over safeguards against *animal disease* introduction in imported veterinary biological products.
Annex V (contd)

It is valid, for evaluation purposes, to seek assurances of effective government controls over veterinary medicines in so far as these relate to the public health risks associated with residues of these chemicals in animals and animal-derived foodstuffs. This process should be consistent with the standards set by the Codex Alimentarius Commission or with alternative requirements set by the importing country where the latter are scientifically justified.

5. Integration between animal health controls and veterinary public health

The existence of any organised programme which incorporates a structured system of information feedback from inspection in establishments producing products of animal origin, in particular meat or dairy products, and applies this in animal health control should be favourably noted. Such programmes should be integrated within a national disease surveillance scheme.

Veterinary Services which direct a significant element of their animal health programmes specifically towards minimising microbial and chemical contamination of animal-derived products in the human food chain should receive favourable recognition in the evaluation. There should be evident linkage between these programmes and the official control of veterinary medicines and relevant agricultural chemicals.

Article 3.2.10.

Performance assessment and audit programmes

1. Strategic plans

The objectives and priorities of the Veterinary Services can be well evaluated if there is a published official strategic plan which is regularly updated. Understanding of functional activities is enhanced if an operational plan is maintained within the context of the strategic plan. The strategic and operational plans, if these exist, should be included in the evaluation.

Veterinary Services which use strategic and operational plans may be better able to demonstrate effective management than countries without such plans.

2. Performance assessment

If a strategic plan is used, it is desirable to have a process which allows the organisation to assess its own performance against its objectives. Performance indicators and the outcomes of any review to measure achievements against pre-determined performance indicators should be available for evaluation. The results should be considered in the evaluation process.

3. Compliance

Matters which can compromise compliance and adversely affect a favourable evaluation include instances of inaccurate or misleading official certification, evidence of fraud, corruption, or interference by higher political levels in international veterinary certification, and lack of resources and poor infrastructure.

It is desirable that the Veterinary Services contain (or have a formal linkage with) an independent internal unit/section/commission the function of which is to critically scrutinise their operations. The aim of this unit should be to ensure consistent and high integrity in the work of the individual officials in the Veterinary Services and of the corporate body itself. The existence of such a body can be important to the establishment of international confidence in the Veterinary Services.
An important feature when demonstrating the integrity of the Veterinary Services is their ability to take corrective action when miscertification, fraud or corruption has occurred.

A supplementary or an alternative process for setting performance standards and application of monitoring and audit is the implementation of formal quality systems to some or all activities for which the Veterinary Services are responsible. Formal accreditation to international quality system standards should be utilised if recognition in the evaluation process is to be sought.

4. Veterinary Services administration

a. Annual reports

Official government annual reports should be published, which provide information on the organisation and structure, budget, activities and contemporary performance of the Veterinary Services. Current and retrospective copies of such reports should be available to counterpart Services in other countries, especially trade partners.

b. Reports of government review bodies

The reports of any periodic or ad hoc government reviews of Veterinary Services or of particular functions or roles of the Veterinary Services should be considered in the evaluation process. Details of action taken as a consequence of the review should also be accessible.

c. Reports of special committees of enquiry or independent review bodies

Recent reports on the Veterinary Services or elements of their role or function, and details of any subsequent implementation of recommendations contained in these reports should be available. The Veterinary Services concerned should recognise that the provision of such information need not be detrimental to the evaluation outcome; in fact, it may demonstrate evidence of an effective audit and response programme. The supplying of such information can reinforce a commitment to transparency.

d. In-service training and development programme for staff

In order to maintain a progressive approach to meeting the needs and challenges of the changing domestic and international role of Veterinary Services, the national administration should have in place an organised programme which provides appropriate training across a range of subjects for relevant staff. This programme should include participation in scientific meetings of animal health organisations. Such a programme should be used in assessing the effectiveness of the Services.

e. Publications

Veterinary Services can augment their reputation by demonstrating that their staff publish scientific articles in refereed veterinary journals or other publications.

f. Formal linkages with sources of independent scientific expertise

Details of formal consultation or advisory mechanisms in place and operating between the Veterinary Services and local and international universities, scientific institutions or recognised veterinary organisations should be taken into consideration. These could serve to enhance the international recognition of the Veterinary Services.
g. Trade performance history

In the evaluation of the Veterinary Services of a country, it is pertinent to examine the recent history of their performance and integrity in trade dealings with other countries. Sources of such historical data may include Customs Services.

Article 3.2.11.

Participation in OIE activities

Questions on a country's adherence to its obligations as a member of the OIE are relevant to an evaluation of the Veterinary Services of the country. Self-acknowledged inability or repeated failure of a Member to fulfil reporting obligations to the OIE will detract from the overall outcome of the evaluation. Such countries, as well as non-member countries, will need to provide extensive information regarding their Veterinary Services and sanitary/zoosanitary status for evaluation purposes.

Article 3.2.12.

Evaluation of veterinary statutory body

1. Scope

In the evaluation of the veterinary statutory body, the following items may be considered, depending on the purpose of the evaluation:

   a. objectives and functions;

   b. legislative basis, autonomy and functional capacity;

   c. the composition and representation of the body's membership;

   d. accountability and transparency of decision-making;

   e. sources and management of funding;

   f. administration of training programmes and continuing professional development for veterinarians and veterinary para-professionals.

2. Evaluation of objectives and functions

The veterinary statutory body should define its policy and objectives, including detailed descriptions of its powers and functions such as:

   a. to regulate veterinarians and veterinary para-professionals through licensing and/or registration of such persons;

   b. to determine the minimum standards of education (initial and continuing) required for degrees, diplomas and certificates entitling the holders thereof to be registered as veterinarians and veterinary para-professionals;

   c. to determine the standards of professional conduct of veterinarians and veterinary para-professionals and to ensure these standards are met.
3. **Evaluation of legislative basis, autonomy and functional capacity**

The *veterinary statutory body* should be able to demonstrate that it has the capacity, supported by appropriate legislation, to exercise and enforce control over all *veterinarians* and *veterinary para-professionals*. These controls should include, where appropriate, compulsory licensing and registration, minimum standards of education (initial and continuing) for the recognition of degrees, diplomas and certificates, setting standards of professional conduct and exercising control and the application of disciplinary procedures.

The *veterinary statutory body* should be able to demonstrate autonomy from undue political and commercial interests.

Where applicable, regional agreements for the recognition of degrees, diplomas and certificates for *veterinarians* and *veterinary para-professionals* should be demonstrated.

4. **Evaluation of membership representation**

Detailed descriptions should be available in respect of the membership of the *veterinary statutory body* and the method and duration of appointment of members. Such information includes:

a. *veterinarians* designated by the *Veterinary Authority*, such as the Chief Veterinary Officer;

b. *veterinarians* elected by members registered by the *veterinary statutory body*;

c. *veterinarians* designated or nominated by the veterinary association(s);

d. representative(s) of veterinary para-professions;

e. representative(s) of veterinary academia;

f. representative(s) of other stakeholders from the private sector;

g. election procedures and duration of appointment;

h. qualification requirements for members.

5. **Evaluation of accountability and transparency of decision-making**

Detailed information should be available on disciplinary procedures regarding the conducting of enquiries into professional misconduct, transparency of decision-making, publication of findings, sentences and mechanisms for appeal.

Additional information regarding the publication at regular intervals of activity reports, lists of registered or licensed persons including deletions and additions should also be taken into consideration.

6. **Evaluation of financial sources and financial management**

Information regarding income and expenditure, including fee structure(s) for the licensing/registration of persons should be available.
7. Evaluation of training programmes and programmes for continuing professional development, for veterinarians and veterinary para-professionals

Descriptive summary of continuing professional development, training and education programmes should be provided, including descriptions of content, duration and participants; documented details of quality manuals and standards relating to Good Veterinary Practice should be provided.

Article 3.2.13.

1. The Veterinary Services of a country may undertake self-evaluation against the above criteria for such purposes as national interest, improvement of internal efficiency or export trade facilitation. The way in which the results of self-evaluation are used or distributed is a matter for the country concerned.

2. A prospective importing country may undertake an evaluation of the Veterinary Services of an exporting country as part of a risk analysis process, which is necessary to determine the sanitary or zoosanitary measures which the country will use to protect human or animal life or health from disease or pest threats posed by imports. Periodic evaluation reviews are also valid following the commencement of trade.

3. In the case of evaluation for the purposes of international trade, the authorities of an importing country should use the principles elaborated above as the basis for the evaluation and should attempt to acquire information according to the model questionnaire outlined in Article 3.2.14. The Veterinary Services of the importing country are responsible for the analysis of details and for determining the outcome of the evaluation after taking into account all the relevant information. The relative ranking of importance ascribed, in the evaluation, to the criteria described in this chapter will necessarily vary according to case-by-case circumstances. This ranking should be established in an objective and justifiable way. Analysis of the information obtained in the course of an evaluation study should be performed in as objective a manner as possible. The validity of the information should be established and reasonableness should be employed in its application. The assessing country should be willing to defend any position taken on the basis of this type of information, if challenged by the other party.

Article 3.2.14.

This article outlines appropriate information requirements for the self-evaluation or evaluation of the Veterinary Services of a country.

1. Organisation and structure of Veterinary Services

   a. National Veterinary Authority

       Organisational chart including numbers, positions and numbers of vacancies.

   b. Sub-national components of the Veterinary Authority

       Organisational charts including numbers, positions and number of vacancies.

   c. Other providers of veterinary services

       Description of any linkage with other providers of veterinary services.
2. **National information on human resources**

   a. **Veterinarians**

      i. Total numbers of veterinarians registered/licensed by the *Veterinary statutory body* of the country.

      ii. Numbers of:

         - full time government veterinarians: national and sub-national;
         - part time government veterinarians: national and sub-national;
         - private veterinarians authorised by the *Veterinary Services* to perform official veterinary functions [Describe accreditation standards, responsibilities and/or limitations applying to these private veterinarians.];
         - other veterinarians.

      iii. **Animal health:**

         Numbers associated with farm livestock sector on a majority time basis in a veterinary capacity, by geographical area [Show categories and numbers to differentiate staff involved in field service, laboratory, administration, import/export and other functions, as applicable.]:

         - full time government veterinarians: national and sub-national;
         - part time government veterinarians: national and sub-national;
         - other veterinarians.

      iv. **Veterinary public health:**

         Numbers employed in food inspection on a majority time basis, by commodity [Show categories and numbers to differentiate staff involved in inspection, laboratory and other functions, as applicable.]:

         - full time government veterinarians: national and sub-national;
         - part time government veterinarians: national and sub-national;
         - other veterinarians.

      v. Numbers of veterinarians relative to certain national indices:

         - per total human population;
         - per farm livestock population, by geographical area;
         - per livestock farming unit, by geographical area.
Annex V (contd)

vi. Veterinary education:
   - number of veterinary schools;
   - length of veterinary course (years);
   - international recognition of veterinary degree.

vii. Veterinary professional associations.

b. Graduate personnel (non-veterinary)

Details to be provided by category (including biologists, biometricians, economists, engineers, lawyers, other science graduates and others) on numbers within the Veterinary Authority and available to the Veterinary Authority.

c. Veterinary para-professionals employed by the Veterinary Services
   i. Animal health:
      - Categories and numbers involved with farm livestock on a majority time basis:
         - by geographical area;
         - proportional to numbers of field Veterinary Officers in the Veterinary Services, by geographical area.
      - Education/training details.
   
   ii. Veterinary public health:
      - Categories and numbers involved in food inspection on a majority time basis:
         - meat inspection: export meat establishments with an export function and domestic meat establishments (no export function);
         - dairy inspection;
         - other foods.
      - Numbers in import/export inspection.
      - Education/training details.
   
   d. Support personnel

      Numbers directly available to Veterinary Services per sector (administration, communication, transport).

   e. Descriptive summary of the functions of the various categories of staff mentioned above
   
   f. Veterinary, veterinary para-professionals, livestock owner, farmer and other relevant associations
   
   g. Additional information and/or comments.
3. **Financial management information**

   a. Total budgetary allocations to the *Veterinary Authority* for the current and past two fiscal years:

      i. for the national *Veterinary Authority*;
      
      ii. for each of any sub-national components of the *Veterinary Authority*;
      
      iii. for other relevant government-funded institutions.

   b. Sources of the budgetary allocations and amount:

      i. government budget;
      
      ii. sub-national authorities;
      
      iii. taxes and fines;
      
      iv. grants;
      
      v. private services.

   c. Proportional allocations of the amounts in a) above for operational activities and for the programme components of *Veterinary Services*.

   d. Total allocation proportionate of national public sector budget. *(This data may be necessary for comparative assessment with other countries which should take into account the contexts of the importance of the livestock sector to the national economy and of the animal health status of the country.)*

   e. Actual and proportional contribution of animal production to gross domestic product.

4. **Administration details**

   a. Accommodation

      Summary of the numbers and distribution of official administrative centres of the *Veterinary Services* (national and sub-national) in the country.

   b. Communications

      Summary of the forms of communication systems available to the *Veterinary Services* on a nation-wide and local area bases.

   c. Transport

      i. Itemised numbers of types of functional transport available on a full-time basis for the *Veterinary Services*. In addition provide details of transport means available part-time.

      ii. Details of annual funds available for maintenance and replacement of motor vehicles.
5. Laboratory services

a. Diagnostic laboratories (laboratories engaged primarily in diagnosis)

i. Descriptive summary of the organisational structure and role of the government veterinary laboratory service in particular its relevance to the field Veterinary Services.

ii. Numbers of veterinary diagnostic laboratories operating in the country:
   - government operated laboratories;
   - private laboratories accredited by government for the purposes of supporting official or officially-endorsed animal health control or public health testing and monitoring programmes and import/export testing.

iii. Descriptive summary of accreditation procedures and standards for private laboratories.

iv. Human and financial resources allocated to the government veterinary laboratories, including staff numbers, graduate and post-graduate qualifications and opportunities for further training.

v. List of diagnostic methodologies available against major diseases of farm livestock (including poultry).

vi. Details of collaboration with external laboratories including international reference laboratories and details on numbers of samples submitted.

vii. Details of quality control and assessment (or validation) programmes operating within the veterinary laboratory service.

viii. Recent published reports of the official veterinary laboratory service which should include details of specimens received and foreign animal disease investigations made.

ix. Details of procedures for storage and retrieval of information on specimen submission and results.

x. Reports of independent reviews of the laboratory service conducted by government or private organisations (if available).

xi. Strategic and operational plans for the official veterinary laboratory service (if available).

b. Research laboratories (laboratories engaged primarily in research)

i. Numbers of veterinary research laboratories operating in the country:
   - government operated laboratories;
   - private laboratories involved in full time research directly related to animal health and veterinary public health matters involving production animal species.

ii. Summary of human and financial resources allocated by government to veterinary research.

iii. Published programmes of future government sponsored veterinary research.

iv. Annual reports of the government research laboratories.
6. **Veterinary legislation, regulations and functional capabilities**

   a. Animal health and veterinary public health

      i. Assessment of the adequacy and implementation of relevant legislation (national or sub-national) concerning the following:

         - animal and veterinary public health controls at national frontiers;
         - control of endemic animal diseases, including zoonoses;
         - emergency powers for control of exotic disease outbreaks, including zoonoses;
         - inspection and registration of facilities;
         - animal feeding;
         - veterinary public health controls of the production, processing, storage and marketing of meat for domestic consumption;
         - veterinary public health controls of the production, processing, storage and marketing of fish, dairy products and other foods of animal origin for domestic consumption;
         - registration and use of veterinary pharmaceutical products including vaccines;
         - animal welfare.

      ii. Assessment of ability of *Veterinary Services* to enforce legislation.

   b. Export/import inspection

      i. Assessment of the adequacy and implementation of relevant national legislation concerning:

         - veterinary public health controls of the production, processing, storage and transportation of meat for export;
         - veterinary public health controls of production, processing, storage and marketing of fish, dairy products and other foods of animal origin for export;
         - animal health and veterinary public health controls of the export and import of animals, animal genetic material, animal products, animal feedstuffs and other products subject to veterinary inspection;
         - animal health controls of the importation, use and bio-containment of organisms which are aetiological agents of animal diseases, and of pathological material;
         - animal health controls of importation of veterinary biological products including vaccines;
         - administrative powers available to *Veterinary Services* for inspection and registration of facilities for veterinary control purposes (if not included under other legislation mentioned above);
         - documentation and compliance.

      ii. Assessment of ability of *Veterinary Services* to enforce legislation.
Annex V (contd)

7. Animal health and veterinary public health controls

a. Animal health

i. Description of and sample reference data from any national animal disease reporting system controlled and operated or coordinated by the Veterinary Services.

ii. Description of and sample reference data from other national animal disease reporting systems controlled and operated by other organisations which make data and results available to Veterinary Services.

iii. Description and relevant data of current official control programmes including:

- epidemiological surveillance or monitoring programmes;
- officially approved industry administered control or eradication programmes for specific diseases.

iv. Description and relevant details of animal disease emergency preparedness and response plans.

v. Recent history of animal disease status:

- animal diseases eradicated nationally or from defined sub-national zones in the last ten years;
- animal diseases of which the prevalence has been controlled to a low level in the last ten years;
- animal diseases introduced to the country or to previously free sub national regions in the last ten years;
- emerging diseases in the last ten years;
- animal diseases of which the prevalence has increased in the last ten years.

b. Veterinary public health

i. Food hygiene

- Annual national slaughter statistics for the past three years according to official data by species of animals (bovine, ovine, porcine, caprine, poultry, farmed game, wild game, equine, other).
- Estimate of total annual slaughterings which occur but are not recorded under official statistics.
- Proportion of total national slaughter which occurs in registered export establishments, by category of animal.
- Proportion of total national slaughter which occurs under veterinary control, by category of animal.
- Numbers of commercial *fresh meat* establishments in the country which are registered for export by the *Veterinary Authority*:
  - *slaughterhouses* (indicate species of *animals*);
  - cutting/packing plants (indicate *meat* type);
  - *meat* processing establishments (indicate *meat* type);
  - cold stores.

- Numbers of commercial *fresh meat* establishments in the country approved by other *importing countries* which operate international assessment inspection programmes associated with approval procedures.

- Numbers of commercial *fresh meat* establishments under direct public health control of the *Veterinary Services* (including details of category and numbers of inspection staff associated with these premises).

- Description of the veterinary public health programme related to production and processing of animal products for human consumption (including *fresh meat*, *poultry meat*, *meat products*, *game meat*, dairy products, fish, fishery products, molluscs and crustaceans and other foods of animal origin) especially including details applying to exports of these *commodities*.

- Descriptive summary of the roles and relationships of other official organisations in public health programmes for the products listed above if the *Veterinary Authority* does not have responsibility for those programmes which apply to national production destined to domestic consumption and/or exports of the *commodities* concerned.

ii. Zoonoses

- Descriptive summary of the numbers and functions of staff of the *Veterinary Authority* involved primarily with monitoring and control of zoonotic diseases.

- Descriptive summary of the role and relationships of other official organisations involved in monitoring and control of *zoonoses* to be provided if the *Veterinary Authority* does not have these responsibilities.

iii. Chemical residue testing programmes

- Descriptive summary of national surveillance and monitoring programmes for environmental and chemical residues and contaminants applied to animal-derived foodstuffs, *animals* and animal feedstuffs.

- Role and function in these programmes of the *Veterinary Authority* and other *Veterinary Services* to be described in summary form.

- Descriptive summary of the analytical methodologies used and their consistency with internationally recognised standards.
iv. Veterinary medicines
   - Descriptive summary of the administrative and technical controls involving registration, supply and use of veterinary pharmaceutical products especially including biological products. This summary should include a focus on veterinary public health considerations relating to the use of these products in food-producing animals.
   - Role and function in these programmes of the Veterinary Authority and other Veterinary Services to be described in summary form.

8. Quality systems
   a. Accreditation
      Details and evidence of any current, formal accreditation by external agencies of the Veterinary Services of any components thereof.
   b. Quality manuals
      Documented details of the quality manuals and standards which describe the accredited quality systems of the Veterinary Services.
   c. Audit
      Details of independent (and internal) audit reports which have been undertaken of the Veterinary Services of components thereof.

9. Performance assessment and audit programmes
   a. Strategic plans and review
      i. Descriptive summary and copies of strategic and operational plans of the Veterinary Services organisation.
      ii. Descriptive summary of corporate performance assessment programmes which relate to the strategic and operational plans - copies of recent review reports.
   b. Compliance
      Descriptive summary of any compliance unit which monitors the work of the Veterinary Services (or elements thereof).
   c. Annual reports of the Veterinary Authority
      Copies of official annual reports of the national (sub-national) Veterinary Authority.
   d. Other reports
      i. Copies of reports of official reviews into the function or role of the Veterinary Services which have been conducted within the past three years.
      ii. Descriptive summary (and copy of reports if available) of subsequent action taken on recommendations made in these reviews.
e. Training

i. Descriptive summary of in-service and development programmes provided by the Veterinary Services (or their parent Ministries) for relevant staff.

ii. Summary descriptions of training courses and duration.

iii. Details of staff numbers (and their function) who participated in these training courses in the last three years.

f. Publications

Bibliographical list of scientific publications by staff members of Veterinary Services in the past three years.

g. Sources of independent scientific expertise

List of local and international universities, scientific institutions and recognised veterinary organisations with which the Veterinary Services have consultation or advisory mechanisms in place.

10. Membership of the OIE

State if country is a member of the OIE and period of membership.

11. Other assessment criteria

— text deleted
CHAPTER 3.3.

VETERINARY LEGISLATION

Article 3.3.1.

General principles

1. Respect of the hierarchy of Acts

Veterinary legislation should scrupulously respect the separation between the primary legislation, represented by primary acts (laws), and the secondary legislation derived from regulations or rule books as laid down in the Constitution or fundamental texts of the country.

2. Legal basis

The competent authorities should have the necessary primary and secondary legislation adopted for their activities at all levels of their functional or territorial organisation.

3. Inventory of the veterinary legislation

The competent authorities should establish and maintain a complete and up to date inventory of veterinary legislation.

The use of computerised databases is recommended, on the condition that their completeness, currency, accessibility and continuity can be guaranteed.

4. Communication

The competent authorities should ensure communication of veterinary legislation and subsequent documentation to stakeholders.

5. Codification

Veterinary legislation should be collected and codified so as to make it readily accessible and intelligible and provide the capability for updating and modification as appropriate.

6. Participation in the process of developing legislation

The drafting of new and updated veterinary texts should involve the competent authorities that are responsible for the scientific and technical content, together with the necessary legal expertise to ensure that the resulting texts are legally sound.

Conversely, the competent authorities should be consulted on all proposals to develop or modify texts that have a bearing on veterinary legislation.

7. Consistency of the legislation

Veterinary legislation should be consistent with civil, penal and administrative laws and the associated procedures as appropriate.
The form of veterinary legislation

1. **Normative character**

   Veterinary legislation should be normative and should be drafted in a manner that prevents ambiguity in interpretation.

2. **Style and precision**

   The syntax and vocabulary should be clear and consistent so as to avoid any ambiguity.

   Precision and accuracy should take precedence over style even if this results in repetition and a cumbersome style.

3. **Definitions**

   Definitions should refer to the precise subjects and texts to which they pertain.

   Definitions in secondary legislation should not create any conflict or ambiguity with definitions in primary legislation.

4. **Competent authority**

   The definition of ‘competent authority’ or ‘competent authorities’ should be consistent with the OIE standards in order to assure an efficient chain of command and reliability in the provision of veterinary certification.

5. **Objectives of veterinary legislation**

   Veterinary legislation should include a clear statement of scope.

   The legislation should as a minimum include relevant guidelines in order to protect:

   a. animal health and food security;

   b. food safety;

   c. public health (zoonotic diseases) and security (stray animals);

   d. animal welfare, as defined by the OIE.

6. **Penalties and sanctions**

   Veterinary legislation should provide for penalties and sanctions at the level required for proper implementation of the overall strategy, as follows:

   a. penal sanctions, to be applied by the competent jurisdictions according to current penal procedures;

   b. administrative sanctions that are designed for immediate application in the case of activities posing a risk to animal health, animal welfare or public health.
Veterinary legislation should distinguish between significant penalties established in primary legislation and those less strong that depend on secondary legislation.

Veterinary legislation should include additional specific sanctions which would be applied on the basis of a decision from the court, notably a ban on the use of animals or the conduct of activities posing a risk to public or animal health or animal welfare.

7. **Powers of the competent authority**

Where official veterinary matters are the responsibility of more than one administration (multiple competent authorities), a reliable system of coordination and cooperation between the different authorities should be put in place.

The competent authorities should be organised in such a way as to provide for taking action quickly and coherently when such action is key to success, notably in case of implementation of animal health emergency measures or veterinary public health crises.

The legislation should provide for a chain of command that is as effective as possible (i.e. short, with all responsibilities clearly defined).

For this purpose, the responsibilities and power of the competent authorities, from the central level to those responsible for the implementation of legislation in the field, should be clearly defined.

If they are not under the responsibility of a unique competent authority, the responsibility for each element of the public veterinary domain should be attributed to a specific competent authority.

8. **Interventions by inspectors**

The competent authority should appoint technically qualified inspectors to take any actions needed for implementation or verification of compliance with the veterinary legislation.

The veterinary legislation should ensure that:

a. inspectors have the legal authority to intervene in accordance with the legislation and the penal procedures in force in the State;

b. the field of competence and the role of each inspector are prescribed according to their technical qualifications;

c. inspectors are protected against legal action and physical harm.

9. **Powers**

The rights of inspectors should be explicitly and thoroughly listed to protect the rights of stakeholders against any abuse of authority.

The powers of inspectors and rules of inspections should be prescribed, notably the authorisation and conditions for obtaining access to professional and private premises and to vehicles.

Inspectors should have powers and procedures to:

a. gain access to documents;

b. take samples;
c. retain (set aside) animals and goods, pending a decision on final disposition.

10. Obligations

The obligation of inspectors to respect confidentiality should be defined.

When attributing a field of competence or sector of responsibility, the competent authority should respect the principles of independence and impartiality prescribed in the OIE Terrestrial Animal Health Code (the Terrestrial Code) (see Article 3.1.2).

11. Administrative and enforcement actions

For the purposes of administrative and enforcement actions the following elements should be prescribed in the veterinary legislation:

a. seizure of animals, products and food of animal origin;

b. suspension of one or more activities of an inspected establishment;

c. the temporary, partial or complete closure of inspected establishments;

d. suspension or withdrawal of authorisations or approvals.

Means of compulsion enabling inspection to be performed should be provided for.

The rights of appeal against an action or a decision of an inspector should be established according to the laws of the State.

12. Financing

Veterinary legislation should provide for the sources, levels and conditions of financing required for the execution of all the activities of the competent authority, notably inspection, sampling and analysis and the procedures of authorisation or approval in all domains covered by the veterinary legislation.

Article 3.3.3.

Veterinary and para-veterinary professions

1. Veterinary medicine

In order to ensure the quality of veterinary medicine, the veterinary legislation should:

a. provide an official definition of veterinary medicine;

b. define the prerogatives of the professionals involved in the practice of veterinary medicine;

c. define the minimum initial and continuous educational requirements for the professionals;

d. prescribe the conditions for recognition of diplomas for veterinarians and para-veterinarians;

e. define the conditions for the exercise of veterinary and para-veterinary professions;

f. define the professional responsibilities of veterinarians and persons working under their control;
Annex V (contd)

g. prescribe the situations where persons other than qualified veterinarians can undertake activities that are normally to be carried out by veterinarians e.g. in exceptional circumstances such as epizootics.

2. **The control of the professions**

   In order to control the veterinary and para-veterinary professions, the veterinary legislation should:

   a. describe the general system of control in terms of the political, administrative and geographic configuration of the State;

   b. provide for the possibility of the delegation of powers to a professional organisation such as a veterinary statutory body;

   c. where powers have been so delegated, describe the prerogatives, the functioning and responsibilities of the mandated professional organisation;

   d. prescribe the disciplinary powers that apply to the relevant professions.

   Article 3.3.4.

**Laboratories in the veterinary field**

1. **Facilities**

   Veterinary legislation should define the role, responsibilities, obligations and quality requirements for:

   a. reference laboratories, which are responsible for controlling the veterinary diagnostic and analytical network, including the maintenance of reference methods;

   b. laboratories designated by the State for carrying out the analysis of official samples;

   c. laboratories recognised by the State as fit to conduct compulsory analyses by the private sector.

   The veterinary legislation should define the conditions for the classification, approval, operations and supervision of laboratories at each level.

2. **Laboratory reagents**

   Veterinary legislation should address the elements listed below:

   a. procedures for authorising the reagents that are used to perform official analyses;

   b. surveillance of marketing of reagents, where these can affect the quality of analyses required by the veterinary legislation;

   c. quality assurance of reagents by manufacturers.
Delegation of powers

1. **General principles**

   The veterinary legislation should provide for the possibility of the competent authorities delegating specific tasks related to official activities.

   The specific tasks delegated, the body(ies) to which the tasks are delegated and the conditions of supervision by the competent authority should be defined.

2. **Animal health delegation**

   The veterinary legislation should provide for the possibility of the competent authority delegating specific tasks in the sector of animal health to individual professional veterinarians who are not civil servants.

   For that purpose the veterinary legislation should:
   
   a. define the field of activities and the specific tasks covered by the delegation;
   
   b. provide for the control, supervision and financing of the delegation;
   
   c. define the procedures for making delegations;
   
   d. define the competencies to be held by persons receiving delegation;
   
   e. define the conditions of withdrawals of delegations.

3. **Delegation of functions relating to veterinary certification**

   Veterinary legislation should conform with Section 5 of the OIE Terrestrial Code concerning certification procedures, especially on the:

   a. conditions of appointment or recognition of certifying officials;
   
   b. role and responsibilities of the certifying officials;
   
   c. conditions of certification;
   
   d. means of supervision and financing of certification;
   
   e. define the conditions of withdrawal of the delegation.

4. **Delegation of functions relating to the identification of animals and traceability**

   a. Veterinary legislation should provide for the possibility of delegating operations, under the supervision of the competent authority, to the operators that are best placed to carry out and manage the identification systems.

   b. Veterinary legislation should define the conditions of withdrawal of the delegation.
5. Relationships with stakeholders

To ensure transparency and facilitate implementation of the veterinary legislation, the competent authority should establish relationships with stakeholders, including by:

a. taking steps to ensure that stakeholders participate in the development of significant legislation and required follow up;

b. supporting, as appropriate, participation of stakeholders in international discussions.

Article 3.3.6.

Health provisions relating to animal production

1. Identification and traceability

Veterinary legislation should address the following elements:

a. the objectives and scope of animal identification;

b. the possibility to make animal identification compulsory for certain species, regions or function;

c. the power of the competent authority to control movements of animals and changes of ownership;

d. identification includes the marking of animals or groups of animals and the recording of corresponding data;

e. the use of identification data for veterinary matters;

f. the equipment and methods to be used and the qualifications of operators for the marking or tracing of animals as appropriate to each situation;

g. the type of data to be recorded and the responsibilities of each party, notably those of animal keepers;

h. for the conduct of checks and corrections, as may be required to ensure the reliability of information in the database, notably in respect of animals that have died or have been slaughtered for any reason;

i. respect for constitutional liberties by restricting the use, security and confidentiality of data.

2. Animal markets and other gatherings

Veterinary legislation should address the following elements:

a. registration of all permanent or temporary animal markets and other animal gatherings;

b. health measures to prevent disease transmission, including procedures for cleaning and disinfection, and animal welfare measures;

c. provision for compulsory veterinary checks at animal gatherings.
3. **Animal reproduction**

Except where the animals or reproductive material are only used in a single holding, the veterinary legislation should address the elements listed below:

a. the health regulation of animal reproduction as appropriate;

b. health regulations may be implemented at the level of animals, genetic material, establishments or operators.

4. **Animal feed**

Veterinary legislation should address the elements listed below:

a. standards for the production and composition of animal feed;

b. registration and, if necessary, approval of establishments and the provision of health requirements for relevant operations;

c. recall from the market of any product likely to present a hazard to human health or animal health.

5. **Animal by-products (i.e. products not used for human consumption)**

Veterinary legislation should address the elements listed below:

a. definition of the animal by-products subject of the legislation;

b. rules for collection, processing methods and authorised uses of animal by-products;

c. registration and, if necessary, approval of establishments and the provision of health requirements for relevant operations;

d. definition of the rules to be applied by animal owners as appropriate.

6. **Disinfection**

Veterinary legislation should address the following elements:

a. the regulation of products and methods that are used for disinfection relating to animal diseases;

b. the use of disinfection at all critical points, notably during the transportation of animals.

Article 3.3.7.

**Animal diseases**

1. **Surveillance**

Veterinary legislation should address the following elements:

a. collection, transmission and utilisation of epidemiological data relevant to listed diseases;

b. an early warning system.
2. **Disease prevention**

Veterinary legislation should address the following elements:

a. specific rules for each listed disease;

b. support to stakeholders in proposing joint programmes;

c. the direct control by the competent authority of some disease prevention programmes;

d. compulsory programmes for some disease prevention when necessary.

3. **Disease control**

a. Veterinary legislation should address the following elements:

   i. different lists of diseases, with provision (as appropriate) for:
      
      i. emergency measures in accordance with established contingency plans;
      
      ii. measures for prevention, control or eradication;
      
      iii. surveillance measures;

   ii. the specification of mandatory control measures for certain diseases;

   iii. arrangements for the declaration of animal diseases including on the grounds of suspicion;

   iv. immediate technical measures including on the grounds of suspicion;

   v. measures for official disease surveillance;

   vi. conditions for confirmation of diseases;

   vii. precautionary measures.

b. Veterinary legislation should provide for the following general measures:

   i. definition of areas in which health measures are applied;

   ii. official publicising of measures;

   iii. listing of all measures requiring a legal basis;

   iv. measures to be implemented by the public force;

   v. epidemiological investigations;

   vi. provisions for wild or protected animals;

   vii. conditions for restocking;

   viii. commercial restrictions.
c. Contingency plan should be developed for certain diseases and, in addition to the general measures, should provide for:

   i. administrative and logistic organisation;

   ii. exceptional powers of the competent authority;

   iii. special and temporary measures to address all identified risks to human or animal health.

d. Veterinary legislation should provide for the financing of animal disease control measures, notably:

   i. operational expenses;

   ii. production losses;

   iii. owners compensation in the event of killing or slaughtering of animals, seizure or destruction of carcasses, meat, animal feed or other things.

   Article 3.3.8.

Animal welfare measures

1. General provisions

   Veterinary legislation should address the elements listed below:

   a. general principles to ensure the protection of animals against cruelty, abuse, abandonment and avoidable suffering, in line with the OIE Terrestrial Code;

   b. legal definition of cruelty as an offense, subject to penal action;

   c. direct intervention of the competent authority in the case of neglect by animal keepers;

   d. accepted practices for livestock, pets, animals used in scientific experiments, sport and leisure, and for wild animals, notably in relation to:

      i. transport and handling;

      ii. animal production and housing;

      iii. slaughtering and killing;

      iv. scientific experiments;

      v. use in games, shows, exhibitions and zoos;

   e. certain activities relating to animals may be restricted to the holders of appropriate qualifications or approvals.

2. Free-roaming and stray domestic animals

   Veterinary legislation should address the elements listed below:

   a. prohibition of abandonment of animals and of allowing animals to stray;
Annex V (contd)

b. establishments where stray animals can be held and the conditions governing their operation;

c. the circumstances and the conditions of capture and of holding of stray animals;

d. the outcomes for these animals, including arrangements for veterinary interventions (including euthanasia in compliance with OIE standards), and for the transfer of ownership.

Article 3.3.9.

Veterinary products

1. Objectives

Veterinary legislation should address the following elements:

a. avoiding the presence of harmful residues in the food chain;

b. ensuring that the use of veterinary products does not give rise to human health risks.

2. General measures

Veterinary legislation should address the elements listed below:

a. definition of veterinary products, including any specific exclusions;

b. regulation of the importation, manufacture, distribution and usage of, and commerce in, veterinary products.

3. Raw materials and veterinary products

Veterinary legislation should address the elements listed below:

a. quality standards for raw materials used in the manufacture or composition of veterinary products and arrangements for checking quality;

b. establishment of the withdrawal periods and maximum residue limits for veterinary products as appropriate;

c. requirements for any substances that may interfere with the conduct of veterinary checks.

4. Authorisation of veterinary products

a. Veterinary legislation should ensure that only authorised veterinary products may be placed on the market.

b. Special provisions should be made for:

i. veterinary products that do not present any risk of residues or interference with the conduct of disease prevention and control programmes;

ii. medicated feed;

iii. products prepared by veterinarians or pharmacists;
iv) emergencies and temporary situations.

c. Veterinary legislation should address the technical, administrative and financial conditions associated with the granting, renewal, refusal and withdrawal of authorisations.

d. In defining the procedures for seeking and granting authorisations, the legislation should:

   i) describe the functioning of the competent authority concerned;

   ii) establish rules providing for the transparency of decisions.

e. Veterinary legislation may provide for the possibility of recognition of the equivalence of authorisations made by other countries.

5. Quality of veterinary products

   To give effect to the objectives identified above, veterinary legislation should address the elements listed below:

   a. the conduct of clinical and non-clinical trials to verify all claims made by the manufacturer, including analysis and dosage methods;

   b. conditions for the conduct of trials;

   c. qualifications of experts involved in trials;

   d. surveillance for adverse effects arising from the use of veterinary products.

6. Establishments producing, storing and selling veterinary products

   Veterinary legislation should address the following elements:

   a. registration or authorisation of all operators importing, storing, processing, selling or otherwise distributing veterinary products or raw materials for use in making veterinary products;

   b. definition of the responsibilities of operators;

   c. good manufacturing practices as appropriate;

   d. arrangements for informing the competent authority about traceability of products and adverse effects.

7. Commerce, distribution, use and traceability of veterinary products

   Veterinary legislation should address the following elements:

   a. control over the circulation and distribution of veterinary products and arrangement for traceability and condition of use;

   b. establishment of rules of prescription and provision of veterinary products to the end user;

   c. restricting to authorised professionals all commerce in veterinary products that are subject to prescription;
Annex V (contd)

d. the supervision by an authorised professional of organisations approved for holding and use of veterinary products;

e. the regulation of advertising claims and other marketing and promotional activities.

Article 3.3.10.

Safeguards for the food production chain and traceability

1. Objectives

Veterinary legislation should address the following elements:

a. the control of the manufacturing process at all relevant levels in the food production chain;

b. requirements to assure food safety for the purpose of (i).

In addition, procedures may be implemented to allow food production appropriate to the economic situation.

2. General

Veterinary legislation should address the following elements in order to ensure the food safety of animal products:

a. recording all significant health events that occur during primary production;

b. prohibition of the marketing of infected products or products likely to be contaminated or hazardous for the consumer or for animal health;

c. inspection for food safety and food composition;

d. inspection of premises;

e. controls over the implementation of the legislation at all stages of the production, processing and distribution of food of animal origin;

f. establish that operators of food production premises have the primary responsibility for food safety;

g. obligations for producers to withdraw from the marketplace all products likely to be hazardous for human or animal health.

3. Products of animal origin intended for human or animal consumption

Veterinary legislation should address the following elements:

a. arrangements for inspection;

b. the conduct of inspection on the basis of veterinary expertise;

c. relevant health standards;

d. application of health identification marks, which are visible to the intermediary or final user.
The competent authority should have the necessary powers and means to rapidly withdraw any products deemed to be hazardous from the food chain or to prescribe uses or treatments that ensure the safety of such products for human or animal health.

4. **Premises and establishments pertaining to the food chain**

Veterinary legislation should address the following elements as appropriate:

a. recording the coordinates of operators working within the food chain;

b. the implementation by operators of procedures based on HACCP principles;

c. prior authorisation of operators whose activities are likely to constitute a significant risk to human or animal health.

Article 3.3.11.

**International movements and trade**

1. **Importation**

Veterinary legislation should address the following elements:

a. the coordinates of importers and, as appropriate, their approval by the competent authority of the importing country;

b. the establishment by the competent authority of:

i. the list of goods to be subject to veterinary checks;

ii. the importation check points officially designated for each kind of goods;

iii. the kinds and procedures of checks to be performed;

iv. the standards with which animals and commodities proposed for importation must comply;

c. prevention of entry of listed goods and consignments into the country unless such goods have been subjected to the required veterinary checks;

d. objectivity and independence of inspectors.

2. **Exports**

Veterinary legislation should specify the conditions governing the provision of veterinary certification and any prohibitions, in conformity with relevant provisions of the OIE and of the Codex Alimentarius Commission.

It should also include provisions ensuring national involvement to relevant activities of the work of the OIE and the Codex Alimentarius and, if necessary, interministerial coordination allowing the harmonization of the positions taken by the country in these international organizations.
CHAPTER 4.2.

DESIGN AND IMPLEMENTATION OF IDENTIFICATION SYSTEMS TO ACHIEVE ANIMAL TRACEABILITY

Article 4.2.1.

Introduction and objectives

These recommendations are based on the general principles presented in Article 4.1.1. The recommendations outline for Members the basic elements that need to be taken into account in the design and implementation of an animal identification system to achieve animal traceability. Whatever animal identification system the country adopts, it should comply with relevant OIE standards, including Chapters 5.10. to 5.12. for animals and animal products intended for export. Each country should design a programme in accordance with the scope and relevant performance criteria to ensure that the desired animal traceability outcomes can be achieved.

Article 4.2.2.

Definitions

For the purpose of this chapter:

Desired outcomes: describe the overall goals of a programme and are usually expressed in qualitative terms, e.g. ‘to help ensure that animals and/or animal products are safe and suitable for use’. Safety and suitability for use could be defined in terms such as animal health, food safety, trade and aspects of animal husbandry.

Performance criteria: are specifications for performance of a programme and are usually expressed in quantitative terms, such as ‘all animals can be traced to the establishment of birth within 48 hours of an enquiry’.

Reporting: means advising the Veterinary Authority and other partner organisations as appropriate in accordance with the procedures listed in the programme.

Scope: specifies the targeted species, population and/or production/trade sector within a defined area (country, zone) or compartment that is the subject of the identification and traceability programme.

Transhumance: periodic/seasonal movements of animals between different pastures within or between countries.

Article 4.2.3.

Key elements of the animal identification system

1. Desired outcomes

Desired outcomes should be defined through consultation between the Veterinary Authority and other parties, which should include (depending on scope) animal producers and food processors, private sector veterinarians, scientific research organisations and other government agencies. Desired outcomes may be defined in terms of any or all of the following:

a) animal health (e.g. disease surveillance and notification; detection and control of disease; vaccination programmes);

b) public health (e.g. surveillance and control of zoonotic diseases and food safety);
Annex VI (contd)

c) management of emergencies e.g. natural catastrophies or man-made events;

d) trade (support for inspection and certification activities of Veterinary Services, as described in Chapters 5.10. to 5.12. which reproduce model international veterinary certificates);

e) aspects of animal husbandry such as animal performance, and genetic data.

2. Scope

Scope should also be defined through consultation between the Veterinary Authority and other parties, as discussed above. The scope of animal identification systems is often based on the definition of a species and sector, to take account of particular characteristics of the farming systems e.g. pigs in pork export production; poultry in a defined compartment; cattle within a defined FMD free zone. Different systems will be appropriate according to the production systems used in countries and the nature of their industries and trade.

3. Performance criteria

Performance criteria are also designed in consultation with other parties, as discussed above. The performance criteria depend on the desired outcomes and scope of the programme. They are usually described in quantitative terms according to the epidemiology of the disease. For example, some countries consider it necessary to trace susceptible animals within 24-48 hours when dealing with highly contagious diseases such as FMD and avian influenza. For food safety, animal tracing to support investigation of incidents may also be urgent. For chronic animal diseases that are not zoonoses, it may be considered appropriate that animals can be traced over a longer period.

4. Preliminary studies

In designing animal identification systems it is useful to conduct preliminary studies, which should take into account:

a) animal populations, species, distribution, herd management,
b) farming and industry structures, production and location,
c) animal health,
d) public health,
e) trade issues,
f) aspects of animal husbandry,
g) zoning and compartmentalisation,
h) animal movement patterns (including transhumance),
i) information management and communication,
j) availability of resources (human and financial),
k) social and cultural aspects,
l) stakeholder knowledge of the issues and expectations,
m) gaps between current enabling legislation and what is needed long term,
n) international experience,
o) national experience,
p) available technology options,
q) existing identification system(s),
r) expected benefits from the animal identification systems and animal traceability and to whom they accrue,
s) issues pertaining to data ownership and access rights,
t) reporting requirements.

Pilot projects may form part of the preliminary study to test the animal identification system and animal traceability and to gather information for the design and the implementation of the programme.

Economic analysis may consider costs, benefits, funding mechanisms and sustainability.

5. Design of the programme

a) General provisions

The programme should be designed in consultation with the stakeholders to facilitate the implementation of the animal identification system and animal traceability. It should take into account the scope, performance criteria and desired outcomes as well as the results of any preliminary study.

All the specified documentation should be standardised as to format, content and context.

To protect and enhance the integrity of the system, procedures should be incorporated into the design of the programme to prevent, detect and correct errors e.g. use of algorithms to prevent duplication of identification numbers and to ensure plausibility of data.

b) Means of animal identification

The choice of a physical animal or group identifier should consider elements such as the durability, human resources, species and age of the animals to be identified, required period of identification, cultural aspects, animal welfare, technology, compatibility and relevant standards, farming practices, production systems, animal population, climatic conditions, resistance to tampering, trade considerations, cost, and retention and readability of the identification method.

The Veterinary Authority is responsible for approving the materials and equipment chosen, to ensure that these means of animal identification comply with technical and field performance specifications, and for the supervision of their distribution. The Veterinary Authority is also responsible for ensuring that identifiers are unique and are used in accordance with the requirements of the animal identification system.

The Veterinary Authority should establish procedures for animal identification and animal traceability including:

i) the establishment of birth, and time period within which an animal is born;

ii) when animals are introduced into an establishment;
Annex VI (contd)

iii) when an animal loses its identification or the identifier becomes unusable;

iv) arrangements and rules for the destruction and/or reuse of identifiers;

v) penalties for the tampering and/or removal of official animal identification devices.

Where group identification without a physical identifier is adequate, documentation should be created specifying at least the number of animals in the group, the species, the date of identification, the person legally responsible for the animals and/or establishment. This documentation constitutes a unique group identifier and it should be updated to be traceable if there are any changes.

Where all animals in the group are physically identified with a group identifier, documentation should also specify the unique group identifier.

c) Registration

Procedures need to be incorporated into the design of the programme in order to ensure that relevant events and information are registered in a timely and accurate manner.

Depending on the scope, performance criteria and desired outcomes, records as described below should specify, at least, the species, the unique animal or group identifier, the date of the event, the identifier of the establishment where the event took place, and the code for the event itself.

i) Establishments/owners or responsible keepers

Establishments where animals are kept should be identified and registered, including at least their physical location (such as geographical coordinates or street address), the type of establishment and the species kept. The register should include the name of the person legally responsible for the animals at the establishment.

The types of establishments that may need to be registered include holdings (farms), assembly centres (e.g. agriculture shows and fairs, sporting events, transit centres, breeding centres), markets, abattoirs, rendering plants, dead stock collection points, transhumance areas, centres for necropsy and diagnosis, research centres, zoos, border posts, quarantine stations.

In cases where the registration of establishments is not applicable e.g. some transhumance systems, the animal owner, the owner’s place of residence and the species kept should be recorded.

ii) Animals

Animal identification and species should be registered for each establishment/owner. Other relevant information about the animals at each establishment/owner may also be recorded (e.g. date of birth, production category, sex, breed, number of animals of each species, animal identification of the parents).

iii) Other events

The registration of animal movements is necessary to achieve animal traceability. When an animal is introduced into or leaves an establishment, these events constitute a movement.

Some countries classify birth, slaughter and death of the animal as movements. When establishments are not registered as part of the animal identification system, ownership and location changes constitute a movement record.
The information registered should include the date of the movement, the establishment from which the animal or group of animals was dispatched, the number of animals moved, the destination establishment, and any establishment used in transit. The movement recording may also include a description of the means of transport and the identification of the vehicle/vessel identifier.

Procedures should be in place to maintain animal traceability during transport and when animals arrive at and leave an establishment.

The following events may also be registered:

- birth, slaughter and death of the animal (when not classified as a movement),
- attachment of the unique identifier to an animal,
- change of owner or keeper regardless of change of establishment,
- observation of an animal on an establishment (testing, health investigation, health certification, etc.),
- animal imported: a record of the animal identification from the exporting country should be kept and linked with the animal identification assigned in the importing country,
- animal exported: a record of the animal identification from the exporting country should be provided to the Veterinary Authority in the importing country,
- animal identifier lost or replaced,
- animal missing (lost, stolen, etc.),
- animal identifier retired (at slaughter, following loss of the identifier or death of the animal on a farm, at diagnostic laboratories, etc.).

d) Documentation

Documentation requirements should be clearly defined and standardised, according to the scope, performance criteria and desired outcomes and supported by the legal framework.

e) Reporting

Depending on the scope, performance criteria and desired outcomes, relevant information (such as animal identification, movement, events, changes in numbers of livestock, establishments) should be reported to the Veterinary Authority by the person responsible for the animals.

f) Information system

An information system should be designed according to the scope, performance criteria and desired outcomes. This may be paper based or electronic. The system should provide for the collection, compilation, storage and retrieval of information on matters relevant to registration. The following considerations are important:

- have the potential for linkage to traceability in the other parts of the food chain;
- minimize duplication;
Annex VI (contd)

- relevant components, including databases, should be compatible;
- confidentiality of data;
- appropriate safeguards to prevent the loss of data, including a system for backing up the data.

The Veterinary Authority should have access to this information system as appropriate to meet the scope, performance criteria and desired outcomes.

g) Laboratories

The results of diagnostic tests should record the animal identifier or the group identifier, the date of sample was taken from the animal and the establishment where the sample was collected.

h) Abattoirs, rendering plants, dead stock collection points, markets and assembly centres

Abattoirs, rendering plants, dead stock collection points, markets and assembly centres should document arrangements for the maintenance of animal identification and animal traceability in compliance with the legal framework.

These establishments are critical points for control of animal health and food safety.

Animal identification should be recorded on documents accompanying samples collected for analysis.

The components of the animal identification system operating within abattoirs should complement and be compatible with arrangements for tracking animal products throughout the food chain. At an abattoir, animal identification should be maintained during the processing of the animal’s carcass until the carcass is deemed fit for human consumption.

The animal identification and the establishment from which the animal was dispatched should be registered by the abattoir, rendering plant and dead stock collection points.

Abattoirs, rendering plants and dead stock collection points should ensure that identifiers are collected and disposed of according to the procedures established and regulated within the legal framework. These procedures should minimize the risk of unauthorized reuse and, if appropriate, should establish arrangements and rules for the reuse of identifiers.

Reporting of movement by abattoirs, rendering plants and dead stock collection points should occur according to the scope, performance criteria and desired outcomes and the legal framework.

i) Penalties

Different levels and types of penalties should be defined in the programme and supported by the legal framework.

6. Legal framework

The Veterinary Authority, with other relevant governmental agencies and in consultation with stakeholders, should establish a legal framework for the implementation and enforcement of animal identification system and animal traceability in the country. The structure of this framework will vary from country to country.

Animal identification, animal traceability and animal movement should be under the responsibility of the Veterinary Authority.

This legal framework should address:
i) desired outcomes and scope;

ii) obligations of the Veterinary Authority and other parties;

iii) organisational arrangements, including the choice of technologies and methods used for the animal identification system and animal traceability;

iv) management of animal movement;

v) confidentiality of data;

vi) data access / accessibility;

vii) checking, verification, inspection and penalties;

viii) where relevant, funding mechanisms;

ix) where relevant, arrangements to support a pilot project.

7. Implementation

a) Action plan

For implementing the animal identification system, an action plan should be prepared specifying the timetable and including the milestones and performance indicators, the human and financial resources, and checking, enforcement and verification arrangements.

The following activities should be addressed in the action plan:

i) Communication

The scope, performance criteria, desired outcomes, responsibilities, movement and registration requirements and sanctions need to be communicated to all parties.

Communication strategies need to be targeted to the audience, taking into account elements such as the level of literacy (including technology literacy) and spoken languages.

ii) Training programmes

It is desirable to implement training programmes to assist the Veterinary Services and other parties.

iii) Technical support

Technical support should be provided to address practical problems.

b) Checking and verification

Checking activities should start at the beginning of the implementation to detect, prevent and correct errors and to provide feedback on programme design.

Verification should begin after a preliminary period as determined by the Veterinary Authority in order to determine compliance with the legal framework and operational requirements.
Annex VI (contd)

c) Auditing

Auditing should be carried out under the authority of the Veterinary Authority to detect any problems with the animal identification system and animal traceability and to identify possible improvements.

d) Review

The programme should be subject to periodic review, taking into account the results of checking, verification and auditing activities.


**CHAPTER 4.3.**

**ZONING AND COMPARTMENTALISATION**

**Article 4.3.1.**

**Introduction**

For the purposes of the *Terrestrial Code*, ‘zoning’ and ‘regionalisation’ have the same meaning.

Establishing and maintaining a disease free-status throughout the country should be the final goal for OIE Members. However, given the difficulty of establishing and maintaining a *disease* free status for an entire territory, especially for *diseases* the entry of which is difficult to control through measures at national boundaries, there may be benefits to a Member in establishing and maintaining a *subpopulation* with a distinct health status within its territory. *Subpopulations* may be separated by natural or artificial geographical barriers or, in certain situations, by the application of appropriate management practices.

Zoning and compartmentalisation are procedures implemented by a Member under the provisions of this chapter with a view to defining *subpopulations* of distinct health status within its territory for the purpose of *disease* control and/or *international trade*. While zoning applies to an animal *subpopulation* defined primarily on a geographical basis (using natural, artificial or legal boundaries), compartmentalisation applies to an animal *subpopulation* defined primarily by management and husbandry practices related to biosecurity. In practice, spatial considerations and good management including *biosecurity plans* play important roles in the application of both concepts.

A particular application of the concept of zoning is the establishment of a *containment zone*. In the event of limited *outbreaks* of a specified *disease* within an otherwise free country or zone, a single *containment zone*, which includes all *cases*, can be established for the purpose of minimizing the impact on the entire country or zone.

This chapter is to assist OIE Members wishing to establish and maintain different *subpopulations* within their territory using the principles of compartmentalisation and zoning. These principles should be applied in accordance with the measures recommended in the relevant *disease* chapter(s). This chapter also outlines a process through which trading partners may recognise such *subpopulations*. This process is best implemented by trading partners through establishing parameters and gaining agreement on the necessary measures prior to *outbreaks* of *disease*.

Before trade in *animals* or their products may occur, an *importing country* needs to be satisfied that its *animal health status* will be appropriately protected. In most cases, the import regulations developed will rely in part on judgements made about the effectiveness of sanitary procedures undertaken by the *exporting country*, both at its borders and within its territory.

As well as contributing to the safety of *international trade*, zoning and compartmentalisation may assist *disease* control or eradication within a Member's territory. *Zoning* may encourage the more efficient use of resources within certain parts of a country and compartmentalisation may allow the functional separation of a *subpopulation* from other domestic or wild animals through biosecurity measures, which a *zone* (through geographical separation) would not achieve. Following a *disease outbreak*, the use of compartmentalisation may allow a Member to take advantage of epidemiological links among *subpopulations* or common practices relating to biosecurity, despite diverse geographical locations, to facilitate *disease* control and/or the continuation of trade.

Zoning and compartmentalisation cannot be applied to all *diseases* but separate requirements will be developed for each *disease* for which the application of zoning or compartmentalisation is considered appropriate.
Annex VII (contd)

To regain free status following a disease outbreak in a zone or compartment, Members should follow the recommendations in the relevant disease chapter in the Terrestrial Code.

Article 4.3.2.

General considerations

The Veterinary Services of an exporting country which is establishing a zone or compartment within its territory for international trade purposes should clearly define the subpopulation in accordance with the recommendations in the relevant chapters in the Terrestrial Code, including those on surveillance, and the identification and traceability of live animals. The Veterinary Services of an exporting country should be able to explain to the Veterinary Services of an importing country the basis for claiming a distinct animal health status for the given zone or compartment under consideration.

The procedures used to establish and maintain the distinct animal health status of a zone or compartment should be appropriate to the particular circumstances, and will depend on the epidemiology of the disease, in particular, the presence and importance role of susceptible wildlife species, and environmental factors, and appropriate on the application of biosecurity measures.

The authority, organisation and infrastructure of the Veterinary Services, including laboratories, should be clearly documented in accordance with the chapter on the evaluation of Veterinary Services of the Terrestrial Code, to provide confidence in the integrity of the zone or compartment. The final authority of the zone or compartment, for the purposes of domestic and international trade, lies with the Veterinary Authority.

In the context of maintaining the health status of a population, references to ‘import’, ‘importation’ and ‘imported animals/products’ found in the Terrestrial Code apply both to importation into a country and to the movement of animals and their products into zones and compartments. Such movements should be the subject of appropriate measures to preserve the animal health status of the zone/compartment.

The exporting country should be able to demonstrate, through detailed documentation provided to the importing country, that it has implemented the recommendations in the Terrestrial Code for establishing and maintaining such a zone or compartment.

An importing country should recognise the existence of this zone or compartment when the appropriate measures recommended in the Terrestrial Code are applied and the Veterinary Authority of the exporting country certifies that this is the case.

The exporting country should conduct an assessment of the resources needed and available to establish and maintain a zone or compartment for international trade purposes. These include the human and financial resources, and the technical capability of the Veterinary Services (and of the relevant industry and production system, in the case of a compartment) including disease surveillance and diagnosis.

Biosecurity and surveillance are essential components of zoning and compartmentalisation, and the arrangements should be developed through cooperation of industry and Veterinary Services.

Industry’s responsibilities include the application of biosecurity measures, documenting and recording movements of animals and personnel, quality assurance schemes, monitoring the efficacy of the measures, documenting corrective actions, conducting surveillance, rapid reporting and maintenance of records in a readily accessible form.

The Veterinary Services should provide movement certification, and carry out documented periodic inspections of facilities, biosecurity measures, records and surveillance procedures. Veterinary Services should conduct or audit surveillance, reporting and laboratory diagnostic examinations.
Principles for defining and establishing a zone or compartment, including protection and containment zones

In conjunction with the above considerations, the following principles should apply when Members define a zone or a compartment.

1. The extent of a zone and its geographical limits should be established by the Veterinary Authority on the basis of natural, artificial and/or legal boundaries, and made public through official channels.

2. A protection zone may be established to preserve the health status of animals in a free country or zone, from adjacent countries or zones of different animal health status. Measures should be implemented based on the epidemiology of the disease under consideration to prevent introduction of the pathogenic agent and to ensure early detection. These measures should include intensified movement control and surveillance and may include:

   a) animal identification and animal traceability to ensure that animals in the protection zone are clearly distinguishable from other populations;
   b) vaccination of all or at risk susceptible animals;
   c) testing and/or vaccination of animals moved;
   d) specific procedures for sample handling, sending and testing;
   e) enhanced biosecurity including cleansing – disinfection procedures for transport means, and possible compulsory routes;
   f) specific surveillance of susceptible wildlife species and relevant vectors;
   g) awareness campaigns to the public or targeted at breeders, traders, hunters, veterinarians.

The application of these measures can be in the entire free zone or in a defined area within and/or outside the free zone.

3. In the event of limited outbreaks in a country or zone previously free of a disease, a containment zone may be established for the purposes of trade. Establishment of a containment zone should be based on a rapid response including:

   a) appropriate standstill of movement of animals and other commodities upon notification of suspicion of the specified disease and the demonstration that the outbreaks are contained within this zone through epidemiological investigation (trace-back, trace-forward) after confirmation of infection. The primary outbreak has been identified and investigations on the likely source of the outbreak have been carried out should be identified and all cases shown to be epidemiologically linked.
   b) A stamping-out policy or another effective control strategy aimed at eradicating the disease should be applied and the susceptible animal population within the containment zones should be clearly identifiable as belonging to the containment zone. Increased passive and targeted surveillance in accordance with Chapter 1.4, in the rest of the country or zone should be carried out and has not detected any evidence of infection.
Annex VII (contd)

c) Measures consistent with the disease specific chapter should be in place to prevent spread of the infection from the containment zone to the rest of the country or zone, including ongoing surveillance in the containment zone.

d) For the effective establishment of a containment zone, it is necessary to demonstrate that there have been no new cases in the containment zone within a minimum of two incubation periods from the last detected case.

e) The free status of the areas outside the containment zone would be suspended pending the establishment of the containment zone. The free status of these areas could be reinstated, once the containment zone is clearly established, irrespective of the provisions of the disease specific chapter.

f) The containment zone should be managed in such a way that it can be demonstrated that commodities for international trade can be shown to have originated outside the containment zone.

g) The recovery of the free status of the containment zone should follow the provisions of the disease specific chapter.

4. The factors defining a compartment should be established by the Veterinary Authority on the basis of relevant criteria such as management and husbandry practices related to biosecurity, and made public through official channels.

5. Animals and herds belonging to such subpopulations need to be recognisable as such through a clear epidemiological separation from other animals and all things presenting a disease risk. For a zone or compartment, the Veterinary Authority should document in detail the measures taken to ensure the identification of the subpopulation and the establishment and maintenance of its health status through a biosecurity plan. The measures used to establish and maintain the distinct animal health status of a zone or compartment should be appropriate to the particular circumstances, and will depend on the epidemiology of the disease, environmental factors, the health status of animals in adjacent areas, applicable biosecurity measures (including movement controls, use of natural and artificial boundaries, the spatial separation of animals, and commercial management and husbandry practices), and surveillance.

6. Relevant animals within the zone or compartment should be identified in such a way that their history can be audited movements are traceable. Depending on the system of production, identification may be done at the herd, flock lot or individual animal level. Relevant animal movements into and out of the zone or compartment should be well documented, controlled and supervised. The existence of a valid animal identification system is a prerequisite to assess the integrity of the zone or compartment.

7. For a compartment, the biosecurity plan should describe the partnership between the relevant industry and the Veterinary Authority, and their respective responsibilities. It should also describe the routine operating procedures to provide clear evidence that the surveillance conducted, the live animal identification and traceability system, and the management practices are adequate to meet the definition of the compartment. In addition to information on animal movement controls, the plan should include herd or flock production records, feed sources, surveillance results, birth and death records, visitor logbook, morbidity and mortality history, medications, vaccinations, documentation of training of relevant personnel and any other criteria necessary for evaluation of risk mitigation. The information required may vary according to the species and disease(s) under consideration. The biosecurity plan should also describe how the measures will be audited to ensure that the risks are regularly re-assessed and the measures adjusted accordingly.
CHAPTER 4.4.

APPLICATION OF COMPARTMENTALISATION

Article 4.4.1.

Introduction and objectives

The recommendations in this chapter provide a structured framework for the application and recognition of compartments within countries or zones, based on the provisions of Chapter 4.3. with the objective to facilitate trade in animals and products of animal origin and as a tool for disease management.

Establishing and maintaining a disease free-status throughout the country should be the final goal for OIE Members. However, establishing and maintaining a disease-free status for an entire country may be difficult, especially in the case of diseases that can easily cross international boundaries. For many diseases, OIE Members have traditionally applied the concept of zoning to establish and maintain an animal subpopulation with a different animal health status within national boundaries.

The essential difference between zoning and compartmentalisation is that the recognition of zones is based on geographical boundaries whereas the recognition of compartments is based on management practices and biosecurity. However, spatial considerations and good management practices play a role in the application of both concepts.

Compartmentalisation is not a new concept for Veterinary Services; in fact, it has been applied for a long time in many disease control programmes that are based on the concept of disease-free herds/flocks.

The fundamental requirement for compartmentalisation is the implementation and documentation of management and biosecurity measures to create a functional separation of subpopulations.

For example, an animal production operation in an infected country or zone might have biosecurity measures and management practices that result in negligible risk from diseases or agents. The concept of a compartment extends the application of a ‘risk boundary’ beyond that of a geographical interface and considers all epidemiological factors that can help to create an effective disease-specific separation between subpopulations.

In disease-free countries or zones, compartments preferably should be defined prior to the occurrence of a disease outbreak. In the event of an outbreak or in infected countries or zones, compartmentalisation may be used to facilitate trade.

For the purpose of international trade, compartments should be under the responsibility of the Veterinary Authority in the country. For the purposes of this chapter, compliance by the Members with Chapters 1.1. and 3.1. is an essential prerequisite.

Article 4.4.2.

Principles for defining a compartment

A compartment may be established with respect of a specific disease or diseases. A compartment should be clearly defined, indicating the location of all its components including establishments, as well as related functional units (such as feed mills, slaughterhouses, rendering plants, etc.), their interrelationships and their contribution to an epidemiological separation between the animals in a compartment and subpopulations with a different health status. The definition of compartment may revolve around disease specific epidemiological factors, animal production systems, biosecurity practices infrastructural factors and surveillance.
Annex VII (contd)

Article 4.4.3.

Separation of a compartment from potential sources of infection

The management of a compartment should provide to the Veterinary Authority documented evidence on the following:

1. **Physical or spatial factors that affect the status of biosecurity in a compartment**

   While a compartment is primarily based on management and biosecurity measures, a review of geographical factors is needed to ensure that the functional boundary provides adequate separation of a compartment from adjacent animal populations with a different health status. The following factors should be taken into consideration in conjunction with biosecurity measures and, in some instances, may alter the degree of confidence achieved by general biosecurity and surveillance measures:

   a) disease status in adjacent areas and in areas epidemiologically linked to the compartment;

   b) location, disease status and biosecurity of the nearest epidemiological units or other epidemiologically relevant premises. Consideration should be given to the distance and physical separation from:

      i) flocks or herds with a different health status in close proximity to the compartment, including wildlife and their migratory routes;

      ii) slaughterhouses, rendering plants or feed mills;

      iii) markets, fairs, agricultural shows, sporting events, zoos, circuses and other points of animal concentration.

2. **Infrastructural factors**

   Structural aspects of the establishments within a compartment contribute to the effectiveness of its biosecurity. Consideration should be given to:

   a) fencing or other effective means of physical separation;

   b) facilities for people entry including access control, changing area and showers;

   c) vehicle access including washing and disinfection procedures;

   d) unloading and loading facilities;

   e) isolation facilities for introduced animals;

   f) facilities for the introduction of material and equipment;

   g) infrastructure to store feed and veterinary products;

   h) disposal of carcasses, manure and waste;

   i) water supply;

   j) measures to prevent exposure to living mechanical or biological vectors such as insects, rodents and wild birds;
k) air supply;
l) feed supply/source.

More detailed recommendations for certain establishments can be found in Sections 4 and 6 of the Terrestrial Code.

3. **Biosecurity plan**

The integrity of the compartment relies on effective biosecurity. The management of the compartment should develop, implement and monitor a comprehensive biosecurity plan.

The biosecurity plan should describe in detail:

a) potential pathways for introduction and spread into the compartment of the agents for which the compartment was defined, including animal movements, rodents, fauna, aerosols, arthropods, vehicles, people, biological products, equipment, fomites, feed, waterways, drainage or other means. Consideration should also be given to the survivability of the agent in the environment;

b) the critical control points for each pathway;

c) measures to mitigate exposure for each critical control point;

d) standard operating procedures including:

i) implementation, maintenance, monitoring of the measures,

ii) application of corrective actions,

iii) verification of the process,

iv) record keeping;

e) contingency plan in the event of a change in the level of exposure;

f) reporting procedures to the Veterinary Authority;

g) the programme for educating and training workers to ensure that all persons involved are knowledgeable and informed on biosecurity principles and practices;

h) the surveillance programme in place.

In any case, sufficient evidence should be submitted to assess the efficacy of the biosecurity plan in accordance with the level of risk for each identified pathway. This evidence should be structured in line with the principles of Hazard Analysis and Critical Control Point (HACCP). The biosecurity risk of all operations of the compartment should be regularly re-assessed and documented at least on a yearly basis. Based on the outcome of the assessment, concrete and documented mitigation steps should be taken to reduce the likelihood of introduction of the disease agent into the compartment.

4. **Traceability system**

A prerequisite for assessing the integrity of a compartment is the existence of a valid traceability system. All animals within a compartment should be individually identified and registered in such a way that their history and movements can be documented and audited. In cases where individual identification may not be feasible, such as broilers and day-old chicks, the Veterinary Authority should provide sufficient assurance of traceability.
Annex VII (contd)

All animal movements into and out of the compartment should be recorded at the compartment level, and when needed, based on a risk assessment, certified by the Veterinary Authority. Movements within the compartment need not be certified but should be recorded at the compartment level.

Article 4.4.4.

Documentation

Documentation should provide clear evidence that the biosecurity, surveillance, traceability and management practices defined for a compartment are effectively and consistently applied. In addition to animal movement information, the necessary documentation should include herd or flock production records, feed sources, laboratory tests, birth and death records, the visitor logbook, morbidity history, medication and vaccination records, biosecurity plans, training documentation and any other criteria necessary for the evaluation of disease exclusion.

The historical status of a compartment for the disease(s) for which it was defined should be documented and demonstrate compliance with the requirements for freedom in the relevant Terrestrial Code chapter.

In addition, a compartment seeking recognition should submit to the Veterinary Authority a baseline animal health report indicating the presence or absence of OIE listed diseases. This report should be regularly updated to reflect the current animal health situation of the compartment.

Vaccination records including the type of vaccine and frequency of administration should be available to enable interpretation of surveillance data.

The time period for which all records should be kept may vary according to the species and disease(s) for which the compartment was defined.

All relevant information should be recorded in a transparent manner and be easily accessible so as to be auditable by the Veterinary Authority.

Article 4.4.5.

Surveillance for the agent or disease

The surveillance system should comply with Chapter 1.4. on surveillance and the specific recommendations for surveillance for the disease(s) for which the compartment was defined, if available.

If there is an increased risk of exposure to the agent for which the compartment has been defined, the sensitivity of the internal and external surveillance system should be reviewed and, where necessary, increased. At the same time, biosecurity measures in place should be reassessed and increased if necessary.

1. **Internal surveillance**

   Surveillance should involve the collection and analysis of disease/infection data so that the Veterinary Authority can certify that the animal subpopulation contained in all the establishments comply with the defined status of that compartment. A surveillance system that is able to ensure early detection in the event that the agent enters a subpopulation is essential. Depending on the disease(s) for which the compartment was defined, different surveillance strategies may be applied to achieve the desired confidence in disease freedom.
2. **External surveillance**

The biosecurity measures applied in a compartment should be appropriate to the level of exposure of the compartment. External surveillance will help identify a significant change in the level of exposure for the identified pathways for disease introduction into the compartment.

An appropriate combination of active and passive surveillance is necessary to achieve the goals described above. Based on the recommendations of Chapter 1.4., targeted surveillance based on an assessment of risk factors may be the most efficient surveillance approach. Targeted surveillance should in particular include epidemiological units in close proximity to the compartment or those that have a potential epidemiological link with it.

**Article 4.4.6.**

**Diagnostic capabilities and procedures**

Officially-designated laboratory facilities complying with the OIE standards for quality assurance, as defined in Chapter 1.1.3. of the Terrestrial Manual, should be available for sample testing. All laboratory tests and procedures should comply with the recommendations of the laboratory for the specific disease. Each laboratory that conducts testing should have systematic procedures in place for rapid reporting of disease results to the Veterinary Authority. Where appropriate, results should be confirmed by an OIE Reference Laboratory.

**Article 4.4.7.**

**Emergency response and notification**

Early detection, diagnosis and notification of disease are critical to minimize the consequences of outbreaks.

In the event of suspicion of occurrence of the disease for which the compartment was defined, the free status of the compartment should be immediately suspended. If confirmed, the status of the compartment should be immediately revoked and importing countries should be notified following the provisions of Chapter 1.1.

In case of an occurrence of any infectious disease not present according to the baseline animal health report of the compartment referred to in Article 4.4.4., the management of the compartment should notify the Veterinary Authority, and initiate a review to determine whether there has been a breach in the biosecurity measures. If a significant breach in biosecurity, even in the absence of outbreak, is detected, export certification as a free compartment should be suspended. Disease free status of the compartment may only be reinstated after the compartment has adopted the necessary measures to re-establish the original biosecurity level and the Veterinary Authority re-approves the status of the compartment.

In the event of a compartment being at risk from a change, in the surrounding area, in the disease situation for which the compartment was defined, the Veterinary Authority should re-evaluate without delay the status of the compartment and consider whether any additional biosecurity measures are needed to ensure that the integrity of the compartment is maintained.

**Article 4.4.8.**

**Supervision and control of a compartment**

The authority, organisation, and infrastructure of the Veterinary Services, including laboratories, should be clearly documented in accordance with the chapter on the evaluation of Veterinary Services of the Terrestrial Code, to provide confidence in the integrity of the compartment.
Annex VII (contd)

The Veterinary Authority has the final authority in granting, suspending and revoking the status of a compartment. The Veterinary Authority should continuously supervise compliance with all the requirements critical to the maintenance of the compartment status described in this chapter and ensure that all the information is readily accessible to the importing countries. Any significant change should be notified to the importing country.
CHAPTER 4.6.

COLLECTION AND PROCESSING OF BOVINE, SMALL RUMINANT AND PORCINE SEMEN

Article 4.6.1.

General considerations

The purposes of official sanitary control of semen production are to:

1. maintain the health of animals on an artificial insemination centre at a level which permits the international distribution of semen with a negligible risk of infecting other animals or humans with pathogens transmissible by semen;

2. ensure that semen is hygienically collected, processed and stored.

Artificial insemination centres should comply with recommendations in Chapter 4.5.

Standards for diagnostic tests are described in the Terrestrial Manual.

Article 4.6.2.

Conditions applicable to testing of bulls and teaser animals

Bulls and teaser animals should enter an artificial insemination centre only when they fulfil the following requirements.

1. Prior to entering pre-entry isolation facility

The animals should comply with the following requirements prior to entry into isolation at the pre-entry isolation facility where the country or zone of origin is not free from the diseases in question.

a) Bovine brucellosis – Point 3 or 4 of Article 11.3.5.

b) Bovine tuberculosis – Point 3 or 4 of Article 11.6.5.

c) Bovine viral diarrhoea-mucosal disease (BVD-MD)

The animals should be subjected to:

i) a virus isolation test or a test for virus antigen, with negative results; and

ii) a serological test to determine the serological status of every animal.

d) Infectious bovine rhinotracheitis-infectious pustular vulvovaginitis

If the artificial insemination centre is to be considered as infectious bovine rhinotracheitis-infectious pustular vulvovaginitis free (IBR/IPV), the animals should either:

i) come from an IBR/IPV free herd as defined in Article 11.11.3.; or
Annex VIII (contd)

ii) be subjected, with negative results, to a serological test for IBR/IPV on a blood sample.

c) Bluetongue

The animals should comply with Articles 8.3.7. or 8.3.8., depending on the bluetongue status of the country or zone of origin of the animals.

2. Testing in the pre-entry isolation facility prior to entering the semen collection facilities

Prior to entering the semen collection facilities of the artificial insemination centre, bulls and teaser animals should be kept in a pre-entry isolation facility for at least 28 days. The animals should be tested as described below a minimum of 21 days after entering the pre-entry isolation facility, except for Campylobacter fetus subsp. venerealis and Tritrichomonas foetus, for which testing may commence after 7 days in pre-entry isolation. All the results should be negative except in the case of BVD-MD antibody serological testing (see point 2b)i) below).

a) Bovine brucellosis

The animals should be subjected to a serological test with negative results.

b) BVD-MD

i) All animals should be tested for viraemia as described in point 1c) above.

Only when all the animals in pre-entry isolation test negative for viraemia, may the animals enter the semen collection facilities upon completion of the 28-day pre-entry isolation period.

ii) After 21 days in pre-entry isolation, all animals should be subjected to a serological test to determine the presence or absence of BVD-MD antibodies.

iii) Only if no sero-conversion occurs in the animals which tested seronegative before entry into the pre-entry isolation facility, may any animal (seronegative or seropositive) be allowed entry into the semen collection facilities.

iv) If sero-conversion occurs, all the animals that remain seronegative should be kept in pre-entry isolation until there is no more seroconversion in the group for a period of 3 weeks. Serologically positive animals may be allowed entry into the semen collection facilities.

c) Campylobacter fetus subsp. venerealis

i) Animals less than 6 months old or kept since that age only in a single sex group prior to pre-entry isolation should be tested once on a preputial specimen, with a negative result.

ii) Animals aged 6 months or older that could have had contact with females prior to pre-entry isolation should be tested three times at weekly intervals on a preputial specimen, with a negative result in each case.

d) Tritrichomonas foetus

i) Animals less than 6 months old or kept since that age only in a single sex group prior to pre-entry isolation, should be tested once on a preputial specimen, with a negative result.

ii) Animals aged 6 months or older that could have had contact with females prior to pre-entry isolation should be tested three times at weekly intervals on a preputial specimen, with a negative result in each case.
Annex VIII (contd)

e) IBR-IPV

If the artificial insemination centre is to be considered as IBR/IPV free, the animals should be subjected, with negative results, to a diagnostic test for IBR/IPV on a blood sample. If any animal tests positive, the animal should be removed immediately from the pre-entry isolation facility and the other animals of the same group should remain in pre-entry isolation and be retested, with negative results, not less than 21 days after removal of the positive animal.

f) Bluetongue

The animals should comply with the provisions referred to in Articles 8.3.6., 8.3.7. or 8.3.8., depending on the bluetongue status of the country or zone where the pre-entry isolation facility is located.

3. Testing programme for bulls and teasers resident in the semen collection facilities

All bulls and teasers resident in the semen collection facilities should be tested at least annually for the following diseases, with negative results, where the country or zone where the semen collection facilities are located is not free:

a) Bovine brucellosis

b) Bovine tuberculosis

c) BVD-MD

Animals negative to previous serological tests should be retested to confirm absence of antibodies.

Should an animal become serologically positive, every ejaculate of that animal collected since the last negative test should be either discarded or tested for virus with negative results.

d) campylobacter fetus subsp. venerealis

i) A preputial specimen should be tested.

ii) Only bulls on semen production or having contact with bulls on semen production need to be tested. Bulls returning to collection after a lay off of more than 6 months should be tested not more than 30 days prior to resuming production.

e) Bluetongue

The animals should comply with the provisions referred to in Article 8.3.11.

f) Tritrichomonas foetus

i) A preputial specimen should be cultured.

ii) Only bulls on semen production or having contact with bulls on semen production need to be tested. Bulls returning to collection after a lay off of more than 6 months should be tested not more than 30 days prior to resuming production.

g) IBR-IPV

If the artificial insemination centre is to be considered as IBR/IPV free, the animals should comply with the provisions in point 2)c) of Article 11.11.3.
Annex VIII (contd)

4. **Testing for BVD-MD prior to the initial dispatch of semen from each serologically positive bull**

Prior to the initial dispatch of semen from BVD-MD serologically positive bulls, a semen sample from each animal should be subjected to a virus isolation or virus antigen test for BVD-MD. In the event of a positive result, the bull should be removed from the centre and all of its semen destroyed.

5. **Testing of frozen semen for IBR/IPV in artificial insemination centres not considered as IBR/IPV free**

Each aliquot of frozen semen should be tested as per Article 11.11.7.

Article 4.6.3.

**Conditions applicable to testing of rams/bucks and teaser animals**

Rams/bucks and teaser animals should only enter an artificial insemination centre if they fulfil the following requirements.

1. **Prior to entering pre-entry isolation facility**

The animals should comply with the following requirements prior to entry into isolation at the pre-entry isolation facility where the country or zone of origin is not free from the diseases in question.

a) Caprine and ovine brucellosis – Article 14.1.6.

b) Ovine epididymitis – Article 14.7.3.

c) Contagious agalactia – Points 1 and 2 of Article 14.3.1.

d) Peste des petits ruminants – Points 1, 2, and 4 or 5 of Article 14.8.7.

e) Contagious caprine pleuropneumonia – Article 14.4.7., depending on the CCPP status of the country or zone of origin of the animals.

f) Paratuberculosis – Free from clinical signs for the past 2 years.

g) Scrapie – Comply with Article 14.9.8. if the animals do not originate from a scrapie free country or zone as defined in Article 14.9.3.

h) Maedi-visna – Article 14.6.2.

i) Caprine arthritis/encephalitis – Article 14.2.2. in the case of goats.

j) Bluetongue

The animals should comply with Articles 8.3.7. or 8.3.8., depending on the bluetongue status of the country or zone of origin of the animals.

k) Tuberculosis — In the case of goats, a single or comparative tuberculin test, with negative results.

2. **Testing in the pre-entry isolation facility prior to entering the semen collection facilities**

Prior to entering the semen collection facilities of the artificial insemination centre, rams/bucks and teasers should be kept in a pre-entry isolation facility for at least 28 days. The animals should be tested as described below a minimum of 21 days after entering the pre-entry isolation facility, with negative results.
Annex VIII (contd)

a) Caprine and ovine brucellosis – Point 1c) of Article 14.1.8.

b) Ovine epididymitis – Points 1d) and 2 of Article 14.7.4.

c) Maedi-visna and caprine arthritis/encephalitis – Test on animals and semen.

d) Bluetongue

The animals should comply with the provisions referred to in Articles 8.3.6., 8.3.7. or 8.3.8., depending on the bluetongue status of the country or zone where the pre-entry isolation facility is located.

3. Testing programme for rams/bucks and teasers resident in the semen collection facilities

All rams/bucks and teasers resident in the semen collection facilities should be tested at least annually for the following diseases, with negative results, where the country or zone where the semen collection facilities are located is not free:

a) Caprine and ovine brucellosis;

b) Ovine epididymitis;

c) Maedi-visna and caprine arthritis/encephalitis;

d) Tuberculosis (for goats only);

e) Bluetongue.

The animals should comply with the provisions referred to in Article 8.3.11.

Article 4.6.4.

Conditions applicable to testing of boars

Boars should only enter an artificial insemination centre if they fulfil the following requirements.

1. Prior to entering pre-entry isolation facility

The animals should be clinically healthy, physiologically normal and comply with the following requirements within 30 days prior to entry into isolation at the pre-entry isolation facility where the country or zone of origin is not free from the diseases in question.

a) Porcine brucellosis – Article 15.3.3.

b) Foot and mouth disease – Articles 8.5.12., 8.5.13. or 8.5.14.

c) Aujeszky’s disease – Article 8.2.8. or Article 8.2.9.

d) Teschovirus encephalomyelitis – Article 15.5.4. or Article 15.5.6.

e) Transmissible gastroenteritis – Article 15.6.2.

f) Swine vesicular disease – Article 15.4.5. or Article 15.4.7.

g) African swine fever – Article 15.1.5. or Article 15.1.6.
Annex VIII (contd)

h) Classical swine fever – Article 15.2.5. or Article 15.2.6.


2. Testing in the pre-entry isolation facility prior to entering the semen collection facilities

Prior to entering the semen collection facilities of the artificial insemination centre, boars should be kept in a pre-entry isolation facility for at least 28 days. The animals should be subjected to diagnostic tests as described below a minimum of 21 days after entering the pre-entry isolation facility, with negative results.

a) Porcine brucellosis – Article 15.3.5.

b) Foot and mouth disease – Articles 8.5.15., 8.5.16., 8.5.17. or 8.5.18.

c) Aujeszky’s disease – Articles 8.2.12., 8.2.13. or 8.2.14.

d) Teschovirus encephalomyelitis – Article 15.5.8. or Article 15.5.9.

e) Transmissible gastroenteritis – Article 15.6.4.

f) Swine vesicular disease – Article 15.4.9. or Article 15.4.10.

g) African swine fever – Article 15.1.8. or Article 15.1.9.

h) Classical swine fever – Article 15.2.8. or Article 15.2.9.


3. Testing programme for boars resident in the semen collection facilities

All boars resident in the semen collection facilities should be tested at least annually for the following diseases, with negative results, where the country or zone where the semen collection facilities are located is not free:

a) Porcine brucellosis – Article 15.3.5.

b) Foot and mouth disease – Articles 8.5.15., 8.5.16., 8.5.17. or 8.5.18.

c) Aujeszky’s disease – Articles 8.2.12., 8.2.13. or 8.2.14.

d) Teschovirus encephalomyelitis – Article 15.5.8. or Article 15.5.9.

e) Transmissible gastroenteritis – Article 15.6.4.

f) Swine vesicular disease – Article 15.4.9. or Article 15.4.10.

g) African swine fever – Article 15.1.8. or Article 15.1.9.

h) Classical swine fever – Article 15.2.8. or Article 15.2.9.

Annex VIII (contd)

Article 4.6.5.

General considerations for hygienic collection and handling of semen

Observation of the recommendations described in the Articles below will very significantly reduce the likelihood of the semen being contaminated with common bacteria which are potentially pathogenic.

Article 4.6.6.

Conditions applicable to the collection of semen

1. The floor of the mounting area should be clean and provide safe footing. A dusty floor should be avoided.

2. The hindquarters of the teaser, whether a dummy or a live teaser animal, should be kept clean. A dummy should be cleaned completely after each period of collection. A teaser animal should have its hindquarters cleaned carefully before each collecting session. The dummy or hindquarters of the teaser animals should be sanitized after the collection of each ejaculate. Disposable plastic covers may be used.

3. The hand of the person collecting the semen should not come into contact with the animal’s penis. Disposable gloves should be worn by the collector and changed for each collection.

4. The artificial vagina should be cleaned completely after each collection where relevant. It should be dismantled, its various parts washed, rinsed and dried, and kept protected from dust. The inside of the body of the device and the cone should be disinfected before re-assembly using approved disinfection techniques such as those involving the use of alcohol, ethylene oxide or steam. Once re-assembled, it should be kept in a cupboard which is regularly cleaned and disinfected.

5. The lubricant used should be clean. The rod used to spread the lubricant should be clean and should not be exposed to dust between successive collections.

6. The artificial vagina should not be shaken after ejaculation, otherwise lubricant and debris may pass down the cone to join the contents of the collecting tube.

7. When successive ejaculates are being collected, a new artificial vagina should be used for each mounting. The vagina should also be changed when the animal has inserted its penis without ejaculating.

8. The collecting tubes should be sterile, and either disposable or sterilised by autoclaving or heating in an oven at 180°C for at least 30 minutes. They should be kept sealed to prevent exposure to the environment while awaiting use.

9. After semen collection, the tube should be left attached to the cone and within its sleeve until it has been removed from the collection room for transfer to the laboratory.

Article 4.6.7.

Conditions applicable to the handling of semen and preparation of semen samples in the laboratory

1. Diluents
   a) All receptacles used should have been sterilised.
   b) Buffer solutions employed in diluents prepared on the premises should be sterilized by filtration (0.22 µm) or by autoclaving (121°C for 30 minutes) or be prepared using sterile water before adding egg yolk (if applicable) or equivalent additive and antibiotics.
Annex VIII (contd)

c) If the constituents of a diluent are supplied in commercially available powder form, the water used should have been distilled or demineralised, sterilized (121°C for 30 minutes or equivalent), stored correctly and allowed to cool before use.

d) Whenever milk, egg yolk or any other animal protein is used in preparing the semen diluent, the product should be free of pathogens or sterilised; milk heat-treated at 92°C for 3-5 minutes, eggs from SPF flocks when available. When egg yolk is used, it should be separated from eggs using aseptic techniques. Alternatively, commercial egg yolk prepared for human consumption or egg yolk treated by, for example, pasteurisation or irradiation to reduce bacterial contamination, may be used. Other additives should also be sterilized before use.

e) Diluent should not be stored for more than 72 hours at +5°C before use. A longer storage period is permissible for storage at -20°C. Storage vessels should be stoppered.

f) A mixture of antibiotics should be included with a bactericidal activity at least equivalent to that of the following mixtures in each ml of frozen semen: gentamicin (250 µg), tylosin (50 µg), lincomycin-spectinomycin (150/300 µg); penicillin (500 IU), streptomycin (500 µg), lincomycin-spectinomycin (150/300 µg); or amikacin (75µg), divekacin (25µg).

The names of the antibiotics added and their concentration should be stated in the international veterinary certificate.

2. Procedure for dilution and packing

a) The tube containing freshly collected semen should be sealed as soon as possible after collection, and kept sealed until processed.

b) After dilution and during refrigeration, the semen should also be kept in a stoppered container.

c) During the course of filling receptacles for dispatch (such as insemination straws), the receptacles and other disposable items should be used immediately after being unpacked. Materials for repeated use should be disinfected with alcohol, ethylene oxide, steam or other approved disinfection techniques.

d) If sealing powder is used, care should be taken to avoid its being contaminated.

3. Conditions applicable to the storage of semen

Semen for export should be stored separately from other genetic material not meeting the requirements of this chapter with fresh liquid nitrogen in sterilised/sanitised flasks before being exported.

Semen straws should be sealed and code marked in line with the international standards of the International Committee for Animal Recording (ICAR)^1.

Prior to export, semen straws or pellets should clearly and permanently be identified and placed into new liquid nitrogen in a new or sterilised flask or container under the supervision of an Official Veterinarian. The contents of the container or flask should be verified by the Official Veterinarian prior to sealing with an official numbered seal before export and accompanied by an international veterinary certificate listing the contents and the number of the official seal.
4. **Sperm sorting**

   Equipment used for sex-sorting sperm should be clean and disinfected between *animals* according to the recommendations of the licencer of the system.

   Where seminal plasma, or components thereof, is added to sorted semen prior to cryopreservation and storage, it should be derived from *animals* of same or better health status.

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1. The ICAR international standards on straws are contained in Recording Guidelines - Appendices to the international agreement of recording practices. The text of this document is available at the following web site: www.icar.org
CHAPTER 5.2.

CERTIFICATION PROCEDURES

Article 5.2.1.

Protection of the professional integrity of the certifying veterinarian

Certification should be based on the highest possible ethical standards, the most important of which is that the professional integrity of the certifying veterinarian should be respected and safeguarded according to Chapters 3.1. and 3.2.

It is essential to include in any requirements only those specific statements that can be accurately and honestly signed by a certifying veterinarian. For example, these requirements should not include certification of an area as being free from diseases other than those notifiable diseases, or the occurrence of which the signing veterinarian is not necessarily informed about. It is unacceptable to ask for certification for events which will take place after the document is signed when these events are not under the direct control and supervision of the signing veterinarian.

Certification of freedom from diseases based on purely clinical freedom and herd history is of limited value. This is also true of diseases for which there is no specific diagnostic test, or the value of the test as a diagnostic aid is limited.

The note of guidance referred to in Article 5.1.1. is not only to inform the signing veterinarian but also to safeguard professional integrity.

Article 5.2.2.

Certifying veterinarians

Certifying veterinarian should:

1. be authorised by the Veterinary Authority of the exporting country to sign international veterinary certificates;

2. only certify matters that are within their own knowledge at the time of signing the certificate, or that have been separately attested by another competent party;

3. sign only at the appropriate time certificates that have been completed fully and correctly; where a certificate is signed on the basis of supporting documentation, the certifying veterinarian should have verified or be in possession of that documentation before signing;

4. have no conflict of interest in the commercial aspects of the animals or animal products being certified and be independent from the commercial parties.

Article 5.2.3.

Preparation of international veterinary certificates

Certificates should be drawn up in accordance with the following principles:
Annex IX (contd)

1. Certificates should be designed so as to minimize the potential for fraud including use of a unique identification number, or other appropriate means to ensure security. Paper certificates should bear the signature of the certifying veterinarian and the official identifier (stamp) of the issuing Veterinary Authority. Each page of a multiple page certificate should bear the unique certificate number and a number indicating the number of the page out of the total number of pages. Electronic certification procedures should include equivalent safeguards.

2. Certificates should be written using terms that are simple, unambiguous and as easy to understand as possible, without losing their legal meaning.

3. If so required, certificates should be written in the language of the importing country. In such circumstances, they should also be written in a language understood by the certifying veterinarian.

4. Certificates should require appropriate identification of animals and animal products except where this is impractical (e.g. day-old birds).

5. Certificates should not require a veterinarian to certify matters that are outside his/her knowledge or which he/she cannot ascertain and verify.

6. Where appropriate, when presented to the certifying veterinarian, certificates should be accompanied by notes of guidance indicating the extent of enquiries, tests or examinations expected to be carried out before the certificate is signed.

7. The text of a certificate should not be amended except by deletions which should be signed and stamped by the certifying veterinarian.

8. The signature and stamp should be in a colour different from that of the printing of the certificate. The stamp may be embossed instead of being a different colour.

9. Replacement certificates may be issued by a Veterinary Authority to replace certificates that have been, for example, lost, damaged, contain errors, or where the original information is no longer correct. These replacements should be provided by the issuing authority and be clearly marked to indicate that they are replacing the original certificate. A replacement certificate should reference the number and the issue date of the certificate that it supersedes. The superseded certificate should be cancelled and where possible, returned to the issuing authority.

10. Only original certificates are acceptable.

   Article 5.2.4.

Electronic certification

1. Certification may be provided by electronic documentation sent directly from the Veterinary Authority of the exporting country to the Veterinary Authority of the importing country. Such systems also normally provide an interface with the commercial organisation marketing the commodity for provision of information to the certifying authority. The certifying veterinarian should have access to all information such as laboratory results and animal identification data.
2. Electronic certificates may be in a different format but should carry the same information as conventional paper certificates.

3. The Veterinary Authority should have in place systems for the security of electronic certificates against access by unauthorised persons or organisations.

4. The certifying veterinarian should be officially responsible for the secure use of his/her electronic signature.

— text deleted
CHAPTER 6.3.
THE CONTROL OF HAZARDS OF
ANIMAL HEALTH AND
PUBLIC HEALTH IMPORTANCE IN ANIMAL FEED

Introduction

Animal feed is a critical component of the food chain that has a direct impact on animal health and welfare and also on food safety and public health.

Historically, the OIE primarily addressed animal feed as an important pathway for the entry and spread of contagious epidemic diseases, such as foot and mouth disease, swine vesicular disease and avian influenza. In recent years, the role of feed as a vector for disease agents, including zoonotic organisms, was a focus of standards development in regards to bovine spongiform encephalopathy. Animal feed and feed ingredients are widely traded internationally and trade disruptions have the potential to impact economies in both developed and developing countries. Since 2002 the OIE has expanded its zoonotic disease mandate to encompass animal production food safety, working in collaboration with the Codex Alimentarius Commission (CAC) and other international organisations. In 2006 the International Committee resolved that the OIE should develop guidance on foodborne zoonoses and animal feeding, complementing relevant CAC texts.

Objective and scope

The objective of this chapter is to provide guidance on animal feeding in relation to animal health and to complement the guidance provided by the Codex Code of Practice on Good Animal Feeding (CAC/RCP 54-2004) which deals primarily with food safety, and related other Codex texts covering animal feeding, e.g. Code of Practice for Source Directed Measures to Reduce Contamination of Food with Chemicals (CAC/RCP 49-2001).

This chapter aims at ensuring the control of animal and public health hazards through adherence to recommended practices during the production (growing, procurement, handling, storage, processing and distribution) and use of both commercial and on-farm produced animal feed and feed ingredients for terrestrial animals.

This chapter applies to the production and use of all products destined for animal feed and feed ingredients at all levels whether produced commercially or on farm. It also includes grazing or free-range feeding, forage crop production and water for drinking. Swill feeding is a particular aspect of on-farm practice that is specifically addressed because of its recognised role in disease transmission.

This chapter deals with feed for terrestrial animals (except bees).

Definitions

Contamination: means the unwanted presence of a material, infectious agent or product in a feed or feed ingredient that is potentially harmful to animal or public health or restricted under current regulations.
Annex X (contd)

Feed: means any material (single or multiple), whether processed, semi-processed or raw, which is intended to be fed directly to terrestrial animals (except bees).

Feed additive: means any intentionally added ingredient not normally consumed as feed by itself, whether or not it has nutritional value or other effect on the animal, which affects the characteristics of feed or of the animal products. Microorganisms, enzymes, pH regulators, trace elements, vitamins and other products fall within the scope of this definition depending on the purpose of use and method of administration. This excludes veterinary drugs.

Feed ingredient: means a component part or constituent of any combination or mixture making up a feed, whether or not it has a nutritional value in the animal's diet, including feed additives. Ingredients are of plant (including aquatic plants) or terrestrial or aquatic animal origin, or other organic or inorganic substances.

General principles

1. Roles and responsibilities

The Competent Authority has the legal power to set and enforce regulatory animal feeding requirements, and has final responsibility for verifying that these requirements are met. The Competent Authority may establish regulatory requirements for relevant parties to provide it with information and assistance. Refer to Chapters 3.1. and 3.2. of the Terrestrial Code.

Those involved in the production and use of animal feed and feed ingredients have the responsibility to ensure that these products meet regulatory requirements. Appropriate contingency plans should be in place to enable tracing and recall of non-compliant products. All personnel involved in the manufacture, storage and handling of feed and feed ingredients should be adequately trained and aware of their role and responsibility in preventing the introduction or spread of hazards. Manufacturing equipment, storage and transport facilities should be adequate and maintained in good working order and in a sanitary condition.

Those providing specialist services to producers and to the feed industry (e.g. private veterinarians, nutritionists and laboratories) may be required to meet specific regulatory requirements pertaining to the services they provide (e.g. disease reporting, quality standards, transparency).

2. Regulatory safety standards

All feed and feed ingredients should meet regulatory safety standards. Scientific evidence, including the sensitivity of analytical methods and on the characterisation of risks, should be taken into account in defining limits and tolerances for hazards.

3. Risk analysis (risk assessment, risk management and risk communication)

Internationally accepted principles and practices on risk analysis (Section 2 of the Terrestrial Code and relevant Codex texts) should be used in developing and applying the regulatory framework.

Application of a generic framework should provide a systematic and consistent process for managing all biosecurity risks, while recognising the different risk assessment methodologies used in animal and public health.

4. Good practices

Where national guidelines exist, good agricultural practices and good manufacturing practices (including good hygienic practices) should be followed. Countries without such guidelines are encouraged to develop them or adopt suitable international standards or recommendations.
Where appropriate, Hazard Analysis and Critical Control Point (HACCP) principles should be followed to control hazards that may occur in the manufacture, distribution and feeding of feed and feed additives and feed ingredients.

5. **Geographic and environmental considerations**

Epidemiological links between potential sources of hazards for animal health or food safety should be considered when assessing water sources, land or facilities for suitability for the production of animal feed and feed ingredients. Animal health considerations include factors such as disease status, location of quarantined premises and existence of zones/compartments of specified health status. Food safety considerations include factors such as industrial operations that generate pollutants and waste treatment plants.

6. **Zoning and compartmentalisation**

Feed is an important component of biosecurity and needs to be considered when defining a compartment or zone in accordance with Chapter 4.3. of the Terrestrial Code.

7. **Sampling and analysis**

Sampling and analysis should be based on scientifically recognised principles and procedures.

8. **Labelling**

Labelling should be informative, unambiguous, legible and conspicuously placed on the package if sold in package form and on the waybill and other sales documents if sold in bulk, un-packaged form, and should comply with regulatory requirements and Section 4.2.10 Labelling of Codex Code of Practice on Good Animal Feeding (CAC/RCP 54-2004), including listing of ingredients and instructions on the handling, storing and use. All claims made on a label should be able to be substantiated.

9. **Design and management of inspection programmes**

In meeting animal and public health objectives prescribed in national legislation or required by importing countries, Competent Authorities contribute through the inspection or through the auditing of animal and public health activities conducted by other agencies or the private sector.

Feed and feed ingredients business operators and other relevant parts of industry should practice self-regulation to secure compliance with required standards for procurement, handling, storage, processing, distribution and use. Operators have full responsibility for implementing systems for quality control. The Competent Authority should verify that process control systems and safety standards achieve all regulatory requirements.

10. **Assurance and certification**

Feed business operators are responsible for demonstrating the safety of the establishments under their control. Competent Authorities are responsible for providing assurances domestically and to trading partners that regulatory safety standards have been met. For international trade in animal product based feeds, Veterinary Services are required to provide international veterinary certificates.

11. **Hazards associated with animal feed**

a) **Biological hazards**

Biological hazards that may occur in feed and feed ingredients include agents such as bacteria, viruses, prions, fungi and parasites.
Annex X (contd)

b) **Chemical hazards**

Chemical hazards that may occur in feed and feed ingredients include naturally occurring chemicals (such as mycotoxins and gossypol), industrial and environmental contaminants (such as dioxins and PCBs), residues of veterinary drugs and pesticides and also radionuclides.

c) **Physical hazards**

Physical hazards that may occur in feed and feed ingredients include foreign objects (such as pieces of glass, metal, plastic or wood).

12. **Contamination**

Procedures to minimise the risk of contamination during the manufacture, production, processing, storage, distribution (including transport) and the use of feed and feed ingredients should be included in current regulations and standards. Scientific evidence, including the sensitivity of analytical methods and on the characterisation of risks, should be drawn upon in developing this framework.

Procedures, such as flushing, sequencing and physical clean-out, should be used to reduce the likelihood of contamination between batches of feed or feed ingredients.

13. **Antimicrobial resistance**

Concerning the use of antimicrobials in animal feed refer to Chapters 6.7. to 6.10. of the Terrestrial Code.

14. **Management of information**

The Competent Authority should establish clear requirements for the provision of information by the private sector as this relates to regulatory requirements.

Records should be maintained in a readily accessible form regarding the production, distribution and use of feed and feed ingredients. These records are required to facilitate the prompt trace-back of feed and feed ingredients to the immediate previous source, and trace-forward to the next subsequent recipients, to address identified animal health or public health concerns (see Section 4.3. of CAC/RCP 54-2004).

*Animal identification* and *animal traceability* are tools for addressing animal health (including zoonoses), and food safety risks arising from animal feed (see Chapters 4.1. and 4.2. of the Terrestrial Code).
CHAPTER 5.X.

CONTROL OF OIE LISTED DISEASES IN HEAT TREATED, SHELF STABLE PET FOOD

Article 5.X.1.

Objective and scope

The objective of this chapter is to provide specific guidance on preventing the transfer of OIE listed diseases (Chapter 1.2.) through international trade in pet food. The chapter should be read in conjunction with Chapter 6.3. of the Terrestrial Code.

Pet food means any commercial feed prepared and distributed for consumption by dogs or cats. This chapter refers to heat-treated, shelf stable pet food (hereafter referred to as pet food). The finished product, in an unopened container, can exist at room temperature for an extended time period.

The chapter aims at ensuring the control of OIE listed diseases through adherence to recommended measures during the production pet food, including pet treats (snacks) and pet chews.

For the purpose of this chapter, “pets” are limited to dogs or cats.

Article 5.X.2.

Pet food specific measures

An important consideration with pet food is that ingredients from multiple animal species, often sourced from multiple countries, zones or compartments are combined into the final product. However, as the products covered in this chapter have been heat-treated, the products themselves would not pose significant animal health risk when compared to unprocessed products coming from the same countries, zones or compartments.

When determining the appropriate import requirements, the potential animal health concerns of all species and ingredients of animal origin need to be addressed.

The Competent Authority should take into account the following factors:

1. Sanitary measures should be based on the relevant chapters of the Terrestrial Code and according to the animal health status of the country, zone or compartment of origin of the animal-derived ingredients. The source of all animal-derived ingredients should be considered. All ingredients should meet OIE requirements, taking into account the end use.

2. When the ingredients cannot be certified as originating from a safe source, thermal treatment can be used for risk mitigation. The table in Article 3 can be used to determine the appropriate disease risk mitigation measures. These treatments should not be cumulative, only the most stringent treatment should apply and will address all identified animal health risks.

3. Quality assurance in the processing facility should be sufficient to verify that the product has been treated as required. The facility should maintain processing records, and the system should provide alert if minimum processing is not accomplished.

4. After processing, the product should be handled in a manner designed to prevent contamination of finished product by unprocessed materials.

5. Processing facilities should have procedures in place to enable tracing and recall of non-compliant products.
Annex XI (contd)

Article 5.X.3.

Elimination of biological hazards from pet food

Biological hazards in pet food may be avoided or eliminated by a number of treatments such as those listed in Table 1. However, other processes determined to be equivalent should be accepted.

Table 1. Risk mitigation measures for processing of pet foods containing ingredients of animal origin (under study)

<table>
<thead>
<tr>
<th>Biological Hazard</th>
<th>Bovine</th>
<th>Ovine</th>
<th>Caprine</th>
<th>Porcine</th>
<th>Equine</th>
<th>Poultry</th>
<th>Egg</th>
<th>Milk</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bluetongue</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
</tr>
<tr>
<td>Foot and mouth disease</td>
<td>70°C/30min (Article 8.3.34)</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>(Article 8.5.26)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Rift Valley fever</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
</tr>
<tr>
<td>Rinderpest</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
</tr>
<tr>
<td>Vesicular stomatitis</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
</tr>
<tr>
<td>Avian influenza</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>60°C/507 sec</td>
<td>70°C/3.5 sec</td>
<td>74°C/0.51 sec (Article 10.4.26)</td>
</tr>
<tr>
<td>Newcastle disease</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>65°C/14 min</td>
<td>74°C/5 min (Article 10.13.21)</td>
<td>57°C/26.6 min (Article 10.13.2)</td>
</tr>
<tr>
<td>Infectious bursal disease</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
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<td>NR</td>
<td>NR</td>
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<tr>
<td>Bovine spongiform encephalopathy</td>
<td>Safe commodities (Article 11.6.1)</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
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<tr>
<td>Contagious bovine pleuropneumonia</td>
<td>(Article 11.8.2)</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
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<tr>
<td>African horse sickness</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
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<tr>
<td>Pest des petits ruminants</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
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<tr>
<td>African swine fever</td>
<td>NR</td>
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<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
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<tr>
<td>Classical swine fever</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>NR</td>
<td>70°C/0°C/ internal pH&lt;6 (Article 15.2.2)</td>
<td>NR</td>
<td>NR</td>
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<tr>
<td>Swine vesicular disease</td>
<td>NR</td>
<td>NR</td>
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NR means no sanitary measures should be imposed.
CHAPTER 6.4.

BIOSECURITY PROCEDURES IN POULTRY PRODUCTION

Introduction

This chapter provides recommended biosecurity procedures in poultry production and is not specifically related to trade.

Infectious agents of poultry are a threat to poultry health and, at times, human health and have significant social and economic implications. In poultry production, especially under intensive conditions, prevention is the most viable and economically feasible approach to the control of infectious agents.

Biosecurity procedures should be implemented with the objective of preventing the introduction and dissemination of infectious agents in the poultry production chain. Biosecurity will be enhanced with the adoption and implementation of the principles of Good Agricultural Practices and the Hazard Analysis Critical Control Point (HACCP) system will help to achieve these objectives.

Purpose and scope

This chapter deals with biosecurity procedures in poultry production. It should be read in conjunction with the Codex Alimentarius Code of Hygienic Practice for Meat (CAC/RCP 58-2005) and Code of Hygienic Practice for Eggs and Egg Products (CAC/RCP 15-1976 Revision 2007).

This chapter provides general recommendations for infectious agents of poultry. Recommendations on specific diseases may be found in relevant disease chapters in the Terrestrial Code.

This chapter identifies several relevant biosecurity measures. The choice of measures to be implemented will vary according to national conditions, including poultry disease status, the risk of introduction and dissemination of infectious agents and the cost effectiveness of control measures.

Recommendations on specific infectious agents may be found in relevant disease chapters in the Terrestrial Code.

Definitions (for this Chapter only)

Breeders: means poultry destined for the production of fertile eggs for incubation for the purpose of producing day-old birds.

Culling: means the depopulation of a flock before the end of its normal production period.

Live bird markets: means markets where live birds from various sources are sold for slaughter, or further rearing or production.
Recommendations on the location and construction of poultry establishments

1. All establishments (poultry farms and hatcheries)
   a) A suitably isolated geographical location is recommended, taking into account factors to consider include the direction of the prevailing winds, location of other poultry and livestock establishments, wild bird concentrations and the distance from roads used to transport poultry.
   b) Poultry establishments should be located and constructed to provide adequate drainage away from the site. Run-off or untreated site wastewater should not discharge into waterfowl habitats.
   c) Poultry houses and hatcheries should be designed and constructed (preferably of smooth impervious materials) so that cleaning and disinfection can be carried out effectively. Ideally, the area immediately surrounding the poultry houses and hatcheries should be paved with concrete or other impervious material to facilitate cleaning and disinfection.
   d) The establishment should be surrounded by a security fence to prevent the entry of unwanted animals and people.
   e) A sign indicating restricted entry should be posted at the entrance to the farm establishment.

2. Additional measures for poultry farms
   a) Establishments should be designed for use with to house a single species and a single production type purpose. Whenever possible, the design should also consider the ‘all-in all-out’ single age group principle should be used. If this is not feasible and several flocks are maintained on one establishment, the establishment should be designed so that each flock can be managed as a separate epidemiological unit.
   b) Poultry houses, and buildings used to store feed, or eggs, or other materials, should be constructed and maintained to prevent the entry of wild birds, rodents and insects arthropods.
   c) Where feasible, the floors of poultry houses should be constructed using concrete or other impervious materials and designed so that cleaning and disinfection can be carried out effectively.
   d) Where feasible, feed should be delivered into the farm from outside the security fence.

3. Additional measures for hatcheries
   a) The design of the hatchery should take account of work flow and air circulation needs, with ‘one way flow’ movement of eggs and day-old birds and one way air flow in the same direction.
   b) The hatchery buildings should include physical separation of areas used for the following:
      i) personnel changing, showering and sanitary facilities;
      ii) receipt, storage and transfer of eggs;
      iii) incubation;
Annex XII (contd)

iv) hatching;
v) sorting, sexing and other handling placing of day-old birds in boxes;
vi) storage of egg boxes and chick boxes for day-old birds, egg flats, chick box pads, liners, chemicals and other items;
vii) washing equipment washing;
viii) waste disposal;
ix) dining facilities for personnel;
x) office space.

Article 64.5.

Recommendations applicable to the operation of poultry establishments

1. All establishments (poultry farms and hatcheries)

a) All establishments should have a written biosecurity plan. Personnel in the establishments should have access to basic training in biosecurity relevant to poultry production and understand the implications to animal health, human health and food safety.

b) There should be good communication between all those personnel involved in the poultry production chain from breeding to production and consumption to ensure that steps are taken to minimise the introduction and dissemination of infectious disease agents. Personnel should have access to basic training in biosecurity relevant to poultry production and food safety.

c) Traceability at all levels of the poultry production chain should be possible.

d) Records of production should be maintained, on an individual flock basis and include data on bird health, production. On farm, this includes cleaning and disinfection, treatment medications, vaccination, flock history, mortality and disease surveillance data. This should be maintained on an individual flock basis. In hatcheries, relevant records should include data on fertility, hatchability, vaccination and treatments. Records should be readily available for inspection on site.

e) A veterinarian should be responsible for monitoring of poultry health on the establishment should be under the supervision of a veterinarian.

f) Access to the establishment should be controlled to ensure only authorised persons and vehicles enter the site.

g) Establishments should be free from control unwanted vegetation and be free from debris.

h) Procedures for the prevention of entry of wild birds into poultry houses and buildings, and the control of vermin such as rodents and arthropods should be implemented on a routine basis.

i) Access to the establishment should be controlled to ensure only authorised persons and vehicles enter the site.

j) All personnel and visitors entering an establishment should follow a biosecurity procedure. The preferred procedure is for visitors and personnel entering the establishment to shower and change into clean clothes and footwear provided by the establishment. Where this is not practical, clean outer garments (coveralls or overalls, head covering hats and footwear) should be provided.
Before entering and after leaving a poultry house, personnel and visitors should wash their hands with soap and water using a properly maintained disinfectant footbath. The disinfectant solution in the footbath should be changed on a regular basis to ensure its efficacy, according to the manufacturer's instructions.

i) Personnel and visitors should not have had recent contact with other poultry, poultry waste, or poultry processing plant(s). This time period should be based on the level of risk of transmission of infectious disease agents. This will depend on the poultry production purpose, biosecurity procedures and disease infection status (e.g. the time between visiting a breeder flock and then a broiler flock would be less than the time between visiting a broiler flock and then a breeder flock).

j) Delivery vehicles should be cleaned, and disinfected before loading each consignment of hatching eggs, day-old birds or poultry.

2. Additional measures for all poultry farms

a) Whenever possible, the 'all-in all-out' single age group principle should be used. If this is not feasible, and several flocks are maintained on one establishment, each flock should be managed as a separate epidemiological unit.

b) All personnel and visitors entering a poultry house should wash their hands with soap and water or sanitize them using a disinfectant. Personnel and visitors should also change footwear, use a boot spray or use a properly maintained disinfectant footbath. The disinfectant solution in the footbath should be changed on a regular basis to ensure its efficacy, according to the manufacturer's instructions.

c) Animals, other than poultry of the appropriate (resident) species and age, should not be permitted access to poultry houses. No animals should have access to other buildings (e.g. those used to store feed, eggs or other material).

d) The drinking water supply to poultry houses should be potable according to the World Health Organization or to the relevant national standard, and microbiological quality should be monitored if there is any reason to suspect contamination. The water delivery system should be cleaned and disinfected between flocks when the poultry house is empty.

e) Birds used to stock a poultry house should preferably be obtained from breeder flocks and hatcheries that are free from vertically transmitted infectious agents.

f) Heat treated feeds with or without the addition of other bacteriostatic or bacteriocidal treatments (e.g. addition of organic acids) is recommended (e.g. organic acids). Where heat treatment is not possible, the use of bacteriostatic or bactericidal treatments is recommended. Feed should be stored in a manner to prevent access by wild birds and rodents. Spilled feed should be cleaned up immediately to remove attractants for wild birds and rodents. The movement of feed between flocks should be avoided.

g) The litter in the poultry house should be kept dry and in good condition.

h) Dead birds should be removed from poultry houses as quickly as possible but at least daily. These should be disposed of in a safe and effective manner.

i) Personnel involved in the catching of birds should be adequately trained in bird handling and basic biosecurity procedures.
h) To minimise stress, poultry should be transported in well ventilated containers and should not be overcrowded. Exposure to extreme temperatures should be avoided.

i) Containers should be cleaned and disinfected between each use.

j) When a poultry house is depopulated, it is recommended that all faeces and litter be removed from the house and disposed of in a safe manner to minimise the risk of dissemination of infectious agents approved by the Veterinary Services.

If litter is not removed and replaced between flocks then the litter should be treated in a manner to inactivate infectious disease agents, to prevent minimise the risk of dissemination of infectious disease agents from one flock to the next.

After removal of faeces and litter, cleaning and disinfection of the poultry house building and equipment should be done in accordance with Chapter 4.13.

All litter removed from a poultry house should be disposed of in a safe manner to prevent the dissemination of infectious agents.

km) For poultry flocks that are allowed to range outdoors, feeders, feed and other items which may attract wild birds should be kept indoors. Attractants to wild birds should be minimised e.g. feeders should be kept inside the poultry house. Poultry should not be allowed access to sources of contamination (e.g. household waste, litter storage areas, other farm animals, stagnant water and water of unknown quality and litter storage areas). The nesting area should be inside the poultry house.

ln) To avoid the development of antimicrobial resistance, antimicrobials should be used according to relevant directions of the Veterinary Services and manufacturer’s instructions and in accordance with Terrestrial Code Chapters 6.8, 6.9, 6.10, and 6.11.

3. Additional measures for layers

Refer to Section 3 of the Codex Alimentarius Code of Hygienic Practice for Eggs and Egg Products (CAC/RCP 15-1976).

44. Additional measures for breeders farm

a) Nest box litter and liners should be kept clean.

b) Hatching eggs should be collected at frequent intervals, at least daily, and placed in new or clean and disinfected packaging material.

c) Grossly dirty, broken, cracked, broken, or leaking eggs should be collected separately and should not be used as hatching eggs.

d) Hatching eggs should be cleaned and sanitized as soon as possible after collection using an approved sanitising agent, in accordance with the manufacturer’s instructions.

e) Hatching eggs or their packaging materials should be marked to assist traceability and veterinary investigations.
Annex XII (contd)

f) The sanitised hatching eggs should be stored in a dedicated room as soon as possible after cleaning and sanitisation collection. Storage conditions should minimise the potential for microbial contamination and growth and ensure maximum hatchability. The room should be well ventilated, kept clean, and regularly disinfected using disinfectants approved for this purpose.

45. Additional measures for hatcheries

a) Dead in shell embryos should be removed from hatcheries as soon as they are found and disposed of in a safe and effective manner.

b) All hatchery waste, garbage and discarded equipment should be contained or at least covered while on site and removed from the hatchery and its environs as soon as possible.

c) After use, hatchery equipment, tables and surfaces should be promptly and thoroughly cleaned and disinfected with an approved disinfectant.

d) Egg handlers, chick sexers and chick handlers of day-old birds should wash their hands with soap and water before commencing work and between working with batches of hatching eggs or day-old birds from different breeder flocks.

e) Hatching eggs and day-old birds from different breeder flocks should be kept separate identifiable during incubation, hatching, sorting and transportation.

f) Day-old birds should be delivered to the farm in new containers or in clean, disinfected containers.

Article 64.6.

Prevention of further dissemination of infectious disease agents of poultry

When a flock is suspected to be infected or determined to be infected, in addition to the general biosecurity measures described previously, management procedures should be adjusted to effectively isolate the infected flock from other flocks on the establishment and other epidemiologically related establishments. The following measures are recommended:

1. Personnel should be trained in the management of suspected or infected flocks to prevent minimise the risk of the dissemination of infectious disease agents to other flocks and establishments, and to humans. Relevant measures include handling of an infected flock separately, last in sequence and the use of dedicated personnel, and clothing and equipment.

2. A veterinarian should be consulted immediately.

3. When infection has been confirmed, Epidemiological investigations should be carried out to determine the origin and route of transmission of the infectious disease agent.

Poultry litter/faeces and other potentially contaminated farm waste should be disposed of in a safe manner to prevent minimise the risk of dissemination of infectious disease agents. The disposal method used will depend on the infectious agent involved.
Depending on the epidemiology of the infectious agent, the results of a risk assessment, and public and animal health policies, culling, destruction or slaughter of a flock before the end of the normal production period may be used to manage infected flocks. When infected flocks are destroyed or slaughtered they should be processed in a manner to minimise exposure of humans and other flocks to the infectious agent, and in accordance with recommendations of the Veterinary Service and relevant Chapters in the Terrestrial Code. Based on risk assessment, non-infected, high risk flocks may be culling, destroyed or slaughtered before the end of their normal production period. Movement of culled poultry should only be allowed for slaughter or destruction.

Before restocking, the poultry house including equipment or establishment should be cleaned, disinfected and tested to verify that the cleaning has been effective. Special attention should be paid to feed equipment and water systems.

Microbiological monitoring of the efficacy of disinfection procedures is recommended when pathogenic agents have been detected in the previous flock.

Depending on the epidemiology of the infectious agent, risk assessment, vaccine availability and public and animal health policies, vaccination is an option to minimise the dissemination of the infectious agent. When used, vaccines poultry should be administered vaccinated in accordance with the directions of the Veterinary Services and the manufacturer's instructions. Recommendations in the Terrestrial Manual should be followed as appropriate.

Article 6.4.7.

Recommendations to prevent the dissemination of infectious disease agents to and from live bird markets

1. Personnel should be educated on the significance of infectious agents and the need to apply biosecurity practices to prevent dissemination of these agents. Education should be targeted to personnel at all levels of operations in these markets (e.g. drivers, owners, handlers, processors).

Programmes should be implemented to raise consumer awareness of consumers about the risks associated with activities of live bird markets

2. Personnel should wash their hands with soap and water before and after handling birds.

3. Birds from diseased flocks should not be transported to live bird markets.

4. All containers and vehicles should be cleaned and disinfected every time they leave the market.

5. Live birds that leave the market and go to a farm should be housed separately from other birds for a period of time to minimise the potential dissemination of infectious agents of poultry.

6. Periodically the market should be emptied, cleaned and disinfected. This is of particular importance when an infectious agent of poultry deemed significant by the Veterinary Services has been identified in the market or the region.
Where feasible, surveillance should be carried out in these markets to detect infectious disease agents of poultry, especially those agents of zoonotic significance. The surveillance programme should be determined by the Veterinary Services, and in accordance with recommendations in relevant disease specific chapters of the Terrestrial Code.

Attempts should be made to ensure the possibility of tracing all birds entering and leaving the markets.
CHAPTER 6.5.

PREVENTION, DETECTION AND CONTROL OF SALMONELLA IN POULTRY

Article 6.5.1.

Introduction

This Chapter provides recommendations on the prevention, detection and control of Salmonella in poultry.

Salmonellosis is one of the most common foodborne bacterial diseases in the world. The great majority of Salmonella infections in humans are foodborne with Salmonella Enteritidis and Salmonella Typhimurium accounting for a major part of the problem. Salmonella serotypes and prevalence may vary considerably between localities, districts, regions and countries and therefore, surveillance and identification of the prevalent Salmonella serotypes in humans and poultry should be carried out in order to develop a control programme for the area.

In most food animal species, Salmonella can establish a clinically inapparent infection of variable duration, which is significant as a potential zoonosis. Such animals may be important in relation to the spread of infection between flocks and as causes of human foodborne infection. In the latter case, this can occur when meat and eggs, or their products, enter the food chain thus producing contaminated food.

Article 6.5.2.

Purpose and scope

This Chapter deals with methods for on farm prevention, detection and control of Salmonella in poultry, and complements the Codex Alimentarius Code of Hygienic Practice for Meat (CAC/RCP 58-2005) and Code of Hygienic Practice for Eggs and Egg Products (CAC/RCP 15-1976). A pathogen reduction strategy at the farm level is seen as the first step in a continuum that will assist in reducing the presence of foodborne pathogens in eggs and meat.

Hygiene and biosecurity procedures to be implemented in poultry farms and hatcheries are described in Chapter 6.4. Hygiene and Biosecurity Procedures in Poultry Production.

The recommendations presented in this Chapter are relevant to the control of all Salmonella with special attention to S. Enteritidis and S. Typhimurium, as these are common Salmonella serotypes in many countries. It should be noted that the epidemiology of animal and human salmonellosis in a particular locality, district, region or country is important for effective control of Salmonella.

Article 6.5.3.

Definitions (for this Chapter only)

Breeders: means poultry destined for the production of fertile eggs for incubation for the purpose of producing day-old birds.

Competitive exclusion: means the administration of defined or undefined bacterial flora to poultry to prevent gut colonisation by enteropathogens, including Salmonella.
Annex XII (contd)

Culling: means the depopulation destruction or slaughter of a flock before the end of its normal production period.

Layers: means poultry during the period of laying eggs for human consumption.

Article 6.5.4.

Surveillance of poultry flocks for Salmonella

Where justified by risk assessment, surveillance should be carried out to identify infected flocks in order to take measures that will reduce the prevalence in poultry and the risk of transmission of Salmonella to humans. Sampling methods, frequency and type of samples required should be determined by the Veterinary Services based on a risk assessment. Microbiological testing is preferred to serological testing because of its higher sensitivity in broilers flocks and higher specificity in breeders and layer flocks. In the framework of regulatory programmes for the control of Salmonella in poultry and salmonellosis in humans, confirmatory testing may be required.

Sampling

1. Available methods for sampling

Drag swabs: sampling is done by dragging swabs throughout the poultry building house.

Boot swabs: sampling is done by walking throughout the poultry building house with absorbent material placed over the footwear of the sampler.

Dust samples: sampling is done by collecting dust from exhaust fans, screens and other equipment in the poultry building house.

Faecal samples: multiple fresh faecal/caecal samples collected from different areas in the poultry building house.

Meconium, chick box liners papers, dead in shell and culled chicks day-old birds at the hatchery.

Hatchery samples: throughout the hatchery, including inside the incubators.

2. Sample size

Refer to the Terrestrial Manual (under development).

3. Laboratory methods

Refer to the Terrestrial Manual (under development).

4. Time and frequency of testing

Time and frequency of sampling for each poultry type are listed below:
Annex XII (contd)

a) Breeders and hatcheries
   i) Breeder flocks before lay
      • Before the end of the first week of life when the status of the breeding flock and/or the hatchery is not known or does not comply with this chapter.
      • Within the four weeks before being moved to another house, or before going into production if the birds will remain in the same house for the production period.
      • One or more times during the growing period if there is a culling policy in place. The frequency would be determined on commercial considerations.
   ii) Breeder flocks in lay
       • At least at monthly intervals during the laying period.
       • Additional testing should be determined by the Veterinary Services.
   iii) Hatcheries
       • Testing at hatcheries should complement on farm testing.
       • The minimal frequency should be determined by the Veterinary Services.

b) Poultry for the production of eggs for human consumption
   i) Flocks grown to be layers
      • Before the end of the first week of life when the status of the breeding flock and/or the hatchery is not known or does not comply with this chapter.
      • Within the four weeks before being moved to another house, or before going into production if the birds will remain in the same house for the production period.
      • One or more times during the growing period if there is a culling policy in place. The frequency would be determined by commercial considerations.
   ii) Layer flocks
       • At expected peak of lay for each production cycle (the period of time in the laying cycle when the production of the flock is highest).
       • One or more times if there is a culling policy in place or if eggs are diverted to processing for the inactivation of the pathogen. The minimal frequency should be determined by the Veterinary Services.
Annex XII (contd)

c) Poultry for the production of meat

i) Flocks should be sampled at least once before slaughter.

ii) When sampling occurs on farms and when there is a long period (2 weeks or more) between thinning and final depopulation further testing should be considered.

iii) When sampling occurs on farms, flocks should be sampled as late as possible before the first birds are transported to the slaughterhouse. In order to allow for the implementation of control measures during processing, this should be done at a time that ensures the results are available before slaughter.

Whether sampling occurs on the farm which is more appropriate for consequent control measures or at the processing plant, there should be an integrated system in place that allows for investigation of the source of positive flocks.

d) Testing of empty building poultry houses testing

i) Bacteriological monitoring of the efficacy of disinfection procedures is recommended when Salmonella have been detected in the previous flock.

As appropriate, sampling of equipment and surfaces as well as boot swabs or drag swabs of the empty building poultry house after depopulation, cleaning and disinfection.

Results from surveillance may lead to the implementation of additional prevention and control measures to reduce the risk of transmission of Salmonella to humans:

a) In breeders, control measures may be implemented to reduce the transmission of Salmonella to the next generation, especially for trans-ovarian transmitted serotypes such as S. Enteriditis.

b) In layer flocks control measures will reduce and may eliminate contamination of eggs with Salmonella.

c) In poultry for meat production, control measures may be implemented at slaughter or further down the food chain.

Article 6.5.5

Prevention and Control measures

Salmonella prevention and control may be achieved by adopting Good Agricultural Practices and Hazard Analysis Critical Control Point (HACCP), and general measures detailed in Chapter 6.4. Hygiene and Biosecurity Procedures in Poultry Production, in combination with the following additional measures, where appropriate. No single measure used alone will achieve effective Salmonella control.

Additional prevention and control measures include: vaccination, competitive exclusion, flock culling, use of organic acids, culling and product diversion to processing.

Antimicrobials should not be used to control infection with Salmonella in poultry because the effectiveness of the treatment is limited, may mask the infection at sampling, has the potential to produce residues in meat and eggs and can contribute to the development of antimicrobial resistance. Antimicrobials may also reduce normal flora in the gut and increase the likelihood of colonisation with Salmonella. In special circumstances antimicrobials may be used to salvage birds with high genetic value.
1. Day-old birds used to stock a poultry house should be obtained from breeding flocks and hatcheries that have been monitored according to this Chapter and in which no evidence of S. Enteritidis and S. Typhimurium has been detected.

2. Layer and breeder flocks should be stocked from flocks that have been monitored according to this chapter and in which no evidence of S. Enteritidis and S. Typhimurium has been detected.

3. Feed contamination with Salmonella is known to be a source of infection for poultry. Therefore, it is recommended to monitor the Salmonella status of poultry feed, and if found positive to take corrective measures.

   The use of heat treated feeds with or without the addition of or feeds subjected to other bacteriostatic or bactericidal treatments (e.g. addition of organic acids) is recommended. Where heat treatment is not possible, the use of bacteriostatic or bactericidal treatments is recommended.

   Feed should be stored in clean closed containers to prevent access by wild birds and rodents. Spilled feed should be cleaned up immediately to remove attractants for wild birds and rodents.

4. Competitive exclusion may be used in day-old birds to reduce colonisation by Salmonella.

   When used, competitive exclusion should be administered according to the instructions provided by the manufacturer and in accordance with the standards and recommendations of the Veterinary Services.

5. Vaccines are used against Salmonella infections caused by different serotypes in various poultry species, including single or combined vaccines. Vaccines produced according to the Terrestrial Manual should be used.

   If live vaccines are used it is important that field and vaccine strains be easily differentiated in the laboratory. If serology is used as the surveillance method, it may not be possible to distinguish between vaccination and infection with a field strain.

   Vaccination can be used as part of an overall Salmonella control programme. It is recommended that vaccination not be used as the sole control measure.

   When the status of the breeding flock farm and/or the hatchery from which the flock originates is not known or does not comply with this Chapter, vaccination of flocks, starting with day-old birds, against the Salmonella serotypes known to be significant should be considered.

   Vaccination against the Salmonella serotypes known to be significant should be considered when moving day-old birds to a previously contaminated shed so as to minimise the risk of the birds contracting Salmonella infection.

   When used, vaccines should be administered according to the instructions provided by the manufacturer and in accordance with the standards and recommendations of the Veterinary Services.

   Vaccination against S. Enteritidis can cause cross reactions in Salmonella Pullorum/S. Gallinarum serological tests and needs to be considered when implementing measures for these pathogens.

6. Depending on animal health, risk assessment, and public health policies, culling is an option to manage infected breeder and layer flocks. Infected flocks should be destroyed or slaughtered and processed to minimise human exposure to Salmonella.

   If culling is not applied poultry are not culled, eggs for human consumption should be diverted for processing for inactivation of Salmonella.
7. *S. Enteritidis* is characterised by its ovarian transmission pattern. Countries should set targets for eradicating (or significantly reducing) *S. Enteritidis* from egg-producing flocks through a guided policy for eradication from the top of the production pyramid, i.e. from grandparent flocks through breeder flocks to layer flocks.

8. The responsible veterinarian should evaluate the results of surveillance testing for *Salmonella* and supervise the implementation of appropriate control measures. This information should be available to the veterinarian before marketing if a veterinary certificate for flock *Salmonella* status is required. When required by the Competent Authority, the veterinarian or other person responsible for notification should notify the Competent Authority if the presence of *Salmonella* of the relevant serotype is confirmed.

Article 6.5.6.

Prevention of *Salmonella* spread from infected flocks

If a flock is found infected with specific *Salmonella* serotypes of concern, the following actions should be taken in addition to general measures detailed in Chapter 6.4. Hygiene and Biosecurity Procedures in Poultry Production:

1. According to the epidemiological situation, investigations should be carried out to determine the origin of the infection.

2. Movement of poultry flocks at the end of the production cycle should only be allowed for slaughter or destruction. Special precautions should be taken in the transport, slaughter and processing of the birds, e.g. they could be sent to a separate slaughterhouse or processed at the end of a shift before cleaning and disinfection of the equipment.

3. Litter should not be reused. Poultry litter/faeces and other potentially contaminated farm waste should be disposed of in a safe manner to prevent the direct or indirect exposure of humans, livestock and wildlife to *Salmonella*. Particular care needs to be taken in regard to poultry litter/faeces used to fertilise plants intended for human consumption. If litter is not removed then it should be treated in a manner to inactivate infectious agents, to prevent the spread from one flock to the next.

4. Particular care should be taken in cleaning and disinfection of the poultry house and equipment.

5. Before restocking the facility, a bacteriological examination should be carried out as detailed in this Chapter and the Terrestrial Manual.

Article 6.5.7.

Recommendations for importation of live poultry (other than day-old birds)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the poultry originated from an establishment flock that participates in a *Salmonella* surveillance programme in accordance with the recommendations in Article 6.5.4.;

2. the poultry originated from an establishment flock in which no evidence of *S. Enteritidis* and *S. Typhimurium* has been detected prior to shipment and have had no contact with birds or other material from establishment flocks that do not comply with this chapter;

3. the poultry originated from an establishment flock that complies with the recommendations of Chapter 6.4.
Recommendations for importation of day-old birds

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the day-old birds showed no clinical signs of salmonellosis on the day of shipment;

2. the day-old birds originated from a breeder establishment flock and hatchery that participate in a Salmonella surveillance programme in accordance with the recommendations in Article 6.5.4.;

3. the day-old birds originated from a breeder establishment flock and hatchery in which no evidence of S. Enteritidis and S. Typhimurium has been detected and have had no contact during setting, incubation or hatching with batching eggs or other material from an establishment that do not comply with this chapter;

4. the day-old birds originated from a breeder establishment flock and hatchery that complies with the recommendations of Chapter 6.4.;

5. the day-old birds were shipped in new and clean containers.

Recommendations for importation of hatching eggs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the batching eggs originated from a breeder establishment flock that participates in a Salmonella surveillance programme in accordance with the recommendations in Article 6.5.4.;

2. the batching eggs originated from a breeder establishment flock in which no evidence of S. Enteritidis and S. Typhimurium has been detected and have had no contact with poultry or other material from an establishment that do not comply with this Chapter;

3. the batching eggs originated from a breeder establishment flock that complies with the recommendations of Chapter 6.4.;

4. the batching eggs were shipped in new and clean packaging materials.

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CHAPTER 7.3.

TRANSPORT OF ANIMALS BY LAND

Preamble: These recommendations apply to the following live domesticated animals: cattle, buffaloes, camels, sheep, goats, pigs, poultry and equines. They will also be largely applicable to some other animals (e.g., deer, other camelids and ratites). Wild, feral and partly domesticated animals may need different conditions.

Article 7.3.1.

The amount of time animals spend on a journey should be kept to the minimum.

Article 7.3.2.

1. Animal behaviour

Animal handlers should be experienced and competent in handling and moving farm livestock and understand the behaviour patterns of animals and the underlying principles necessary to carry out their tasks.

The behaviour of individual animals or groups of animals will vary depending on their breed, sex, temperament and age and the way in which they have been reared and handled. Despite these differences, the following behaviour patterns, which are always present to some degree in domestic animals, should be taken into consideration in handling and moving the animals.

Most domestic livestock are kept in groups and follow a leader by instinct.

Animals which are likely to harm each other in a group situation should not be mixed.

The desire of some animals to control their personal space should be taken into account in designing loading and unloading facilities, transport vessels and containers.

Domestic animals will try to escape if any person approaches closer than a certain distance. This critical distance, which defines the flight zone, varies among species and individuals of the same species, and depends upon previous contact with humans. Animals reared in close proximity to humans (i.e. tame) have a smaller flight zone, whereas those kept in free range or extensive systems may have flight zones which may vary from one metre to many metres. Animal handlers should avoid sudden penetration of the flight zone which may cause a panic reaction which could lead to aggression or attempted escape and compromise the welfare of the animals.

Animal handlers should use the point of balance at the animal’s shoulder to move animals, adopting a position behind the point of balance to move an animal forward and in front of the point of balance to move it backward.

Domestic animals have a wide-angle vision but only have a limited forward binocular vision and poor perception of depth. This means that they can detect objects and movements beside and behind them, but can only judge distances directly ahead.

Although domestic animals have a highly sensitive sense of smell, they may react differently to the smells encountered during travel. Smells which cause negative responses should be taken into consideration when managing animals.
Domestic animals can hear over a greater range of frequencies than humans and are more sensitive to higher frequencies. They tend to be alarmed by constant loud noises and by sudden noises, which may cause them to panic. Sensitivity to such noises should also be taken into account when handling animals.

An example of a flight zone (cattle)

Handler movement pattern to move cattle forward

2. Distractions and their removal

Design of new loading and unloading facilities or modification of existing facilities should aim to minimise the potential for distractions that may cause approaching animals to stop, baulk or turn back. Below are examples of common distractions and methods for eliminating them:

a) reflections on shiny metal or wet floors - move a lamp or change lighting;

b) dark entrances — illuminate with indirect lighting which does not shine directly into the eyes of approaching animals;

c) animals seeing moving people or equipment up ahead — install solid sides on chutes and races or install shields;

d) dead ends — avoid if possible by curving the passage, or make an illusory passage;
e) chains or other loose objects hanging in chutes or on fences — remove them;

f) uneven floors or a sudden drop in floor levels — avoid uneven floor surfaces or install a solid false floor to provide an illusion of a solid and continuous walking surface;

g) sounds of air hissing from pneumatic equipment — install silencers or use hydraulic equipment or vent high pressure to the external environment using flexible hosing;

h) clanging and banging of metal objects — install rubber stops on gates and other devices to reduce metal to metal contact;

i) air currents from fans or air curtains blowing into the face of animals — redirect or reposition equipment.

Article 7.3.3.

Responsibilities

Once the decision to transport the animals has been made, the welfare of the animals during their journey is the paramount consideration and is the joint responsibility of all people involved. The individual responsibilities of persons involved will be described in more detail in this article.

The roles of each of those responsible are defined below:

1. The owners and managers of the animals are responsible for:

   a) the general health, overall welfare and fitness of the animals for the journey;

   b) ensuring compliance with any required veterinary or other certification;

   c) the presence of an animal handler competent for the species being transported during the journey with the authority to take prompt action; in case of transport by individual trucks, the truck driver may be the sole animal handler during the journey;

   d) the presence of an adequate number of animal handlers during loading and unloading;

   e) ensuring that equipment and veterinary assistance are provided as appropriate for the species and the journey.

2. Business agents or buying/selling agents are responsible for:

   a) selection of animals that are fit to travel;

   b) availability of suitable facilities at the start and at the end of the journey for the assembly; loading, transport, unloading and holding of animals, including for any stops at resting points during the journey and for emergencies.

3. Animal handlers are responsible for the humane handling and care of the animals, especially during loading and unloading, and for maintaining a journey log. To carry out their responsibilities, they should have the authority to take prompt action. In the absence of a separate animal handler, the driver is the animal handler.

4. Transport companies, vehicle owners and drivers are responsible for planning the journey to ensure the care of the animals; in particular they are responsible for:
Annex XIII (contd)

a) choosing appropriate vehicles for the species transported and the journey;

b) ensuring that properly trained staff are available for loading/unloading of animals;

c) ensuring adequate competency of the driver in matters of animal welfare for the species being transported in case a separate animal handler is not assigned to the truck;

d) developing and keeping up-to-date contingency plans to address emergencies (including adverse weather conditions) and minimise stress during transport;

e) producing a journey plan which includes a loading plan, journey duration, itinerary and location of resting places;

f) loading only those animals which are fit to travel, for their correct loading into the vehicle and their inspection during the journey, and for appropriate responses to problems arising; if its fitness to travel is in doubt, the animal should be examined by a veterinarian in accordance with point 3a) of Article 7.3.7.;

g) welfare of the animals during the actual transport.

5. Managers of facilities at the start and at the end of the journey and at resting points are responsible for:

a) providing suitable premises for loading, unloading and securely holding the animals, with water and feed when required, and with protection from adverse weather conditions until further transport, sale or other use (including rearing or slaughter);

b) providing an adequate number of animal handlers to load, unload, drive and hold animals in a manner that causes minimum stress and injury; in the absence of a separate animal handler, the driver is the animal handler;

c) minimising the opportunities for disease transmission;

d) providing appropriate facilities, with water and feed when required;

e) providing appropriate facilities for emergencies;

f) providing facilities for washing and disinfecting vehicles after unloading;

g) providing facilities and competent staff to allow the humane killing of animals when required;

h) ensuring proper rest times and minimal delay during stops.

6. The responsibilities of Competent Authorities include:

a) establishing minimum standards for animal welfare, including requirements for inspection of animals before, during and after their travel, defining ‘fitness to travel’ and appropriate certification and record keeping;

b) setting standards for facilities, containers and vehicles for the transport of animals;

c) setting standards for the competence of animal handlers, drivers and managers of facilities in relevant issues in animal welfare;

d) ensuring appropriate awareness and training of animal handlers, drivers and managers of facilities in relevant issues in animal welfare;
Annex XIII (contd)

e) implementation of the standards, including through accreditation of / interaction with other organisations;

f) monitoring and evaluating the effectiveness of standards of health and other aspects of welfare;

g) monitoring and evaluating the use of veterinary medications;

h) giving animal consignments priority at frontiers in order to allow them to pass without unnecessary delay.

7. All individuals, including veterinarians, involved in transporting animals and the associated handling procedures should receive appropriate training and be competent to meet their responsibilities.

8. The receiving Competent Authority should report back to the sending Competent Authority on significant animal welfare problems which occurred during the journey.

Article 7.3.4.

Competence

1. All people responsible for animals during journeys, should be competent according to their responsibilities listed in Article 7.3.3. Competence may be gained through formal training and/or practical experience.

2. The assessment of the competence of animal handlers should at a minimum address knowledge, and ability to apply that knowledge, in the following areas:

   a) planning a journey, including appropriate space allowance, and feed, water and ventilation requirements;

   b) responsibilities for animals during the journey, including loading and unloading;

   c) sources of advice and assistance;

   d) animal behaviour, general signs of disease, and indicators of poor animal welfare such as stress, pain and fatigue, and their alleviation;

   e) assessment of fitness to travel; if fitness to travel is in doubt, the animal should be examined by a veterinarian;

   f) relevant authorities and applicable transport regulations, and associated documentation requirements;

   g) general disease prevention procedures, including cleaning and disinfection;

   h) appropriate methods of animal handling during transport and associated activities such as assembling, loading and unloading;

   i) methods of inspecting animals, managing situations frequently encountered during transport such as adverse weather conditions, and dealing with emergencies, including humane killing;

   j) species-specific aspects and age-specific aspects of animal handling and care, including feeding, watering and inspection; and

   k) maintaining a journey log and other records.
Planning the journey

1. General considerations

a) Adequate planning is a key factor affecting the welfare of animals during a journey.

b) Before the journey starts, plans should be made in relation to:

i) preparation of animals for the journey;

ii) choice of road, rail, roll-on roll-off vessels or containers;

iii) nature and duration of the journey;

iv) vehicle design and maintenance, including roll-on roll-off vessels;

v) required documentation;

vi) space allowance;

vii) rest, water and feed;

viii) observation of animals en route;

ix) control of disease;

x) emergency response procedures;

xi) forecast weather conditions (e.g. conditions being too hot or too cold to travel during certain periods of the day);

xii) transfer time when changing mode of transport, and

xiii) waiting time at frontiers and inspection points.

c) Regulations concerning drivers (for example, maximum driving periods) should take into account animal welfare whenever possible.

2. Preparation of animals for the journey

a) When animals are to be provided with a novel diet or method of water provision during transport, an adequate period of adaptation should be planned. For all animals it is essential that the rest stops during long journeys are long enough to fulfil each animal’s need for feed and water. Species-specific short period of feed deprivation prior to loading may be desirable.

b) Animals more accustomed to contact with humans and with being handled are likely to be less fearful of being loaded and transported. Animal handlers should handle and load animals in a manner that reduces their fearfulness and improves their approachability.

c) Behaviour-modifying compounds (such as tranquillisers) or other medication should not be used routinely during transport. Such compounds should only be administered when a problem exists in an individual animal, and should be administered by a veterinarian or other person who has been instructed in their use by a veterinarian.
3. **Nature and duration of the journey**

The maximum duration of a *journey* should be determined according to factors such as:

a) the ability of the *animals* to cope with the stress of transport (such as very young, old, lactating or pregnant *animals*);

b) the previous transport experience of the *animals*;

c) the likely onset of fatigue;

d) the need for special attention;

e) the need for feed and water;

f) the increased susceptibility to injury and disease;

g) *space allowance*, *vehicle* design, road conditions and driving quality;

h) weather conditions;

i) *vehicle* type used, terrain to be traversed, road surfaces and quality, skill and experience of the driver.

4. **Vehicle and container design and maintenance**

a) *Vehicles and containers* used for the transport of *animals* should be designed, constructed and fitted as appropriate for the species, size and weight of the *animals* to be transported. Special attention should be paid to avoid injury to *animals* through the use of secure smooth fittings free from sharp protrusions. The avoidance of injury to drivers and *animal handlers* while carrying out their responsibilities should be emphasised.

b) *Vehicles and containers* should be designed with the structures necessary to provide protection from adverse weather conditions and to minimise the opportunity for *animals* to escape.

c) In order to minimise the likelihood of the spread of infectious disease during transport, *vehicles and containers* should be designed to permit thorough cleaning and disinfection, and the containment of faeces and urine during a *journey*.

d) *Vehicles and containers* should be maintained in good mechanical and structural condition.

e) *Vehicles and containers* should have adequate ventilation to meet variations in climate and the thermo-regulatory needs of the animal species being transported; the ventilation system (natural or mechanical) should be effective when the *vehicle* is stationary, and the airflow should be adjustable.

f) *Vehicles* should be designed so that the faeces or urine from *animals* on upper levels do not soil *animals* on lower levels, nor their feed and water. This condition is not applicable for *poultry*. They are generally transported in plastic crates which are designed to let air flow through in all directions to obtain a better ventilation.

g) When *vehicles* are carried on board ferries, facilities for adequately securing them should be available.

h) If feeding or watering while the *vehicle* is moving is required, adequate facilities on the *vehicle* should be available.
i) When appropriate, suitable bedding should be added to vehicle floors to assist absorption of urine and faeces, to minimise slipping by animals, and protect animals (especially young animals) from hard flooring surfaces and adverse weather conditions.

5. Special provisions for transport in vehicles (road and rail) on roll-on/roll-off vessels or for containers

a) Vehicles and containers should be equipped with a sufficient number of adequately designed, positioned and maintained securing points enabling them to be securely fastened to the vessel.

b) Vehicles and containers should be secured to the vessel before the start of the sea journey to prevent them being displaced by the motion of the vessel.

c) Roll-on/roll-off vessels should have adequate ventilation to meet variations in climate and the thermo-regulatory needs of the animal species being transported, especially where the animals are transported in a secondary vehicle/container on enclosed decks.

6. Space allowance

a) The number of animals which should be transported on a vehicle or in a container and their allocation to compartments should be determined before loading.

b) The space required on a vehicle or in a container depends upon whether or not the animals need to lie down (for example, cattle, sheep, pigs, camels and poultry), or to stand (horses). Animals which will need to lie down often stand when first loaded or when the vehicle is driven with too much lateral movement or sudden braking.

c) When animals lie down, they should all be able to adopt a normal lying posture, without being on top of one another, and allowing necessary thermoregulation.

d) When animals are standing, they should have sufficient space to adopt a balanced position as appropriate to the climate and species transported.

e) The amount of headroom necessary depends on the species of animal. Each animal should be able to assume its natural standing position for transport (including during loading and unloading) without coming into contact with the roof or upper deck of the vehicle, and there should be sufficient headroom to allow adequate airflow over the animals. These conditions will not normally apply to poultry except for one day old chicks. However, under tropical and subtropical conditions poultry benefit from having adequate head room to allow head cooling.

f) Calculations for the space allowance for each animal should be carried out using the figures given in a relevant national or international document. The number and size of pens on the vehicle should be varied to where possible accommodate already established groups of animals while avoiding group sizes which are too large.
Other factors which may influence space allowance include:

i) vehicle/container design;

ii) length of journey;

iii) need to provide feed and water on the vehicle;

iv) quality of roads;

v) expected weather conditions;

vi) category and sex of the animals.

Rest, water and feed

i) Suitable water and feed should be available as appropriate and needed for the species, age, and condition of the animals, as well as the duration of the journey, climatic conditions, etc.

ii) Animals should be allowed to rest at resting points at appropriate intervals during the journey. The type of transport, the age and species of the animals being transported, and climatic conditions should determine the frequency of rest stops and whether the animals should be unloaded. Water and feed should be available during rest stops.

7. Ability to observe animals during the journey

a) Animals should be positioned to enable each animal to be observed regularly during the journey to ensure their safety and good welfare. This condition will not normally apply to poultry.

b) If the animals are in crates or on multi-tiered vehicles which do not allow free access for observation, for example where the roof of the tier is too low, animals cannot be inspected adequately, and serious injury or disease could go undetected. In these circumstances, a shorter journey duration should be allowed, and the maximum duration will vary according to the rate at which problems arise in the species and under the conditions of transport.

8. Control of disease

As animal transport is often a significant factor in the spread of infectious diseases, journey planning should take the following into account:

a) mixing of animals from different sources in a single consignment should be minimised;

b) contact at resting points between animals from different sources should be avoided;

c) when possible, animals should be vaccinated against diseases to which they are likely to be exposed at their destination;

d) medications used prophylactically or therapeutically should be approved by the Veterinary Authority of the exporting country and the importing country and should only be administered by a veterinarian or other person who has been instructed in their use by a veterinarian.
Annex XIII (contd)

9. Emergency response procedures

There should be an emergency management plan that identifies the important adverse events that may be encountered during the journey, the procedures for managing each event and the action to be taken in an emergency. For each important event, the plan should document the actions to be undertaken and the responsibilities of all parties involved, including communications and record keeping.

10. Other considerations

a) Extreme weather conditions are hazardous for animals undergoing transport and require appropriate vehicle design to minimise risks. Special precautions should be taken for animals that have not been acclimatised or which are unsuited to either hot or cold conditions. In some extreme conditions of heat or cold, animals should not be transported at all.

b) In some circumstances, transportation during the night may reduce thermal stress or the adverse effects of other external stimuli.

Article 7.3.6.

Documentation

1. Animals should not be loaded until the documentation required to that point is complete.

2. The documentation accompanying the consignment should include:

   a) journey travel plan and emergency management plan;

   b) date, time and place of loading and unloading;

   c) veterinary certification, when required;

   d) animal welfare competencies of the driver (under study);

   e) animal identification to allow animal traceability to the premises of departure and, where possible, to the premises of origin;

   f) details of any animals considered at particular risk of suffering poor welfare during transport (point 3e) of Article 7.3.7.);

   g) documentation of the period of rest, and access to feed and water, prior to the journey;

   h) stocking density estimate for each load in the consignment;

   i) the journey log - daily record of inspection and important events, including records of morbidity and mortality and actions taken, climatic conditions, rest stops, travel time and distance, feed and water offered and estimates of consumption, medication provided, and mechanical defects.

3. When veterinary certification is required to accompany consignments of animals, it should address:

   a) fitness of animals to travel;

   b) animal identification (description, number, etc.);

   c) health status including any tests, treatments and vaccinations carried out;
d) when required, details of disinfection carried out.

At the time of certification, the veterinarian should notify the animal handler or the driver of any factors affecting the fitness of animals to travel for a particular journey.

Article 7.3.7.

Pre-journey period

1. General considerations

a) Pre-journey rest is necessary if the welfare of animals has become poor during the collection period because of the physical environment or the social behaviour of the animals. The need for rest should be judged by a veterinarian or other competent person.

b) Pre-journey assembly/holding areas should be designed to:
   i) securely hold the animals;
   ii) maintain a safe environment from hazards, including predators and disease;
   iii) protect animals from exposure to severe weather conditions;
   iv) allow for maintenance of social groups;
   v) allow for rest, and appropriate water and feed.

c) Consideration should be given to the previous transport experience, training and conditioning of the animals, if known, as these may reduce fear and stress in animals.

d) Feed and water should be provided pre-journey if the journey duration is greater than the normal inter-feeding and drinking interval for the animal. Recommendations for specific-species are described in detail in Article 7.3.12.

e) When animals are to be provided with a novel diet or method of feed or water provision during the journey, an adequate period of adaptation should be allowed.

f) Before each journey, vehicles and containers should be thoroughly cleaned and, if necessary, treated for animal health and public health purposes, using methods approved by the Competent Authority. When cleaning is necessary during a journey, this should be carried out with the minimum of stress and risks to the animals.

g) Where an animal handler believes that there is a significant risk of disease among the animals to be loaded or significant doubt as to their fitness to travel, the animals should be examined by a veterinarian.

2. Selection of compatible groups

Compatible groups should be selected before transport to avoid adverse animal welfare consequences. The following recommendations should be applied when assembling groups of animals:

a) Animals reared together should be maintained as a group; animals with a strong social bond, such as a dam and offspring, should be transported together.
Annex XIII (contd)

b) Animals of the same species can be mixed unless there is a significant likelihood of aggression; aggressive individuals should be segregated (recommendations for specific species are described in detail in Article 7.3.12.). For some species, animals from different groups should not be mixed because poor welfare occurs unless they have established a social structure.

c) Young or small animals should be separated from older or larger animals, with the exception of nursing mothers with young at foot.

d) Animals with horns or antlers should not be mixed with animals lacking horns or antlers unless judged to be compatible.

e) Animals of different species should not be mixed unless they are judged to be compatible.

3. Fitness to travel

a) Each animal should be inspected by a veterinarian or an animal handler to assess fitness to travel. If its fitness to travel is in doubt, the animal should be examined by a veterinarian. Animals found unfit to travel should not be loaded onto a vehicle, except for transport to receive veterinary attention.

b) Humane and effective arrangements should be made by the owner and the agent for the handling and care of any animal rejected as unfit to travel.

c) Animals that are unfit to travel include, but may not be limited to:

i) those that are sick, injured, weak, disabled or fatigued;

ii) those that are unable to stand unaided and bear weight on each leg;

iii) those that are blind in both eyes;

iv) those that cannot be moved without causing them additional suffering;

v) newborn with an unhealed navel;

vi) pregnant animals which would be in the final 10% of their gestation period at the planned time of unloading;

vii) females travelling without young which have given birth within the previous 48 hours;

viii) those whose body condition would result in poor welfare because of the expected climatic conditions.

d) Risks during transport can be reduced by selecting animals best suited to the conditions of travel and those that are acclimatised to expected weather conditions.

e) Animals at particular risk of suffering poor welfare during transport and which require special conditions (such as in the design of facilities and vehicles, and the length of the journey) and additional attention during transport, may include:

i) large or obese individuals;

ii) very young or old animals;

iii) excitable or aggressive animals;
iv) animals which have had little contact with humans;

v) animals subject to motion sickness;

vi) females in late pregnancy or heavy lactation, dam and offspring;

vii) animals with a history of exposure to stressors or pathogenic agents prior to transport;

viii) animals with unhealed wounds from recent surgical procedures such as dehorning.

4. Specific species requirements

Transport procedures should be able to take account of variations in the behaviour of the species. Flight zones, social interactions and other behaviour vary significantly among species and even within species. Facilities and handling procedures that are successful with one species are often ineffective or dangerous with another.

Recommendations for specific species are described in detail in Article 7.3.12.

Article 7.3.8.

Loading

1. Competent supervision

a) Loading should be carefully planned as it has the potential to be the cause of poor welfare in transported animals.

b) Loading should be supervised and/or conducted by animal handlers. The animals are to be loaded quietly and without unnecessary noise, harassment or force. Untrained assistants or spectators should not impede the process.

c) When containers are loaded onto a vehicle, this should be carried out in such a way to avoid poor animal welfare.

2. Facilities

a) The facilities for loading including the collecting area, races and loading ramps should be designed and constructed to take into account the needs and abilities of the animals with regard to dimensions, slopes, surfaces, absence of sharp projections, flooring, etc.

b) Loading facilities should be properly illuminated to allow the animals to be observed by animal handler(s), and to allow the ease of movement of the animals at all times. Facilities should provide uniform light levels directly over approaches to sorting pens, chutes, loading ramps, with brighter light levels inside vehicles/containers, in order to minimise baulking. Dim light levels may be advantageous for the catching of poultry and some other animals. Artificial lighting may be required. Loading ramps and other facilities should have a non-slippery flooring.

c) Ventilation during loading and the journey should provide for fresh air, the removal of excessive heat, humidity and noxious fumes (such as ammonia and carbon monoxide), and the prevention of accumulations of ammonia and carbon dioxide. Under warm and hot conditions, ventilation should allow for the adequate convective cooling of each animal. In some instances, adequate ventilation can be achieved by increasing the space allowance for animals.
Annex XIII (contd)

3. Goads and other aids

When moving animals, their species-specific behaviour should be used (see Article 7.3.12.). If goads and other aids are necessary, the following principles should apply:

a) *Animals* that have little or no room to move should not be subjected to physical force or goads and other aids which compel movement. Electric goads and prods should only be used in extreme cases and not on a routine basis to move animals. The use and the power output should be restricted to that necessary to assist movement of an animal and only when an animal has a clear path ahead to move. Goads and other aids should not be used repeatedly if the animal fails to respond or move. In such cases it should be investigated whether some physical or other impediment is preventing the animal from moving.

b) The use of such devices should be limited to battery-powered goads on the hindquarters of pigs and large ruminants, and never on sensitive areas such as the eyes, mouth, ears, anogenital region or belly. Such instruments should not be used on horses, sheep and goats of any age, or on calves or piglets.

c) Useful and permitted goads include panels, flags, plastic paddles, flappers (a length of cane with a short strap of leather or canvas attached), plastic bags and rattles; they should be used in a manner sufficient to encourage and direct movement of the animals without causing undue stress.

d) Painful procedures (including whipping, tail twisting, use of nose twitches, pressure on eyes, ears or external genitalia), or the use of goads or other aids which cause pain and suffering (including large sticks, sticks with sharp ends, lengths of metal piping, fencing wire or heavy leather belts), should not be used to move animals.

e) Excessive shouting at animals or making loud noises (e.g., through the cracking of whips) to encourage them to move should not occur, as such actions may make the animals agitated, leading to crowding or falling.

f) The use of well trained dogs to help with the loading of some species may be acceptable.

g) *Animals* should be grasped or lifted in a manner which avoids pain or suffering and physical damage (e.g. bruising, fractures, dislocations). In the case of quadrupeds, manual lifting by a person should only be used in young animals or small species, and in a manner appropriate to the species; grasping or lifting animals only by their wool, hair, feathers, feet, neck, ears, tails, head, horns, limbs causing pain or suffering should not be permitted, except in an emergency where animal welfare or human safety may otherwise be compromised.

h) Conscious animals should not be thrown, dragged or dropped.

i) Performance standards should be established in which numerical scoring is used to evaluate the use of such instruments, and to measure the percentage of animals moved with an electric instrument and the percentage of animals slipping or falling as a result of their usage.

Article 7.3.9.

Travel

1. General considerations

a) Drivers and animal handlers should check the load immediately before departure to ensure that the animals have been properly loaded. Each load should be checked again early in the trip and adjustments made as appropriate. Periodic checks should be made throughout the trip, especially at rest or refuelling stops or during meal breaks when the vehicle is stationary.
b) Drivers should utilise smooth, defensive driving techniques, without sudden turns or stops, to minimise uncontrolled movements of the animals.

2. Methods of restraining or containing animals
   a) Methods of restraining animals should be appropriate to the species and age of animals involved and the training of the individual animal.
   b) Recommendations for specific species are described in detail in Article 7.3.12.

3. Regulating the environment within vehicles or containers
   a) Animals should be protected against harm from hot or cold conditions during travel. Effective ventilation procedures for maintaining the environment within vehicles or containers will vary according to whether conditions are cold, hot and dry or hot and humid, but in all conditions a build-up of noxious gases should be prevented.
   b) The environment within vehicles or containers in hot and warm weather can be regulated by the flow of air produced by the movement of the vehicle. In warm and hot weather, the duration of journey stops should be minimised and vehicles should be parked under shade, with adequate and appropriate ventilation.
   c) To minimise slipping and soiling, and maintain a healthy environment, urine and faeces should be removed from floors when necessary and disposed of in such a way as to prevent the transmission of disease and in compliance with all relevant health and environmental legislation.

4. Sick, injured or dead animals
   a) A driver or an animal handler finding sick, injured or dead animals should act according to a predetermined emergency response plan.
   b) Sick or injured animals should be segregated.
   c) Ferries (roll-on roll-off) should have procedures to treat sick or injured animals during the journey.
   d) In order to reduce the likelihood that animal transport will increase the spread of infectious disease, contact between transported animals, or the waste products of the transported animals, and other farm animals should be minimised.
   e) During the journey, when disposal of a dead animal becomes necessary, this should be carried out in such a way as to prevent the transmission of disease and in compliance with all relevant health and environmental legislation.
   f) When killing is necessary, it should be carried out as quickly as possible and assistance should be sought from a veterinarian or other person(s) competent in humane killing procedures. Recommendations for specific species are described in Chapter 7.6. on killing of animals for disease control purposes.

5. Sick, injured or dead animals
   a) If journey duration is such that feeding or watering is required or if the species requires feed or water throughout, access to suitable feed and water for all the animals (appropriate for their species and age) carried in the vehicle should be provided. There should be adequate space for all animals to move to the feed and water sources and due account taken of likely competition for feed.
   b) Recommendations for specific species are described in detail in Article 7.3.12.
Annex XIII (contd)

6. **Rest periods and conditions**
   a) *Animals* that are being transported should be rested at appropriate intervals during the *journey* and offered feed and water, either on the *vehicle* or, if necessary, unloaded into suitable facilities.
   
   b) Suitable facilities should be used en route, when resting requires the *unloading* of the *animals*. These facilities should meet the needs of the particular animal species and should allow access of all *animals* to feed and water.

7. **In-transit observations**
   a) *Animals* being transported by road should be observed soon after a *journey* is commenced and whenever the driver has a rest stop. After meal breaks and refuelling stops, the *animals* should be observed immediately prior to departure.
   
   b) *Animals* being transported by rail should be observed at each scheduled stop. The responsible rail transporter should monitor the progress of trains carrying *animals* and take all appropriate action to minimise delays.
   
   c) During stops, it should be ensured that the *animals* continue to be properly confined, have appropriate feed and water, and their physical condition is satisfactory.

   Article 7.3.10.

**Unloading and post-journey handling**

1. **General considerations**
   a) The required facilities and the principles of animal handling detailed in Article 7.3.8. apply equally to *unloading*, but consideration should be given to the likelihood that the *animals* will be fatigued.
   
   b) *Unloading* should be supervised and/or conducted by an *animal handler* with knowledge and experience of the behavioural and physical characteristics of the species being unloaded. *Animals* should be unloaded from the *vehicle* into appropriate facilities as soon as possible after arrival at the destination but sufficient time should be allowed for *unloading* to proceed quietly and without unnecessary noise, harassment or force.
   
   c) Facilities should provide all *animals* with appropriate care and comfort, adequate space and ventilation, access to feed (if appropriate) and water, and shelter from extreme weather conditions.
   
   d) For details regarding the *unloading* of *animals* at a *slaughterhouse*, see Chapter 7.5. on slaughter of *animals* for human consumption.

2. **Sick or injured animals**
   a) An *animal* that has become sick, injured or disabled during a *journey* should be appropriately treated or humanely killed (see Chapter 7.6. on killing of *animals* for disease control purposes). If necessary, veterinary advice should be sought in the care and treatment of these *animals*. In some cases, where *animals* are non-ambulatory due to fatigue, injury or sickness, it may be in the best *welfare* interests of the *animal* to be treated or killed aboard the *vehicle*. Assistance should be sought from a *veterinarian* or other person(s) competent in humane *killing* procedures.
   
   b) At the destination, the *animal handler* or the driver during transit should ensure that responsibility for the *welfare* of sick, injured or disabled *animals* is transferred to a *veterinarian* or other suitable person.
c) If treatment or humane killing is not possible aboard the vehicle, there should be appropriate facilities and equipment for the humane unloading of animals that are non-ambulatory due to fatigue, injury or sickness. These animals should be unloaded in a manner that causes the least amount of suffering. After unloading, separate pens and other appropriate facilities should be available for sick or injured animals.

d) Feed, if appropriate, and water should be available for each sick or injured animal.

3. Addressing disease risks

The following should be taken into account in addressing the greater risk of disease due to animal transport and the possible need for segregation of transported animals at the destination:

a) increased contact among animals, including those from different sources and with different disease histories;

b) increased shedding of pathogens and increased susceptibility to infection related to stress and impaired defences against disease, including immunosuppression;

c) exposure of animals to pathogens which may contaminate vehicles, resting points, markets, etc.

4. Cleaning and disinfection

a) Vehicles, crates, containers, etc. used to carry the animals should be cleaned before re-use through the physical removal of manure and bedding by scraping, washing and flushing with water and detergent. This should be followed by disinfection when there are concerns about disease transmission.

b) Manure, litter, bedding and the bodies of any animals which die during the journey should be disposed of in such a way as to prevent the transmission of disease and in compliance with all relevant health and environmental legislation.

c) Establishments like livestock markets, slaughterhouses, resting sites, railway stations, etc. where animals are unloaded should be provided with appropriate areas for the cleaning and disinfection of vehicles.

Article 7.3.11.

Actions in the event of a refusal to allow the completion of the journey

1. The welfare of the animals should be the first consideration in the event of a refusal to allow the completion of the journey.

2. When the animals have been refused import, the Competent Authority of the importing country should make available suitable isolation facilities to allow the unloading of animals from a vehicle and their secure holding, without posing a risk to the health of national herd or flock, pending resolution of the situation. In this situation, the priorities should be:

a) the Competent Authority of the importing country should provide urgently in writing the reasons for the refusal;

b) in the event of a refusal for animal health reasons, the Competent Authority of the importing country should provide urgent access to a veterinarian, where possible an OIE veterinarian(s) appointed by the Director General, to assess the health status of the animals with regard to the concerns of the importing country, and the necessary facilities and approvals to expedite the required diagnostic testing;

c) the Competent Authority of the importing country should provide access to allow continued assessment of the health and other aspects of the welfare of the animals;
Annex XIII (contd)

d) if the matter cannot be promptly resolved, the Competent Authorities of the exporting and importing countries should call on the OIE to mediate.

3. In the event that a Competent Authority requires the animals to remain on the vehicle, the priorities should be:

a) to allow provisioning of the vehicle with water and feed as necessary;

b) to provide urgently in writing the reasons for the refusal;

c) to provide urgent access to an independent veterinarian(s) to assess the health status of the animals, and the necessary facilities and approvals to expedite the required diagnostic testing in the event of a refusal for animal health reasons;

d) to provide access to allow continued assessment of the health and other aspects of the welfare of the animals, and the necessary actions to deal with any animal issues which arise.

4. The OIE should utilise its informal procedure for dispute mediation to identify a mutually agreed solution which will address animal health and any other welfare issues in a timely manner.

Species-specific issues

Camelids of the new world in this context comprise llamas, alpacas, guanaco and vicuna. They have good eyesight and, like sheep, can negotiate steep slopes, though ramps should be as shallow as possible. They load most easily in a bunch as a single animal will strive to rejoin the others. Whilst they are usually docile, they have an unnerving habit of spitting in self-defence. During transport, they usually lie down. They frequently extend their front legs forward when lying, so gaps below partitions should be high enough so that their legs are not trapped when the animals rise.

Cattle are sociable animals and may become agitated if they are singled out. Social order is usually established at about two years of age. When groups are mixed, social order has to be re-established and aggression may occur until a new order is established. Crowding of cattle may also increase aggression as the animals try to maintain personal space. Social behaviour varies with age, breed and sex; Bos indicus and B. indicus-cross animals are usually more temperamental than European breeds. Young bulls, when moved in groups, show a degree of playfulness (pushing and shoving) but become more aggressive and territorial with age. Adult bulls have a minimum personal space of six square metres. Cows with young calves can be very protective, and handling calves in the presence of their mothers can be dangerous. Cattle tend to avoid “dead end” in passages.

Goats should be handled calmly and are more easily led or driven than if they are excited. When goats are moved, their gregarious tendencies should be exploited. Activities which frighten, injure or cause agitation to animals should be avoided. Bullying is particularly serious in goats and can reflect demands for personal space. Housing strange goats together could result in fatalities, either through physical violence, or subordinate goats being refused access to food and water.

Horses in this context include donkeys, mules and hinnies. They have good eyesight and a very wide angle of vision. They may have a history of loading resulting in good or bad experiences. Good training should result in easier loading, but some horses can prove difficult, especially if they are inexperienced or have associated loading with poor transport conditions. In these circumstances, two experienced animal handlers can load an animal by linking arms or using a strop below its rump. Blindfolding may even be considered. Ramps should be as shallow as possible. Steps are not usually a problem when horses mount a ramp, but they tend to jump a step when descending, so steps should be as low as possible. Horses benefit from being individually stalled, but may be transported in compatible groups. When horses are to travel in groups, their shoes should be removed. Horses are prone to respiratory disease if they are restricted by period by tethers that prevent the lowering and lifting of their heads.
Pigs have poor eyesight, and may move reluctantly in unfamiliar surroundings. They benefit from well lit loading bays. Since they negotiate ramps with difficulty, these should be as level as possible and provided with secure footholds. Ideally, a hydraulic lift should be used for greater heights. Pigs also negotiate steps with difficulty. A good ‘rule-of-thumb’ is that no step should be higher than the pig’s front knee. Serious aggression may result if unfamiliar animals are mixed. Pigs are highly susceptible to heat stress. Pigs are susceptible to motion sickness when in transit. Feed deprivation prior to loading may be beneficial to prevent motion sickness.

Sheep are sociable animals with good eyesight, a relatively subtle and undemonstrative behaviour and a tendency to “flock together”, especially when they are agitated. They should be handled calmly and their tendency to follow each other should be exploited when they are being moved. Crowding of sheep may lead to damaging aggressive and submissive behaviours as animals try to maintain personal space. Sheep may become agitated if they are singled out for attention, or kept alone, and will strive to rejoin the group. Activities which frighten, injure or cause agitation to sheep should be avoided. They can negotiate steep ramps.
Livestock containers

1. **Design**
   
a) General principles of design

   The *container* should:

   - conform to the size of the standard pallet of the aircraft that will be used to transport *animals*; the common sizes are 224 x 318 cm (88 x 125 in.) and 244 x 318 cm (96 x 125 in.);
   - not be constructed of material that could be harmful to the *animals* health or welfare;
   - allow observation of the *animals* and be marked on opposite sides with the International Air Transport Association (IATA) symbols which indicate *animals* and the upright position;
   - allow emergency access to *animals*;
   - allow the *animal* to stand in its normal position without touching the roof of the *container* or, in the case of open *containers*, the restraining nets, and provide at least 10 cm (4 in.) clearance above the *animal’s* head when standing in its normal position; in the case of horses, provide sufficient space above the horses head (21 cm, 8 in. recommended) to allow for the movement required to maintain the horses balance;
   - protect the *animals* from adverse weather;
   - ensure *animals* stand on a suitable floor to prevent slipping or injury;
   - have adequate strength to ensure the safety of the *animals* and to prevent the *animals* from escaping;
   - ensure doors can be opened and closed easily, but be secured so that they cannot be opened accidentally;
   - be free of any nails, bolts and other protrusions or sharp edges that could cause injuries;
   - be designed to minimise the risk of any opening or space entrapping any portion of the *animals* body;
   - if reusable, crates should be constructed of impermeable material that is easily cleaned and disinfected;
   - ensure faeces and urine cannot escape from the crate; this requires a minimum upturn of 20 cm but it should not block any ventilation openings;
Annex XIII (contd)

- if designated for stacking be stable, not block any ventilation space and prevent urine and faeces from leaking into the containers below when stacked;
- allow for a facility for provision of water and possibly food during transportation of longer than 6 hours duration.

b) Ventilation

The container design should:

- provide adequate ventilation taking into consideration the species stocking densities, maximum temperature and humidity of the points of departure, destination, and any interim technical stops;
- allow the normal resting or sleeping position to be assumed for certain species and juvenile animals;
- ensure there is no dead air space in the container;
- provide ventilation openings on the walls equal to at least 16% of the wall area; this may be reduced if the container has an open top;
- in the case of two-tiered containers, ventilation in the sides should be for cattle equivalent to not less than 20% of the floor area of each deck, and for pigs and sheep up to 40% of the floor area of each deck;
- have ventilation openings on all four sides of the crate except that two walls may have reduced ventilation space and the other walls have increased space where required by the positioning of the crates during transportation and/or the ventilation pattern of the aircraft;
- ensure that any internal supports or dividers do not block the cross ventilation;
- not have a solid wall above the height of the animal’s head in normal resting position;
- in those species where the mouth is normally held near the floor, have at least 25 cm (10 in.) of ventilation space at the level of the animal’s head; this opening should be divided in two with a maximum height for any opening of 13 cm; in all containers, there should be a sufficiently large ventilation opening at a height of 25 cm to 30 cm (10 to 11 in.) above floor level on all four sides to allow for circulation;
- have some physical means of ensuring the ventilation space is not blocked, such as the use of cleats (wedges) or allowing space between the outside of the container and the pallet.

2. Species requirements

In general, fractious animals or animals in late pregnancy should not be transported by air (see Article 7.4.2.).

a) Horses

Should be transported in containers and be separated from each other if they are more than 145 cm (57 in.) in height.
Crates used to transport horses should:

- be strong enough to prevent unruly horses from breaking or escaping from the container under any circumstances;
- in the case of multi-horse containers, have partitions of sufficient strength and size to separate the horses and to support each horse's weight;
- adjust to allow mare and foal to travel together;
- provide the same percentage of open space for ventilation as required in point 1 above, divided between the two side walls; however, if the access doors are constructed in such a manner that they may be left open during the flight, the door space may be included in the ventilation space;
- be constructed to minimise noise;
- allow access to the head during the flight;
- have the front end notched and padded to accept the neck of the animal;
- have a secure point for attaching restraining devices;
- have a front and rear barrier that will restrict the movement of the horse and will ensure that liquids are deflected into the container;
- ensure horses cannot bite other animals;
- be constructed to resist kicking;
- have no fittings or projections in the area likely to be kicked, metal plates should be covered with a protective material;
- ramps shall be non-skid in nature, have foot battens, and be of a maximum slope of 25 degrees when the container is on a standard 50 cm (20 in.) dolly;
- not have a step up or down of more than 25 cm (10 in.).

b) Swine

- Crate design and shipment planning should recognize that swine are extremely susceptible to high heat and humidity and that they normally carry their head near the floor.
- In the use of multi-tiered crates, special attention should be paid to ensure air can move through the crate, in accordance with the aircraft’s ventilation pattern and capacity to remove heat.
- Crate construction should take into consideration the tendency for mature swine to chew.
- Litter should be dust-free, shavings or other non-toxic materials may be used but not sawdust.
- Containers for immature swine should only be constructed when flight is imminent, since rapid growth can result in undersized containers if the flight is delayed.
Annex XIII (contd)

- In order to reduce fighting, swine shipped in group pens should be housed together as a group prior to shipment and not be mixed with other swine before loading on the aircraft.
- Mature boars and incompatible females should be shipped in individual crates.
- Individual crates should be 20 cm (8 in.) longer than the body, 15 cm (6 in.) higher than the loin of the pig and of sufficient width, to allow the pigs to lie on their side.

**c) Cattle**

Crates used to transport cattle should:

- if multi-tiered or roofed, have at least 33% of the roof and four walls as open space;
- have at least one ventilation opening 20-25 cm (8-10 in.) above the floor which is of such width that it will not cause injuries to the feet.

Adult bulls should be transported separately unless they have been accustomed to each other. Cattle with and without horns should be separated from each other.

**d) Poultry**

The most current container requirement published by IATA should be adhered to.

Crates/containers containing poultry should be handled and carried carefully with no unnecessary tilting.

The majority of birds transported by air will be newly hatched chicks. These animals are very vulnerable to sudden changes in temperature.

**e) Other species**

- Animals that normally exhibit a herding instinct, including buffalo and deer, can be shipped in group containers providing the mental and physical characteristics of the species are taken into consideration.
- All crates used to move such animals should have a roof or other method of preventing the animals from escaping.
- Animals in which the horns or antler cannot be removed, should be transported individually.
- Deer should not be transported in velvet nor in rut.

Article 7.4.2.

**Recommendations for pregnant animals**

Heavily pregnant animals should not be carried except under exceptional circumstances. Pregnant animals should not be accepted when the last service or exposure to a male prior to departure has exceeded the following time given here for guidance only:

<table>
<thead>
<tr>
<th>Females</th>
<th>Maximum number of days since the last service</th>
</tr>
</thead>
<tbody>
<tr>
<td>Horses</td>
<td>300</td>
</tr>
<tr>
<td>Cows</td>
<td>250</td>
</tr>
<tr>
<td>Deer (axis, fallow and sika)</td>
<td>170</td>
</tr>
<tr>
<td>(red deer, reindeer)</td>
<td>185</td>
</tr>
<tr>
<td>Ewes (sheep)</td>
<td>115</td>
</tr>
<tr>
<td>Nannies (goats)</td>
<td>115</td>
</tr>
<tr>
<td>Sows (pigs)</td>
<td>90</td>
</tr>
</tbody>
</table>
Where service dates or date of last exposure to a male are not available, the animals should be examined by a veterinarian to ensure that pregnancy is not so advanced that animals are likely to give birth during transport or suffer unnecessarily.

Any animal showing udder engorgement and slackening of the pelvic ligament should be refused.

Article 7.4.3.

Stocking density

The current stocking densities agreed by the International Air Transport Association (IATA) should continue to be accepted. However, the graphs giving the space requirements should be extended to take into account animals larger and smaller than those dealt with currently.

1. General considerations

When calculating stocking rates, the following should be taken into account:

a) it is essential that accurate weights of animals are obtained in view of the limitations imposed by the load capabilities of the aircraft and the space required per animal;

b) in narrow bodied aircraft, there is a loss of floor area in the upper tier of two-tier penning due to the contours of the aircraft;

c) space available should be calculated on the inside measurements of the crates or penning system used, not on the floor space of the aircraft;

d) multi-tiered crates, high outdoor temperatures at departure, arrival or stopover points, or extreme length of the trip will require an increase in the amount of space per animal; a 10% decrease in stocking density is recommended for trips in excess of 24 hours;

e) special attention should be paid to the transport of sheep in heavy wool which require an increase in space allotted per animal and to pigs which have limited ability to dissipate heat;

f) animals confined in groups, especially in pens, should be stocked at a high enough density to prevent injuries at take-off, during turbulence and at landing, but not to the extent that individual animals cannot lie down and rise without risk of injury or crushing;

g) in multi-tiered shipments, it should be recognized that the ventilation and cooling capacity of the aircraft is the limiting factor, especially in narrow bodied aircraft. Ventilation capacity varies on each individual aircraft and between aircraft of the same model.

2. Recommendations for stocking densities

The following table gives stocking density recommendations for different domestic species. The values are expressed in kilograms and metres.
### Annex XIII (contd)

<table>
<thead>
<tr>
<th>Species</th>
<th>Weight</th>
<th>Density</th>
<th>Space/animal</th>
<th>No. of animals per</th>
<th>Animals per single tier pallet</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>kg</td>
<td>kg/m²</td>
<td>m²/animal</td>
<td>10 m²</td>
<td>214x264 cm</td>
</tr>
<tr>
<td>Calves</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>50</td>
<td>220</td>
<td>0.23</td>
<td>43</td>
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<tr>
<td></td>
<td>70</td>
<td>246</td>
<td>0.28</td>
<td>35/6</td>
<td>20</td>
</tr>
<tr>
<td></td>
<td>80</td>
<td>266</td>
<td>0.30</td>
<td>33</td>
<td>18</td>
</tr>
<tr>
<td></td>
<td>90</td>
<td>280</td>
<td>0.32</td>
<td>31</td>
<td>17</td>
</tr>
<tr>
<td>Cattle</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>300</td>
<td>344</td>
<td>0.84</td>
<td>11-12</td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>500</td>
<td>393</td>
<td>1.27</td>
<td>8</td>
<td>4</td>
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<tr>
<td></td>
<td>600</td>
<td>408</td>
<td>1.45</td>
<td>6-7</td>
<td>3</td>
</tr>
<tr>
<td></td>
<td>700</td>
<td>400</td>
<td>1.63</td>
<td>6</td>
<td>3</td>
</tr>
<tr>
<td>Sheep</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>25</td>
<td>147</td>
<td>0.17</td>
<td>59</td>
<td>32</td>
</tr>
<tr>
<td></td>
<td>70</td>
<td>196</td>
<td>0.36</td>
<td>27/8</td>
<td>15</td>
</tr>
<tr>
<td>Pigs</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>25</td>
<td>172</td>
<td>0.15</td>
<td>67</td>
<td>37</td>
</tr>
<tr>
<td></td>
<td>100</td>
<td>196</td>
<td>0.51</td>
<td>20</td>
<td>10</td>
</tr>
</tbody>
</table>

### Preparation for air transport of livestock

1. **Health and customs requirements**
   
   The legal requirements including animal health, welfare and species conservation, should be ascertained from the country of destination and any in transit countries before the animals are assembled or the transportation is arranged.

   Contact the *Veterinary Authorities* in the country of origin regarding veterinary certification.

   Planning of the transportation should take into account weekends, holidays and airport closures.

   Verify that any proposed intransit stops or alternates will not jeopardise the importing or in transit countries health requirements.

   Waiting time at customs (cargo handling and clearance) should be reduced as much as possible to avoid welfare problems.

2. **Environment**

   *Animals* are affected by extremes of temperature. This is especially true of high temperature when compounded by high humidity. Temperature and humidity should therefore be taken into consideration when planning the shipment.

   Times of arrival, departure and stopovers should be planned so that the aircraft lands during the coolest hours.

   At outside temperatures of below 25°C at the landing point, the aircraft doors should be opened to ensure adequate ventilation. Confirmation should be received from government authorities that animal health legislation does not prevent opening of aircraft doors.

   When outside temperatures at any landing point exceed 25°C, prior arrangements should be made to have an adequate air-conditioning unit available when the plane lands.
3. **Facilities and equipment**

Specific arrangements should be made to ensure that holding and loading facilities including ramps, trucks, and air-conditioning units are available at departure, all in transit and arrival airports. This should include identification of specific staff who are responsible and the method of contacting them, e.g. telephone number and address.

Specific notification should be given to all those responsible for providing facilities or equipment at the destination and in transit stops immediately before departure.

*Containers* should be loaded so as to ensure access can be made to the *animals* at all times.

4. **Preparation of animals**

Vaccination should be done far enough in advance of the departure date to allow for immunity to develop.

Veterinary certification and serological testing should be arranged several weeks in advance of livestock shipment.

Many *animals* require acclimatisation before they are transported. *Animals* such as swine and wild herbivores should be separated and held in the groups that will occupy *containers*. Mixing of such *animals* immediately before or during transport is extremely stressing and should be avoided.

Incompatible *animals* should be transported singly.

Article 7.4.5.

**Disinfection and disinfestation**

1. **Disinfection**

   a) Those parts of the interior of the aircraft destined for the carriage of *animals* should be thoroughly cleaned of all foreign matters using methods acceptable to aircraft management before being loaded.

   b) These parts should be sprayed with a disinfectant:

      i) suitable for the *diseases* which could be carried by the *animals*;

      ii) that does not cause problems with the aircraft;

      iii) that will not leave a residue hazardous to the *animals* being transported.

   If in doubt, the airline should be consulted on the suitability of the disinfectant. A mechanical nebuliser should be used to minimise the amount of disinfectant used.

   Suggested disinfectants currently in use are:

      iv) 4% sodium carbonate and 0.1% sodium silicate;

      v) 0.2% citric acid.

   c) All removeable equipment, penning and *containers* including loading ramps should be thoroughly cleaned and disinfected in accordance with the requirements of both the *exporting* and *importing countries*. 
Annex XIII (contd)

d) After disinfection, all equipment to be replaced in the aircraft should be washed with clean water to remove any traces of disinfectant to avoid any damage to the aircraft structures.

2. Disinfestation

Where disinfestation is required, the country requesting the action should be consulted for appropriate procedures.


Article 7.4.6.

Radiation

Radioactive materials should be separated from live animals by a distance of at least 0.5 metre for journeys not exceeding 24 hours, and by a distance of at least 1.0 metre for journeys longer than 24 hours (reference: Technical instructions on storage and loading-separation of the International Civil Aviation Organisation). Special care should be taken with regard to pregnant animals, semen and embryos/ova.

Article 7.4.7.

Tranquilization

Experience has shown that there is considerable risk in sedating animals transported by air. Tranquilizers reduce the ability of the animals to respond to stress during transportation. In addition, the reaction of various species to tranquilization cannot always be foreseen. For these reasons, routine tranquilization is not recommended. Tranquilizers should only be used when a specific problem exists, and should be administered by a veterinarian or by a person who has been instructed in their use. Persons using these drugs should understand the full implications of the effects of the drug in air transport, e.g. certain animals such as horses and elephants should not go down in containers. Drugs should only be administered during the flight with the knowledge and consent of the captain.

In all cases, when tranquilizers are used, a note should be attached to the container stating the weight of the individual animal, the generic name of the drug used, the dose, the method and time of administration.

Article 7.4.8.

Destruction of carcasses

In the event of any animal death on board, the competent authority of the airport of destination should be notified in advance of landing.

Carcasses should be disposed of under the supervision of and to the satisfaction of the Veterinary Authority of the country the aircraft is in.

The method of disposal should be based on the risk of introducing a controlled disease.

For carcasses which represent a high risk of introducing disease, the following is recommended:

1. destruction by incineration, rendering or deep burial under the supervision of the Veterinary Authority;

2. if removed from the airport site, transportation in a closed, leakproof container.
Emergency slaughter

Emergency slaughter of animals in aircraft should, in general, only occur when the safety of the aircraft, crew or other animals are involved.

Every aircraft transporting animals should have a method of killing the animals with minimum pain and someone trained in that method.

In all cases when horses or other large animals are to be carried, the method of killing should be discussed with the airline during the planning stages. Suitable methods are:

1. Captive bolt stunner, followed by an injection of a lethal chemical
   a) Operator should be trained to use the captive bolt stunner on the species or type of animal being transported.
   b) An expert should determine that the type of captive bolt pistol is adequate for all the animals being transported.
   c) Some airlines and countries may prohibit the carriage of captive bolt pistols.
   d) The user should recognise that the noise associated with the captive bolt may excite other animals.
   e) The requirement that the captive bolt pistol is accurately centered may be difficult to achieve with an excited animal.

2. Injection of a chemical
   a) Various chemicals may be used to sedate, immobilize or kill animals.
   b) Central nervous system depressants such as barbiturate euthanasia solutions should be injected directly into a vein to be effective. This is not normally practical for anyone but an experienced veterinarian or an especially trained and experienced attendant, where the animal is sufficiently fractious to require euthanasia.
   c) Sedatives such as promazine and its derivatives may make the animal more fractious (see Article 7.4.7.).
   d) Immobilizing solutions such as succinylcholine are not humane.

3. Firearms
   Airlines do not permit the use of firearms which discharge a free bullet because of the danger to the aircraft.

Handling of food and waste material

Waste material which contains anything of animal origin including food, litter, manure, or animal feed should be handled, collected and disposed of in a manner that ensures it will not be fed to livestock. It should be collected in specified areas, and stored and transported in closed, leakproof containers.

Some importing countries legislation may prohibit or restrict the use of hay or straw during the transportation period. Unloading of hay, straw, other animal feed and litter may be restricted or prohibited by in transit countries.
Disposal of food and waste material

Recommended methods of disposal are:

1. incineration to an ash;
2. heating at an internal temperature of at least of 100°C for 30 minutes, then disposal in a land fill site;
3. controlled burial in a land fill site.
CHAPTER 7.5.

SLAUGHTER OF ANIMALS

Article 7.5.1.

General principles

1. Object

These recommendations address the need to ensure the welfare of food animals during pre-slaughter and slaughter processes, until they are dead.

These recommendations apply to the slaughter in slaughterhouses of the following domestic animals: cattle, buffalo, bison, sheep, goats, camelids, deer, horses, pigs, ratites, rabbits and poultry. Other animals, wherever they have been reared, and all animals slaughtered outside slaughterhouses should be managed to ensure that their transport, lairage, restraint and slaughter is carried out without causing undue stress to the animals; the principles underpinning these recommendations apply also to these animals.

2. Personnel

Persons engaged in the unloading, moving, lairage, care, restraint, stunning, slaughter and bleeding of animals play an important role in the welfare of those animals. For this reason, there should be a sufficient number of personnel, who should be patient, considerate, competent and familiar with the recommendations outlined in the present chapter and their application within the national context.

Competence may be gained through formal training and/or practical experience. This competence should be demonstrated through a current certificate from the Competent Authority or from an independent body accredited by the Competent Authority.

The management of the slaughterhouse and the Veterinary Services should ensure that slaughterhouse staff are competent and carry out their tasks in accordance with the principles of animal welfare.

3. Animal behaviour

Animal handlers should be experienced and competent in handling and moving farm livestock, and understand the behaviour patterns of animals and the underlying principles necessary to carry out their tasks.

The behaviour of individual animals or groups of animals will vary, depending on their breed, sex, temperament and age and the way in which they have been reared and handled. Despite these differences, the following behaviour patterns which are always present to some degree in domestic animals should be taken into consideration in handling and moving the animals.

Most domestic livestock are kept in groups and follow a leader by instinct.

Animals which are likely to harm each other in a group situation should not be mixed at slaughterhouses.

The desire of some animals to control their personal space should be taken into account in designing facilities.
Domestic animals will try to escape if any person approaches closer than a certain distance. This critical distance, which defines the flight zone, varies among species and individuals of the same species, and depends upon previous contact with humans. Animals reared in close proximity to humans i.e. tame have a smaller flight zone, whereas those kept in free range or extensive systems may have flight zones which may vary from one metre to many metres. Animal handlers should avoid sudden penetration of the flight zone which may cause a panic reaction which could lead to aggression or attempted escape.

Animal handlers should use the point of balance at the animal's shoulder to move animals, adopting a position behind the point of balance to move an animal forward and in front of the point of balance to move it backward.

Domestic animals have wide-angle vision but only have limited forward binocular vision and poor perception of depth. This means that they can detect objects and movements beside and behind them, but can only judge distances directly ahead.

Although most domestic animals have a highly sensitive sense of smell, they react in different ways to the smells of slaughterhouses. Smells which cause fear or other negative responses should be taken into consideration when managing animals.

Domestic animals can hear over a greater range of frequencies than humans and are more sensitive to higher frequencies. They tend to be alarmed by constant loud noise and by sudden noises, which may cause them to panic. Sensitivity to such noises should also be taken into account when handling animals.

An example of a flight zone (cattle)
4. **Distractions and their removal**

Distractions that may cause approaching animals to stop, baulk or turn back should be designed out from new facilities or removed from existing ones. Below are examples of common distractions and methods for eliminating them:

a) reflections on shiny metal or wet floors — move a lamp or change lighting;

b) dark entrances to chutes, races, stun boxes or conveyor restrainers — illuminate with indirect lighting which does not shine directly into the eyes of approaching animals;

c) animals seeing moving people or equipment up ahead — install solid sides on chutes and races or install shields;

d) dead ends — avoid if possible by curving the passage, or make an illusory passage;

e) chains or other loose objects hanging in chutes or on fences — remove them;

f) uneven floors or a sudden drop in floor levels at the entrance to conveyor restrainers — avoid uneven floor surfaces or install a solid false floor under the restrainer to provide an illusion of a solid and continuous walking surface;

g) sounds of air hissing from pneumatic equipment — install silencers or use hydraulic equipment or vent high pressure to the external environment using flexible hosing;

h) clanging and banging of metal objects — install rubber stops on gates and other devices to reduce metal to metal contact;
Moving and handling animals

1. General considerations

Each slaughterhouse should have a dedicated plan for animal welfare. The purpose of such plan should be to maintain good level of animal welfare at all stages of the handling of animals until they are killed. The plan should contain standard operating procedures for each step of animal handling as to ensure that animal welfare is properly implemented based on relevant indicators. It also should include specific corrective actions in case of specific risks, like power failures or other circumstances that could negatively affect the welfare of animals.

Animals should be transported to slaughter in a way that minimises adverse animal health and welfare outcomes, and the transport should be conducted in accordance with the OIE recommendations for the transportation of animals (Chapters 7.2. and 7.3.).

The following principles should apply to unloading animals, moving them into lairage pens, out of the lairage pens and up to the slaughter point:

a) The conditions of the animals should be assessed upon their arrival for any animal welfare and health problems.

b) Injured or sick animals, requiring immediate slaughter, should be killed humanely and without delay, in accordance with the recommendations of the OIE.

c) Animals should not be forced to move at a speed greater than their normal walking pace, in order to minimise injury through falling or slipping. Performance standards should be established where numerical scoring of the prevalence of animals slipping or falling is used to evaluate whether animal moving practices and/or facilities should be improved. In properly designed and constructed facilities with competent animal handlers, it should be possible to move 99% of animals without their falling.

d) Animals for slaughter should not be forced to walk over the top of other animals.

e) Animals should be handled in such a way as to avoid harm, distress or injury. Under no circumstances should animal handlers resort to violent acts to move animals, such as crushing or breaking tails of animals, grasping their eyes or pulling them by the ears. Animal handlers should never apply an injurious object or irritant substance to animals and especially not to sensitive areas such as eyes, mouth, ears, anogenital region or belly. The throwing or dropping of animals, or their lifting or dragging by body parts such as their tail, head, horns, ears, limbs, wool, hair or feathers, should not be permitted. The manual lifting of small animals is permissible.

f) When using goads and other aids, the following principles should apply:

i) Animals that have little or no room to move should not be subjected to physical force or goads and other aids which compel movement. Electric goads and prods should only be used in extreme cases and not on a routine basis to move animals. The use and the power output should be restricted to that necessary to assist movement of an animal and only when an animal has a clear path ahead to move. Goads and other aids should not be used repeatedly if the animal fails to respond or move. In such cases it should be investigated whether some physical or other impediment is preventing the animal from moving.
ii) The use of such devices should be limited to battery-powered goads on the hindquarters of pigs and large ruminants, and never on sensitive areas such as the eyes, mouth, ears, anogenital region or belly. Such instruments should not be used on horses, sheep and goats of any age, or on calves or piglets.

iii) Useful and permitted goads include panels, flags, plastic paddles, flappers (a length of cane with a short strap of leather or canvas attached), plastic bags and metallic rattles; they should be used in a manner sufficient to encourage and direct movement of the animals without causing undue stress.

iv) Painful procedures (including whipping, tail twisting, use of nose twitches, pressure on eyes, ears or external genitalia), or the use of goads or other aids which cause pain and suffering (including large sticks, sticks with sharp ends, lengths of metal piping, fencing wire or heavy leather belts), should not be used to move animals.

v) Excessive shouting at animals or making loud noises (e.g. through the cracking of whips) to encourage them to move should not occur, as such actions may make the animals agitated, leading to crowding or falling.

vi) Animals should be grasped or lifted in a manner which avoids pain or suffering and physical damage (e.g. bruising, fractures, dislocations). In the case of quadrupeds, manual lifting by a person should only be used in young animals or small species, and in a manner appropriate to the species; grasping or lifting such animals only by their wool, hair, feathers, feet, neck, ears, tails, head, horns, limbs causing pain or suffering should not be permitted, except in an emergency where animal welfare or human safety may otherwise be compromised.

vii) Conscious animals should not be thrown, dragged or dropped.

viii) Performance standards should be established to evaluate the use of such instruments. Numerical scoring may be used to measure the percentage of animals moved with an electric instrument and the percentage of animals slipping or falling at a point in the slaughterhouse. Any risk of compromising animal welfare, for example slippery floor, should be investigated immediately and the defect rectified to eliminate the problem. In addition to resource-based measures, outcome-based measures (e.g. bruises, lesions, behaviour, and mortality) should be used to monitor the level of welfare of the animals.

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2. Specific considerations for poultry

Stocking density in transport crates should be optimum to suit climatic conditions and to maintain species-specific thermal comfort within containers.

Care is especially necessary during loading and unloading to avoid wings or legs body parts being caught on crates, leading to dislocated or broken wings bones in conscious birds. Such injuries will adversely affect animal welfare, carcass and meat quality.
Modular systems that involve tipping of live birds are not conducive to maintaining good animal welfare. These systems, when used, should be incorporated with a mechanism to facilitate birds sliding out of the transport system, rather than being dropped or dumped on top of each other from heights of more than a metre.

Birds may get trapped or their wings or claws may get caught in the fixtures, mesh or holes in poorly designed and/or constructed transport systems. Under this situation, operators unloading birds should ensure gentle release of trapped birds.

Drawers in modular systems and crates should be stacked and de-stacked carefully so as to avoid injury to birds.

Birds should have sufficient space so that all can lie down at the same time without being on top of each other.

Birds with broken bones and/or dislocated joints should be humanely killed before being hung on shackles for processing.

The number of poultry arriving at the processing plant with broken bones and/or dislocated joints should be recorded in a manner that allows for verification. For poultry, the percentage of chickens with broken or dislocated wings should not exceed 2%, with less than 1% being the goal (under study).

3. Provisions relevant to animals delivered in containers

   a) Containers in which animals are transported should be handled with care, and should not be thrown, dropped or knocked over. Where possible, they should be horizontal while being loaded and unloaded mechanically, and stacked to ensure ventilation. In any case they should be moved and stored in an upright position as indicated by specific marks.

   b) Animals delivered in containers with perforated or flexible bottoms should be unloaded with particular care in order to avoid injury. Where appropriate, animals should be unloaded from the containers individually.

   c) Animals which have been transported in containers should be slaughtered as soon as possible; mammals and ratites which are not taken directly upon arrival to the place of slaughter should have drinking water available to them from appropriate facilities at all times. Delivery of poultry for slaughter should be scheduled such that they are not deprived of water at the premises for longer than 12 hours. Animals which have not been slaughtered within 12 hours of their arrival should be fed, and should subsequently be given moderate amounts of food at appropriate intervals.

4. Provisions relevant to restraining and containing animals

   a) Provisions relevant to restraining animals for stunning or slaughter without stunning, to help maintain animal welfare, include:

      i) provision of a non-slippery floor;

      ii) avoidance of excessive pressure applied by restraining equipment that causes struggling or vocalisation in animals;

      iii) equipment engineered to reduce noise of air hissing and clanging metal;

      iv) absence of sharp edges in restraining equipment that would harm animals;
v) avoidance of jerking or sudden movement of restraining device;

vi) the restrainer should not look like a dead end.

b) Methods of restraint causing avoidable suffering should not be used in conscious animals because they cause severe pain and stress:

i) suspending or hoisting animals (other than poultry) by the feet or legs;

ii) indiscriminate and inappropriate use of stunning equipment;

iii) mechanical clamping of the legs or feet of the animals (other than shackles used in poultry and ostriches) as the sole method of restraint;

iv) breaking legs, cutting leg tendons or blinding animals in order to immobilise them;

v) severing the spinal cord, for example using a puntilla or dagger, to immobilise animals, except for proper stunning.

Article 7.5.3.

**Lairage design and construction**

1. **General considerations**

   The lairage should be designed and constructed to hold an appropriate number of animals in relation to the throughput rate of the slaughterhouse without compromising the welfare of the animals.

   In order to permit operations to be conducted as smoothly and efficiently as possible without injury or undue stress to the animals, the lairage should be designed and constructed so as to allow the animals to move freely in the required direction, using their behavioural characteristics and without undue penetration of their flight zone.

   The following recommendations may help to achieve this.

2. **Design of lairage**

   a) The lairage should be designed to allow a one-way flow of animals from unloading to the point of slaughter, with a minimum number of abrupt corners to negotiate.

   b) In red meat slaughterhouses, pens, passageways and races should be arranged in such a way as to permit inspection of animals at any time, and to permit the removal of sick or injured animals when considered to be appropriate, for which separate appropriate accommodation should be provided.
c) Each animal should have room to stand up and lie down and, when confined in a pen, to turn around, except where the animal is reasonably restrained for safety reasons (e.g. fractious bulls). Fractious animals should be slaughtered as soon as possible after arrival at the slaughterhouse to avoid welfare problems. The lairage should have sufficient accommodation for the number of animals intended to be held. Drinking water should always be available to the animals, and the method of delivery should be appropriate to the type of animal held. Troughs should be designed and installed in such a way as to minimise the risk of fouling by faeces, without introducing risk of bruising and injury in animals, and should not hinder the movement of animals.

d) Holding pens should be designed to allow as many animals as possible to stand or lie down against a wall. Where feed troughs are provided, they should be sufficient in number and feeding space to allow adequate access of all animals to feed. The feed trough should not hinder the movement of animals.

e) Where tethers, ties or individual stalls are used, these should be designed so as not to cause injury or distress to the animals and should also allow the animals to stand, lie down and access any food or water that may need to be provided.

f) Passageways and races should be either straight or consistently curved, as appropriate to the animal species. Passageways and races should have solid sides, but when there is a double race, the shared partition should allow adjacent animals to see each other. For pigs and sheep, passageways should be wide enough to enable two or more animals to walk side by side for as long as possible. At the point where passageways are reduced in width, this should be done by a means which prevents excessive bunching of the animals.

g) Animal handlers should be positioned alongside races and passageways on the inside radius of any curve, to take advantage of the natural tendency of animals to circle an intruder. Where one-way gates are used, they should be of a design which avoids bruising. Races should be horizontal but where there is a slope, they should be constructed to allow the free movement of animals without injury.

h) In slaughterhouses with high throughput, there should be a waiting pen, with a level floor and solid sides, between the holding pens and the race leading to the point of stunning or slaughter, to ensure a steady supply of animals for stunning or slaughter and to avoid having animal handlers trying to rush animals from the holding pens. The waiting pen should preferably be circular, but in any case, so designed that animals cannot be trapped or trampled.

i) Ramps or lifts should be used for the loading and unloading of animals where there is a difference in height or a gap between the floor of the vehicle and the unloading area. Unloading ramps should be designed and constructed so as to permit animals to be unloaded from vehicles on the level or at the minimum gradient achievable. Lateral side protection should be available to prevent animals escaping or falling. They should be well drained, with secure footholds and adjustable to facilitate easy movement of animals without causing distress or injury.

3. Construction of lairage

a) Lairages should be constructed and maintained so as to provide protection from unfavourable climatic conditions, using strong and resistant materials such as concrete and metal which has been treated to prevent corrosion. Surfaces should be easy to clean. There should be no sharp edges or protuberances which may injure the animals.

b) Floors should be well drained and not slippery; they should not cause injury to the feet of the animals. Where necessary, floors should be insulated or provided with appropriate bedding. Drainage grids should be placed at the sides of pens and passageways and not where animals would have to cross them. Discontinuities or changes in floor, wall or gate, colour, patterns or texture which could cause baulking in the movement of animals should be avoided.
c) **Lairages** should be provided with adequate lighting, but care should be taken to avoid harsh lights and shadows, which frighten the *animals* or affect their movement. The fact that *animals* will move more readily from a darker area into a well-lit area might be exploited by providing for lighting that can be regulated accordingly.

d) **Lairages** should be adequately ventilated to ensure that waste gases (e.g. ammonia) do not build up and that draughts at animal height are minimised. Ventilation should be able to cope with the range of expected climatic conditions and the number of *animals* the lairage will be expected to hold.

e) Care should be taken to protect the *animals* from excessively or potentially disturbing noises, for example by avoiding the use of noisy hydraulic or pneumatic equipment, and muffling noisy metal equipment by the use of suitable padding, or by minimising the transmission of such noises to the areas where *animals* are held and slaughtered.

f) Where *animals* are kept in outdoor **lairages** without natural shelter or shade, they should be protected from the effects of adverse weather conditions.

Article 7.5.4.

**Care of animals in lairages**

*Animals* in **lairages** should be cared for in accordance with the following recommendations:

1. As far as possible, established groups of *animals* should be kept together. Each *animal* should have enough space to stand up, lie down and turn around. *Animals* hostile to each other should be separated.

2. Where tethers, ties or individual stalls are used, they should allow *animals* to stand up and lie down without causing injury or distress.

3. Where bedding is provided, it should be maintained in a condition that minimises risks to the health and safety of the *animals*, and sufficient bedding should be used so that *animals* do not become soiled with manure.

4. *Animals* should be kept securely in the **lairage**, and care should be taken to prevent them from escaping and from predators.

5. Suitable drinking water should be available to the *animals* on their arrival and at all times to *animals* in **lairages** unless they are to be slaughtered without delay.

6. If *animals* are not to be slaughtered **within 12 hours of their arrival** as soon as possible, suitable feed should be available to the *animals* on arrival and at intervals appropriate to the species. Unweaned *animals* should be slaughtered as soon as possible.

7. In order to prevent heat stress, *animals* subjected to high temperatures, particularly pigs and **poultry**, should be cooled by the use of water sprays, fans or other suitable means. However, the potential for water sprays to reduce the ability of *animals* to thermoregulate (especially **poultry**) should be considered in any decision to use water sprays. The risk of *animals* being exposed to very cold temperatures or sudden extreme temperature changes should also be considered.

8. The **lairage** area should be well lit in order to enable the *animals* to see clearly without being dazzled. During the night, the lights should be dimmed. Lighting should also be adequate to permit inspection of all *animals*. Subdued lighting, and for example blue light, may be useful in **poultry lairages** in helping to calm birds.
Annex XIII (contd)

9. The condition and state of health of the animals in a lairage should be inspected at least every morning and evening by a veterinarian or, under the veterinarian's responsibility, by another competent person, such as an animal handler. Animals which are sick, weak, injured or showing visible signs of distress should be separated, and veterinary advice should be sought immediately regarding treatment or the animals should be humanely killed immediately if necessary.

10. Lactating dairy animals should be slaughtered as soon as possible. Dairy animals with obvious udder distension should be milked to minimise udder discomfort.

11. Animals which have given birth during the journey or in the lairage should be slaughtered as soon as possible or provided with conditions which are appropriate for suckling for their welfare and the welfare of the newborn. Under normal circumstances, animals which are expected to give birth during a journey should not be transported.

12. Animals with horns, antlers or tusks capable of injuring other animals, if aggressive, should be penned separately.

13. Poultry awaiting slaughter should be protected from adverse weather conditions and provided with adequate ventilation.

14. Waiting time should be minimised and should not exceed 12 hours when no food or water is provided during waiting.

15. Poultry in transport containers should be examined at the time of arrival. Containers should be stacked with sufficient space between the stacks to facilitate inspection of birds and air movement.

16. Forced ventilation or other cooling systems may be necessary under certain conditions to avoid build up of temperature and humidity. Temperature and humidity should be monitored at appropriate intervals.

Recommendations for specific species are described in detail in Articles 7.5.5. to 7.5.9.

Article 7.5.5.

Management of foetuses during slaughter of pregnant animals

Under normal circumstances, pregnant animals that would be in the final 10% of their gestation period at the planned time of unloading at the slaughterhouse should be neither transported nor slaughtered. If such an event occurs, an animal handler should ensure that females are handled separately, and the specific procedures described below are applied. In all cases, the welfare of foetuses and dams during slaughter should be safeguarded.

Foetuses should not be removed from the uterus sooner than 5 minutes after the maternal neck or chest cut, to ensure absence of consciousness. A foetal heartbeat will usually still be present and foetal movements may occur at this stage, but these are only a cause for concern if the exposed foetus successfully breathes air.

If a live mature foetus is removed from the uterus, it should be prevented from inflating its lungs and breathing air (e.g. by clamping the trachea).

When uterine, placental or foetal tissues, including foetal blood, are not to be collected as part of the post-slaughter processing of pregnant animals, all foetuses should be left inside the unopened uterus until they are dead. When uterine, placental or foetal tissues are to be collected, where practical, foetuses should not be removed from the uterus until at least 15–20 minutes after the maternal neck or chest cut.
If there is any doubt about consciousness, the foetus should be killed with a captive bolt of appropriate size or a blow to the head with a suitable blunt instrument.

The above recommendations do not refer to foetal rescue. Foetal rescue, the practice of attempting to revive foetuses found alive at the evisceration of the dam, should not be attempted during normal commercial slaughter as it may lead to serious welfare complications in the newborn animal. These include impaired brain function resulting from oxygen shortage before rescue is completed, compromised breathing and body heat production because of foetal immaturity, and an increased incidence of infections due to a lack of colostrum.

Article 7.5.6.

Summary analysis of handling and restraining methods and the associated animal welfare issues

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<td>Wing shackling</td>
<td>Electrical</td>
<td>Excessive tension applied prior to stunning</td>
<td>Competent animal handlers</td>
</tr>
<tr>
<td></td>
<td>Mechanical - upright</td>
<td>V-restrainer</td>
<td>Electrical methods</td>
<td>Loading of animal and overriding; excessive pressure, size mismatch between restrainer and animal</td>
<td>Proper design and operation of equipment</td>
</tr>
<tr>
<td>Presentation of animals</td>
<td>Specific procedure</td>
<td>Specific purpose</td>
<td>Animal welfare concerns/implications</td>
<td>Key animal welfare requirements</td>
<td>Applicable species</td>
</tr>
<tr>
<td>------------------------</td>
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</tr>
<tr>
<td>Mechanical-upright</td>
<td>Mechanical straddle – band restrainer (moving)</td>
<td>Electrical methods</td>
<td>Loading of animal and overriding, size mismatch between restrainer and animal</td>
<td>Competent animal handlers, proper design and layout of restraint</td>
<td>Cattle, calves, sheep, goats, pigs</td>
</tr>
<tr>
<td>Mechanical-upright</td>
<td>Flat bed/deck Tipped out of containers on to conveyors</td>
<td>Presentation of birds for shackling prior to electrical stunning Gas stunning</td>
<td>Stress and injury due to tipping in dump-module systems height of tipping conscious poultry broken bones and dislocations</td>
<td>Proper design and operation of equipment</td>
<td>Poultry</td>
</tr>
<tr>
<td>Suspension and/or inversion</td>
<td>Poultry shackle</td>
<td>Electrical stunning</td>
<td>Inversion stress; pain from compression on leg bones; Keep restraint as short as possible</td>
<td>Competent animal handlers; proper design and operation of equipment; birds should be hung by both legs</td>
<td>Poultry</td>
</tr>
<tr>
<td>Suspension and/or inversion</td>
<td>Cone</td>
<td>Electrical – head-only Captive bolt Slaughter without stunning</td>
<td>Inversion stress</td>
<td>Competent animal handlers; proper design and operation of equipment</td>
<td>Poultry</td>
</tr>
<tr>
<td>Upright restraint</td>
<td>Mechanical leg clamping</td>
<td>Electrical – head-only</td>
<td>Stress of resisting restraint in ostriches</td>
<td>Competent animal handlers; proper equipment design and operation</td>
<td>Ostriches</td>
</tr>
<tr>
<td>Restraining by inversion</td>
<td>Rotating box Fixed side(s) (e.g. Weinberg pen)</td>
<td>Slaughter without stunning</td>
<td>Inversion stress; stress of resisting restraint, prolonged restraint, inhalation of blood and ingesta Keep restraint as brief as possible</td>
<td>Proper design and operation of equipment</td>
<td>Cattle</td>
</tr>
<tr>
<td></td>
<td>Compressible side(s)</td>
<td>Slaughter without stunning</td>
<td>Inversion stress, stress of resisting restraint, prolonged restraint Preferable to rotating box with fixed sides Keep restraint as brief as possible</td>
<td>Proper design and operation of equipment</td>
<td>Cattle</td>
</tr>
<tr>
<td>Presentation of animals</td>
<td>Specific procedure</td>
<td>Specific purpose</td>
<td>Animal welfare concerns/implications</td>
<td>Key animal welfare requirements</td>
<td>Applicable species</td>
</tr>
</tbody>
</table>
Annex XIII (contd)

<table>
<thead>
<tr>
<th>Presentation of animals</th>
<th>Specific procedure</th>
<th>Specific purpose</th>
<th>Animal welfare concerns/implications</th>
<th>Key animal welfare requirements</th>
<th>Applicable species</th>
</tr>
</thead>
<tbody>
<tr>
<td>Body restraint</td>
<td>Casting/hobbling</td>
<td>Manual</td>
<td>Mechanical stunning methods</td>
<td>Slaughter without stunning</td>
<td>Stress of resisting restraint; animal temperament; bruising. Keep restraint as short as possible</td>
</tr>
<tr>
<td>Leg restraints</td>
<td>Rope casting</td>
<td>Mechanical stunning methods</td>
<td>Slaughter without stunning</td>
<td>Stress of resisting restraint; prolonged restraint, animal temperament; bruising. Keep restraint as short as possible</td>
<td>Competent animal handlers</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Tying of 3 or 4 legs</td>
<td>Mechanical stunning methods</td>
<td>Slaughter without stunning</td>
<td>Stress of resisting restraint; prolonged restraint, animal temperament; bruising. Keep restraint as short as possible</td>
<td>Competent animal handlers</td>
</tr>
</tbody>
</table>

**Stunning methods**

1. **General considerations**

   The competence of the operators, and the appropriateness, and effectiveness of the method used for stunning and the maintenance of the equipment are the responsibility of the management of the slaughterhouse, and should be checked regularly by a Competent Authority.

   Persons carrying out stunning should be properly trained and competent, and should ensure that:
   
   a) the animal is adequately restrained;
   
   b) animals in restraint are stunned as soon as possible;
   
   c) the equipment used for stunning is maintained and operated properly in accordance with the manufacturer’s recommendations, in particular with regard to the species and size of the animal;
   
   d) the equipment is applied correctly;
   
   e) stunned animals are bled out (slaughtered) as soon as possible;
   
   f) animals are not stunned when slaughter is likely to be delayed; and
   
   g) backup stunning devices are available for immediate use if the primary method of stunning fails. Provision of a manual inspection area and simple intervention like captive bolt and cervical dislocation for poultry would help prevent potential welfare problems.

   In addition, such persons should be able to recognise when an animal is not correctly stunned and should take appropriate action.
2. Mechanical stunning

A mechanical device should be applied usually to the front of the head and perpendicular to the bone surface. For a more detailed explanation on the different methods for mechanical stunning, see Chapter 7.6. and Articles 7.6.6., 7.6.7. and 7.6.8. The following diagrams illustrate the proper application of the device for certain species.

**Cattle**

![Diagram of cattle head showing the mechanical stunning point](source)

The optimum position for cattle is at the intersection of two imaginary lines drawn from the rear of the eyes to the opposite horn buds.

**Pigs**

![Diagram of pig head showing the mechanical stunning point](source)

The optimum position for pigs is on the midline just above eye level, with the shot directed down the line of the spinal cord.

*Figure source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).*
Sheep

The optimum position for hornless sheep and goats is on the midline.

Goats

The optimum position for heavily horned sheep and horned goats is behind the poll, aiming towards the angle of the jaw.
Horses

Figure Source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).

The optimum position for horses is at right angles to the frontal surface, well above the point where imaginary lines from eyes to ears cross.

Signs of correct stunning using a mechanical instrument are as follows:

a) the animal collapses immediately and does not attempt to stand up;

b) the body and muscles of the animal become tonic (rigid) immediately after the shot;

c) normal rhythmic breathing stops; and

d) the eyelid is open with the eyeball facing straight ahead and is not rotated.

Poultry

Figure Source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).
Captive bolts powered by cartridges, compressed air or spring can be used for poultry. The optimum position for poultry species is at right angles to the frontal surface.

Firing of a captive bolt according to the manufacturers’ instructions should lead to immediate destruction of the skull and the brain and, as a result, immediate death.

3. Electrical stunning

   a) General considerations

   An electrical device should be applied to the animal in accordance with the following recommendations.

   Electrodes should be designed, constructed, maintained and cleaned regularly to ensure that the flow of current is optimal and in accordance with manufacturing specifications. They should be placed so that they span the brain. The application of electrical currents which bypass the brain is unacceptable unless the animal has been stunned. The use of a single current leg-to-leg is unacceptable as a stunning method.

   If, in addition, it is intended to cause cardiac arrest, the electrodes should either span the brain and immediately thereafter the heart, on the condition that it has been ascertained that the animal is adequately stunned, or span brain and heart simultaneously.

   Electrical stunning equipment should not be applied on animals as a means of guidance, movement, restraint or immobilisation, and shall not deliver any shock to the animal before the actual stunning or killing.

   Electrical stunning apparatus should be tested prior to application on animals using appropriate resistors or dummy loads to ensure the power output is adequate to stun animals.

   The electrical stunning apparatus should incorporate a device that monitors and displays voltage (true RMS) and the applied current (true RMS) and that such devices are regularly calibrated at least annually.
Appropriate measures, such as removing excess wool or wetting the skin only at the point of contact, can be taken to minimise impedance of the skin and facilitate effective stunning.

The stunning apparatus should be appropriate for the species. Apparatus required for electrical stunning should be provided with adequate power to achieve continuously the minimum current level recommended for stunning as indicated in the table below.

In all cases, the correct current level shall be attained within one second of the initiation of stun and maintained at least for between one and three seconds and in accordance with the manufacturer’s instructions. Minimum current levels for head-only stunning are shown in the following table.

<table>
<thead>
<tr>
<th>Species</th>
<th>Minimum current levels for head-only stunning</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cattle</td>
<td>1.5 amps</td>
</tr>
<tr>
<td>Calves (bovines of less than 6 month of age)</td>
<td>1.0 amps</td>
</tr>
<tr>
<td>Pigs</td>
<td>1.25 amps</td>
</tr>
<tr>
<td>Sheep and goats</td>
<td>1.0 amps</td>
</tr>
<tr>
<td>Lambs</td>
<td>0.7 amps</td>
</tr>
<tr>
<td>Ostriches</td>
<td>0.4 amps</td>
</tr>
</tbody>
</table>

b) Electrical stunning of birds using a waterbath

There should be no sharp bends or steep gradients in the shackle line and the shackle line should be as short as possible consistent with achieving acceptable line speeds, and ensuring that birds have settled by the time they reach the water bath. A breast comforter can be used effectively to reduce wing flapping and calm birds. The angle at which the shackle line approaches the entrance to the water bath, and the design of the entrance to the water bath, and the draining of excess 'live' water from the bath are all important considerations in ensuring birds are calm as they enter the bath, do not flap their wings, and do not receive pre-stun electric shocks.

In the case of birds suspended on a moving line, measures should be taken to ensure that the birds are not wing flapping at the entrance of the stunner. The birds should be secure in their shackle, but there should not be undue pressure on their shanks. The shackle size should be appropriate to fit the size of the shanks (metatarsal bones) of birds.

Birds should be hung on shackles by both legs.

Birds with dislocated or broken legs or wings should be humanely killed rather than shackled.

The duration between hanging on shackles and stunning should be kept to the minimum. In any event, the time between shackling and stunning should not exceed one minute.

Waterbaths for poultry should be adequate in size and depth for the type of bird being slaughtered, and their height should be adjustable to allow for the head of each bird to be immersed. The electrode immersed in the bath should extend the full length of the waterbath. Birds should be immersed in the bath up to the base of their wings.

The waterbath should be designed and maintained in such a way that when the shackles pass over the water, they are in continuous contact with the earthed rubbing bar.

The control box for the waterbath stunner should incorporate an ammeter which displays the total current flowing through the birds.
Annex XIII (contd)

The shackle-to-leg contact should be wetted preferably before the birds are inserted in the shackles. In order to improve the electrical conductivity of the water, it is recommended that salt be added in the waterbath as necessary. Additional salt should be added regularly as a solution to maintain suitable constant concentrations in the waterbath.

Using waterbaths, birds are stunned in groups and different birds will have different impedances. The voltage should be adjusted so that the total current is the required current per bird as shown in the table hereafter, multiplied by the number of birds in the waterbath at the same time. The following values have been found to be satisfactory when employing a 50 Hertz sinusoidal alternating current.

Birds should receive the current for at least 4 seconds.

While a lower current may also be satisfactory, the current shall in any case be such as to ensure that unconsciousness occurs immediately and lasts until the bird has been killed by cardiac arrest or by bleeding. When higher electrical frequencies are used, higher currents may be required.

Every effort shall be made to ensure that no conscious or live birds enter the scalding tank.

In the case of automatic systems, until fail-safe systems of stunning and bleeding have been introduced, a manual back-up system should be in place to ensure that any birds which have missed the waterbath stunner and/or the automatic neck-cutter are immediately stunned and/or killed immediately, and they are dead before entering scald tank.

To lessen the number of birds that have not been effectively stunned reaching neck cutters, steps should be taken to ensure that small birds do not go on the line amongst bigger birds and that these small birds are stunned separately. The height of the waterbath stunner should be adjusted according to the size of birds to ensure even the small birds are immersed in the water bath up to the base of the wings.

Waterbath stunning equipment should be fitted with a device which displays and records the details of the electrical key parameter.

Minimum current for stunning poultry when using 50Hz is as follows:

<table>
<thead>
<tr>
<th>Species</th>
<th>Current (milliamperes per bird)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Broilers</td>
<td>100</td>
</tr>
<tr>
<td>Layers (spent hens)</td>
<td>100</td>
</tr>
<tr>
<td>Turkeys</td>
<td>150</td>
</tr>
<tr>
<td>Ducks and geese</td>
<td>130</td>
</tr>
</tbody>
</table>

Minimum current for stunning poultry when using high frequencies is as follows:

<table>
<thead>
<tr>
<th>Frequency (Hz)</th>
<th>Chickens</th>
<th>Turkeys</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 200 Hz</td>
<td>100 mA</td>
<td>250 mA</td>
</tr>
<tr>
<td>From 200 to 400 Hz</td>
<td>150 mA</td>
<td>400 mA</td>
</tr>
<tr>
<td>From 400 to 1500 Hz</td>
<td>200 mA</td>
<td>400 mA</td>
</tr>
</tbody>
</table>
3. **Gas stunning (under study)**

   a) **Stunning of pigs by exposure to carbon dioxide (CO$_2$)**

   The concentration of CO$_2$ for stunning should be preferably 90% by volume but in any case no less than 80% by volume. After entering the stunning chamber, the animals should be conveyed to the point of maximum concentration of the gas as rapidly as possible and be kept until they are dead or brought into a state of insensibility which lasts until death occur due to bleeding. Ideally, pigs should be exposed to this concentration of CO$_2$ for 3 minutes. Sticking should occur as soon as possible after exit from the gas chamber.

   In any case, the concentration of the gas should be such that it minimises as far as possible all stress of the animal prior to loss of consciousness.

   The chamber in which animals are exposed to CO$_2$ and the equipment used for conveying them through it shall be designed, constructed and maintained in such a way as to avoid injury or unnecessary stress to the animals. The animal density within the chamber should be such to avoid stacking animals on top of each other.

   The conveyor and the chamber shall be adequately lit to allow the animals to see their surroundings and, if possible, each other.

   It should be possible to inspect the CO$_2$ chamber whilst it is in use, and to have access to the animals in emergency cases.

   The chamber shall be equipped to continuously measure and display register at the point of stunning the CO$_2$ concentration and the time of exposure, and to give a clearly visible and audible warning if the concentration of CO$_2$ falls below the required level.

   Emergency stunning equipment should be available at the point of exit from the stunning chamber and used on any pigs that do not appear to be dead or completely stunned.

   b) **Inert gas mixtures for stunning pigs**

   Inhalation of high concentration of carbon dioxide is aversive and can be distressing to animals. Therefore, the use of non-aversive gas mixtures is being developed.

   Such gas mixtures include:

   i) a maximum of 2% by volume of oxygen in argon, nitrogen or other inert gases, or

   ii) to a maximum of 30% by volume of carbon dioxide and a maximum of 2% by volume of oxygen in mixtures with carbon dioxide and argon, nitrogen or other inert gases.

   Exposure time to the gas mixtures should be sufficient to ensure that no pigs regain consciousness before death supervenes through bleeding or cardiac arrest is induced.

   c) **Gas stunning of poultry**

   The main objective of gas stunning is to avoid the pain and suffering associated with shackling conscious poultry under water bath stunning and killing systems. Therefore, gas stunning should be limited to birds contained in crates or on conveyors only. The gas mixture should be non-aversive to poultry.
Annex XIII (contd)

Live poultry contained within transport modules or crates may be exposed to gradually increasing concentrations of CO\textsubscript{2} until the birds are properly stunned. No bird should recover consciousness during bleeding.

Gas stunning of poultry in their transport containers will eliminate the need for live birds’ handling at the processing plant and all the problems associated with the electrical stunning. Gas stunning of poultry on a conveyor eliminates the problems associated with the electrical water bath stunning.

Live poultry should be conveyed into the gas mixtures either in transport crates or on conveyor belts.

The following gas procedures have been properly documented for chickens and turkeys but do not necessarily apply for other domestic birds. In any case the procedure should be designed as to ensure that all animals are properly stunned without unnecessary suffering. Some monitoring points for gas stunning could be the following:

- ensure smooth entry and passage of crates or birds through the system;
- avoid crowding of birds in crates or conveyors;
- monitor and maintain gas concentrations continuously during operation;
- provide visible and audible alarm systems if gas concentrations are inappropriate to the species;
- calibrate gas monitors and maintain verifiable records;
- ensure that duration of exposure is adequate to prevent recovery of consciousness;
- make provision to monitor and deal with recovery of consciousness;
- ensure that blood vessels are cut to induce death in unconscious birds;
- ensure that all birds are dead before entering scalding tank;
- provide emergency procedures in the event of system failure.

i) Gas mixtures used for stunning poultry include:

- a minimum of 2 minutes exposure to 40% carbon dioxide, 30% oxygen and 30% nitrogen, followed by a minimum of one minute exposure to 80% carbon dioxide in air; or
- a minimum of 2 minutes exposure to any mixture of argon, nitrogen or other inert gases with atmospheric air and carbon dioxide, provided that the carbon dioxide concentration does not exceed 30% by volume and the residual oxygen concentration does not exceed 2% by volume; or
- a minimum of 2 minutes exposure to argon, nitrogen, other inert gases or any mixture of these gases in atmospheric air with a maximum of 2% residual oxygen by volume; or
- a minimum of 2 minutes exposure to a minimum of 55% carbon dioxide in air; or
- a minimum of one minute exposure to 30% carbon dioxide in air, followed by a minimum of one minute exposure to at least 60% carbon dioxide in air.
ii) Requirements for effective use are as follows:

- Compressed gases should be vaporised prior to administration into the chamber and should be at room temperature to prevent any thermal shock; under no circumstances, should solid gases with freezing temperatures enter the chamber.
- Gas mixtures should be humidified.
- Appropriate gas concentrations of oxygen and carbon dioxide should be monitored and displayed continuously at the level of the birds inside the chamber to ensure that anoxia ensues.

Under no circumstances, should birds exposed to gas mixtures be allowed to regain consciousness. If necessary, the exposure time should be extended.

4. Bleeding

From the point of view of animal welfare, animals which are stunned with a reversible method should be bled without delay. Maximum stun-stick interval depends on the parameters of the stunning method applied, the species concerned and the bleeding method used (full cut or chest stick when possible). As a consequence, depending on those factors, the slaughterhouse operator should set up a maximum stun-stick interval that ensures that no animals recover consciousness during bleeding. In any case the following time limits should be applied.

All animals should be bled out by incising both carotid arteries, or the vessels from which they arise (e.g. chest stick). However, when the stunning method used causes cardiac arrest, the incision of all of these vessels is not necessary from the point of view of animal welfare.

It should be possible for staff to observe, inspect and access the animals throughout the bleeding period. Any animal showing signs of recovering consciousness should be re-stunned.

After incision of the blood vessels, no scalding carcass treatment or dressing procedures should be performed on the animals for at least 30 seconds, or in any case until all brain-stem reflexes have ceased.

<table>
<thead>
<tr>
<th>Stunning method</th>
<th>Maximum delay for bleeding to be started</th>
</tr>
</thead>
<tbody>
<tr>
<td>Electrical methods and non-penetrating captive bolt</td>
<td>20 seconds</td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
</tr>
<tr>
<td>CO₂</td>
<td>60 seconds (after leaving the chamber)</td>
</tr>
</tbody>
</table>

Article 7.5.8.

Summary analysis of stunning methods and the associated animal welfare issues

<table>
<thead>
<tr>
<th>Method</th>
<th>Specific method</th>
<th>Animal welfare concerns/implications</th>
<th>Key animal welfare requirements applicable</th>
<th>Species</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mechanical</td>
<td>Free bullet</td>
<td>Inaccurate targeting and inappropriate ballistics</td>
<td>Operator competence; achieving outright kill with first shot</td>
<td>Cattle, calves, buffalo, deer, horses, pigs (boars and sows)</td>
<td>Personnel safety</td>
</tr>
</tbody>
</table>
### Annex XIII (contd)

<table>
<thead>
<tr>
<th>Method</th>
<th>Specific method</th>
<th>Animal welfare concerns/implications</th>
<th>Key animal welfare requirements applicable</th>
<th>Species</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Captive bolt - penetrating</td>
<td></td>
<td>Inaccurate targeting, velocity and diameter of bolt</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, buffalo, sheep, goats, pigs, camelids, ratites, poultry</td>
<td>(Unsuitable for specimen collection from TSE suspects). A back-up gun should be available in the event of an ineffective shot</td>
</tr>
<tr>
<td>Captive bolt - non-penetrating</td>
<td></td>
<td>Inaccurate targeting, velocity of bolt, potentially higher failure rate than penetrating captive bolt</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, sheep, goats, deer, pigs, camelids, ratites, poultry</td>
<td>Presently available devices are not recommended for young bulls and animals with thick skull. This method should only be used for cattle and sheep when alternative methods are not available.</td>
</tr>
<tr>
<td>Manual percussive blow</td>
<td></td>
<td>Inaccurate targeting; insufficient power; size of instrument</td>
<td>Competent animal handlers; restraint; accuracy. Not recommended for general use</td>
<td>Young and small mammals, ostriches and poultry</td>
<td>Mechanical devices potentially more reliable. Where manual percussive blow is used, unconsciousness should be achieved with single sharp blow delivered to central skull bones</td>
</tr>
<tr>
<td>Electrical</td>
<td>Split application:</td>
<td>Accidental pre-stun electric shocks; electrode positioning; application of a current to the body while animal conscious; inadequate current and voltage</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, sheep, goats and pigs, ratites and poultry</td>
<td>Systems involving repeated application of head-only or head-to-leg with short current durations (&lt;1 second) in the first application should not be used.</td>
</tr>
<tr>
<td>Single application:</td>
<td>1. across head then head to chest; 2. across head then across chest;</td>
<td>Accidental pre-stun electric shocks; inadequate current and voltage; wrong electrode positioning; recovery of consciousness</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, sheep, goats, pigs, ratites, poultry</td>
<td></td>
</tr>
<tr>
<td>Waterbath</td>
<td></td>
<td>Restraint, accidental pre-stun electric shocks; inadequate current and voltage; recovery of consciousness</td>
<td>Competent operation and maintenance of equipment</td>
<td>Poultry only</td>
<td></td>
</tr>
<tr>
<td>Gaseous</td>
<td>CO₂ air/O₂ mixture; CO₂ inert gas mixture</td>
<td>Aversiveness of high CO₂; respiratory distress; inadequate exposure</td>
<td>Concentration; duration of exposure; design, maintenance and operation of equipment; stocking density management</td>
<td>Pigs, poultry</td>
<td></td>
</tr>
<tr>
<td>Inert gases</td>
<td></td>
<td>Recovery of consciousness</td>
<td>Concentration; duration of exposure; design, maintenance and operation of equipment; stocking density management</td>
<td>Pigs, poultry</td>
<td></td>
</tr>
</tbody>
</table>
### Summary analysis of slaughter methods and the associated animal welfare issues

<table>
<thead>
<tr>
<th>Slaughter methods</th>
<th>Specific method</th>
<th>Animal welfare concerns/implications</th>
<th>Key requirements</th>
<th>Species</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Bleeding out by severance of blood vessels in the neck</strong></td>
<td>Full frontal cutting across the throat</td>
<td>Failure to cut both common carotid arteries; occlusion of cut arteries; pain during and after the cut</td>
<td>Failures to cut both common carotid arteries; occlusion of cut arteries; pain during and after the cut. A very sharp blade or knife of sufficient length so that the point of the knife remains outside the incision during the cut; the point of the knife should not be used to make the incision. The incision should not close over the knife during the throat cut.</td>
<td>Cattle, buffalo, horses, camels, sheep, goats, poultry, reptiles</td>
<td>No further procedure should be carried out before the bleeding out is completed (i.e. at least 30 seconds for mammals). The practice to remove hypothetical blood clots just after the bleeding should be discouraged since this may increase animal suffering.</td>
</tr>
<tr>
<td><strong>Bleeding out by severance of blood vessels in the neck</strong></td>
<td>Neck stab followed by forward cut</td>
<td>Ineffective stunning; failure to cut both common carotid arteries; impaired blood flow; delay in cutting after reversible stunning</td>
<td>Prompt and accurate cutting</td>
<td>Camels, sheep, goats, poultry, reptiles</td>
<td></td>
</tr>
<tr>
<td><strong>Bleeding out by severance of blood vessels in the neck</strong></td>
<td>Neck stab alone</td>
<td>Ineffective stunning; failure to cut both common carotid arteries; impaired blood flow; delay in cutting after reversible stunning</td>
<td>Prompt and accurate cutting</td>
<td>Camels, sheep, goats, poultry, reptiles</td>
<td></td>
</tr>
<tr>
<td><strong>Bleeding out by severance of blood vessels in the neck</strong></td>
<td>Chest stick into major arteries or hollow-tube knife into heart</td>
<td>Ineffective stunning; inadequate size of stick wound; inadequate length of sticking knife; delay in sticking after reversible stunning</td>
<td>Prompt and accurate sticking</td>
<td>Cattle, sheep, goats, pigs</td>
<td></td>
</tr>
<tr>
<td><strong>Bleeding out by severance of blood vessels in the neck</strong></td>
<td>Neck skin cut followed by severance of vessels in the neck</td>
<td>Ineffective stunning; inadequate size of stick wound; inadequate length of sticking knife; delay in sticking after reversible stunning</td>
<td>Prompt and accurate cutting of vessels</td>
<td>Cattle</td>
<td></td>
</tr>
<tr>
<td><strong>Bleeding out by severance of blood vessels in the neck</strong></td>
<td>Automated mechanical cutting</td>
<td>Ineffective stunning; failure to cut and misplaced cuts. Recovery of consciousness following reversible stunning systems</td>
<td>Design, maintenance and operation of equipment; accuracy of cut; manual back-up</td>
<td>Poultry only</td>
<td></td>
</tr>
<tr>
<td><strong>Bleeding out by severance of blood vessels in the neck</strong></td>
<td>Manual neck cut on one side</td>
<td>Ineffective stunning; recovery of consciousness following reversible stunning systems</td>
<td>Prior non-reversible stunning</td>
<td>Poultry only</td>
<td>N.B. slow induction of unconsciousness under slaughter without stunning</td>
</tr>
</tbody>
</table>
Annex XIII (contd)

<table>
<thead>
<tr>
<th>Slaughter methods</th>
<th>Specific method</th>
<th>Animal welfare concerns/ implications</th>
<th>Key requirements</th>
<th>Species</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bleeding with prior stunning (contd)</td>
<td>Oral cut</td>
<td>Ineffective stunning; recovery of consciousness following reversible stunning systems</td>
<td>Prior non-reversible stunning</td>
<td>Poultry only</td>
<td>N.B. slow induction of unconsciousness in non-stun systems</td>
</tr>
<tr>
<td>Other methods without stunning</td>
<td>Decapitation with a sharp knife</td>
<td>Pain due to loss of consciousness not being immediate</td>
<td></td>
<td>Sheep, goats, poultry</td>
<td>This method is only applicable to Jhatka slaughter</td>
</tr>
<tr>
<td></td>
<td>Manual neck dislocation and decapitation</td>
<td>Pain due to loss of consciousness not being immediate; difficult to achieve in large birds</td>
<td>Neck dislocation should be performed in one stretch to sever the spinal cord</td>
<td>Poultry only</td>
<td>Slaughter by neck dislocation should be performed in one stretch to sever the spinal cord. Acceptable only when slaughtering small numbers of small birds.</td>
</tr>
<tr>
<td>Cardiac arrest in a waterbath electric stunner</td>
<td>Bleeding by evisceration</td>
<td>Induction of cardiac arrest</td>
<td></td>
<td>Quail</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Bleeding by neck cutting</td>
<td></td>
<td></td>
<td>Poultry</td>
<td></td>
</tr>
</tbody>
</table>

Article 7.5.10.

Methods, procedures or practices unacceptable on animal welfare grounds

1. The restraining methods which work through immobilisation by injury such as breaking legs, leg tendon cutting, and severing the spinal cord (e.g. using a puntilla or dagger) cause severe pain and stress in animals. Those methods are not acceptable in any species.

2. The use of the electrical stunning method with a single application leg to leg is ineffective and unacceptable in any species.

3. The slaughter method of brain stem severance by piercing through the eye socket or skull bone without prior stunning is not acceptable in any species.

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text deleted
CHAPTER 7.6.

KILLING OF ANIMALS FOR DISEASE CONTROL PURPOSES

Article 7.6.1.

General principles

These recommendations are based on the premise that a decision to kill the animals has been made, and address the need to ensure the welfare of the animals until they are dead.

1. All personnel involved in the humane killing of animals should have the relevant skills and competencies. Competence may be gained through formal training and/or practical experience.

2. As necessary, operational procedures should be adapted to the specific circumstances operating on the premises and should address, apart from animal welfare, aesthetics of the method of euthanasia, cost of the method, operator safety, biosecurity and environmental aspects.

3. Following the decision to kill the animals, killing should be carried out as quickly as possible, and normal husbandry should be maintained until the animals are killed.

4. The handling and movement of animals should be minimised and when done, it should be carried out done in accordance with the recommendations described below.

5. Animal restraint should be sufficient to facilitate effective killing, and in accordance with animal welfare and operator safety requirements; when restraint is required, killing should follow with minimal delay.

6. When animals are killed for disease control purposes, methods used should result in immediate death or immediate loss of consciousness lasting until death; when loss of consciousness is not immediate, induction of unconsciousness should be non-aversive or the least aversive possible and should not cause avoidable anxiety, pain, distress or suffering in animals.

7. For animal welfare considerations, young animals should be killed before older animals; for biosecurity considerations, infected animals should be killed first, followed by in-contact animals, and then the remaining animals.

8. There should be continuous monitoring of the procedures by the Competent Authorities to ensure they are consistently effective with regard to animal welfare, operator safety and biosecurity.

9. When the operational procedures are concluded, there should be a written report describing the practices adopted and their effect on animal welfare, operator safety and biosecurity.

10. These general principles should also apply when animals need to be killed for other purposes such as after natural disasters or for culling animal populations.

Article 7.6.2.

Organisational structure

Disease control contingency plans should be in place at a national level and should contain details of management structure, disease control strategies and operational procedures; animal welfare considerations should be addressed within these disease control contingency plans. The plans should also include a strategy to ensure that an adequate number of personnel competent in the humane killing of animals is available. Local level plans should be based on national plans and be informed by local knowledge.
Annex XIII (contd)

Disease control contingency plans should address the animal welfare issues that may result from animal movement controls.

The operational activities should be led by an official Veterinarian who has the authority to appoint the personnel in the specialist teams and ensure that they adhere to the required animal welfare and biosecurity standards. When appointing the personnel, he/she should ensure that the personnel involved have the required competencies.

The official Veterinarian should be responsible for all activities across one or more affected premises and should be supported by coordinators for planning (including communications), operations and logistics to facilitate efficient operations.

The official Veterinarian should provide overall guidance to personnel and logistic support for operations on all affected premises to ensure consistency in adherence to the OIE animal welfare and animal health recommendations.

A specialist team, led by a team leader answerable to the official Veterinarian, should be deployed to work on each affected premises. The team should consist of personnel with the competencies to conduct all required operations; in some situations, personnel may be required to fulfil more than one function. Each team should contain a veterinarian or have access to veterinary advice at all times.

In considering the animal welfare issues associated with killing animals, the key personnel, their responsibilities and competencies required are described in Article 7.6.3.

Article 7.6.3.

Responsibilities and competencies of the specialist team

1. Team leader
   a) Responsibilities
      i) plan overall operations on affected premises;
      ii) determine and address requirements for animal welfare, operator safety and biosecurity;
      iii) organise, brief and manage team of people to facilitate humane killing of the relevant animals on the premises in accordance with national regulations and these recommendations;
      iv) determine logistics required;
      v) monitor operations to ensure animal welfare, operator safety and biosecurity requirements are met;
      vi) report upwards on progress and problems;
      vii) provide a written report at the conclusion of the killing, describing the practices adopted and their effect on the animal welfare, operator safety and biosecurity outcomes.

   b) Competencies
      i) appreciation of normal animal husbandry practices;
      ii) appreciation of animal welfare and the underpinning behavioural, anatomical and physiological processes involved in the killing process;
      iii) skills to manage all activities on premises and deliver outcomes on time;
      iv) awareness of psychological effects on farmer, team members and general public;
v) effective communication skills;
vi) appreciation of the environmental impacts caused by their operation.

2. **Veterinarian**
   a) **Responsibilities**
      i) determine and supervise the implementation of the most appropriate killing method to ensure that animals are killed without avoidable pain and distress;
      ii) determine and implement the additional requirements for animal welfare, including the order of killing;
      iii) ensure that confirmation of the death of the animals is carried out by competent persons at appropriate times after the killing procedure;
      iv) minimise the risk of disease spread within and from the premises through the supervision of biosecurity procedures;
      v) continuously monitor animal welfare and biosecurity procedures;
      vi) in cooperation with the leader, prepare a written report at the conclusion of the killing, describing the practices adopted and their effect on animal welfare.
   b) **Competencies**
      i) ability to assess animal welfare, especially the effectiveness of stunning and killing and to correct any deficiencies;
      ii) ability to assess biosecurity risks.

3. **Animal handlers**
   a) **Responsibilities**
      i) review on-site facilities in terms of their appropriateness;
      ii) design and construct temporary animal handling facilities, when required;
      iii) move and restrain animals;
      iv) continuously monitor animal welfare and biosecurity procedures.
   b) **Competencies**
      i) animal handling in emergency situations and in close confinement is required;
      ii) an appreciation of biosecurity and containment principles.

4. **Animal killing personnel**
   a) **Responsibilities**
      Humane killing of the animals through effective stunning and killing should be ensured.
Annex XIII (contd)

b) Competencies
   i) when required by regulations, licensed to use necessary equipment;
   ii) competent to use and maintain relevant equipment;
   iii) competent to use techniques for the species involved;
   iv) competent to assess effective stunning and killing.

5. Carcass disposal personnel
   a) Responsibilities
      An efficient carcass disposal (to ensure killing operations are not hindered) should be ensured.
   b) Competencies
      The personnel should be competent to use and maintain available equipment and apply techniques for the species involved.

6. Farmer/owner/manager
   a) Responsibilities
      i) assist when requested.
   b) Competencies
      ii) specific knowledge of his/her animals and their environment.

Considerations in planning the humane killing of animals

Many activities will need to be conducted on affected premises, including the humane killing of animals. The team leader should develop a plan for humbly killing animals on the premises which should include consideration of:

1. minimising handling and movement of animals;
2. killing the animals on the affected premises; however, there may be circumstances where the animals may need to be moved to another location for killing; when the killing is conducted at an abattoir, the recommendations in Chapter 7.5. on the slaughter of animals should be followed;
3. the species, number, age and size of animals to be killed, and the order of killing them;
4. methods of killing the animals, and their cost;
5. housing, husbandry, location of the animals as well as accessibility of the farm;
6. the availability and effectiveness of equipment needed for killing of the animals, as well as the time necessary to kill the required number of animals using such methods;
7. the facilities available on the premises that will assist with the killing including any additional facilities that may need to be brought on and then removed from the premises;
8. biosecurity and environmental issues;
9. the health and safety of personnel conducting the killing;
10. any legal issues that may be involved, for example where restricted veterinary drugs or poisons may be used, or where the process may impact on the environment;
11. the presence of other nearby premises holding animals;
12. possibilities for removal, disposal and destruction of carcasses.

The plan should minimise the negative welfare impacts of the killing by taking into account the different phases of the procedures to be applied for killing (choice of the killing sites, killing methods, etc.) and the measures restricting the movements of the animals.

Competences and skills of the personnel handling and killing animals.

In designing a killing plan, it is essential that the method chosen be consistently reliable to ensure that all animals are humanely and quickly killed.

Article 7.6.5.

Table summarising killing methods described in Articles 7.6.6.-7.6.18.

The methods are described in the order of mechanical, electrical and gaseous, not in an order of desirability from an animal welfare viewpoint.

<table>
<thead>
<tr>
<th>Species</th>
<th>Age range</th>
<th>Procedure</th>
<th>Restraint necessary</th>
<th>Animal welfare concerns with inappropriate application</th>
<th>Article reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cattle</td>
<td>all</td>
<td>free bullet</td>
<td>no</td>
<td>non-lethal wounding</td>
<td>7.6.6.</td>
</tr>
<tr>
<td></td>
<td>all except neonates</td>
<td>penetrating captive bolt - followed by pithing or bleeding</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>7.6.7.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>non-penetrating captive bolt, followed by bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before killing</td>
<td>7.6.8.</td>
</tr>
<tr>
<td></td>
<td>calves only</td>
<td>electrical, two-stage application</td>
<td>yes</td>
<td>pain associated with cardiac arrest after ineffective stunning</td>
<td>7.6.10.</td>
</tr>
<tr>
<td></td>
<td>calves only</td>
<td>electrical, single application (method 1)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>7.6.11.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>injection with barbiturates and other drugs</td>
<td>yes</td>
<td>non-lethal dose, pain associated with injection site</td>
<td>7.6.15.</td>
</tr>
<tr>
<td>Sheep and goats</td>
<td>all</td>
<td>free bullet</td>
<td>no</td>
<td>non-lethal wounding</td>
<td>7.6.6.</td>
</tr>
<tr>
<td>Sheep and goats (contd)</td>
<td>all except neonates</td>
<td>penetrating captive bolt, followed by pithing or bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before death</td>
<td>7.6.7.</td>
</tr>
<tr>
<td></td>
<td>all except neonates</td>
<td>non-penetrating captive bolt, followed by bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before death</td>
<td>7.6.8.</td>
</tr>
<tr>
<td></td>
<td>neonates</td>
<td>non-penetrating captive bolt</td>
<td>yes</td>
<td>non-lethal wounding</td>
<td>7.6.8.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>electrical, two-stage application</td>
<td>yes</td>
<td>pain associated with cardiac arrest after ineffective stunning</td>
<td>7.6.10.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>electrical, single application (method 1)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>7.6.11.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>CO₂ / air mixture</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>7.6.12.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>nitrogen and/or inert gas mixed with CO₂</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>7.6.13.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>nitrogen and/or inert gases</td>
<td>yes</td>
<td>slow induction of unconsciousness</td>
<td>7.6.14.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>injection of barbiturates and other drugs</td>
<td>yes</td>
<td>non-lethal dose, pain associated with injection site</td>
<td>7.6.15.</td>
</tr>
</tbody>
</table>
### Annex XIII (contd)

<table>
<thead>
<tr>
<th>Species</th>
<th>Age range</th>
<th>Procedure</th>
<th>Restraint necessary</th>
<th>Animal welfare concerns with inappropriate application</th>
<th>Article reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Pigs</strong></td>
<td>all, except neonates</td>
<td>free bullet</td>
<td>no</td>
<td>non-lethal wounding</td>
<td>7.6.6.</td>
</tr>
<tr>
<td></td>
<td>all except neonates</td>
<td>penetrating captive bolt, followed by pithing or bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before death</td>
<td>7.6.7.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>non-penetrating captive bolt</td>
<td>yes</td>
<td>non-lethal wounding</td>
<td>7.6.8.</td>
</tr>
<tr>
<td></td>
<td>all&lt;sup&gt;1&lt;/sup&gt;</td>
<td>electrical, two-stage application</td>
<td>yes</td>
<td>pain associated with cardiac arrest after ineffective stunning</td>
<td>7.6.10.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>electrical, single application (method 1)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>7.6.11.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>CO&lt;sub&gt;2&lt;/sub&gt; / air mixture</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>7.6.12.</td>
</tr>
<tr>
<td><strong>Pigs</strong> (contd)</td>
<td>neonates only</td>
<td>nitrogen and/or inert gas mixed with CO&lt;sub&gt;2&lt;/sub&gt;</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>7.6.13.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>nitrogen and/or inert gases</td>
<td>yes</td>
<td>slow induction of unconsciousness</td>
<td>7.6.14.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>injection with barbiturates and other drugs</td>
<td>yes</td>
<td>non-lethal dose, pain associated with injection site</td>
<td>7.6.15.</td>
</tr>
<tr>
<td><strong>Poultry</strong></td>
<td>adults only</td>
<td>non-penetrating captive bolt</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>7.6.8.</td>
</tr>
<tr>
<td></td>
<td>day-olds and eggs only</td>
<td>maceration</td>
<td>no</td>
<td>non-lethal wounding, non-immediacy</td>
<td>7.6.9.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>electrical, single application (method 2)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>7.6.11.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>electrical, single application, followed by killing (method 3)</td>
<td>yes</td>
<td>ineffective stunning; regaining of consciousness before death</td>
<td>7.6.11.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>CO&lt;sub&gt;2&lt;/sub&gt; / air mixture Method 1 Method 2</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>7.6.12.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>nitrogen and/or inert gas mixed with CO&lt;sub&gt;2&lt;/sub&gt;</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>7.6.13.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>nitrogen and/or inert gases</td>
<td>yes</td>
<td>slow induction of unconsciousness</td>
<td>7.6.14.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>injection of barbiturates and other drugs</td>
<td>yes</td>
<td>non-lethal dose, pain associated with injection site</td>
<td>7.6.15.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>addition of anaesthetics to feed or water, followed by an appropriate killing method</td>
<td>no</td>
<td>ineffective or slow induction of unconsciousness</td>
<td>7.6.16.</td>
</tr>
</tbody>
</table>

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**Free bullet**

1. **Introduction**
   
   a) A free bullet is a projectile fired from a shotgun, rifle, handgun or purpose-made humane killer.
   
   b) The most commonly used firearms for close range use are:
      
      i) humane killers (specially manufactured/adapted single-shot weapons);
      
      ii) shotguns (12, 16, 20, 28 bore and .410);
      
      iii) rifles (.22 rimfire);
iv) handguns (various calibres from .32 to .45).

c) The most commonly used firearms for long range use are rifles (.22, .243, .270 and .308).

d) A free bullet used from long range should be aimed to penetrate the skull or soft tissue at the top of the neck of the animals (high neck shot) and to cause irreversible concussion and death and should only be used by properly trained and competent marksmen.

2. Requirements for effective use

a) The marksman should take account of human safety in the area in which he/she is operating. Appropriate vision and hearing protective devices should be worn by all personnel involved.

b) The marksman should ensure that the animal is not moving and in the correct position to enable accurate targeting and the range should be as short as possible (5–50 cm for a shotgun) but the barrel should not be in contact with the head of the animals.

c) The correct cartridge, calibre and type of bullet for the different species age and size should be used. Ideally, the ammunition should expand upon impact and dissipate its energy within the cranium.

d) Shot animals should be checked to ensure the absence of brain stem reflexes.

3. Advantages

a) Used properly, a free bullet provides a quick and effective method for killing.

b) It requires minimal or no restraint and can be used to kill from a distance by properly trained and competent marksmen.

c) It is suitable for killing agitated animals in open spaces.

4. Disadvantages

a) The method is potentially dangerous to humans and other animals in the area.

b) It has the potential for non-lethal wounding.

c) Destruction of brain tissue may preclude diagnosis of some diseases.

d) Leakage of bodily fluids may present a biosecurity risk.

e) Legal requirements may preclude or restrict use.

f) There is a limited availability of competent personnel.

5. Conclusion

The method is suitable for cattle, sheep, goats and pigs, including large animals in open spaces.

**Figure 1.** The optimum shooting position for cattle is at the intersection of two imaginary lines drawn from the rear of the eyes to the opposite horn buds.
Annex XIII (contd)

Figure 2. The optimum position for hornless sheep and goats is on the midline.

Figure 3. The optimum shooting position for heavily horned sheep and horned goats is behind the poll aiming towards the angle of the jaw.
Figure 4. The optimum shooting position for pigs is just above eye level, with the shot directed down the line of the spinal cord.

Figure source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).

Article 7.6.7.

Penetrating captive bolt

1. Introduction

A penetrating captive bolt is fired from a gun powered by either compressed air or a blank cartridge. There is no free projectile.

The captive bolt should be aimed on the skull in a position to penetrate the cortex and mid-brain of the animal. The impact of the bolt on the skull produces unconsciousness. Physical damage to the brain caused by penetration of the bolt may result in death; however, pithing or bleeding should be performed as soon as possible after the shot to ensure the death of the animal. Shooting poultry species with the captive bolts results in immediate destruction of the skull and brain, causing death. For a detailed description on the use of this method, see Chapter 7.5. of the Terrestrial Code.

2. Requirements for effective use

a) For cartridge powered and compressed air guns, the bolt velocity and the length of the bolt should be appropriate to the species and type of animal, in accordance with the recommendations of the manufacturer.

b) Captive bolt guns should be frequently cleaned and maintained in good working condition.

c) More than one gun may be necessary to avoid overheating, and a back-up gun should be available in the event of an ineffective shot.

d) Animals should be restrained; at a minimum, they should be penned for cartridge powered guns and in a race for compressed air guns.
Annex XIII (contd)

e) The operator should ensure that the head of the animal is accessible.

f) The operator should fire the captive bolt at right angles to the skull in the optimal position (see figures 1, 3 & 4. The optimum shooting position for hornless sheep is on the highest point of the head, on the midline and aim towards the angle of the jaw).

g) To ensure the death of the animal, pithing or bleeding should be performed as soon as possible after stunning.

h) Animals should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

3. Advantages
   a) Mobility of cartridge powered equipment reduces the need to move animals.
   b) The method induces an immediate onset of a sustained period of unconsciousness.

4. Disadvantages
   a) Poor gun maintenance and misfiring, and inaccurate gun positioning and orientation may result in poor animal welfare.
   b) Post stun convulsions may make pithing difficult and hazardous.
   c) The method is difficult to apply in agitated animals.
   d) Repeated use of a cartridge powered gun may result in over-heating.
   e) Leakage of bodily fluids may present a biosecurity risk.
   f) Destruction of brain tissue may preclude diagnosis of some diseases.

5. Conclusions
   The method is suitable for poultry, cattle, sheep, goats and pigs (except neonates), when followed by pithing or bleeding.

   Article 7.6.8.

Non-penetrating captive bolt

1. Introduction
   A non-penetrating captive bolt is fired from a gun powered by either compressed air or a blank cartridge. There is no free projectile.

   The gun should be placed on the front of the skull to deliver a percussive blow which produces unconsciousness in cattle (adults only), sheep, goats and pigs, and death in poultry and neonate sheep, goats and pigs. Bleeding should be performed as soon as possible after the blow to ensure the death of the animal.

2. Requirements for effective use
   a) For cartridge powered and compressed air guns, the bolt velocity should be appropriate to the species and type of animal, in accordance with the recommendations of the manufacturer.
   b) Captive bolt guns should be frequently cleaned and maintained in good working condition.
c) More than one gun may be necessary to avoid overheating, and a back-up gun should be available in the event of an ineffective shot.

d) Animals should be restrained; at a minimum mammals should be penned for cartridge powered guns and in a race for compressed air guns; birds should be restrained in cones, shackles, crushes or by hand.

e) The operator should ensure that the head of the animal is accessible.

f) The operator should fire the captive bolt at right angles to the skull in the optimal position (figures 1–4).

g) To ensure death in non-neonate mammals, bleeding should be performed as soon as possible after stunning.

h) Animals should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

3. **Advantages**

   a) The method induces an immediate onset of unconsciousness, and death in birds and neonates.

   b) Mobility of equipment reduces the need to move animals.

4. **Disadvantages**

   a) As consciousness can be regained quickly in non-neonate mammals, they should be bled as soon as possible after stunning.

   b) Laying hens in cages have to be removed from their cages and most birds have to be restrained.

   c) Poor gun maintenance and misfiring, and inaccurate gun positioning and orientation may result in poor animal welfare.

   d) Post stun convulsions may make bleeding difficult and hazardous.

   e) Difficult to apply in agitated animals; such animals may be sedated in advance of the killing procedure.

   f) Repeated use of a cartridge powered gun may result in over-heating.

   g) Bleeding may present a biosecurity risk.

5. **Conclusions**

   The method is suitable for killing poultry, and neonate sheep, goats and pigs up to a maximum weight of 10 kg.

   **Article 7.6.9.**

**Maceration**

1. **Introduction**

   Maceration, utilising a mechanical apparatus with rotating blades or projections, causes immediate fragmentation and death in day-old poultry and embryonated eggs.
Annex XIII (contd)

2. **Requirements**
   a) Maceration requires specialised equipment which should be kept in excellent working order.
   b) The rate of introducing the birds should not allow the equipment to jam, birds to rebound from the blades or the birds to suffocate before they are macerated.

3. **Advantages**
   a) Procedure results in immediate *death*.
   b) Large numbers can be killed quickly.

4. **Disadvantages**
   a) Specialised equipment is required.
   b) Macerated tissues may present biosecurity or human health risks.
   c) The cleaning of the equipment can be a source of contamination.

5. **Conclusion**
   The method is suitable for *killing* day-old poultry and embryonated eggs.

Article 7.6.10.

**Electrical – two-stage application**

1. **Introduction**
   A two-stage application of electric current comprises firstly an application of current to the head by scissor-type tongs, immediately followed by an application of the tongs across the chest in a position that spans the heart.

   The application of sufficient electric current to the head will induce ‘tonic/clonic’ epilepsy and unconsciousness. Once the *animal* is unconscious, the second stage will induce ventricular fibrillation (cardiac arrest) resulting in *death*. The second stage (the application of low frequency current across the chest) should only be applied to unconscious *animals* to prevent unacceptable levels of pain.

2. **Requirements for effective use**
   a) The stunner control device should generate a low frequency (AC sine wave 50 Hz) current with a minimum voltage and current as set out in the following table:

<table>
<thead>
<tr>
<th>Animal</th>
<th>Minimum voltage (V)</th>
<th>Minimum current (A)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cattle</td>
<td>220</td>
<td>1.5</td>
</tr>
<tr>
<td>Sheep</td>
<td>220</td>
<td>1.0</td>
</tr>
<tr>
<td>Pigs over 6 weeks of age</td>
<td>220</td>
<td>1.3</td>
</tr>
<tr>
<td>Pigs less than 6 weeks of age</td>
<td>125</td>
<td>0.5</td>
</tr>
</tbody>
</table>

   b) Appropriate protective clothing (including rubber gloves and boots) should be worn.
   c) *Animals* should be restrained, at a minimum free-standing in a pen, close to an electrical supply.
d) Two team members are required, the first to apply the electrodes and the second to manipulate the position of the animal to allow the second application to be made.

e) A stunning current should be applied via scissor-type stunning tongs in a position that spans the brain for a minimum of 3 seconds; immediately following the application to the head, the electrodes should be transferred to a position that spans the heart and the electrodes applied for a minimum of 3 seconds.

f) Electrodes should be cleaned regularly and after use, to enable optimum electrical contact to be maintained.

g) Animals should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

h) Electrodes should be applied firmly for the intended duration of time and pressure not released until the stun is complete.

3. Advantages
   a) The application of the second stage minimises post-stun convulsions and therefore the method is particularly effective with pigs.

   b) Non-invasive technique minimises biosecurity risk.

4. Disadvantages
   a) The method requires a reliable supply of electricity.

   b) The electrodes should be applied and maintained in the correct positions to produce an effective stun and kill.

   c) Most stunner control devices utilise low voltage impedance sensing as an electronic switch prior to the application of high voltages; in unshorn sheep, contact impedance may be too high to switch on the required high voltage (especially during stage two).

   d) The procedure may be physically demanding, leading to operator fatigue and poor electrode placement.

5. Conclusion
   The method is suitable for calves, sheep and goats, and especially for pigs (over one week of age).

Article 7.6.11.

Electrical – single application

1. Method 1

   Method 1 comprises the single application of sufficient electrical current to the head and back, to simultaneously stun the animal and fibrillate the heart. Provided sufficient current is applied in a position that spans both the brain and heart, the animal will not recover consciousness.
Annex XIII (contd)

a) Requirements for effective use

i) The stunner control device should generate a low frequency (30–60 Hz) current with a minimum voltage of 250 volts true RMS under load.

ii) Appropriate protective clothing (including rubber gloves and boots) should be worn.

iii) Animals should be individually and mechanically restrained close to an electrical supply as the maintenance of physical contact between the stunning electrodes and the animal is necessary for effective use.

iv) The rear electrode should be applied to the back, above or behind the heart, and then the front electrode in a position that is forward of the eyes, with current applied for a minimum of 3 seconds.

v) Electrodes should be cleaned regularly between animals and after use, to enable optimum electrical contact to be maintained.

vi) Water or saline may be necessary to improve electrical contact with sheep.

viii) An effective stun and kill should be verified by the absence of brain stem reflexes.

b) Advantages

i) Method 1 stuns and kills simultaneously.

ii) It minimises post-stun convulsions and therefore is particularly effective with pigs.

iii) A single team member only is required for the application.

iv) Non-invasive technique minimises biosecurity risk.

c) Disadvantages

i) Method 1 requires individual mechanical animal restraint.

ii) The electrodes should be applied and maintained in the correct positions to produce an effective stun and kill.

iii) Method 1 requires a reliable supply of electricity.

d) Conclusion

Method 1 is suitable for calves, sheep, goats, and pigs (over one week of age).

2. Method 2

Method 2 stuns and kills by drawing inverted and shackled poultry through an electrified waterbath stunner. Electrical contact is made between the ‘live’ water and earthed shackle and, when sufficient current is applied, poultry will be simultaneously stunned and killed.

a) Requirements for effective use

i) A mobile waterbath stunner and a short loop of processing line are required.
ii) A low frequency (50–60 Hz) current applied for a minimum of 3 seconds is necessary to stun and kill the birds.

iii) Poultry need to be manually removed from their cage, house or yard, inverted and shackled onto a line which conveys them through a waterbath stunner with their heads fully immersed.

iv) The required minimum currents to stun and kill dry birds are:
   - Quails – 100 mA/bird
   - Chickens – 160 mA/bird
   - Ducks & geese – 200 mA/bird
   - Turkeys – 250 mA/bird.
   A higher current is required for wet birds.

v) An effective stun and kill should be verified by the absence of brain stem reflexes.

b) Advantages
   i) Method 2 stuns and kills simultaneously.
   ii) It is capable of processing large numbers of birds reliably and effectively.
   iii) This non-invasive technique minimises biosecurity risk.

c) Disadvantages
   i) Method 2 requires a reliable supply of electricity.
   ii) Handling, inversion and shackling of birds are required.

d) Conclusion
   Method 2 is suitable for large numbers of poultry.

3. Method 3
   Method 3 comprises the single application of sufficient electrical current to the head of poultry in a position that spans the brain, causing unconsciousness; this is followed by a killing method (see Article 7.6.17.).

   a) Requirements for effective use
   i) The stunner control device should generate sufficient current (more than 600 mA/duck and more than 300 mA/bird) to stun.
   ii) Appropriate protective clothing (including rubber gloves and boots) should be worn.
   iii) Birds should be restrained, at a minimum manually, close to an electrical supply.
   iv) Electrodes should be cleaned regularly and after use, to enable optimum electrical contact to be maintained.
Annex XIII (contd)

v) Birds should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

b) Advantages

Non-invasive technique (when combined with cervical dislocation) minimises biosecurity risk.

c) Disadvantages

i) Method 3 requires a reliable supply of electricity and is not suitable for large-scale operations.

ii) The electrodes should be applied and maintained in the correct position to produce an effective stun.

iii) Birds should be individually restrained.

iv) It should be followed by a killing method.

d) Conclusion

Method 3 is suitable for small numbers of poultry.

CO2 / air mixture

1. Introduction

Controlled atmosphere killing is performed by exposing animals to a predetermined gas mixture, either by placing them in a gas-filled container or apparatus (Method 1) or by placing transport modules or crates containing birds in a gas tight container and introducing a gas mixture (Method 2) or by the gas being introduced into a poultry house (Method 3). Method 2 should be used whenever possible, as it eliminates welfare issues resulting from the need to manually remove live birds. Although Method 2 requires handling and crating of the birds, it benefits bird welfare overall (in comparison with Method 1) as it reduces the risk of death by smothering or suffocation.

Inhalation of carbon dioxide (CO2) induces respiratory and metabolic acidosis and hence reduces the pH of cerebrospinal fluid (CSF) and neurones thereby causing unconsciousness and, after prolonged exposure, death. Exposure to carbon dioxide does not induce immediate loss of consciousness, therefore the aversive nature of gas mixtures containing high concentrations of CO2 and the respiratory distress occurring during the induction phase are important considerations for animal welfare.

2. Method 1

The animals are placed in a gas-filled container or apparatus.

a) Requirements for effective use in a container or apparatus

i) Containers or apparatus should allow the required gas concentration to be maintained and accurately measured.

ii) When animals are exposed to the gas individually or in small groups in a container or apparatus, the equipment used should be designed, constructed, and maintained in such a way as to avoid injury to the animals and allow them to be observed.

iii) Animals can also be introduced to low concentrations (as low concentrations are not aversive) and the concentration could be increased afterwards and the animals then held in the higher concentration until death is confirmed.
Annex XIII (contd)

iv) Team members should ensure that there is sufficient time allowed for each batch of animals to die before subsequent ones are introduced into the container or apparatus.

v) Containers or apparatus should not be overcrowded and measures are needed to avoid animals suffocating by climbing on top of each other.

b) Advantages

i) $\text{CO}_2$ is readily available.

ii) Application methods are simple.

iii) The volume of gas required can be readily calculated.

iv) As the units are operated outdoor, the gas is dispersed quickly at the end of each cycle by opening the door, improving operator's health and safety.

v) The system uses skilled catching teams and equipment in daily use by the industry.

vi) Metal containers can be readily cleansed and disinfected.

c) Disadvantages

i) The need for properly designed container or apparatus.

ii) The aversive nature of high $\text{CO}_2$ concentrations.

iii) No immediate loss of consciousness.

iv) The risk of suffocation due to overcrowding.

v) Difficulty in verifying death while the animals are in the container or apparatus.

d) Conclusion

Method 1 is suitable for use in poultry, and neonatal sheep, goats and pigs.

3. Method 2

In this method, the crates or modules holding the birds are loaded into a chamber into which gas is introduced. As illustrated in the example below, a containerised gassing unit (CGU) typically comprises a gas-tight chamber designed to accommodate poultry transport crates or a single module. The chamber is fitted with gas lines and diffusers, with silencers that are connected via a system of manifolds and gas regulators to gas cylinders. There is a hole at the top to permit displaced air to escape when the container is filling with gas.

The procedures for the operation of CGU include (a) position the container on level, solid, open ground; (b) connect the gas cylinder to the container (c) load birds into the container (d) shut and secure the door, (e) deliver the gas until a concentration of $45\%$ by volume of carbon dioxide has been achieved at the top of the container, (f) allow time for the birds to become unconscious and die (g) open the door and allow gas to be dispersed in the air (h) remove the module (i) check each drawer for survivors (j) humanely kill any survivors; and (k) dispose of carcasses appropriately.
Annex XIII (contd)

a) Requirements for effective use of containerised gassing units (CGU)

i) The birds should be caught gently and placed in crates or modules of appropriate size and at appropriate stocking densities to allow all birds to sit down.

ii) The crates or module full of birds should be placed inside the container and the door shut only when the operator is ready to administer the gas.

iii) Ensure the container door is locked and administer the gas until a minimum concentration of 45% carbon dioxide is achieved at the top of the crates.

iv) An appropriate gas meter should be used to ensure the appropriate concentration of carbon dioxide is achieved and maintained until it can be confirmed that the birds have been killed.

v) Sufficient exposure time should be allowed for birds to die before the door is opened. In the absence of a viewing window that allows direct observation of birds during killing, cessation of vocalisation and convulsive wing flapping sounds, which can be listened to by standing near the container, can be used to determine that the birds are unconscious and that death is imminent. Remove the crates or modules from the container and leave them in the open air.

vi) Each crate or module should be examined and birds checked to ensure they are dead. Dilated pupils and absence of breathing indicate death.

vii) Any survivors should be humanely killed.

viii) Ducks and geese are resilient to the effects of carbon dioxide and therefore require a minimum of 80% CO₂ and a longer period of exposure to die.

b) Advantages

i) The gas is introduced quickly and quietly resulting in less turbulence and disturbance to the birds.

ii) Gradual increase in the concentration of CO₂ minimises the aversive nature of this method for inducing unconsciousness.

iii) The use of transport crates or modules to move birds minimises handling. Birds should be handled by trained, experienced catching teams at the time of depopulation of the poultry house.

iv) The modules are loaded mechanically into the CGU and a lethal mixture of gas is rapidly introduced into the chamber immediately after sealing.

v) CO₂ is readily available.

vi) Birds are exposed to gas more uniformly and they do not smother each other when compared with Method 1.

vii) The volume of gas required can be readily calculated.

viii) As the units are operated outdoors, the gas is dispersed quickly at the end of each cycle by opening the door, improving operator’s health and safety.

ix) The system uses skilled catching teams and equipment in daily use by the industry.

x) Metal containers can be readily cleansed and disinfected.
c) Disadvantages

i) Requires trained operators, trained catchers, transport modules and fork lift. However, this equipment and suitable areas with hard surfaces are usually available.

ii) The main limiting factors are speed of catching birds.

iii) In the absence of a viewing window, visual confirmation of death while the birds are still in the container is difficult. However, cessation of vocalisation and convulsive wing flapping sounds can be used to determine onset of death.

iv) The need for properly designed container or apparatus

v) No immediate loss of consciousness.

vi) The risk of suffocation due to overcrowding.

d) Conclusion

i) Method 2 is suitable for use in a wide range of poultry systems, providing there is access to vehicles to carry the containers and equipment.

ii) Birds should be introduced into the container or apparatus, which is then sealed and filled as quickly as possible with the required gas concentrations, i.e. more than 40% CO$_2$. Birds are held in this atmosphere until death is confirmed.

iii) Method 2 is suitable for use in poultry, and neonatal sheep, goats and pigs. However, CO$_2$ is likely to cause a period of distress in the animals before they lose consciousness.

4. Method 3

The gas is introduced into a poultry house.

a) Requirements for effective use in a poultry house

i) Prior to introduction of the CO$_2$, the poultry house should be appropriately sealed to allow control over the gas concentration. The interval between sealing and gas administration should be kept to the minimum so as to avoid overheating.

Forced ventilation systems, where fitted, should only be switched off immediately prior to gas administration.

The main water supply to the poultry house may have to be turned off and water drained to avoid freezing and bursting of water pipes.

Feeders and water troughs should be lifted to avoid obstruction of the gas entry and prevent injury to birds.

ii) Gas delivery pipes or lancets should be positioned appropriately such that birds are not hit directly by very cold gas delivered at high pressures. It may be necessary to exclude birds from the area in front of the delivery pipes, for a distance of about 20 meters, by partitioning the house with nets, wire mesh or similarly perforated materials.

iii) The house should be gradually filled with CO$_2$ so that all birds are exposed to a concentration of >40% until they are dead; a vaporiser may be required to prevent freezing.
Annex XIII (contd)

iv) Devices should be used to accurately measure the gas concentration at the maximum height accommodation of birds.

b) Advantages

i) Applying gas to birds in situ eliminates the need to manually remove live birds.

ii) CO₂ is readily available.

iii) Gradual raising of CO₂ concentration minimises the aversiveness of the induction of unconsciousness.

c) Disadvantages

i) It is difficult to determine volume of gas required to achieve adequate concentrations of CO₂ in some poultry houses.

ii) It is difficult to verify death while the birds are in the poultry house.

The extremely low temperature of liquid CO₂ entering the house and formation of solid CO₂ (dry ice) may cause concern for bird welfare.

d) Conclusion

Method 32 is suitable for use in poultry in closed-environment sheds. This method could be developed for killing pigs. However, CO₂ is likely to cause a period of distress in the birds before they lose consciousness.

Article 7.6.13.

Nitrogen and/or inert gas mixed with CO₂

1. Introduction

CO₂ may be mixed in various proportions with nitrogen or an inert gas (e.g. argon), and the inhalation of such mixtures leads to hypercapnic-hypoxia and death when the oxygen concentration by volume is <2%. Various mixtures of CO₂ and nitrogen or an inert gas can be administered to kill birds using Methods 1 and 32 described under 7.6.12. Whole house gassing with mixtures of CO₂ and nitrogen, or an inert gas, has not been tested owing to the complex issues presented by mixing gases in large quantities. Such mixtures however do not induce immediate loss of consciousness, therefore the aversiveness of various gas mixtures containing high concentrations of CO₂ and the respiratory distress occurring during the induction phase, are important animal welfare considerations.

Pigs and poultry appear not to find low concentrations of CO₂ strongly aversive, and a mixture of nitrogen or argon with <30% CO₂ by volume and <2% O₂ by volume can be used for killing poultry, neonatal sheep, goats and pigs.

2. Method 1

The animals are placed in a gas-filled container or apparatus.

a) Requirements for effective use

i) Containers or apparatus should allow the required gas concentrations to be maintained, and the O₂ and CO₂ concentrations accurately measured during the killing procedure.
ii) When animals are exposed to the gases individually or in small groups in a container or apparatus, the equipment used should be designed, constructed, and maintained in such a way as to avoid injury to the animals and allow them to be observed.

iii) Animals should be introduced into the container or apparatus after it has been filled with the required gas concentrations (with <2% O₂), and held in this atmosphere until death is confirmed.

iv) Team members should ensure that there is sufficient time allowed for each batch of animals to die before subsequent ones are introduced into the container or apparatus.

v) Containers or apparatus should not be overcrowded and measures are needed to avoid animals suffocating by climbing on top of each other.

b) Advantages

i) Low concentrations of CO₂ cause little aversiveness and, in combination with nitrogen or an inert gas, produces a fast induction of unconsciousness.

ii) The volume of gas required can be readily calculated.

iii) As the units are operated outdoors, the gas is dispersed quickly at the end of each cycle by opening the door, improving operator’s health and safety.

iv) Metal containers can be readily cleansed and disinfected.

v) Mixtures containing up to 20% carbon dioxide in argon are readily available as welding gas cylinders.

c) Disadvantages

i) A properly designed container or apparatus is needed.

ii) It is difficult to verify death while the animals are in the container or apparatus.

iii) There is no immediate loss of consciousness.

iv) Exposure times required to kill are considerable.

v) The risk of suffocation due to overcrowding.

d) Conclusion

The method is suitable for poultry, and for neonatal sheep, goats and pigs.

3. **Method 2**

In this method, the crates or modules holding the birds are loaded into a container and gas is introduced into the container (refer to Figures under Article 7.6.12.). As shown in the example below, each containerised gassing unit (CGU) typically comprises a gas-tight chamber designed to accommodate poultry transport crates or a module. The container or chamber is fitted with gas lines and diffusers, with silencers, which in turn are connected via a system of manifolds and gas regulators to gas cylinders. There is a hole at the top of the unit to permit displaced air to escape when filling the container with gas.
Procedures involved in the operation of CGU includes (a) position the container on a level, solid, open ground; (b) connect gas cylinder to the container (c) load a module of birds into the container, (d) shut and secure the door, (e) deliver the gas to the point where less than 2% by volume of oxygen is found at the top of the container, (f) allow time for the birds to become unconscious and die, (g) open the door and allow the gas to be dispersed in air, (h) remove the module, (i) check each drawer for survivors; (j) humanely kill survivors, if any; and (k) dispose carcasses appropriately.

a) Requirements for effective use of containerised gassing units (CGU)

   i) The birds should be caught gently and placed in crates or modules of appropriate size and at appropriate stocking densities to allow all birds to sit down.

   ii) The crates or module of birds should be placed inside the container and the door shut only when the operator is ready to administer the gas mixture.

   iii) Ensure the container door is locked and administer the gas mixture until <2% residual oxygen is achieved at the top of the crates.

   iv) An appropriate gas meter should be used to ensure a concentration of oxygen <2% is achieved and maintained until it can be confirmed that the birds have been killed.

   v) Sufficient exposure time should be allowed for birds to die before the door is opened. In the absence of a viewing window, which allows direct observation of birds during killing, cessation of vocalisation and wing flapping sounds can be observed by standing close to the container and used to determine the onset of death in birds. Remove the crates or modules from the container and leave them in the open air.

   vi) Each crate or module should be examined and birds checked to ensure they are dead. Dilated pupils and absence of breathing movements indicate death.

   vii) Any survivors should be humanely killed.

   viii) Ducks and geese do not appear to be resilient to the effects of a mixture of 20% carbon dioxide and 80% nitrogen or argon.

b) Advantages

   i) The gas mixture is introduced quickly and quietly resulting in less turbulence and disturbance to the birds.

   ii) The use of transport crates or modules to move birds minimises handling. Birds should be handled by trained, experienced catching teams at the time of depopulation of the poultry house.

   iii) The modules are loaded mechanically into the CGU and a lethal mixture of gas is rapidly introduced into the chamber immediately after sealing.

   iv) Mixtures containing up to 20% carbon dioxide in argon are readily available as welding gas cylinders.

   v) Birds are exposed to gas in a more uniform manner and they do not smother each other when compared with Method 1.

   vi) Two CGU can be operated in tandem and throughputs of up to 4,000 chickens per hour are possible.
vii) The volume of gas required can be readily calculated.

viii) As the units are operated outdoor the gas is dispersed quickly at the end of each cycle by opening the door, improving operators’ health and safety.

ix) The system uses skilled catching teams and equipment in daily use by the industry.

x) Metal containers can be readily cleansed and disinfected.

c) Disadvantages

i) Requires trained operators, trained catchers, transport modules and a fork lift. However, such equipment and suitable outdoor areas with a hard surface are usually available.

ii) The main limiting factors are speed of catching birds and availability of gas mixtures.

iii) In the absence of a viewing window, visual confirmation of death while the birds are still in the container is difficult. However, cessation of vocalisation and convulsive wing flapping can be used to determine the onset of death.

iv) CGU could be used to kill poultry on small to medium farms, e.g. up to 25 thousand birds on a single farm.

d) Conclusion

i) Method 2 is suitable for use in poultry and in neonatal sheep, goats and pigs.

ii) Method 2 is suitable for use in poultry in a wide range of poultry systems providing that these have access to vehicles to carry containers and equipment.

iii) Animals should be introduced into the container or apparatus, which is then sealed and filled as quickly as possible with the gas mixture. A residual oxygen concentration of less than 2% should be achieved and maintained and birds should be held in this atmosphere until death is confirmed.
Nitrogen and/or inert gases

1. Introduction

This method involves the introduction of animals into a container or apparatus containing nitrogen or an inert gas such as argon. The controlled atmosphere produced leads to unconsciousness and death from hypoxia.

Research has shown that hypoxia is not aversive to pigs and poultry, and it does not induce any signs of respiratory distress prior to loss of consciousness.

2. Requirements for effective use

a) Containers or apparatus should allow the required gas concentrations to be maintained, and the O2 concentration accurately measured.

b) When animals are exposed to the gases individually or in small groups in a container or apparatus, the equipment used should be designed, constructed, and maintained in such a way as to avoid injury to the animals and allow them to be observed.

c) Animals should be introduced into the container or apparatus after it has been filled with the required gas concentrations (with <2% O2), and held in this atmosphere until death is confirmed.

d) Team members should ensure that there is sufficient time allowed for each batch of animals to die before subsequent ones are introduced into the container or apparatus.

e) Containers or apparatus should not be overcrowded, and measures are needed to avoid animals suffocating by climbing on top of each other.

3. Advantages

Animals are unable to detect nitrogen or inert gases, and the induction of hypoxia by this method is not aversive to animals.

4. Disadvantages

a) A properly designed container or apparatus is needed.

b) It is difficult to verify death while the animals are in the container or apparatus.

c) There is no immediate loss of consciousness.

d) Exposure times required to kill are considerable.

5. Conclusion

The method is suitable for poultry and neonatal sheep, goats and pigs.

Lethal injection

1. Introduction

A lethal injection using high doses of anaesthetic and sedative drugs causes CNS depression, unconsciousness and death. In practice, barbiturates in combination with other drugs are commonly used.
Annex XIII (contd)

2. Requirements for effective use
   a) Doses and routes of administration that cause rapid loss of consciousness followed by death should be used.
   b) Prior sedation may be necessary for some animals.
   c) Intravenous administration is preferred, but intraperitoneal or intramuscular administration may be appropriate, especially if the agent is non-irritating.
   d) Animals should be restrained to allow effective administration.
   e) Animals should be monitored to ensure the absence of brain stem reflexes.

3. Advantages
   a) The method can be used in all species.
   b) Death can be induced smoothly.

4. Disadvantages
   a) Restraint and/or sedation may be necessary prior to injection.
   b) Some combinations of drug type and route of administration may be painful, and should only be used in unconscious animals.
   c) Legal requirements and skill/training required may restrict use to veterinarians.
   d) Contaminated carcasses may present a risk to other wild or domestic animals.

5. Conclusion
   The method is suitable for killing small numbers of cattle, sheep, goats, pigs and poultry.

Article 7.6.16.

Addition of anaesthetics to feed or water

1. Introduction
   An anaesthetic agent which can be mixed with poultry feed or water may be used to kill poultry in houses. Poultry which are only anaesthetised need to be killed by another method such as cervical dislocation.

2. Requirements for effective use
   a) Sufficient quantities of anaesthetic need to be ingested rapidly for effective response.
   b) Intake of sufficient quantities is facilitated if the birds are fasted or water is withheld.
   c) Should be followed by killing (see Article 7.6.17.) if birds are anaesthetised only.

3. Advantages
   a) Handling is not required until birds are anaesthetised.
   b) There may be biosecurity advantages in the case of large numbers of diseased birds.
4. **Disadvantages**
   a) Non-target *animals* may accidentally access the medicated feed or water when provided in an open environment.
   b) Dose taken is unable to be regulated and variable results may be obtained.
   c) *Animals* may reject adulterated feed or water due to illness or adverse flavour.
   d) The method may need to be followed by *killing*.
   e) Care is essential in the preparation and provision of treated feed or water, and in the disposal of uneaten treated feed/water and contaminated carcasses.

5. **Conclusion**
   The method is suitable for *killing* large numbers of poultry in houses. However, a back-up method should be available to kill birds that are anaesthetized but not killed.

   **Article 7.6.17.**

**Cervical dislocation and decapitation**

1. **Cervical dislocation (manual and mechanical)**
   a) **Introduction**

   Unconscious poultry may be killed by either manual cervical dislocation (stretching the neck). This method results in *death* from cerebral anoxia due to cessation of breathing and/or blood supply to the brain.

   When the number of birds to be killed is small, and other methods of *killing* are not available, conscious birds of less than 3 kilograms may be killed using cervical dislocation in such a way that the blood vessels of the neck are severed and *death* is instantaneous.

   b) **Requirements for effective use**

   i) *Killing* should be performed either by manually or mechanically stretching the neck to sever the spinal cord or by using mechanical pliers to crush the cervical vertebrae with consequent major damage to the spinal cord.

   ii) Consistent results require strength and skill so team members should be rested regularly to ensure consistently reliable results.

   iii) Birds should be monitored continuously until *death* to ensure the absence of brain stem reflexes.

   c) **Advantages**

   i) It is a non-invasive *killing* method.

   ii) It can be performed manually on small birds.

   d) **Disadvantages**

   i) Operator fatigue.

   ii) The method is more difficult in larger birds.
Annex XIII (contd)

iii) Requires trained personnel to perform humanely.

iv) Human health and safety concerns due to handling of the birds.

v) Additional stress to the animals from handling.

2. **Decapitation**
   a) **Introduction**
      Decapitation results in *death* by cerebral ischaemia using a guillotine or knife.
   b) **Requirements for effective use**
      The required equipment should be kept in good working order.
   c) **Advantages**
      The technique is effective and does not require monitoring.
   d) **Disadvantages**
      i) The working area is contaminated with body fluids, which increases biosecurity risks.
      ii) Pain if consciousness is not lost immediately.

**Pithing and bleeding**

1. **Pithing**
   a) **Introduction**
      Pithing is a method of *killing animals* which have been stunned by a penetrating captive bolt, without immediate *death*. Pithing results in the physical destruction of the brain and upper regions of the spinal cord, through the insertion of a rod or cane through the bolt hole.
   b) **Requirements for effective use**
      i) Pithing cane or rod is required.
      ii) An access to the head of the animal and to the brain through the skull is required.
      iii) *Animals* should be monitored continuously until *death* to ensure the absence of brain stem reflexes.
   c) **Advantages**
      The technique is effective in producing immediate *death*.
   d) **Disadvantages**
      i) A delayed and/or ineffective pithing due to convulsions may occur.
      ii) The working area is contaminated with body fluids, which increases biosecurity risks.
2. **Bleeding**

   a) **Introduction**

   Bleeding is a method of *killing animals* through the severance of the major blood vessels in the neck or chest that results in a rapid fall in blood pressure, leading to cerebral ischaemia and *death*.

   b) **Requirements for effective use**

      i) A sharp knife is required.

      ii) An access to the neck or chest of the *animal* is required.

      iii) *Animals* should be monitored continuously until *death* to ensure the absence of brain stem reflexes.

   c) **Advantages**

   The technique is effective in producing *death* after an effective *stunning* method which does not permit pithing.

   d) **Disadvantages**

      i) A delayed and/or ineffective bleeding due to convulsions may occur.

      ii) The working area is contaminated with body fluids, which increases biosecurity risks.

   

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The only preclusion against the use of this method for neonates is the design of the stunning tongs that may not facilitate their application across such a small-sized head/body.
Preamble: The scope of these recommendations is to deal with stray and feral dogs, which pose serious human health, animal health and welfare problems and have a socio-economic, environmental, political and religious impact in many countries. Whilst acknowledging human health is a priority including the prevention of zoonotic diseases notably rabies (Chapter 8.10.), dog population management is an integral part of rabies control programmes, the OIE recognises the importance of controlling dog populations without causing unnecessary or avoidable animal suffering. Veterinary Services should play a lead role in preventing zoonotic diseases and ensuring animal welfare and should be involved in dog population control, coordinating their activities with other competent public institutions and/or agencies.

Article 7.7.1.

Guiding principles

The following recommendations are based on those laid down in Chapter 7.1. Some additional principles are relevant to these recommendations:

1. The promotion of responsible dog ownership can significantly reduce the numbers of stray dogs and the incidence of zoonotic diseases.

2. Because dog ecology is linked with human activities, control of dog populations has to be accompanied by changes in human behaviour to be effective.

Article 7.7.2.

Definitions

Carrying capacity: means the upper limit of the dog population density that could be supported by the habitat based on the availability of resources (food, water, shelter), and human acceptance.

Dog population control programme: means a programme with the aim of reducing a stray dog population to a particular level and/or maintaining it at that level and/or managing it in order to meet a predetermined objective (see Article 7.7.3).

Euthanasia: means the act of inducing death in a humane manner.

Owned dog: means a dog with a person that claims responsibility.

Person: this can include more than one individual, and could comprise family/household members or an organisation.

Responsible dog ownership: means the situation whereby a person (as defined above) accepts and commits to perform various duties according to the legislation in place and focused on the satisfaction of the behavioural, environmental and physical needs of a dog and to the prevention of risks (aggression, disease transmission or injuries) that the dog may pose to the community, other animals or the environment.

Stray dog: means any dog not under direct control by a person or not prevented from roaming. Types of stray dog:

1. free-roaming owned dog not under direct control or restriction at a particular time;
2. free-roaming dog with no owner;

3. feral dog: domestic dog that has reverted to the wild state and is no longer directly dependent upon humans for successful reproduction.

Article 7.7.3.

Dog population control programme objectives

The objectives of a programme to control the dog population may include the following:

1. improve health and welfare of owned and stray dog population;
2. reduce numbers of stray dogs to an acceptable level;
3. promote responsible ownership;
4. assist in the creation and maintenance of a rabies immune or rabies free dog population;
5. reduce the risk of zoonotic diseases other than rabies;
6. manage other risks to human health (e.g. parasites);
7. prevent harm to the environment and other animals;
8. prevent illegal trade and trafficking.

Article 7.7.4.

Responsibilities and competencies

1. Veterinary Authority

The Veterinary Authority is responsible for the implementation of animal health and animal welfare legislation, in coordination with other competent government agencies and institutions. Control of endemic zoonotic diseases such as rabies and parasitic infections (e.g. Echinococcus spp.) would require technical advice from the Veterinary Authority, as animal health and some aspects of public health are within this Authority’s competence but organising and/or supervising dog control schemes can be the responsibility of non-governmental organisations and governmental agencies other than the Veterinary Authority.

2. Other government agencies

The responsibilities of other government agencies will depend on the risk being managed and the objective/nature of the dog population control measures employed.

The ministry or other agency responsible for public health would normally play a leadership role and may have legislative authority in dealing with zoonotic diseases. Control of stray dogs with regard to other human health risks (e.g. stray dogs on roads; dog attacks within communities) may fall within the responsibility of the public health agency but is more likely to be the responsibility of the local government authorities or other agencies for public safety/security operating at the state/provincial or municipal level.
Environment protection agencies may take responsibility for control problems associated with stray dogs when they present a hazard to the environment (e.g. control of feral dogs in national parks; prevention of dog attacks on wildlife or transmission of diseases to wildlife) or where a lack of environmental controls is giving rise to stray dog populations that threaten human health or access to amenities. For example, environmental protection agencies may regulate and enforce measures to prevent dogs from accessing waste or human sewage.

3. Private sector veterinarians

The private sector veterinarian is responsible for providing advice to dog owners or handlers consulting the veterinarian for advice or treatment of a dog. The private sector veterinarian can play an important role in disease surveillance because he/she might be the first to see a dog suffering from a notifiable disease such as rabies. It is necessary that the private sector veterinarian follow the procedure established by the Veterinary Authority for responding to and reporting a suspected rabies case or a dog that is suffering from any other notifiable disease. Private sector veterinarians also play an important role (often in liaison with the police and/or local authorities) in dealing with cases of neglect that can lead to problems with stray and mismanaged dogs.

The private veterinarian has competence and will normally be involved in dog health programmes and population control measures, including health testing, vaccination, identification, kennelling during the absence of the owner, sterilisation and euthanasia. Two-way communication between the private sector veterinarian and Veterinary Authority, often via the medium of a veterinary professional organisation, is very important and the Veterinary Authority is responsible for setting up appropriate mechanisms for this action.

4. Non governmental organisations

Non governmental organisations (NGOs) are potentially important partners of the Veterinary Services in contributing to public awareness and understanding and helping to obtain resources to contribute in a practical way to the design and successful implementation of dog control programmes. NGOs can supply local knowledge on dog populations and features of ownership, as well as expertise in handling and kennelling dogs and the implementation of sterilisation programmes. NGOs can also contribute, together with veterinarians and the authorities in educating the public in responsible dog ownership.

5. Local government authorities

Local government authorities are responsible for many services and programmes that relate to health, safety and public good within their jurisdiction. In many countries the legislative framework gives authority to local government agencies in regard to aspects of public health, environmental health/hygiene and inspection/compliance activities.

In many countries local government agencies are responsible for the development and enforcement of legislation relating to dog ownership (e.g. registration, microchipping, vaccination, leash laws, abandonment), the control of stray dogs (e.g. dog catching and shelters) and the alleviation of the problems stray dogs cause in their jurisdiction. This would normally be done with advice from a higher level (national or state/provincial) authority with specialised expertise in regard to public health and animal health. Collaboration with the private sector veterinarians (e.g. in programmes to sterilise and vaccinate stray dogs) and NGOs is a common feature of dog control programmes. Regardless of the legislative basis, it is essential to have the co-operation of local government authorities in the control of stray dogs.
Annex XIII (contd)

6. **Dog owners**

When a person takes on the ownership of a dog, there should be an immediate acceptance of responsibility for that dog, and for any offspring it may produce, for the duration of its life or until a subsequent owner is found. The owner should ensure that the welfare of the dog, including behavioural needs, are respected and the dog is protected, as far as possible, from infectious diseases (e.g. through vaccination and parasite control) and from unwanted reproduction (e.g. through contraception or sterilisation). Owners should ensure that the dog’s ownership is clearly identified (preferably with permanent identification such as a tattoo or microchip) and, where required by legislation, registered on a centralised database. All reasonable steps should be taken to ensure that the dog does not roam out of control in a manner that would pose a problem to the community and/or the environment.

**Article 7.7.5.**

In the development of a dog population control programme it is recommended that the authorities establish an advisory group, which should include veterinarians, experts in dog ecology, dog behaviour and zoonotic diseases, and representatives of relevant stakeholders (local authorities, human health services/authorities, environmental control services/authorities, NGOs and the public). The main purpose of this advisory group would be to analyse and quantify the problem, identify the causes, obtain public opinion on dogs and propose the most effective approaches to use in the short and long term.

Important considerations are as follows:

1. **Identifying the sources of stray dogs**
   a) owned dogs that roam freely;
   b) dogs that have been abandoned by their owner, including puppies resulting from uncontrolled breeding of owned dogs;
   c) unowned dogs that reproduce successfully.

2. **Estimating the existing number, distribution and ecology**

   Practical tools that are available include registers of dogs, population estimates, and surveys of dogs, owners, dog shelters and veterinarians. The important factors relevant to the dog carrying capacity of the environment include food, shelter, water and human attitudes and behaviour.

   A methodology could be established to make an estimate of the total dog population. An overview of appropriate methodologies may be found in Article 7.7.8. The same methodology could be used at appropriate intervals to assess population trends.

3. **Regulatory framework**

   A regulatory framework that would help authorities establish successful dog control programmes could include the following key elements:

   a) registration and identification of dogs and licensing of dog breeders;
   b) vaccination against rabies and other preventive measures against zoonotic disease, as appropriate;
   c) veterinary procedures (e.g. surgical procedures);
   d) control of dog movement (national and international);
e) control of dangerous dogs;
f) regulations on the breeding and sale of dogs;
g) environmental controls (e.g. abattoirs, rubbish dumps, dead stock facilities);
h) regulations for dog shelters;
i) animal welfare obligations of owners and authorities.

4. Resources available to authorities
   a) Human resources;
   b) financial resources;
   c) technical tools;
   d) infrastructure;
   e) cooperative activities;
   f) public-private-NGO partnerships;
   g) central-state or province-local partnerships.

Control measures

The following control measures could be implemented according to the national context and local circumstances. Measures may be used in combination. Euthanasia of dogs, used alone, is not an effective control measure. If used, it should be done humanely (see point 11 of Article 7.7.6.) and in combination with other measures to achieve effective long term control. It is also important that authorities gain an understanding of people’s attitudes towards dog ownership so that they can develop a cooperative approach to the control of dog populations.

1. Education and legislation for responsible ownership

   Encouraging dog owners to be more responsible will reduce the number of dogs allowed to roam, improve the health and welfare of dogs, and minimise the risk that dogs pose to the community. The promotion of responsible dog ownership through legislation and education is a necessary part of a dog population control programme. Collaboration with local government authorities, animal welfare NGOs, kennel clubs, private veterinarians and veterinary organisations will assist Veterinary Authorities in establishing and maintaining programmes.

   Education on responsible dog ownership (for the currently owned dog and any offspring it produces) should address the following elements:

   a) the importance of proper selection for behaviour and care to ensure the welfare of the dog and any offspring; the latter may include preparing the dog to cope with its environment through attention to socialisation and training;
   b) registration and identification of dogs (see point 2 of Article 7.7.6.);
   c) disease prevention, in particular zoonotic disease, e.g. through regular vaccination in rabies endemic areas;
Annex XIII (contd)

d) preventing negative impacts of dogs on the community, via pollution (e.g. faeces and noise), risks to human health through biting or traffic accidents and risks to other dogs, wildlife, livestock and other companion animal species;

e) control of dog reproduction.

In order to achieve a shift towards responsible ownership, a combination of legislation, public awareness, education, and promotion of these elements will be required. It may also be necessary to improve access to resources supporting responsible ownership, such as veterinary care, identification and registration services and measures for control of zoonotic diseases.

2. Registration and identification of dogs (licensing)

A core component of dog population control by the Competent Authorities is the registration and identification of owned dogs. This may include granting licences to owners and breeders. Registration and identification may be emphasized as part of responsible dog ownership and are often linked to animal health programmes, for example, mandatory rabies vaccination and traceability.

Registration of animals in a centralised database can be used to support the enforcement of legislation and the reuniting of lost animals with owners. The control of dog reproduction by sterilisation can be encouraged through financial incentives presented by differential licensing fees.

3. Reproductive control

Controlling reproduction in dogs prevents the birth of unwanted puppies and can help address the balance between demand for dogs and the size of the population. It is advisable to focus efforts to control reproduction on those individuals or groups in the dog population identified as the most productive and the most likely to be the sources of unwanted and stray dogs, to ensure best use of resources. Methods of controlling reproduction will require direct veterinary input to individual animals. Involvement of both private and public veterinary sectors may be required to meet demand for services. Subsidisation of sterilisation programmes by government or other organisations may be considered to encourage uptake. The control of reproduction is essentially the responsibility of owners and can be incorporated into education on responsible ownership (see point 1 of Article 7.7.6.). Methods for controlling reproduction in dogs include:

a) surgical sterilisation;

b) chemical sterilisation;

c) chemical contraception;

d) separation of female dogs during oestrus from unsterilised males.

Surgical sterilisation should be carried out by a veterinarian and include appropriate anaesthesia and pain management.

Any chemicals or drugs used in controlling reproduction should be shown to have appropriate safety, quality and efficacy for the function required and used according to the manufacturer’s and Competent Authority’s regulations. In the case of chemical sterilants and contraceptives, research and field trials may need to be completed before use.
4. **Removal and handling**

The Competent Authority should collect dogs that are not under direct supervision and verify their ownership. Capture, transport, and holding of the dogs should be done humanely. The Competent Authority should develop and implement appropriate legislation and training to regulate these activities. Capture should be achieved with the minimum force required and equipment should be used that supports humane handling. Uncovered wire loops should not be used for capture.

5. **Capture and return, rehoming or release**

Competent Authorities have the responsibility to develop minimum standards for the housing (physical facilities) and care of these dogs. There should be provision for holding the dogs for a reasonable period of time to allow for reunion with the owner and, as appropriate, for rabies observation.

a) Minimum standards for housing should include the following provisions:

i) site selection: access to drainage, water and electricity are essential and environmental factors such as noise and pollution should be taken into account;

ii) kennel size, design and occupancy taking exercise into account;

iii) disease control measures including isolation and quarantine facilities.

b) Management should address:

i) adequate fresh water and nutritious food;

ii) regular hygiene and cleaning;

iii) routine inspection of the dogs;

iv) monitoring of health and provision of required veterinary treatments;

v) policies and procedures for rehoming (adoption), sterilisation and euthanasia;

vi) training of staff in safe and appropriate handling of dogs;

vii) record keeping and reporting to authorities.

Dogs that are removed from a community may be reunited with the owner or offered to new owners for rehoming. This provides an opportunity to promote responsible ownership and good animal health care (including rabies vaccination). Prior to rehoming, authorities may consider sterilisation of dogs as a population control measure. The suitability of new owners to adopt dogs should be assessed and owners matched with available animals. The effectiveness of rehoming may be limited due to the suitability and number of dogs.

Dogs that are removed from a community may in some cases be provided with health care (including rabies vaccination), sterilised, and released to their local community at or near the place of capture. This method is more likely to be accepted in the situation where the presence of stray dogs is considered to be inevitable and is well tolerated by the local community.
This method is not applicable in all situations and may be illegal in countries or regions where legislation prohibits the abandonment of dogs. Problems caused by dogs, such as noise, faecal pollution, bite injuries and traffic accidents, would not be alleviated as dogs are returned to the local community and their movements are not restricted. If the local community has owned dogs, and sterilised dogs are released, consideration should be given to the risk that this could encourage abandonment of unwanted dogs. In the situation where many dogs are owned, a population control programme that focuses on neutering and responsible ownership may be more appropriate.

It is recommended that before adopting this approach, a cost-benefit analysis is conducted. Factors such as the monetary costs, impact on culture of ownership and public safety should be assessed as well as the benefits for disease control and animal welfare as well as any societal benefits.

c) If this method is adopted, the following factors should be addressed:

i) raising awareness of the programme within the local community to ensure understanding and support;

ii) use of humane methods for catching, transporting and holding dogs;

iii) correct surgical technique, anaesthesia and analgesia, followed by post-operative care;

iv) disease control may include blanket vaccination (e.g. rabies) and treatments and testing for diseases (e.g. leishmaniasis) followed, as appropriate by treatment or euthanasia of the dog;

v) behavioural observation may be used to assess if dogs are suitable for release; if not suitable for release or rehoming, euthanasia should be considered;

vi) permanent marking (e.g. tattoo or microchip) to indicate that the animal has been sterilised; individual identification also allows for tracking of vaccination status and treatment history and identification of a level of ‘ownership’ by the organisation/authority responsible for carrying out this intervention; a visible identification (e.g. collar) may also be used to prevent unnecessary recapture;

vii) the dog should be returned to a place that is as near as possible to the place of capture;

viii) the welfare of dogs after release should be monitored and action taken if required.

Dogs that are removed from a community may be too numerous or may be unsuitable for any rehoming scheme. If euthanasia of these unwanted animals is the only option, the procedure should be conducted in accordance with the regulations of the Competent Authority (see point 11 of Article 7.7.6.).

6. Environmental controls

Steps should be taken to exclude dogs from sources of food (e.g. rubbish dumps and abattoirs, and installing animal-proof rubbish containers).

This should be linked to a reduction in the dog population by other methods, to avoid animal welfare problems.

7. Control of dog movement – international (export/import)

Chapter 8.10. provides recommendations on the international movement of dogs, with respect to provision for between rabies-free countries and countries considered to be infected with rabies.
8. Control of dog movements – within country (e.g., leash laws, roaming restrictions)

Measures for the control of dog movement in a country are generally invoked for the following reasons:

a) for rabies control when the disease is present in a country;

b) for public safety reasons;

c) for the safety of ‘owned dogs’ in an area or locality when a stray dog control programme is in place;

d) to protect wildlife and livestock.

It is necessary to have a regulatory framework and a national or local infrastructure comprising organisation, administration, staff and resources to encourage the finders of stray dogs to report to the Competent Authority.

9. Regulation of commercial dog dealers

Dog breeders and dealers should be encouraged to form or join an appropriate association. Such associations should encourage a commitment to the raising and selling of physically and psychologically healthy dogs, as unhealthy dogs may be more likely to be abandoned to become part of the stray population. They should encourage breeders and dealers to provide advice on proper care to all new owners of dogs. Regulations covering commercial dog breeders and dealers should include specific requirements for accommodation, provision of suitable food, drink and bedding, adequate exercise, veterinary care and disease control and may require breeders and dealers to allow regular inspection, including veterinary inspection.

10. Reduction in dog bite incidence

The most effective means of reducing prevalence of dog bites are education and placing responsibility on the owner. Dog owners should be educated in principles of responsible dog ownership as described in point 1 of Article 7.7.6.) Legal mechanisms that enable the Competent Authorities to impose penalties or otherwise deal with irresponsible owners are necessary. Mandatory registration and identification schemes will facilitate the effective application of such mechanisms. Young children are the group at highest risk for dog bites. Public education programmes focused on appropriate dog-directed behaviour have been demonstrated to be effective in reducing dog bite prevalence and these programmes should be encouraged. Authorities should seek advice from dog behaviour experts in developing dog safety education programmes.

11. Euthanasia

When euthanasia is practised, the general principles in the Terrestrial Code should be followed, with the emphasis on using the most practical, rapid and humane methods and ensuring operator safety. Regardless of the method used, it is important to minimise distress, anxiety and pain by ensuring that operators are appropriately trained.

Table 1 shows a summary analysis of methods for the euthanasia of dogs.
Annex XIII (contd)

Comments on methods for the euthanasia of dogs:

a) Restraint

When a dog needs to be restrained for any procedure, including euthanasia, this should always be done with full regard for operator security and animal welfare. Some euthanasia methods should be used in association with sedation or anaesthesia in order to be considered humane.

b) Special equipment

When special equipment is needed to perform euthanasia (e.g. gas chamber), the system should be designed for the purpose and regularly maintained in order to achieve operator security and animal welfare.

c) The following methods, procedures and practices are unacceptable on animal welfare grounds:

i) Chemical methods:

- Embutramide +Mebezonium +Tetracaine without sedation or by other than IV injection
- Chloral hydrate
- Nitrous oxide: may be used with other inhalants to speed the onset of anaesthesia, but alone it does not induce anaesthesia in dogs
- Ether
- Chloroform
- Cyanide
- Strychnine
- Neuromuscular blocking agents (nicotine, magnesium sulphate, potassium chloride, all curariform agents): when used alone, respiratory arrest occurs before loss of consciousness, so the dog may perceive pain
- Formalin
- Household products and solvents.

ii) Mechanical methods:

- Air embolism on conscious animal
- Burning
- Exsanguination of conscious animal
- Decompression: expansion of gas trapped in body cavities may be very painful
- Drowning
- Hypothermia, rapid freezing
- Stunning: stunning is not a euthanasia method, it should always be followed by a method which ensures death
- Kill-trapping
- Electrocution of conscious animal.

Because neonatal animals and adults with impaired breathing or low blood pressure are resistant to hypoxia, methods that depend upon achieving a hypoxic state (e.g. CO2, CO, N2, Ar) should not be used. These methods should not be used in animals aged less than 2 months, except to produce loss of consciousness and should be followed by another method to cause death. Concussion and cervical dislocation may be used in very small neonatal dogs and only in cases of emergency.
Operators should be well trained in the use of physical techniques to ensure that they are correctly and humanely carried out. The dog should be exsanguinated immediately after concussion or cervical dislocation.

d) Confirmation of death

For all methods of euthanasia used, death should be confirmed before animals are disposed of or left unattended. If an animal is not dead, another method of euthanasia should be performed.

e) Carcass disposal

Carcasses should be disposed of in a manner that complies with legislation. Attention should be paid to the risk of residues occurring in the carcass. Incineration is generally the safest way of carcass disposal.

<table>
<thead>
<tr>
<th>Euthanasia method</th>
<th>Specific method</th>
<th>Animal welfare concerns/implications</th>
<th>Key animal welfare requirements</th>
<th>Considerations relating to operator security</th>
<th>Advantages</th>
<th>Disadvantages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chemical via infection</td>
<td>Embutramide +Mebezonium +Tetracaine</td>
<td>Muscle paralysis may occur before loss of consciousness if injection given rapidly.</td>
<td>Use slow IV injection with sedation to permit slow rate of injection.</td>
<td>Correct restraint is needed. To be administered under veterinary supervision and by trained personnel.</td>
<td>Quite low cost.</td>
<td>Unavailable/unlicensed in some countries.</td>
</tr>
<tr>
<td></td>
<td>Anaesthetic agent overdose (thiopentone or propofol)</td>
<td>Underdosing may lead to recovery.</td>
<td>IV injection of a sufficient dose.</td>
<td>Correct restraint is needed. To be administered under veterinary supervision and by trained personnel.</td>
<td>Generally quick action and minimal discomfort to animal.</td>
<td>Large volume required (cost implications).</td>
</tr>
<tr>
<td></td>
<td>Barbiturates</td>
<td>Correct restraint is needed. IP is slow and may be irritant. IC injection is a painful procedure.</td>
<td>Recommend to use IV injection. When using IP injection, the solution may be diluted or local anaesthetic agent used in conjunction. IC should only be performed on unconscious animal and by skilled operator.</td>
<td>Correct restraint is needed. Administered under veterinary supervision and requires trained personnel.</td>
<td>Speed of action generally depends on the dose, concentration, route and rate of injection. Barbiturates induce euthanasia smoothly, with minimal discomfort to the animal. Barbiturates are less expensive than many other euthanasia agents.</td>
<td>These drugs persist in the carcass and may cause sedation or death in animals that consume the cadaver.</td>
</tr>
</tbody>
</table>
### Annex XIII (contd)

<table>
<thead>
<tr>
<th>Euthanasia method</th>
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<th>Disadvantages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Potassium chloride (KCl)</td>
<td>K+ is cardiotoxic and very painful if used without anaesthetic agent.</td>
<td>Only use on anaesthetised animals, IV injection.</td>
<td>Requires trained personnel.</td>
<td>Readily available without veterinary control.</td>
<td>Prior need for anaesthetic (cost and availability implications).</td>
<td></td>
</tr>
<tr>
<td>Mechanical</td>
<td>Free bullet</td>
<td>Can be inhumane if shot is inaccurate and dog is only wounded; dog may also escape.</td>
<td>Skilled operator essential.</td>
<td>Risk of injury to operators and spectators.</td>
<td>Not necessary to handle or capture dog.</td>
<td>Brain tissue may be unavailable for rabies diagnosis. Risk of injury to bystanders. Legal constraints on use of firearms.</td>
</tr>
<tr>
<td>Mechanical (contd)</td>
<td>Penetrating captive bolt followed by pithing where necessary to ensure death</td>
<td>Can be inhumane if shot is inaccurate and dog is only wounded.</td>
<td>Skilled operator essential.</td>
<td>Animal should be restrained. Skilled operator essential.</td>
<td>No risk to operator (see free bullet) unless risk of dog infected with rabies, due to potential contact with brain tissue.</td>
<td>Brain tissue may be unavailable for rabies diagnosis. Legal constraints on use of firearms. May raise aesthetic objections.</td>
</tr>
<tr>
<td>Exsanguination</td>
<td>Onset of hypovolaemia may cause dog to become anxious.</td>
<td>Only use on unconscious animal.</td>
<td>Danger to operator through use of sharp instrument.</td>
<td>Material requirements minimal.</td>
<td>Should be done on unconscious animal. Need to render animal unconscious. Aesthetically objectionable.</td>
<td></td>
</tr>
<tr>
<td>Carbon monoxide (CO)</td>
<td>Inadequate concentration of CO is not lethal and can cause suffering. Signs of distress (convulsions, vocalization and agitation) may occur.</td>
<td>Compressed CO in cylinders should be used to achieve and maintain adequate concentration, which should be monitored. Note: fumes from gasoline engines are an irritant and this source of CO is not recommended.</td>
<td>Very hazardous for operator - gas is odourless and causes toxicity at both acute high levels and chronic low levels.</td>
<td>Dog dies quite rapidly if concentration of 4 to 6% used.</td>
<td>No odour (therefore no aversive effect). Gas is not flammable or explosive except at concentration greater than 10%.</td>
<td></td>
</tr>
<tr>
<td>Gaseous</td>
<td>Carbon dioxide (CO2)</td>
<td>Gas is aversive. Inadequate concentration of CO2 is not lethal and can cause suffering. CO2 is heavier than air, so when incomplete filling of the chamber occurs, dogs may raise their head and avoid exposure. Few studies on adequate concentration and animal welfare.</td>
<td>Compressed CO2 gas chamber is the only acceptable method because the concentration can be monitored and regulated.</td>
<td>Minimal hazard to operator when properly designed equipment used.</td>
<td>Dog is not flammable or explosive and causes quite rapid anaesthesia when correct concentrations used. Low cost. Readily available as compressed gas.</td>
<td>Unconsciousness can occur in minutes, but death may take some time. Likelihood of suffering before unconsciousness.</td>
</tr>
<tr>
<td>Euthanasia method</td>
<td>Specific method</td>
<td>Animal welfare concerns/ implications</td>
<td>Key animal welfare requirements</td>
<td>Considerations relating to operator security</td>
<td>Advantages</td>
<td>Disadvantages</td>
</tr>
<tr>
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</tr>
<tr>
<td>Gaseous (contd)</td>
<td>Inert gas (nitrogen, N₂ argon, Ar)</td>
<td>Loss of consciousness is preceded by hypoxemia and ventilatory stimulation, which may be distressing to the dog. Re-establishing a low concentration of O₂ (i.e. greater than or equal to 6%) in the chamber before death will allow immediate recovery.</td>
<td>Concentration above 98% should be achieved rapidly and maintained. Properly designed equipment should be used.</td>
<td>Minimal hazard to operator when properly designed equipment used.</td>
<td>Gas is not flammable or explosive and is odourless. Readily available as compressed gas.</td>
<td>High cost. Little data on animal welfare implications in dogs.</td>
</tr>
<tr>
<td></td>
<td>Anaesthetic gas overdose (halothane or enflurane)</td>
<td>Animal may struggle and become anxious during induction. Vapours may be irritating and can induce excitement.</td>
<td>Supplementation with air or O₂ required to avoid hypoxemia during induction phase.</td>
<td>Some gases may be hazardous, especially for pregnant women. General recommendation: avoid human exposure to greater than or equal to 2 ppm to avoid narcosis.</td>
<td>Gas is not flammable or explosive. Valuable for use with small animals (&lt;7 kgs) and animals that are already anaesthetised with gas.</td>
<td>High cost. Anaesthetic and euthanasia properties of the gas used should be known. Isoflurane has a pungent odour. Methoxyflurane’s action is slow and dog may become agitated.</td>
</tr>
<tr>
<td>Electrical</td>
<td>Electrocution</td>
<td>Cardiac fibrillation occurs before onset of unconsciousness, causing severe pain if dog is conscious. Pain can also be caused by violent extension of the limbs, head and neck. Method may not be effective if insufficient current applied.</td>
<td>Only use on unconscious dog. Dogs should be unconscious before being electrocuted. This can be accomplished by electrical stunning (current through the brain to produce an instantaneous stun) or anaesthesia. Electrodes should span the brain in order that the current passed through the brain in order to achieve an effective stun. Death would result from current passed through the heart of an unconscious animal. Proper equipment and trained operator is essential.</td>
<td>May be hazardous for operator, who should use protective equipment (boots and gloves).</td>
<td>Low cost.</td>
<td>Need to render animal unconscious. Inhumane if performed on conscious dog. May raise aesthetic objections.</td>
</tr>
</tbody>
</table>

**KEY to abbreviations used in Table 1:**

IV: intravenous
IP: intraperitoneal
IC: intracardiac
Annex XIII (contd)

Article 7.7.7.

Monitoring and evaluation of dog population control programmes

1. Monitoring and evaluation allows for comparison of important indicators against the baselines measured during initial assessment (see Article 7.7.5.). The three main reasons for carrying out monitoring and evaluation are:
   a. to help improve performance, by highlighting both problems and successful elements of interventions;
   b. for accountability, to demonstrate that the programme is achieving its aims;
   c. assuming methods are standardised, to compare the success of strategies used in different locations and situations.

2. Monitoring is a continuous process that aims to check the programme progress against targets and allows for regular adjustments. Evaluation is a periodic assessment, usually carried out at particular milestones to check the programme is having the desired and stated impact. These procedures involve the measurement of ‘indicators’ that are chosen because they reflect important components of the programme at different stages. Selection of suitable indicators requires clear planning of what the programme is aiming to achieve, the best selection of indicators will be one that reflects the interest of all relevant stakeholders. Standardised methodology will facilitate comparison of data from subsequent evaluations and performance between different projects. Indicators can be direct measurements of an area targeted to change (e.g. population of free roaming dogs on public property) or indirect measures that reflect change in a targeted area.

3. Elements that should generally be monitored and evaluated include:
   a. dog population size, separated into sub-populations according to ownership and restriction of movement (i.e. roaming unrestricted or restricted by an owner);
   b. dog welfare, in the target population (e.g. body condition score, skin conditions and injuries or lameness) and as a result of the programme (if interventions involve direct handling of dogs, the welfare of the dogs as result of this handling should be monitored);
   c. prevalence of zoonotic diseases, such as rabies, in both the animal and human population;
   d. responsible animal ownership, including measures of attitudes and understanding of responsible ownership and evidence that this is translating into responsible behaviour.

4. There are many sources of information for monitoring and evaluation purposes, including:
   a. feedback from the local community (e.g. through the use of structured questionnaires, focus groups or ‘open format’ consultation processes);
   b. records and opinions obtained from relevant professionals (e.g. veterinarians, medical doctors, law enforcement agencies, educators);
   c. animal based measurements (e.g. direct observation surveys of population size and welfare status).
5. The output of activities against budget should be carefully recorded in order to evaluate the effort (or cost) against the outcomes and impact (or benefit) that are reflected in the results of monitoring and evaluation.

Article 7.7.8.

An overview of appropriate methods for estimating the size of dog populations

Population estimates are necessary for making realistic plans for dog population management and zoonosis control, and for monitoring the success of such interventions. However, for designing effective management plans, data on population sizes alone are insufficient. Additional information is required, such as degrees of supervision of owned dogs, the origin of ownerless dogs, accessibility, etc.

The term ‘owned’ may be restricted to a dog that is registered with licensing authorities, or it may be expanded to unregistered animals that are somewhat supervised and receive shelter and some form of care in individual households. Owned dogs may be well supervised and restrained at all times, or they may be left without control for various time periods and activities. Dogs without owners that claim responsibility may still be accepted or tolerated in the neighbourhood, and individuals may provide food and protection. Such animals are sometimes called ‘community owned dogs’ or ‘neighbourhood dogs’. For an observer it is frequently impossible to decide if a free roaming dog belongs to someone or not.

The choice of methods for assessing the size of a dog population depends on the ratio of owned versus ownerless dogs, which may not always be easy to judge. For populations with a large proportion of owned dogs it may be sufficient to consult dog registration records or to conduct household surveys. These surveys should establish the number of owned dogs and the dog to human ratio in the area. In addition, questions on dog reproduction and demographics, care provided, zoonosis prevention, dog bite incidence, etc. may be asked. Sample questionnaires can be found in the “Guidelines for Dog Population Management” (WHO/WSPA 1990). Standard polling principles should be applied.

If the proportion of ownerless dogs is high or difficult to assess, then one should resort to more experimental approaches. Methods borrowed from wildlife biology can be applied. These methods are described WHO/WSPA’s “Guidelines for Dog Population Management” (1990), and in more detail in numerous professional publications and handbooks, such as Bookhout (1994) and Sutherland (2006).

Being generally diurnal and tolerant to human proximity, dogs lend themselves to direct observation and the application of mark-recapture techniques. Nevertheless, a number of caveats and limitations have to be taken into account. Firstly, the risk of zoonotic disease transmission is increased through close physical contact. Also, the methods are relatively labour intensive, they require some understanding of statistics and population biology, and most importantly, they are difficult to apply to very large areas. One should take into account that dog distribution is non-random, that their populations are not static, and that individual dogs are fairly mobile.

Counting of dogs visible in a defined area is the simplest approach to getting information on population size. One has to take into account that the visibility of dogs depends on the physical environment, but also on dog and human activity patterns. The visibility of animals changes with the time of the day and with seasons as a function of food availability, shelter (shade), disturbance, etc. Repeated standardized counting of dogs visible within defined geographical localities (e.g. wards) and specific times will provide indications of population trends. Direct counting is most reliable if it is applied to small and relatively confined dog populations, e.g. in villages, where it might be possible to recognize individual dogs based on their physical appearance.

Methods using mark-recapture procedures are often considered more reliable. However, they also produce trustworthy results only when a number of preconditions are met. Mortality, emigration and recruitment into the population should be minimal during the census period. One may be able to incorporate corrective factors into the calculations.
Annex XIII (contd)

It is therefore important that the recommended census procedures are applied at times of low dispersal and that one selects study plots of shape and size that minimize the effect of dog movements in and out of the observation area. Census surveys should be completed within a few days to a maximum of two weeks in order to reduce demographic changes. In addition, all individuals in the population should have an equal chance of being counted. This is a highly improbable condition for dogs, whose visibility depends on ownership status and degrees of supervision. It is therefore recommended that the investigator determines what fraction of the total population he/she might cover with an observational method and how much this part overlaps with the owned dog segment that he/she assesses with household surveys.

There are essentially two ways to obtain a population estimate if it is possible, in a defined area and within a few days, to tag a large number of dogs with a visible mark, e.g. a distinctive collar or a paint smudge. The first method requires that the capture (marking) effort remains reasonably constant for the whole length of the study. By plotting the daily number of dogs marked against the accumulated total of marked dogs for each day one can extrapolate the value representing the total number of dogs in the area. More commonly used in wildlife studies are mark recapture methods (Peterson Jackson, Lincoln indices). Dogs are marked (tagged) and released back into the population. The population is subsequently sampled by direct observation. The number of marked and unmarked dogs is recorded. One multiplies the number of dogs that were initially marked and released by the number of subsequently observed dogs divided by the number of dogs seen as marked during the re-observation to obtain a total population estimate. Examples for the two methods are given in WHO/WSPA’s “Guidelines for Dog Population Management” (1990).

Since the dog populations of entire countries, states, provinces or even cities are much too large for complete assessment, it is necessary to apply the methods summarized above to sample areas. These should be selected (using common sense) so that results can be extrapolated to larger areas.


**Preamble:** The purpose of this chapter is to provide advice and assistance for OIE Members to follow when formulating regulatory requirements, or other form of oversight, for the use of live animals in research and education. A system of animal use oversight should be implemented in each country. The system will, in practice, vary from country to country and according to cultural, economic, religious and social factors. However, the OIE recommends that Members address all the essential elements identified in this chapter in formulating a regulatory framework that is appropriate to their local conditions. This framework may be delivered through a combination of national, regional and institutional jurisdictions and both public sector and private sector responsibilities should be clearly defined.

The OIE recognises the vital role played by the use of live animals in research and education. The OIE Guiding Principles for Animal Welfare state that such use makes a major contribution to the wellbeing of people and animals and emphasise the importance of the Three Rs (see Article 7.8.3.). Most scientists and members of the public agree that the animals should only be used when necessary; ethically justified (thereby avoiding unnecessary duplication of animal-based research); and when no other alternative methods, not using live animals, are available; that the minimum number of animals should be used to achieve the scientific or educational goals; and that such use of animals should cause as little pain and/or distress as possible. In addition, animal suffering is often recognised separately from pain and distress and should be considered alongside any lasting harm which is expected to be caused to animals.

The OIE emphasises the need for humane treatment of animals and that good quality science depends upon good animal welfare. It is the responsibility of all involved in the use of animals to ensure that they give due regard to these recommendations. In keeping with the overall approach to animal welfare detailed in the Guiding Principles, the OIE stresses the importance of standards based on outcomes for the animals.

The OIE recognises the significant role of veterinarians in animal-based research. Given their unique training and skills, they are essential members of a team including scientists and animal care technicians. This team approach is based on the concept that everyone involved in the use of animals has an ethical responsibility for the animals' welfare. The approach also ensures that animal use leads to high quality scientific and educational outcomes and optimum welfare for the animals used.

The OIE recommends that records on animal use should be maintained at an institutional level, as appropriate to the institution and project proposals and species used. Key events and interventions should be recorded to aid decision making and promote good science and welfare. A summary of these records may be gathered on a national basis and be published to provide a degree of public transparency, without compromising personnel or animal safety, or releasing proprietary information.

**Article 7.8.1.**

**Definitions**

**Biocontainment:** means the system and procedures designed to prevent the accidental release of biological material including allergens.

**Bioexclusion:** means the prevention of the unintentional transfer of adventitious organisms with subsequent infection of animals, resulting in adverse effects on their health or suitability for research.

**Biosecurity:** means a continuous process of risk assessment and risk management designed to minimise or eliminate microbiological infection with adventitious organisms that can cause clinical disease in the infected animals or humans, or make animals unsuitable for biomedical research.
Annex XIII (contd)

**Cloned animal:** means a genetic copy of another living or dead animal produced by somatic cell nuclear transfer or other reproductive technology.

**Distress:** means the state of an animal, that has been unable to adapt to stressors, and that manifests as abnormal physiological or behavioural responses. It can be acute or chronic and may result in pathological conditions.

**Endangered species:** means a population of organisms which is at risk of becoming extinct because it is either few in numbers, or threatened by changing environmental or predation parameters.

**Environmental enrichment:** means increasing the complexity (e.g. with toys, cage furniture, foraging opportunities, social housing, etc.) in a captive animal's environment to foster the expression of non-injurious species-typical behaviours and reduce the expression of maladaptive behaviours, as well as provide cognitive stimulation.

**Ethical review:** means consideration of the validity and justification for using animals including: an assessment and weighing of the potential harms for animals and likely benefits of the use and how these balance (see harm-benefit analysis below); and consideration of experimental design; implementation of the Three Rs; animal husbandry and care and other related issues such as personnel training. Ethical judgements are influenced by prevailing societal attitudes.

**Euthanasia:** means the act of inducing death using a method that causes a rapid and irreversible loss of consciousness with minimum pain and distress to the animal.

**Harm-benefit analysis:** means the process of weighing the likely adverse effects (harms) to the animals against the benefits likely to accrue as a result of the proposed project.

**Humane endpoint:** means the point in time at which an experimental animal's pain and/or distress is avoided, terminated, minimised or reduced, by taking actions such as giving treatment to relieve pain and/or distress, terminating a painful procedure, removing the animal from the study, or humanely killing the animal.

**Operant conditioning:** means the association that an animal makes between a particular response (such as pressing a bar) and a particular reinforcement that may be positive (for example, a food reward) or negative (e.g. a mild electric shock). As a result of this association, the occurrence of a specific behaviour of the animal can be modified (e.g. increased or decreased in frequency or intensity).

**Pain:** means an unpleasant sensory and emotional experience associated with actual or potential tissue damage. It may elicit protective actions, result in learned avoidance and distress and may modify species-specific traits of behaviour, including social behaviour.

**Project proposal (sometimes called protocol):** means a written description of a study or experiment, programme of work, or other activities that includes the goals of the work, characterises the use of the animals, and includes ethical considerations.

**Suffering:** means an unpleasant, undesired state of being which is the outcome of the impact on an animal of a variety of noxious stimuli and/or the absence of important positive stimuli. It is the opposite of good welfare.
Article 7.8.2.

Scope

This chapter applies to animals as defined in the Terrestrial Code (excluding bees) bred, supplied and/or used in research (including testing) and higher education. Animals to be used for production of biologicals and/or humanely killed for harvesting their cells, tissues and organs for scientific purposes are also covered. Members should consider both the species and the developmental stage of the animal when implementing these standards.

Article 7.8.3.

The Three Rs

The internationally accepted tenet, the ‘Three Rs’, comprises the following alternatives:

1. replacement refers to the use of methods utilizing cells, tissues or organs of animals (relative replacement), as well as those that do not require the use of animals to achieve the scientific aims (absolute replacement);

2. reduction refers to the use of methods that enable researchers to obtain comparable levels of information from fewer animals or to obtain more information from the same number of animals;

3. refinement refers to the use of methods that prevent, alleviate or minimise pain, suffering, distress or lasting harm and/or enhance welfare for the animals used. Refinement includes the appropriate selection of relevant species with a lesser degree of structural and functional complexity in their nervous systems and a lesser apparent capacity for experiences that derive from this complexity. Opportunities for refinement should be considered and implemented throughout the lifetime of the animal and include, for example, housing and transportation as well as procedures and euthanasia.

Article 7.8.4.

The oversight framework

The role of a Competent Authority is to implement a system (governmental or other) for verification of compliance by institutions. This usually involves a system of authorisation (such as licensing or registering of institutions, scientists, and/or projects) and compliance which may be assessed at the institutional, regional and/or national level.

The oversight framework encompasses both ethical review of animal use and considerations related to animal care and welfare. This may be accomplished by a single body or distributed across different groups. Different systems of oversight may involve animal welfare officers, regional, national or local committees or bodies. An institution may utilise a local committee (often referred to as Animal Care and Use Committee, Animal Ethics Committee, Animal Welfare Body or Animal Care Committee) to deliver some or all of this oversight framework. It is important that the local committee reports to senior management within the institution to ensure it has appropriate authority, resources and support. Such a committee should undertake periodic review of its own policies, procedures and performance.

Ethical review of animal use may be undertaken by regional, national or local ethical review bodies or committees. Consideration should be given on how to ensure impartiality and independence from all those serving on the committees.

In providing this oversight and ensuring the implementation of the Three Rs, the following expertise should be included as a minimum:
Annex XIII (contd)

a) one scientist with experience in animal research, whose role is to ensure that protocols are designed and implemented in accordance with sound science;

b) one veterinarian, with the necessary expertise to work with research animals, whose specific role is to provide advice on the care, use and welfare of such animals;

c) one public member to represent general community interests who is independent of the science and care of the animals and is not involved in the use of animals in research.

Additional expertise may be sought from the animal care staff, as these professional and technical staff are centrally involved in ensuring the welfare of animals used. Other participants, especially in relation to ethical review, may include statisticians, information scientists and ethicists and biosafety specialists, as appropriate to the studies conducted. It may be appropriate, in teaching institutions, to involve student representation.

Oversight responsibilities include three key elements:

1. Project proposal review

The purpose of the project proposal is to enable assessment of the quality of, and justification for, the study, work or activity.

Project proposals, or significant amendments to these, should be reviewed and approved prior to commencement of the work. The proposal should identify the person with primarily responsibility for the project and should include a description of the following elements, where relevant:

a) the scientific or educational aims, including consideration of the relevance of the experiment to human or animal health or welfare, the environment, or the advancement of biological knowledge;

b) an informative, non-technical (lay) summary may enhance understanding of the project and facilitate the ethical review of the proposal by allowing full and equitable participation of members of the oversight body or committees who may be dealing with matters outside their specific field. Subject to safeguarding confidential information, such summaries may be made publicly available;

c) the experimental design, including justification for choice of species, source and number of animals, including any proposed reuse;

d) the experimental procedures;

e) methods of handling and restraint and consideration of refinements such as animal training and operant conditioning;

f) the methods to avoid or minimise pain, discomfort, distress, suffering or lasting impairment of physical or physiological function, including the use of anaesthesia and/or analgesia and other means to limit discomfort such as warmth, soft bedding and assisted feeding;

g) application of humane endpoints and the final disposition of animals, including methods of euthanasia;

h) consideration of the general health, husbandry and care of the species proposed to be used, including environmental enrichment and any special housing requirements;

i) ethical considerations such as the application of the Three Rs and a harm/benefit analysis; the benefits should be maximised and the harms, in terms of pain and distress, should be minimized;

j) an indication of any special health and safety risks; and
k) resources/infrastructure necessary to support the proposed work (e.g. facilities, equipment, staff trained and found competent to perform the procedures described in the proposed project).

The oversight body has a critical responsibility in determining the acceptability of project proposals, taking account of the animal welfare implications, the advancement of knowledge and scientific merit, as well as the societal benefits, in a risk-based assessment of each project using live animals.

Following approval of a project proposal, consideration should be given to implementing an independent (of those managing the projects) oversight method to ensure that animal activities conform with those described in the approved project proposal. This process is often referred to as post approval monitoring. Such monitoring may be achieved through animal observations made during the conduct of routine husbandry and experimental procedures; observations made by the veterinary staff during their rounds; or by inspections by the oversight body, which may be the local committee, animal welfare officer, compliance/quality assurance officer or government inspector.

2. **Facility inspection**

   There should be regular inspections of the facilities, at least annually. These inspections should include the following elements:
   
   a) the animals and their records, including cage labels and other methods of animal identification;
   
   b) husbandry practices;
   
   c) maintenance, cleanliness and security of the facility;
   
   d) type and condition of caging and other equipment;
   
   e) environmental conditions of the animals at the cage and room level;
   
   f) procedure areas such as surgery; necropsy and animal research laboratories;
   
   g) support areas such as washing equipment; animal feed, bedding and drug storage locations;
   
   h) occupational health and safety concerns.

   Principles of risk management should be followed when determining the frequency and nature of inspections.

3. **Ethical evaluation**

   The ethical evaluation reflects the policies and practices of the institution in complying with regulations and relevant guidance. It should include consideration of the functioning of the local committee; training and competency of staff; veterinary care; husbandry and operational conditions, including emergency plans; sourcing and final disposition of animals; and occupational health and safety. The programme should be reviewed regularly. A requirement for the components of such a programme should be included in relevant regulations to empower the Competent Authority to take appropriate action to ensure compliance.

   Article 7.8.5.

**Assurance of training and competency**

An essential component of the animal care and use programme is the assurance that the personnel working with the animals are appropriately trained and competent to work with the species used and the procedures to be performed, including ethical considerations. A system (institutional, regional or national) to assure competency should be in place, which includes supervision during the training period until competence has been demonstrated. Continuing professional and paraprofessional educational opportunities should be made available to relevant staff. Senior management, given their overarching responsibility for the animal care and use programme, should be knowledgeable about issues related to the competence of staff.
Annex XIII (contd)

1. **Scientific staff**

Researchers using animals have a direct ethical and legal responsibility for all matters relating to the welfare of the animals in their care. Due to the specialised nature of animal research, focused training should be undertaken to supplement educational and experiential backgrounds of scientists (including visiting scientists) before initiating a study. Focused training may include such topics as the national and/or local regulatory framework and institutional policies. The laboratory animal veterinarian is often a resource for this and other training. Scientific staff should have demonstrated competency in procedures related to their research (e.g. surgery, anaesthesia, sampling and administration, etc.).

2. **Veterinarians**

It is important that veterinarians working in an animal research environment have veterinary medical knowledge and experience in the species used, including the normal behaviour of the species, and they should understand research methodology. Relevant approvals issued by the veterinary statutory body and appropriate national or regional schemes (where these exist) should be adopted as the reference for veterinary training.

3. **Animal care staff**

Animal care staff should receive training that is consistent with the scope of their work responsibilities and have demonstrated competency in the performance of these tasks.

4. **Students**

Students should learn scientific and ethical principles using non-animal methods (videos, computer models, etc.) when such methods can effectively reduce or replace the use of live animals and still meet learning objectives. Wherever it is necessary for students to participate in classroom or research activities involving live animals, they should receive appropriate supervision in the use of animals until such time that they have demonstrated competency in the related procedure(s).

5. **Members of the local oversight committee or others involved with oversight**

Continuing education about the use of animals in research and education, including associated ethics, regulatory requirements and their institutional responsibility, should be provided.

Occupational health and safety training for research animal related risks should be provided as part of the assurance of training and competency for personnel. This might include consideration of human infectious diseases which may infect research animals and thus compromise research results, as well as possible zoonoses. Personnel should understand that there are two categories of hazards, those that are intrinsic to working in an animal facility and those associated with the research. Specific training may be required for particular species, for specific procedures, and for the use of appropriate protective measures for personnel who may be exposed to animal allergens. Research materials, such as chemicals of unknown toxicity, biological agents and radiation sources, may present special hazards.

Article 7.8.6.

**Provision of veterinary care**

Adequate veterinary care includes responsibility for promoting an animal's health and welfare before, during and after research procedures and providing advice and guidance based on best practice. Veterinary care includes attention to the physical and behavioural status of the animal. The veterinarian should have authority and responsibility for making judgements concerning animal welfare. Veterinary advice and care should be available at all times.
1. **Clinical responsibilities**

Preventive medicine programmes that include vaccinations, ectoparasite and endoparasite treatments and other disease control measures should be initiated according to currently acceptable veterinary medical practices appropriate to the particular animal species and source. Disease surveillance is a major responsibility of the veterinarian and should include routine monitoring of colony animals for the presence of parasitic, bacterial and viral agents that may cause overt or sub clinical diseases. The veterinarian should have the authority to use appropriate treatment or control measures, including euthanasia if indicated, and access to appropriate resources, following diagnosis of an animal disease or injury. Where possible, the veterinarian should discuss the situation with the scientist to determine a course of action consistent with experimental goals. Controlled drugs prescribed by the veterinary staff should be managed in accordance with applicable regulations.

2. **Post-mortem examinations**

In the case of unexpected diseases or deaths, the veterinarian should provide advice based on post-mortem examination results. As part of health monitoring, a planned programme of post-mortem examinations may be considered.

3. **Veterinary medical records**

Veterinary medical records, including post-mortem records, are considered to be a key element of a programme of adequate veterinary care for animals used in research and education. Application of performance standards within the veterinary medical record programme allows the veterinarian to effectively employ professional judgment, ensuring that the animal receives the highest level of care available.

4. **Advice on zoonotic risks and notifiable diseases**

The use of some species of animals poses a significant risk of the transmission of zoonotic disease (e.g. some nonhuman primates). The veterinarian should be consulted to identify sources of animals that minimise these risks and to advice on measures that may be taken in the animal facility to minimize the risk of transmission (e.g. personal protective equipment, appropriate disinfection procedures, air pressure differentials in animal holding rooms, etc.). Animals brought into the institution may carry diseases that require notification to government officials. It is important that the veterinarian be aware of, and comply with, these requirements.

5. **Advice on surgery and postoperative care**

A programme of adequate veterinary care includes input into the review and approval process of preoperative, surgical and postoperative procedures by an appropriately qualified veterinarian. A veterinarian's inherent responsibility includes providing advice concerning preoperative procedures, aseptic surgical techniques, the competence of staff to perform surgery and the provision of postoperative care. Veterinary oversight should include the detection and resolution of emerging patterns of surgical and post procedural complications.

6. **Advice on analgesia, anaesthesia and euthanasia**

Adequate veterinary care includes providing advice on the proper use of anaesthetics, analgesics, and methods of euthanasia.

7. **Advice on humane endpoints**

Humane endpoints should be established prior to commencement of a study in consultation with the veterinarian who also plays an important role in ensuring that approved humane endpoints are followed during the course of the study. It is essential that the veterinarian has the authority to ensure euthanasia or other measures are carried out as required to relieve pain and distress unless the project proposal approval specifically does not permit such intervention on the basis of the scientific purpose and the ethical evaluation.
Annex XIII (contd)

Ideal humane endpoints are those that can be used to end a study before the onset of pain and/or distress, without jeopardising the study’s objectives. In consultation with the veterinarian, humane endpoints should be described in the project proposal and, thus, established prior to commencement of the study. They should form part of the ethical review. Endpoint criteria should be easy to assess over the course of the study. Except in rare cases, death (other than euthanasia) as a planned endpoint is considered ethically unacceptable.

Article 7.8.7.

Source of animals

Animals to be used for research should be of high quality to ensure the validity of the data.

1. Animal procurement

Animals should be legally acquired. It is preferable that animals are purchased from recognised sources producing or securing high quality animals.

Purpose bred animals should be used whenever these are available and animals that are not bred for the intended use should be avoided unless there is compelling scientific justification or are the only available and suitable source. In the case of farm animals, non traditional breeds and species, and animals captured in the wild, non purpose bred animals are often used to achieve specific study goals. The use of wild caught nonhuman primates is generally discouraged.

2. Documentation

Relevant documentation related to the source of the animals, such as health and other veterinary certification, breeding records, genetic status and animal identification, should accompany the animals.

3. Animal health status

The health status of animals can have a significant impact on scientific outcomes. There also may be occupational health and safety concerns related to animal health status. Animals should have appropriate health profiles for their intended use. The health status of animals should be known before initiating research.

4. Genetically defined animals

A known genetic profile of the animals used in a study can reduce variability in the experimental data resulting from genetic drift and increase the reproducibility of the results. Genetically defined animals are used to answer specific research questions and are the product of sophisticated and controlled breeding schemes which should be validated by periodic genetic monitoring. Detailed and accurate documentation of the colony breeding records should be maintained.

5. Genetically altered (also genetically modified or genetically engineered) or cloned animals (also genetically modified animal and genetically engineered animal).

A genetically altered or cloned animal is one that has had undergone genetic modification of its nuclear or mitochondrial genomes through a deliberate human intervention, or the progeny of such an animal(s), where they have inherited the modification. If genetically altered or cloned animals are used, such use should be conducted in accordance with relevant regulatory guidance. With such animals, as well as harmful mutant lines arising from spontaneous mutations and induced mutagenesis, consideration should be given to addressing and monitoring special husbandry and welfare needs associated with abnormal phenotypes. Records should be kept of biocontainment requirements, genetic and phenotypic information, and individual identification, and be communicated by the animal provider to the recipient. Archiving and sharing of genetically altered lines is recommended to facilitate the sourcing of these customised animals.
6. **Animals captured in the wild**

If wild animals are to be used, the capture technique should be humane and give due regard to human and animal health, welfare and safety. Field studies have the potential to cause disturbance to the habitat thus adversely affecting both target and non-target species. The potential for such disturbance should be assessed and minimised. The effects of a series of stressors, such as trapping, handling, transportation, sedation, anaesthesia, marking and sampling, can be cumulative, and may produce severe, possibly fatal, consequences. An assessment of the potential sources of stress and management plans to eliminate or minimise distress should form part of the project proposal.

7. **Endangered species**

Endangered species should only be used in exceptional circumstances where there is strong scientific justification that the desired outcomes cannot be achieved using any other species.

8. **Transport, importation and exportation**

Animals should be transported under conditions that are appropriate to their physiological and behavioural needs and pathogen status, with care to ensure appropriate physical containment of the animals as well as exclusion of contaminants. The amount of time animals spend on a journey should be kept to a minimum. It is important to ensure that there is a well constructed journey plan, with key staff identified who have responsibility for the animals and that relevant documentation accompanies animals during transport to avoid unnecessary delays during the journey from the sender to the receiving institution.

9. **Risks to biosecurity**

In order to minimise the risk of contamination of animals with unwanted infectious microorganisms or parasites that may compromise the health of animals or make them unsuitable for use in research, the microbiological status of the animals should be determined and regularly assessed. Appropriate biocontainment and bioexclusion measures should be practised to maintain their health status and, if appropriate, measures taken to prevent their exposure to certain human or environmental commensals.

**Article 7.8.8.**

**Physical facility and environmental conditions**

A well-planned, well-designed, well-constructed, and properly maintained facility should include animal holding rooms as well as areas for support services such as for procedures, surgery and necropsy, cage washing and appropriate storage. An animal facility should be designed and constructed in accordance with all applicable building standards. The design and size of an animal facility depend on the scope of institutional research activities, the animals to be housed, the physical relationship to the rest of the institution, and the geographic location. For indoor housing, non-porous, non-toxic and durable materials should be used which can be easily cleaned and sanitised. Animals should normally be housed in facilities designed for that purpose. Security measures (e.g. locks, fences, cameras, etc.) should be in place to protect the animals and prevent their escape. For many species (e.g. rodents), environmental conditions should be controllable to minimise physiological changes which may be potentially confounding scientific variables and of welfare concern.

Important environmental parameters to consider include ventilation, temperature and humidity, lighting and noise:
1. **Ventilation**

   The volume and physical characteristics of the air supplied to a room and its diffusion pattern influence the ventilation of an animal’s primary enclosure and are thus important determinants of its microenvironment. Factors to consider when determining the air exchange rate include range of possible heat loads; the species, size, and number of animals involved; the type of bedding or frequency of cage changing; the room dimensions; and the efficiency of air distribution from the secondary to the primary enclosure. Control of air pressure differentials is an important tool for biocontainment and bioexclusion.

2. **Temperature and humidity**

   Environmental temperature is a physical factor which has a profound effect on the welfare of animals. Typically, animal room temperature should be monitored and controlled. The range of daily fluctuations should be appropriately limited to avoid repeated demands on the animals’ metabolic and behavioural processes to compensate for large changes in the thermal environment as well as to promote reproducible and valid scientific data. Relative humidity may also be controlled where appropriate for the species.

3. **Lighting**

   Light can affect the physiology, morphology and behaviour of various animals. In general, lighting should be diffused throughout an animal holding area and provide appropriate illumination for the welfare of the animals while facilitating good husbandry practices, adequate inspection of animals and safe working conditions for personnel. It may also be necessary to control the light/dark cycle.

4. **Noise**

   Separation of human and animal areas minimises disturbance to animal occupants of the facility. Noisy animals, such as dogs, pigs, goats and nonhuman primates, should be housed in a manner which ensures they do not adversely affect the welfare of quieter animals, such as rodents, rabbits and cats. Consideration should be given to insulating holding rooms and procedure rooms to mitigate the effects of noise sources. Many species are sensitive to high frequency sounds and thus the location of potential sources of ultrasound should be considered.

**Husbandry**

Good husbandry practices enhance the health and welfare of the animals used and contributes to the scientific validity of animal research. Animal care and accommodation should, as a minimum, demonstrably conform to relevant published animal care, accommodation and husbandry guidelines and regulations.

The housing environment and husbandry practices should take into consideration the normal behaviour of the species, including their social behaviour and age of the animal, and should minimise stress to the animal. During the conduct of husbandry procedures, personnel should be keenly aware of their potential impact on the animal’s welfare.

1. **Transportation**

   Transportation is a typically stressful experience. Therefore, every precaution should be taken to avoid unnecessary stress through inadequate ventilation, exposure to extreme temperatures, lack of feed and water, long delays, etc. Consignments of animals should be accepted into the facility without avoidable delay and, after inspection, should be transferred to clean cages or pens and be supplied with feed and water as appropriate. Social animals should be transported in established pairs or groups and maintained in these on arrival.
2. **Acclimatisation**

Newly received *animals* should be given a period for physiological and behavioural stabilisation before their use. The length of time for stabilisation will depend on the type and duration of transportation, the age and species involved, place of origin, and the intended use of the *animals*. Facilities should be available to isolate *animals* showing signs of ill health.

3. **Cages and pens**

Cages and pens should be made out of material that can be readily cleaned and decontaminated. Their design should be such that the *animals* are unlikely to injure themselves. Space allocations should be reviewed and modified as necessary to address individual housing situations and animal needs (for example, for prenatal and postnatal care, obese *animals*, and group or individual housing). Both the quantity and quality of space provided is important. Whenever it is appropriate, social *animals* should be housed in pairs or groups, rather than individually, provided that such housing is not contraindicated by the protocol in question and does not pose an undue risk to the *animals*.

4. **Enrichment**

*Animals* should be housed with a goal of maximising species appropriate behaviours and avoiding or minimising stress induced behaviours. One way to achieve this is to enrich the structural and social environment of the *animals* and to provide opportunities for physical and cognitive activity. Such provision should not compromise the health and safety of the *animals* or people, nor interfere with the scientific goals.

5. **Feeding**

Provision should be made for each *animal* to have access to feed to satisfy its physiological needs. Precautions should be taken in packing, transporting, storing and preparing feed to avoid chemical, physical and microbiological contamination, deterioration or destruction. Utensils used for feeding should be regularly cleaned and, if necessary, sterilised.

6. **Water**

Uncontaminated potable drinking water should normally be available at all times. Watering devices, such as drinking tubes and automatic watering systems, should be checked daily to ensure their proper maintenance, cleanliness, and operation.

7. **Bedding**

*Animals* should have appropriate bedding provided, with additional nesting material if appropriate to the species. Animal bedding is a controllable environmental factor that can influence experimental data and animal welfare. Bedding should be dry, absorbent, non-dusty, non-toxic and free from infectious agents, vermin or chemical contamination. Soiled bedding should be removed and replaced with fresh material as often as is necessary to keep the animals clean and dry.

8. **Hygiene**

The successful operation of a facility depends very much on good hygiene. Special care should be taken to avoid spreading infection between animals through fomites, including through personnel traffic between animal rooms. Adequate routines and facilities for the cleaning, washing, decontamination and, when necessary, sterilisation of cages, cage accessories and other equipment should be established. A very high standard of cleanliness and organisation should also be maintained throughout the facility.
Annex XIII (contd)

9. Identification

Animal identification is an important component of record keeping. Animals may be identified individually or by group. Where it is desirable to individually identify animals, this should be done by a reliable and the least painful method.

10. Handling

Staff dealing with animals should have a caring and respectful attitude towards the animals and be competent in handling and restraint. Familiarising animals to handling during routine husbandry and procedures reduces stress both to animals and personnel. For some species, for example dogs and non-human primates, a training programme to encourage cooperation during procedures can be beneficial to the animals, the animal care staff and the scientific programme. For certain species, social contact with humans should be a priority. However, in some cases handling should be avoided. This may be particularly the case with wild animals. Consideration should be given to setting up habituation and training programmes suitable for the animals, the procedures and length of projects.

1. Wherever the term “research” is used, it includes basic and applied research, testing and the production of biological materials; “education” includes teaching and training.
CHAPTER 8.1

ANTHRAX

Article 8.1.1.

General provisions

This chapter is intended to manage the human and animal health risks associated with the presence of Bacillus anthracis in commodities and the environment.

There is no evidence that anthrax is transmitted by animals before the onset of clinical and pathological signs. Early detection of outbreaks, quarantine of affected premises, destruction of diseased animals and fomites, and implementation of appropriate sanitary procedures at abattoirs and dairy factories will ensure the safety of products of animal origin intended for human consumption.

For the purposes of the Terrestrial Code, the incubation period for anthrax shall be 20 days.

Anthrax should be notifiable in the whole country.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

When authorising import or transit of commodities covered in the chapter, with the exception of those listed in Article 8.1.2., Veterinary Authorities should require the conditions prescribed in this chapter.

Article 8.1.2.

Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any anthrax related conditions: semen and in vivo derived cattle embryos collected and handled in accordance with Chapters 4.5., 4.6. and 4.7., as relevant.

Article 8.1.3.

Recommendations for the importation of ruminants, equines and pigs

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of anthrax on the day of shipment;

AND

2. were kept for the 20 days prior to shipment in an establishment where no case of anthrax was officially declared during that period; or

3. were vaccinated, not less than 20 days and not more than 6 months prior to shipment in accordance with the Terrestrial Manual.
Annex XIV (contd)

Article 8.1.4.

Recommendations for the importation of fresh meat and meat products destined for human consumption

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products originate from animals which:

1. have shown no sign of anthrax during ante-mortem and post-mortem inspections; and

2. were not vaccinated against anthrax using live vaccine during the 21 days prior to slaughter or a longer period depending on the manufacturer’s recommendations; and

3. come from establishments which are not placed under movement restriction on account of anthrax and in which there has been no case of anthrax during the 20 days prior to slaughter.

Article 8.1.5.

Recommendations for the importation of hides, skins and hair (from ruminants, equines and pigs)

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products originate from animals which:

1. originate from animals which:
   1. have shown no sign of anthrax during ante-mortem and post-mortem inspections; and
   2. come from establishments which are not placed under movement restriction on account of anthrax;
   OR
   2. for hair, have been treated in accordance with the recommendations in Article 8.1.11.

Article 8.1.6.

Recommendations for the importation of wool

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products originate from live animals and

1. originate from establishments where no case of anthrax has been reported since the previous shearing of all animals which, at the time of shearing, were part of a flock that was not subject to restrictions imposed for the control of anthrax;

OR

2. have been treated in accordance with the recommendations in Article 8.1.11.

Article 8.1.7.

Recommendations for the importation of milk and milk products intended for human consumption

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that:
1. the milk originates from animals showing no clinical signs of anthrax at the time of milking;

2. if the milk originates from herds or flocks that have had a case of anthrax within the previous 20 days, it has been chilled promptly and processed using a heat treatment at least equivalent to pasteurisation.

Article 8.1.8.

Recommendations for the importation of bristles (from pigs)

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products originate from animals which:

1. have shown no sign of anthrax during ante-mortem and post-mortem inspections; and

2. come from establishments which are not placed under movement restriction on account of anthrax control;

OR

3. have been processed to ensure the destruction of B. anthracis by:
   a) boiling for 60 minutes; and
   b) drying in hot air.

Article 8.1.9.

Procedures for the inactivation of B. anthracis spores in skins and trophies from wild animals

In situations in which skins and trophies from wild animals may be contaminated with B. anthracis spores, the following disinfection procedure is recommended:

1. fumigation with ethylene oxide 500 mg/L, at relative humidity 20 – 40%, at 55°C for 30 minutes; or

2. fumigation with formaldehyde 400 mg/m³ at relative humidity 30%, at >15°C for 4 hours; or

3. gamma irradiation with a dose of 40 kGy.

Article 8.1.10.

Procedures for the inactivation of B. anthracis spores in bone-meal and meat-and-bone meal

In situations where raw materials used to produce bone meal or meat-and-bone meal may be contaminated with B. anthracis spores, the following inactivation procedure should be used to inactivate any B. anthracis spores which may be present during the production of bone meal or meat and bone meal from ruminants, equines and pigs:

1. the raw material should be reduced to a maximum particle size of 50 mm before heating; and

2. the raw material should be heated under saturated steam conditions to a temperature of not less than 133°C for a minimum of 20 minutes at an absolute pressure of 3 bar. Other industrial processes demonstrating equivalent efficacy is also acceptable, subjected to moist heat at one of the following temperature and time regimes:
   a) 105 °C for at least 8 minutes; or
   b) 100°C for at least 10 minutes; or
Annex XIV (contd)

c) 95 °C for at least 25 minutes; or

d) 90°C for at least 45 minutes; or

e) an industrial process demonstrated to be of equivalent efficacy.

Article 8.1.11.

Procedures for the inactivation of *B. anthracis* spores in wool and hair

In situations in which wool or hair may be contaminated with *B. anthracis* spores, the following five-step disinfection procedures are recommended:

1. gamma irradiation with a dose of 50 kGy; or

2. a five step washing procedure:

   a) immersion in 0.25 – 0.3% soda liquor for 10 minutes at 40.5°C;

   b) immersion in soap liquor for 10 minutes at 40.5°C;

   c) immersion in 2% formaldehyde solution for 10 minutes at 40.5°C;

   d) a second immersion in 2% formaldehyde solution for 10 minutes at 40.5°C;

   e) rinsing on cold water followed by drying in hot air.

Article 8.1.12.

Procedures for the inactivation of *B. anthracis* spores in manure, dung and bedding

In situations in which manure, dung or bedding may be contaminated with *B. anthracis* spores, the following are recommended:

1. small volumes by incineration; or

2. chemothermal treatment by composting as follows:

   a) mix with one of the following at a rate of 1 – 1.5L/m³;

      i) 10% formaldehyde (approximately 30% formalin), or

      ii) 4% gluteraldehyde (pH 8.0 – 8.5);

   b) turn the material after 5 weeks;

   c) leave for a further 5 weeks.

[Note: spontaneous combustion of the composting pile is possible.]
Procedures for the inactivation of *B. anthracis* spores in liquid manure (slurry)

In situations in which liquid manure (slurry) may be contaminated with *B. anthracis* spores, disinfection with formalin (35% aqueous solution of formaldehyde) with stirring for one hour daily is recommended:

1. for slurry up to 5% dry matter, 50 kg formalin per m³ for 4 days;
2. for slurry >5% and <10% dry matter, 100 kg formalin per m³ for 4 days.

Procedures for the disinfection of surfaces in animal houses, buildings contaminated with *B. anthracis*

In situations in which surfaces in animal houses, stables, vehicles, etc. may be contaminated with *B. anthracis* spores, the following three-step approach is recommended:

1. a preliminary disinfection should be carried out using one of the following disinfectants at a rate of 1 – 1.5 L/m³ for 2 hours;
   a) 10% formaldehyde (approximately 30% formalin); or
   b) 4% glutaraldehyde (pH 8.0 – 8.5);
2. all surfaces should be washed and scrubbed using ample hot water and, when cleaned and waste water is free from dirt particles, dried;
3. a final disinfection step should be carried out using one of the following disinfectants applied at a rate of 0.4 L/m³ for 2 hours;
   a) 10% formaldehyde (approximately 30% formalin), repeated after one hour; or
   b) 4% glutaraldehyde (pH 8.0 – 8.5), repeated after one hour; or
   c) 3% hydrogen peroxide; or
   d) 1% peracetic acid, repeated after one hour.

[Note: Formaldehyde and glutaraldehyde should not be used at temperatures below 10°C. Hydrogen peroxide and peracetic acid are not suitable in the presence of blood.]

Procedures for the fumigation of rooms contaminated with *B. anthracis*

Contaminated rooms which cannot be cleared before cleaning and disinfection can be fumigated to eliminate *B. anthracis* spores. The following procedure is recommended:

1. all windows, doors and vents to the outside should be sealed with heavy adhesive tape; and
Annex XIV (contd)

2. for rooms up to 30 m³, 4 L of water containing 400 ml of concentrated formalin (37% w/v formaldehyde) in an electric kettle (with a timing switch to turn it off) should be boiled away and the room left overnight. Room temperature should be >15°C.

[Note: Formaldehyde fumigation is hazardous and proper respirators should be on hand for operator safety. The effectiveness of the fumigation process should be verified by exposing dried discs of filter paper which have been dipped in a suspension of spores of B. subtilis var globigii or B. cereus or Sterne vaccine strain of B. anthracis and placed in the room before fumigation is started. At the end of fumigation, the discs should be placed on nutrient agar plates containing 0.1% histidine and incubated overnight at 37°C. If fumigation has been effective, there will be no bacterial growth.]
General provisions

The Aujeszky's disease (AD) free or provisionally free status of a country or zone can only be determined if the following conditions are fulfilled:

1. a risk assessment has been conducted identifying all potential factors for AD occurrence and their historic perspective;
2. AD is notifiable in the whole country, and all clinical cases suggestive of AD are subjected to field and laboratory investigations;
3. an on-going awareness programme is in place to encourage reporting of all cases suggestive of AD in susceptible species;
4. the Veterinary Authority has current knowledge of, and authority over, all establishments containing pigs in the whole country;
5. domestic pigs are properly identified when leaving their establishment of origin with an indelible mark giving the identification number of their herd of origin; a reliable tracing back procedure is in place for all pigs leaving their establishment of origin.

An AD infected establishment means an establishment in which the virus has been isolated or identified, or a positive serological result (total or gE antibodies) has been confirmed in a laboratory.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 8.2.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the AD status of the exporting country or zone.

Safe commodities

When authorising import or transit of the following commodities and any products made from these, Veterinary Authorities should not require any AD related conditions, regardless of the AD status of the the exporting country or zone.

1. fresh meat of domestic and wild pigs not containing offal (head, and thoracic and abdominal viscera);
2. meat products of domestic and wild pigs not containing offal (head, and thoracic and abdominal viscera);
3. products of animal origin not containing offal (head, and thoracic and abdominal viscera).
Annex XV (contd)

AD free country or zone

1. Qualification

a) A country or zone may be considered free from the disease without formally applying a specific surveillance programme (historical freedom) if the disease has not been reported for at least 25 years, and if for at least the past 10 years:

   a) it has been a notifiable disease;

   b) an early detection system has been in place;

   c) measures to prevent the introduction of the AD virus into the country or zone have been in place;

   d) no vaccination against the disease has been carried out;

   e) infection is not known to be established in wild swine, or measures have been implemented to prevent any transmission of the AD virus from wild swine to domestic pigs.

b) A country or zone which does not meet the conditions of the above paragraph may be considered free from AD when:

   i) animal health regulations to control the movement of commodities with the exception of those listed in Article 8.2.2. in order to prevent the introduction of infection into the establishments of the country or zone have been in place for at least 2 years;

   ii) vaccination against AD has been banned for all domestic pigs in the country or zone for at least 2 years;

   iii) if AD has never been reported in the country or zone, serological surveys, with negative results, have been conducted on a representative sample of all pig establishments in conformity with the recommendations in Chapter X.X. 14. (under study) no more than 3 years prior to qualification; the serological surveys should be directed at the detection of antibodies to the whole virus, and based on the breeding pig population or, for establishments that contain no breeding pigs, on a comparable number of fattening pigs; or

   iv) if AD has been reported in the country or zone, a surveillance and control programme has been in place to detect every infected establishment and eradicate AD from it; the surveillance programme should be carried out in conformity with the recommendations in Chapter X.X. 14. (under study) and demonstrate that no establishments within the country or zone have had any clinical, virological or serological evidence of AD for at least 2 years.

In order for a country to reach free status, all of its zones should have reached AD free status.

v) In countries or zones with wild swine, measures should be implemented to prevent any transmission of the AD virus from wild swine to domestic pigs.
2. **Maintenance of free status**

In order to maintain its free status, a country or zone should comply with the following requirements:

- **Fa** periodic serological surveys directed at the detection of antibodies to the whole AD virus should be carried out on a statistically significant number of breeding pigs, in conformity with the recommendations in Chapter X.X. 1.4 (under study);

- **Gb** the importation of the commodities with the exception of those listed in Article 8.2.2. into the country or zone is carried out in conformity with the import conditions contained in the relevant Articles of the present chapter;

- **Hc** the ban on AD vaccination remains in force;

- **Id** measures aimed at preventing the transmission of the AD virus from wild swine to domestic pigs remain in force.

3. **Recovery of free status**

Should an AD outbreak occur in an establishment of a free country or zone, the status of the country or zone may be restored if either:

- **a** all the pigs in the outbreak have been slaughtered; and, during and after the application of this measure, an epidemiological investigation including clinical examination, and serological and/or virological testing has been carried out in all pig establishments which have been directly or indirectly in contact with the infected establishment and in all pig establishments located within a 5 kilometre prescribed radius of from the outbreak, demonstrating that these establishments are not infected; or

- **b** vaccination with gE- deleted vaccines has been applied and:
  
  - **i** a serological testing procedure (differential ELISA) has been implemented in the establishments where vaccination has been applied to demonstrate the absence of infection;
  
  - **ii** the movement of pigs from these establishments has been banned, except for immediate slaughter, until the above procedure has demonstrated the absence of infection;
  
  - **iii** all vaccinated animals have been slaughtered;
  
  - **iv** during and after the application of the measures described in points i) to iii) above, a thorough epidemiological investigation including clinical examination and serological and/or virological testing has been carried out in all pig establishments which have been directly or indirectly in contact with the infected establishment and in all pig establishments located within a 5 kilometre prescribed radius of from the outbreak, demonstrating that these establishments are not infected.

**Article 8.2.4.**

**AD provisionally free country or zone**

1. **Qualification**

A country or zone may be considered as provisionally free from AD if the following conditions are complied with:
Annex XV (contd)

a) animal health regulations to control the movement of commodities with the exception of those listed in Article 8.2.2. in order to prevent the introduction of infection into the establishments of the country or zone have been in place for at least 2 years;

b) if AD has never been reported in the country or zone, a serological survey, with negative results, has been conducted on a representative sample of all pig establishments in conformity with the recommendations in Chapter X.X. 1.4 (under study) (at a level of confidence not sufficient to meet requirements for freedom); the serological survey should be directed at the detection of antibodies to the whole virus, and based on the breeding pig population or, for establishments that contain no breeding pigs, on a comparable number of fattening pigs; or

c) if AD has been reported in the country or zone, a surveillance and control programme has been in place to detect infected establishments and eradicate AD from these establishments, the herd prevalence rate in the country or zone has not exceeded 1% for at least 3 years (the sampling procedure described in point 1e) of the definition of ‘AD free establishment’ should be applied within the establishments of the country or zone), and at least 90% of the establishments in the country or zone are qualified free;

d) in countries or zones with wild swine, measures should be taken to prevent any transmission of the AD virus between wild swine and domestic pigs.

2. Maintenance of provisionally free status

In order to maintain its provisionally free status, a country or zone should comply with the following requirements:

a) the measures described in points 1b) and 1d) above should be continued;

b) the percentage of infected establishments remains <1%;

c) the importation of the commodities with the exception of those listed in Article 8.2.2. into the country or zone is carried out in conformity with the import conditions contained in the relevant Articles of the present chapter.

3. Recovery of provisionally free status

Should the percentage of infected establishments exceed 1% in a provisionally free country or zone, the status of the country or zone is cancelled and may be restored only once the percentage of infected establishments has remained <1% for at least 6 months, and this result is confirmed by a serological survey conducted in conformity with point 1c) above.

Article 8.2.5.

AD infected country or zone

For the purpose of this chapter, countries and zones which do not fulfil the conditions to be considered free or provisionally free of AD should be considered as infected.

Article 8.2.6.

AD free establishment

1. Qualification

To qualify as free from AD, an establishment should satisfy the following conditions:
a) it is under the control of the Veterinary Authority;

b) no clinical, virological or serological evidence of AD has been found for at least one year;

c) the introduction of pigs, semen and embryos/ova into the establishment is carried out in conformity with the import conditions for these commodities contained in the relevant articles of the present chapter;

d) vaccination against AD has not been carried out in the establishment for at least 12 months, and any previously vaccinated pigs are free from gE antibodies;

e) a number of breeding pigs from the establishment has been subjected, with negative results, to serological tests to the whole AD virus, applying a sampling procedure set out in conformity with the recommendations in Chapter XX.1.4 (under study); these tests should have been carried out on two occasions, at an interval of 2 months; for establishments that contain no breeding pigs, the tests should be carried out only once on a comparable number of fattening or weaning pigs;

f) a surveillance and control programme has been in place to detect infected establishments located within a 5 kilometre prescribed radius of from the establishment and no establishment is known to be infected within this zone.

2. Maintenance of free status

For establishments located in an infected country or infected zone, the testing procedure described in point 1e) above should be carried out every 4 months.

For establishments located in a provisionally free country or zone, the testing procedure described in point 1e) above should be carried out every year.

3. Recovery of free status

Should a free establishment become infected, or should an outbreak occur within a 5 kilometre prescribed radius of from a free establishment, the free status of the establishment should be suspended until the following conditions are met:

a) in the infected establishment:

i) all the pigs in the establishment have been slaughtered, or

ii) at least 30 days after removal of all infected animals, all breeding animals have been subjected to a serological test to the whole AD virus, with negative results, on two occasions, at an interval of 2 months;

b) in other establishments located within the 5 kilometre prescribed radius zone; a number of breeding pigs from each establishment has been subjected, with negative results, to serological tests to the whole AD virus (non vaccinated establishments) or to gE antibodies (vaccinated establishments), applying the sampling procedure described in point 1e above.

Article 8.2.7.

Recommendations for importation from AD free countries or zones

for domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:
Annex XV (contd)

1. showed no clinical sign of AD on the day of shipment;
2. come from an establishment located in an AD free country or zone;
3. have not been vaccinated against AD.

Article 8.2.8.

Recommendations for importation from AD provisionally free countries or zones

for domestic pigs for breeding or rearing

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of AD on the day of shipment;
2. have been kept exclusively in AD free establishments since birth;
3. have not been vaccinated against AD;
4. were subjected to a serological test to the whole AD virus, with negative results, within 15 days prior to shipment.

Article 8.2.9.

Recommendations for importation from AD infected countries or zones

for domestic pigs for breeding or rearing

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of AD on the day of shipment;
2. were kept exclusively in AD free establishments since birth;
3. have not been vaccinated against AD;
4. were isolated in the establishment of origin or a quarantine station, and were subjected to a serological test to the whole AD virus, with negative results, on two occasions, at an interval of not less than 30 days between each test, the second test being performed during the 15 days prior to shipment.

Article 8.2.10.

Recommendations for importation from AD provisionally free countries or zones or AD infected countries or zones

for domestic pigs for slaughter

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. a surveillance and control programme is in place in the country or zone to detect infected establishments and eradicate AD;
2. the animals.
Annex XV (contd)

a) are not being eliminated as part of an eradication programme;
b) showed no clinical sign of AD on the day of shipment;
c) have been kept exclusively in AD free establishments since birth; or
d) have been vaccinated against AD at least 15 days prior to shipment.

[Note: Appropriate precautions should be taken both by the exporting country and the importing country to ensure that the pigs are transported directly from the place of shipment to the abattoir for immediate slaughter.]

Article 8.2.11.

Recommendations for importation from AD free countries or zones

for wild swine

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of AD on the day of shipment;
2. were captured in an AD free country or zone;
3. have not been vaccinated against the disease;
4. were isolated in a quarantine station, and were subjected to a serological test to the whole AD virus, with negative results, on two occasions, at an interval of not less than 30 days between each test, the second test being performed during the 15 days prior to shipment.

Article 8.2.12.

Recommendations for importation from AD free countries or zones

for semen of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of AD on the day of collection of the semen;
   b) were kept in an establishment or artificial insemination centre located in an AD free country or zone at the time of semen collection;
2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.6. and 4.5.

Article 8.2.13.

Recommendations for importation from AD provisionally free countries or zones

for semen of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:
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1. the donor animals:
   a) have been kept for at least 4 months prior to semen collection in an artificial insemination centre which has the status of AD free establishment, and where all boars are subjected to a serological test to the whole AD virus, with negative results, every 4 months;
   b) showed no clinical sign of AD on the day of collection;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.6. and 4.5.

Article 8.2.14.

Recommendations for importation from AD infected countries or zones

for semen of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in an AD free establishment for at least 6 months prior to entering the artificial insemination centre;
   b) have been kept for at least 4 months prior to semen collection in the artificial insemination centre which has the status of AD free establishment, and where all boars are subjected to a serological test to the whole AD virus, with negative results, every 4 months;
   c) were subjected to a serological test to the whole AD virus, with negative results, within 10 days prior to or 21 days after semen collection;
   d) showed no clinical sign of AD on the day of collection;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.6. and 4.5.

Article 8.2.15.

Recommendations for importation from AD free countries or zones

for in vivo derived embryos of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) showed no clinical sign of AD on the day of collection of the embryos;
   b) were kept in an establishment located in an AD free country or zone prior to collection;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.

Article 8.2.16.

Recommendations for importation from AD provisionally free countries or zones
for in vivo derived embryos of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) showed no clinical sign of AD on the day of collection of the embryos;
   b) were kept in an AD free establishment for at least 3 months prior to collection;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.

Article 8.2.17.

Recommendations for importation from AD infected countries or zones

for in vivo derived embryos of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) showed no clinical sign of AD on the day of collection of the embryos;
   b) were kept in an AD free establishment for at least 3 months prior to collection;
   c) were subjected to a serological test to the whole AD virus, with negative results, within 10 days prior to collection;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.

Article 8.2.18.

Recommendations for importation from AD free countries or zones

for offal (head, and thoracic and abdominal viscera) of pigs or products containing pig offal

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of offal or products containing pig offal comes from animals which come from establishments located in an AD free country or zone.

Article 8.2.19.

Recommendations for importation from AD provisionally free countries or zones or from AD infected countries or zones

for offal (head, and thoracic and abdominal viscera) of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of offal comes from animals:

1. which have been kept in an AD free establishment since birth;
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2. which have not been in contact with animals from establishments not considered free from AD during their transport to the approved abattoir and therein.

Article 8.2.20.

Recommendations for importation from AD provisionally free countries or zones or from AD infected countries or zones

for products containing pig offal (head, and thoracic and abdominal viscera)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. either the entire consignment of offal used to prepare the products complied with the conditions referred to in Article 8.2.19.; or

2. the products have been processed to ensure the destruction of the AD virus; and

3. the necessary precautions were taken after processing to avoid contact of the products with any source of AD virus.

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CHAPTER 8.3.

BLUETONGUE

Article 8.3.1.

General provisions

For the purposes of the Terrestrial Code, the infective period for bluetongue virus (BTV) shall be 60 days.

Historically, the global BTV distribution has been confined between the latitudes of approximately 53°N and north of 34°S with a recent extension in Northern Europe.

In the absence of clinical disease in a country or zone, its BTV status should be determined by an ongoing surveillance programme (in accordance with Articles 8.3.16. to 8.3.21.). The programme may need to be adapted to target parts of the country or zone at a higher risk due to historical, geographical and climatic factors, ruminant population data and Culicoides ecology, or proximity to enzootic or incursional zones as described in Articles 8.3.16. to 8.3.21.

All countries or zones adjacent to a country or zone not having free status should be subjected to similar surveillance. The surveillance should be carried out over a distance of at least 100 kilometres from the border with that country or zone, but a lesser distance could be acceptable if there are relevant ecological or geographical features likely to interrupt the transmission of BTV or a bluetongue surveillance programme (in accordance with Articles 8.3.16. to 8.3.21.) in the country or zone not having free status supports a lesser distance.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 8.3.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the BTV status of the ruminant population of the exporting country or zone.

Article 8.3.2.

Safe trade commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any BTV related conditions regardless of the BTV status of the ruminant population of the exporting country or zone:

1. milk and milk products;
2. meat and meat products;
3. hides and skins;
4. wool and fibre;
5. in vivo derived bovine embryos and oocytes collected, processed and stored in conformity with the provisions of Chapter 4.7. except for BTV8 (under study).

Article 8.3.3.

BTV free country or zone

1. A country or a zone may be considered free from BTV when bluetongue is notifiable in the whole country and either:
Annex XVI (contd)

a) a surveillance programme in accordance with Articles 8.3.16. to 8.3.21. has demonstrated no evidence of BTV in the country or zone during the past 2 years; or

b) a surveillance programme has demonstrated no evidence of Culicoides in the country or zone.

2. A BTV free country or zone in which ongoing vector surveillance, performed according to point 5 of Article 8.3.19., has found no evidence of Culicoides will not lose its free status through the importation of vaccinated, seropositive or infective animals, or semen or embryos/ova from infected countries or infected zones.

3. A BTV free country or zone in which surveillance has found evidence that Culicoides are present will not lose its free status through the importation of vaccinated or seropositive animals from infected countries or infected zones, provided:

a) the animals have been vaccinated, at least 60 days prior to dispatch, in accordance with the Terrestrial Manual with a vaccine which covers all serotypes whose presence in the source population has been demonstrated through a surveillance programme in accordance with Articles 8.3.16. to 8.3.21., and the animals are identified in the accompanying certification as having been vaccinated; or

b) the animals are not vaccinated and, at least 60 days prior to dispatch, are demonstrated to have specific antibodies against the bluetongue virus serotypes whose presence has been demonstrated in the exporting country or zone.

c) the animals are not vaccinated and a surveillance programme in accordance with Articles 8.3.16. to 8.3.21. has been in place in the source population for a period of at least 60 days immediately prior to dispatch and no evidence of BTV transmission has been detected.

4. A BTV free country or zone adjacent to an infected country or infected zone should include a zone as described in Article 8.3.1. in which surveillance is conducted in accordance with Articles 8.3.16. to 8.3.21. Animals within this zone should be subjected to continuing surveillance. The boundaries of this zone should be clearly defined, and should take account of geographical and epidemiological factors that are relevant to BTV transmission.

Article 8.3.4.

BTV seasonally free zone

A BTV seasonally free zone is a part of an infected country or an infected zone for which for part of a year, surveillance demonstrates no evidence either of BTV transmission or of adult Culicoides.

For the application of Articles 8.3.7., 8.3.10. and 8.3.13., the seasonally free period is taken to commence the day following the last evidence of BTV transmission (as demonstrated by the surveillance programme), and of the cessation of activity of adult Culicoides.

For the application of Articles 8.3.7., 8.3.10. and 8.3.13., the seasonally free period is taken to conclude either:

1. at least 28 days before the earliest date that historical data show bluetongue virus activity has recommenced; or

2. immediately if current climatic data or data from a surveillance programme indicate an earlier resurgence of activity of adult Culicoides.

A BTV seasonally free zone in which surveillance has found no evidence that Culicoides are present will not lose its free status through the importation of vaccinated, seropositive or infective animals, or semen or embryos/ova from infected countries or infected zones.
BTV infected country or zone

For the purpose of this chapter, a BTV infected country or infected zone is a clearly defined area where evidence of BTV has been reported during the past 2 years.

Recommendations for importation from BTV free countries or zones

for ruminants and other BTV susceptible herbivores

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the animals were kept in a BTV free country or zone since birth or for at least 60 days prior to shipment; or

2. the animals were kept in a BTV free country or zone for at least 28 days, then were subjected, with negative results, to a serological test to detect antibody to the BTV group according to the Terrestrial Manual and remained in the BTV free country or zone until shipment; or

3. the animals were kept in a BTV free country or zone for at least 7 days, then were subjected, with negative results, to an agent identification test according to the Terrestrial Manual, and remained in the BTV free country or zone until shipment; or

4. the animals:
   a) were kept in a BTV free country or zone for at least 7 days;
   b) were vaccinated, at least 60 days before the introduction into the free country or zone, in accordance with the Terrestrial Manual against all serotypes whose presence in the source population has been demonstrated through a surveillance programme as described in Articles 8.3.16. to 8.3.21.;
   c) were identified as having been vaccinated; and
   d) remained in the BTV free country or zone until shipment;

AND

5. if the animals were exported from a free zone, either:
   a) did not transit through an infected zone during transportation to the place of shipment; or
   b) were protected from attack from Culicoides at all times when transiting through an infected zone; or
   c) had been vaccinated in accordance with point 4 above.

Recommendations for importation from BTV seasonally free zones

for ruminants and other BTV susceptible herbivores

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:
Annex XVI (contd)

1. were kept during the seasonally free period in a BTV seasonally free zone since birth or for at least 60 days prior to shipment; or

2. were kept during the BTV seasonally free period in a BTV seasonally free zone for at least 28 days prior to shipment, and were subjected during the residence period in the zone to a serological test to detect antibody to the BTV group according to the Terrestrial Manual, with negative results, carried out at least 28 days after the commencement of the residence period; or

3. were kept during the BTV seasonally free period in a BTV seasonally free zone for at least 14 days prior to shipment, and were subjected during the residence period in the zone to an agent identification test according to the Terrestrial Manual, with negative results, carried out at least 14 days after the commencement of the residence period; or

4. were kept during the seasonally free period in a BTV seasonally free zone and were vaccinated, at least 60 days before the introduction into the free country or zone, in accordance with the Terrestrial Manual against all serotypes whose presence in the source population has been demonstrated through a surveillance programme in accordance with Articles 8.3.16. to 8.3.21. and were identified as having been vaccinated and remained in the BTV free country or zone until shipment;

AND

5. if the animals were exported from a free zone, either:

   a) did not transit through an infected zone during transportation to the place of shipment; or

   b) were protected from attack from Culicoides at all times when transiting through an infected zone; or

   c) were vaccinated in accordance with point 4 above.

Article 8.3.8.

Recommendations for importation from BTV infected countries or zones

for ruminants and other BTV susceptible herbivores

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. were protected from attack from Culicoides in an insect proof vector protected establishment for at least 60 days prior to shipment and during transportation to the place of shipment; or

2. were protected from attack from Culicoides in an insect proof vector protected establishment for at least 28 days prior to shipment and during transportation to the place of shipment, and were subjected during that period to a serological test according to the Terrestrial Manual to detect antibody to the BTV group, with negative results, carried out at least 28 days after introduction into the insect proof vector protected establishment; or

3. were protected from attack from Culicoides in an insect proof vector protected establishment for at least 14 days prior to shipment and during transportation to the place of shipment, and were subjected during that period to an agent identification test according to the Terrestrial Manual, with negative results, carried out at least 14 days after introduction into the insect proof vector protected establishment; or
Annex XVI (contd)

4. were vaccinated, at least 60 days before shipment, in accordance with the Terrestrial Manual against all serotypes whose presence in the source population has been demonstrated through a surveillance programme in accordance with Articles 8.3.16. to 8.3.21., and were identified in the accompanying certification as having been vaccinated or, if demonstrated to have antibodies, have been protected from vectors for at least 60 days prior to shipment; or

5. demonstrated to have antibodies for at least 60 days prior to dispatch against all serotypes whose presence has been demonstrated in the source population through a surveillance programme in accordance with Articles 8.3.16. to 8.3.21.; or

6. are not vaccinated and a surveillance programme in accordance with Articles 8.3.16. to 8.3.21. has been in place in the source population for a period of at least 60 days immediately prior to dispatch and no evidence of BTV transmission has been detected, and were protected from attack from Culicoides during transportation to the place of shipment.

Article 8.3.9.

Recommendations for importation from BTV free countries or zones

for semen of ruminants and other BTV susceptible herbivores

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in a BTV free country or zone for at least 60 days before commencement of, and during, collection of the semen; or
   b) were subjected to a serological test according to the Terrestrial Manual to detect antibody to the BTV group, between 21 and 60 days after the last collection for this consignment, with negative results; or
   c) were subjected to an agent identification test according to the Terrestrial Manual on blood samples collected at commencement and conclusion of, and at least every 7 days (virus isolation test) or at least every 28 days (PCR test) during, semen collection for this consignment, with negative results;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 8.3.10.

Recommendations for importation from BTV seasonally free zones

for semen of ruminants and other BTV susceptible herbivores

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept during the BTV seasonally free period in a seasonally free zone for at least 60 days before commencement of, and during, collection of the semen; or
   b) were subjected to a serological test according to the Terrestrial Manual to detect antibody to the BTV group, with negative results, at least every 60 days throughout the collection period and between 21 and 60 days after the final collection for this consignment; or
Annex XVI (contd)

c) were subjected to an agent identification test according to the Terrestrial Manual on blood samples collected at commencement and conclusion of, and at least every 7 days (virus isolation test) or at least every 28 days (PCR test) during, semen collection for this consignment, with negative results;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 8.3.11.

Recommendations for importation from BTV infected countries or zones

for semen of ruminants and other BTV susceptible herbivores

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were protected from attack from Culicoides for at least 60 days before commencement of, and during, collection of the semen; or
   b) were subjected to a serological test according to the Terrestrial Manual to detect antibody to the BTV group, with negative results, at least every 60 days throughout the collection period and between 21 and 60 days after the final collection for this consignment; or
   c) were subjected to an agent identification test according to the Terrestrial Manual on blood samples collected at commencement and conclusion of, and at least every 7 days (virus isolation test) or at least every 28 days (PCR test) during, semen collection for this consignment, with negative results;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 8.3.12.

Recommendations for importation from BTV free countries or zones

for in vivo derived embryos of ruminants (other than bovines) and other BTV susceptible herbivores and for in vitro produced bovine embryos

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) were kept in a BTV free country or zone for at least the 60 days prior to, and at the time of, collection of the embryos; or
   b) were subjected to a serological test according to the Terrestrial Manual to detect antibody to the BTV group, between 21 and 60 days after collection, with negative results; or
   c) were subjected to an agent identification test according to the Terrestrial Manual on a blood sample taken on the day of collection, with negative results;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7., 4.8. and 4.9., as relevant.
Annex XVI (contd)

Article 8.3.13.

Recommendations for importation from BTV seasonally free zones

for in vivo derived embryos/oocytes of ruminants (other than bovines) and other BTV susceptible herbivores and for in vitro produced bovine embryos

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) were kept during the seasonally free period in a seasonally free zone for at least 60 days before commencement of, and during, collection of the embryos/oocytes; or
   b) were subjected to a serological test according to the Terrestrial Manual to detect antibody to the BTV group, between 21 and 60 days after collection, with negative results; or
   c) were subjected to an agent identification test according to the Terrestrial Manual on a blood sample taken on the day of collection, with negative results;

2. the embryos/oocytes were collected, processed and stored in conformity with the provisions of Chapters 4.7., 4.8. and 4.9., as relevant.

Article 8.3.14.

Recommendations for importation from BTV infected countries or zones

for in vivo derived embryos/oocytes of ruminants (other than bovines) and other BTV susceptible herbivores and for in vitro produced bovine embryos

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) were protected from attack from Culicoides for at least 60 days before commencement of, and during, collection of the embryos/oocytes; or
   b) were subjected to a serological test according to the Terrestrial Manual to detect antibody to the BTV group, between 21 and 60 days after collection, with negative results; or
   c) were subjected to an agent identification test according to the Terrestrial Manual on a blood sample taken on the day of collection, with negative results;

2. the embryos/oocytes were collected, processed and stored in conformity with the provisions of Chapters 4.7., 4.8. and 4.9., as relevant.

Article 8.3.15.

Protecting animals from Culicoides attack

1. Vector-protected establishment or facility

   The means of protection of the establishment or facility should at least comprise the following:

   a) double-door entry-exit system;

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b) openings of the building are vector screened with mesh of appropriate aperture size (under study) impregnated regularly with an approved insecticide according to manufacturers’ instruction;

c) vector surveillance and control within and around the building;

d) measures to limit breeding sites for vectors in vicinity of the establishment or facility;

e) Standard Operating Procedure, including description of back-up and alarm systems, for operation of the establishment or facility and transport of horses to the place of loading.

2. During transportation

When transporting animals through BTV infected countries or infected zones, Veterinary Authorities should require strategies to protect animals from attack from Culicoides during transport, taking into account the local ecology of the vector.

Potential risk management strategies include:

4a) treating animals with insect repellents prior to and during transportation;

4b) loading, transporting and unloading animals at times of low vector activity (i.e. bright sunshine, low temperature);

4c) ensuring vehicles do not stop en route during dawn or dusk, or overnight, unless the animals are held behind insect proof netting;

4d) darkening the interior of the vehicle, for example by covering the roof and/or sides of vehicles with shadecloth;

4e) surveillance for vectors at common stopping and offloading points to gain information on seasonal variations;

4f) using historical information and/or information from appropriately verified and validated BTV epidemiological models to identify low risk ports and transport routes.

Article 8.3.16.

Surveillance: introduction

Articles 8.3.16. to 8.3.21. define the principles and provide a guide on the surveillance for BT complementary to Chapter 1.4. and for vectors complementary to Chapter 1.5., applicable to Members seeking to determine their BT status. This may be for the entire country or zone. Guidance for Members seeking free status following an outbreak and for the maintenance of BT status is also provided.

BT is a vector-borne infection transmitted by different species of Culicoides insects in a range of ecosystems. An important component of BT epidemiology is vectorial capacity which provides a measure of disease risk that incorporates vector competence, abundance, biting rates, survival rates and extrinsic incubation period. However, methods and tools for measuring some of these vector factors remain to be developed, particularly in a field context. Therefore, surveillance for BT should focus on transmission in domestic ruminants.

The impact and epidemiology of BT differ widely in different regions of the world and therefore it is impossible to provide specific recommendations for all situations. It is incumbent upon Members to provide scientific data that explain the epidemiology of BT in the region concerned and adapt the surveillance strategies for defining their infection status (free, seasonally free or infected country or zone) to the local conditions. There is considerable latitude available to Members to justify their infection status at an acceptable level of confidence.
Surveillance for BT should be in the form of a continuing programme.

Article 8.3.17.

Surveillance: case definition

For the purposes of surveillance, a case refers to an animal infected with BT virus (BTV).

For the purposes of international trade, a distinction should be made between a case as defined below and an animal that is potentially infectious to vectors. The conditions for trade are defined in Articles 8.3.1. to 8.3.15. of this chapter.

The purpose of surveillance is the detection of virus circulation in a country or zone and not determination of the status of an individual animal or herd. Surveillance deals not only with the occurrence of clinical signs caused by BTV, but also with the evidence of infection with BTV in the absence of clinical signs.

The following defines the occurrence of BTV infection:

1. BTV has been isolated and identified as such from an animal or a product derived from that animal, or
2. viral antigen or viral ribonucleic acid (RNA) specific to one or more of the serotypes of BTV has been identified in samples from one or more animals showing clinical signs consistent with BT, or epidemiologically linked to a confirmed or suspected case, or giving cause for suspicion of previous association or contact with BTV, or
3. antibodies to structural or nonstructural proteins of BTV that are not a consequence of vaccination have been identified in one or more animals that either show clinical signs consistent with BT, or epidemiologically linked to a confirmed or suspected case, or give cause for suspicion of previous association or contact with BTV.

Article 8.3.18.

Surveillance: general conditions and methods

1. A surveillance system in accordance with Chapter I.4. should be under the responsibility of the Veterinary Authority. In particular:
   a) a formal and ongoing system for detecting and investigating outbreaks of disease should be in place;
   b) a procedure should be in place for the rapid collection and transport of samples from suspect cases of BT to a laboratory for BT diagnosis as described in the Terrestrial Manual;
   c) a system for recording, managing and analysing diagnostic and surveillance data should be in place.
2. The BT surveillance programme should:
   a) in a country/zone free or seasonally free, include an early warning system for reporting suspicious cases. Farmers and workers, who have regular contact with domestic ruminants, as well as diagnosticians, should report promptly any suspicion of BT to the Veterinary Authority. They should be supported directly or indirectly (e.g. through private veterinarians or Veterinary para-professionals) by government information programmes and the Veterinary Authority. An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is BTV. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. All suspected cases of BT should be investigated immediately and samples should be taken and submitted to a laboratory. This requires that sampling kits and other equipment are available for those responsible for surveillance;
Annex XVI (contd)

b) conduct random or targeted serological and virological surveillance appropriate to the infection status of the country or zone.

Generally, the conditions to prevent exposure of susceptible animals to BTV infected vectors will be difficult to apply. However, under specific situations, in establishments such as artificial insemination centres or quarantine stations exposure to vectors may be preventable. The testing requirements for animals kept in these facilities are described in Articles 8.3.11. and 8.3.14.

Article 8.3.19.

Surveillance strategies

The target population for surveillance aimed at identification of disease and/or infection should cover susceptible domestic ruminants within the country or zone. Active and passive surveillance for BTV infection should be ongoing. Surveillance should be composed of random or targeted approaches using virological, serological and clinical methods appropriate for the infection status of the country or zone.

The strategy employed may be based on surveillance using randomised sampling that would demonstrate the absence of BTV infection at an acceptable level of confidence. The frequency of sampling should be dependent on the epidemiological situation. Random surveillance is conducted using serological tests described in the Terrestrial Manual. Positive serological results may be followed up with virological methods as appropriate.

Targeted surveillance (e.g. based on the increased likelihood of infection in particular localities or species) may be an appropriate strategy. Virological and serological methods may be used concurrently to define the BTV status of targeted populations.

A Member should justify the surveillance strategy chosen as being adequate to detect the presence of BTV infection in accordance with Chapter 1.4. and the prevailing epidemiological situation. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clinical signs (e.g. sheep). Similarly, virological and serological testing may be targeted to species that rarely show clinical signs (e.g. cattle).

In vaccinated populations, serological and virological surveillance is necessary to detect the BTV types circulating to ensure that all circulating types are included in the vaccination programme.

If a Member wishes to declare freedom from BTV infection in a specific zone, the design of the surveillance strategy would need to be aimed at the population within the zone.

For random surveys, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect evidence of infection if it were to occur at a predetermined minimum rate. The sample size and expected prevalence determine the level of confidence in the results of the survey. The Member should justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Chapter 1.4. Selection of the design prevalence in particular needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey approach selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and the different species in the target population.
Irrespective of the testing system employed, *surveillance* system design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of *infection* or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as those which may be epidemiologically linked to it.

The principles involved in *surveillance* for disease/infection are technically well defined. The design of *surveillance* programmes to prove the absence of BTV infection/circulation needs to be carefully followed to avoid producing results that are either insufficiently reliable to be accepted by international trading partners, or excessively costly and logistically complicated. The design of any *surveillance* programme, therefore, requires inputs from professionals competent and experienced in this field.

1. **Clinical surveillance**

   Clinical *surveillance* aims at the detection of clinical signs of BT at the flock/ herd level. Whereas significant emphasis is placed on the diagnostic value of mass serological screening, *surveillance* based on clinical inspection should not be underrated, particularly during a newly introduced *infection*. In sheep and occasionally goats, clinical signs may include oedema, hyperaemia of mucosal membranes, coronitis and cyanotic tongue.

   BT suspects detected by clinical *surveillance* should always be confirmed by laboratory testing.

2. **Serological surveillance**

   An active programme of *surveillance* of host populations to detect evidence of BTV transmission is essential to establish BTV status in a country or zone. Serological testing of ruminants is one of the most effective methods of detecting the presence of BTV. The species tested depends on the epidemiology of BTV infection, and the species available, in the local area. Cattle are usually the most sensitive indicator species. Management variables that may influence likelihood of *infection*, such as the use of insecticides and animal housing, should be considered.

   *Surveillance* may include serological surveys, for example abattoir surveys, the use of cattle as sentinel animals (which should be individually identifiable), or a combination of methods. *Surveillance* may also be conducted by sampling and testing of bulk milk using an ELISA, as prescribed in the *Terrestrial Manual*.

   The objective of serological *surveillance* is to detect evidence of BTV circulation. Samples should be examined for antibodies against BTV using tests prescribed in the *Terrestrial Manual*.

   Positive BTV antibody tests results can have four possible causes:

   a) natural *infection* with BTV,
   b) vaccination against BTV,
   c) maternal antibodies,
   d) positive results due to the lack of specificity of the test.

   It may be possible to use sera collected for other survey purposes for BTV *surveillance*. However, the principles of survey design described in these recommendations and the requirements for a statistically valid survey for the presence of BTV infection should not be compromised.
Annex XVI (contd)

The results of random or targeted serological surveys are important in providing reliable evidence that no BTV infection is present in a country or zone. It is, therefore, essential that the survey is thoroughly documented. It is critical to interpret the results in light of the movement history of the animals being sampled.

Serological surveillance in a free zone should target those areas that are at highest risk of BTV transmission, based on the results of previous surveillance and other information. This will usually be towards the boundaries of the free zone. In view of the epidemiology of BTV infection, either random or targeted sampling is suitable to select birds and/or animals for testing.

A protection zone within a free country or zone should separate it from a potentially infected country or infected zone. Serological surveillance in a free country or zone should be carried out over an appropriate distance from the border with a potentially infected country or infected zone, based upon geography, climate, history of infection and other relevant factors.

Serological surveillance in infected zones will identify changes in the boundary of the zone, and can also be used to identify the BTV types circulating. In view of the epidemiology of BTV infection, either random or targeted sampling is suitable.

3. Virological surveillance

Isolation and genetic analysis of BTV from a proportion of infected animals is beneficial in terms of providing information on serotype and genetic characteristics of the viruses concerned.

Virological surveillance using tests described in the Terrestrial Manual can be conducted:

a) to identify virus circulation in at risk populations,

b) to confirm clinically suspect cases,

c) to follow up positive serological results,

d) to better characterize the genotype of circulating virus in a country or zone.

4. Sentinel animals

Sentinel animals are a form of targeted surveillance with a prospective study design. They are the preferred strategy for BTV surveillance. They comprise groups of unexposed animals managed at fixed locations and sampled regularly to detect new BTV infections.

The primary purpose of a sentinel animal programme is to detect BTV infections occurring at a particular place, for instance sentinel groups may be located on the usual boundaries of infected zones to detect changes in distribution of BTV. In addition, sentinel animal programmes allow the timing and dynamics of infections to be observed.

A sentinel animal programme should use animals of known source and history of exposure, control management variables such as use of insecticides and animal housing (depending on the epidemiology of BTV in the area under consideration), and be flexible in its design in terms of sampling frequency and choice of tests.
Care is necessary in choosing the sites for the sentinel groups. The aim is to maximise the chance of detecting BTV activity at the geographical location for which the sentinel site acts as a sampling point. The effect of secondary factors that may influence events at each location, such as climate, may also be analysed. To avoid bias, sentinel groups should comprise animals selected to be of similar age and susceptibility to BTV infection. Cattle are the most appropriate sentinels but other domestic ruminant species may be used. The only feature distinguishing groups of sentinels should be their geographical location.

Sera from sentinel animal programmes should be stored methodically in a serum bank to allow retrospective studies to be conducted in the event of new serotypes being isolated.

The frequency of sampling will depend on the reason for choosing the sampling site. In endemic areas, virus isolation will allow monitoring of the serotypes and genotypes of BTV circulating during each time period. The borders between infected and non infected areas can be defined by serological detection of infective period. Monthly sampling intervals are frequently used. Sentinels in declared free zones add to confidence that BTV infections are not occurring unobserved. In such cases, sampling prior to and after the possible period of transmission is sufficient.

Definitive information on BTVs circulating in a country or zone is provided by isolation and identification of the viruses. If virus isolation is required, sentinels should be sampled at sufficiently frequent intervals to ensure that samples are collected during the period of viraemia.

5. **Vector surveillance**

BTV is transmitted between ruminant hosts by species of *Culicoides* which vary across the world. It is therefore important to be able to identify potential vector species accurately although many such species are closely related and difficult to differentiate with certainty.

The main purpose of vector surveillance is to determine areas of different levels of risk and local details of seasonality by determining the various vector species present in an area, their respective seasonal occurrence, and abundance. Vector surveillance has particular relevance to potential areas of spread. Long term surveillance can also be used to assess vector suppression measures.

The most effective way of gathering this information should take account of the biology and behavioural characteristics of the local vector species of *Culicoides* and may include the use of Onderstepoort-type light traps or similar, operated from dusk to dawn in locations adjacent to domestic ruminants, or the use of drop traps over ruminant animals.

*Vector surveillance* should be based on scientific sampling techniques. The choice of the number and type of traps to be used in vector surveillance and the frequency of their use should take into account the size and ecological characteristics of the area to be surveyed.

The operation of vector surveillance sites at the same locations as sentinel animals is advisable.

The use of a vector surveillance system to detect the presence of circulating virus is not recommended as a routine procedure as the typically low vector infection rates mean that such detections can be rare. Other surveillance strategies (e.g. the use of sentinel animals of domestic ruminants) are preferred to detect virus circulation.
Annex XVI (contd)

Article 8.3.20.

Documentation of BTV infection free status

1. **Members declaring freedom from BTV infection for the country or zone: additional surveillance procedures**

   In addition to the general conditions described in the above-mentioned articles, a Member declaring freedom from BTV infection for the entire country or a zone should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and should be planned and implemented according to general conditions and methods described in this chapter, to demonstrate absence of BTV infection during the preceding 24 months in susceptible domestic ruminant populations. This requires the support of a laboratory able to undertake identification of BTV infection through virus detection and antibody tests described in the *Terrestrial Manual*. This surveillance should be targeted to non-vaccinated animals. Clinical surveillance may be effective in sheep while serological surveillance is more appropriate in cattle.

2. **Additional requirements for countries or zones that practise vaccination**

   Vaccination to prevent the transmission of BTV may be part of a disease control programme. The level of flock or herd immunity required to prevent transmission will depend on the flock or herd size, composition (e.g. species) and density of the susceptible population. It is therefore impossible to be prescriptive. The vaccine should also comply with the provisions stipulated for BTV vaccines in the *Terrestrial Manual*. Based on the epidemiology of BTV infection in the country or zone, it may be that a decision is reached to vaccinate only certain species or other subpopulations.

   In countries or zones that practise vaccination, there is a need to perform virological and serological tests to ensure the absence of virus circulation. These tests should be performed on non-vaccinated subpopulations or on sentinels. The tests have to be repeated at appropriate intervals according to the purpose of the surveillance programme. For example, longer intervals may be adequate to confirm endemicity, while shorter intervals may allow on-going demonstration of absence of transmission.

   **Article 8.3.21.**

The use and interpretation of serological and virus detection tests

1. **Serological testing**

   Ruminants infected with BTV produce antibodies to structural and non-structural viral proteins, as do animals vaccinated with current modified live virus vaccines. Antibodies to the BTV serogroup antigen are detected with high sensitivity and specificity by competitive ELISA (c-ELISA) and to a lesser extent by AGID as described in the *Terrestrial Manual*. Positive c-ELISA results can be confirmed by neutralization assay to identify the infecting serotype(s); however, BTV infected ruminants can produce neutralizing antibodies to serotypes of BTV other than those to which they were exposed (false positive results), especially if they have been infected with multiple serotypes.

2. **Virus detection**

   The presence of BTV in ruminant blood and tissues can be detected by virus isolation or polymerase chain reaction (PCR) as described in the *Terrestrial Manual*.

   Interpretation of positive and negative results (both true and false) differs markedly between these tests because they detect different aspects of BTV infection, specifically (1) infectious BTV (virus isolation) and (2) nucleic acid (PCR). The following are especially relevant to interpretation of PCR assays:
a) The nested PCR assay detects BTV nucleic acid in ruminants long after the clearance of infectious virus. Thus positive PCR results do not necessarily coincide with active infection of ruminants. Furthermore, the nested PCR assay is especially prone to template contamination, thus there is considerable risk of false positive results.

b) PCR procedures other than real time PCR allow sequence analysis of viral amplicons from ruminant tissues, insect vectors or virus isolates. These sequence data are useful for creating data bases to facilitate important epidemiological studies, including the possible distinction of field and vaccine virus strains of BTV, genotype characterization of field strains of BTV, and potential genetic divergence of BTV relevant to vaccine and diagnostic testing strategies.

It is essential that BTV isolates are sent regularly to the OIE Reference Laboratories for genetic and antigenic characterization.

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CHAPTER 8.5.

FOOT AND MOUTH DISEASE

Article 8.5.1.

Introduction

For the purposes of the Terrestrial Code, the incubation period for foot and mouth disease (FMD) shall be 14 days.

For the purposes of this Chapter, ruminants include animals of the family of Camelidae (except Camelus dromedarius).

For the purposes of this Chapter, a case includes an animal infected with FMD virus (FMDV).

For the purposes of international trade, this Chapter deals not only with the occurrence of clinical signs caused by FMDV, but also with the presence of infection with FMDV in the absence of clinical signs.

The following defines the occurrence of FMDV infection:

1. FMDV has been isolated and identified as such from an animal or a product derived from that animal; or

2. viral antigen or viral ribonucleic acid (RNA) specific to one or more of the serotypes of FMDV has been identified in samples from one or more animals, whether showing clinical signs consistent with FMD or not, or epidemiologically linked to a confirmed or suspected outbreak of FMD, or giving cause for suspicion of previous association or contact with FMDV; or

3. antibodies to structural or non-structural proteins of FMDV that are not a consequence of vaccination, have been identified in one or more animals showing clinical signs consistent with FMD, or epidemiologically linked to a confirmed or suspected outbreak of FMD, or giving cause for suspicion of previous association or contact with FMDV.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 8.5.2.

FMD free country where vaccination is not practised

Susceptible animals in the FMD free country where vaccination is not practised should be protected from neighbouring infected countries by the application of animal health measures that effectively prevent the entry of the virus, taking into consideration physical or geographical barriers. These measures may include a protection zone.

To qualify for inclusion in the existing list of FMD free countries where vaccination is not practised, a Member should:

1. have a record of regular and prompt animal disease reporting;

2. send a declaration to the OIE stating that:
   a) there has been no outbreak of FMD during the past 12 months;
   b) no evidence of FMDV infection has been found during the past 12 months;
   c) no vaccination against FMD has been carried out during the past 12 months;
d) no vaccinated animal has been introduced since the cessation of vaccination;

3. supply documented evidence that:
   a) surveillance for FMD and FMDV infection in accordance with Articles 8.5.42. to 8.5.48. is in operation;
   b) regulatory measures for the early detection, prevention and control of FMD have been implemented;

4. describe in detail the boundaries and measures of a protection zone, if applicable.

The Member will be included in the list only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information in points 2, 3 and 4 above be re-submitted annually and changes in the epidemiological situation or other significant events including those relevant to points 3b) and 4 should be reported to the OIE according to the requirements in Chapter 1.1.

Article 8.5.3.

FMD free country where vaccination is practised

Susceptible animals in the FMD free country where vaccination is practised should be protected from neighbouring infected countries by the application of animal health measures that effectively prevent the entry of the virus, taking into consideration physical or geographical barriers. These measures may include a protection zone.

To qualify for inclusion in the list of FMD free countries where vaccination is practised, a Member should:

1. have a record of regular and prompt animal disease reporting;

2. send a declaration to the OIE stating that:
   a) there has been no outbreak of FMD during the past 2 years;
   b) no evidence of FMDV circulation has been found during the past 12 months;

3. supply documented evidence that:
   a) surveillance for FMD and FMDV circulation in accordance with Articles 8.5.42. to 8.5.48. is in operation;
   b) regulatory measures for the early detection, prevention and control of FMD have been implemented;
   c) routine vaccination is carried out for the purpose of the prevention of FMD;
   d) the vaccine used complies with the standards described in the Terrestrial Manual and is appropriate for the strains of virus currently circulating;

4. describe in detail the boundaries and measures of a protection zone, if applicable.

The Member will be included in the list only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information in point 2, 3 and 4 above be re-submitted annually and changes in the epidemiological situation or other significant events including those relevant to points 3b) and 4 should be reported to the OIE according to the requirements in Chapter 1.1.
If a Member that meets the requirements of a FMD free country where vaccination is practised wishes to change its status to FMD free country where vaccination is not practised, the status of this country remains unchanged for a period of at least 12 months after vaccination has ceased. Evidence should also be provided showing that FMDV infection has not occurred during that period.

Article 8.5.4.

FMD free zone where vaccination is not practised

An FMD free zone where vaccination is not practised can be established in either an FMD free country where vaccination is practised or in a country of which parts are infected. In defining such zones the principles of Chapter 4.3. should be followed. Susceptible animals in the FMD free zone should be protected from the rest of the country and from neighbouring countries if they are of a different animal health status by the application of animal health measures that effectively prevent the entry of the virus, taking into consideration physical or geographical barriers. These measures may include a protection zone.

To qualify for inclusion in the list of FMD free zones where vaccination is not practised, a Member should:

1. have a record of regular and prompt animal disease reporting;
2. send a declaration to the OIE stating that within the proposed FMD free zone:
   a) there has been no outbreak of FMD during the past 12 months;
   b) no evidence of FMDV infection has been found during the past 12 months;
   c) no vaccination against FMD has been carried out during the past 12 months;
   d) no vaccinated animal has been introduced into the zone since the cessation of vaccination, except in accordance with Article 8.5.10.;
3. supply documented evidence that:
   a) surveillance for FMD and FMDV infection in accordance with Articles 8.5.42. to 8.5.48. is in operation;
   b) regulatory measures for the early detection, prevention and control of FMD have been implemented;
4. describe in detail and supply documented evidence that these are properly implemented and supervised:
   a) the boundaries of the proposed FMD free zone,
   b) the boundaries and measures of a protection zone, if applicable,
   c) the system for preventing the entry of the virus (including the control of the movement of susceptible animals) into the proposed FMDV free zone (in particular if the procedure described in Article 8.5.10. is implemented).

The proposed free zone will be included in the list of FMD free zones where vaccination is not practised only after the submitted evidence has been accepted by the OIE.

The information required in points 2, 3 and 4b)-c) above should be re-submitted annually and changes in the epidemiological situation or other significant events including those relevant to points 3b) and 4b) should be reported to the OIE according to the requirements in Chapter 1.1.
Annex XVII (contd)

Article 8.5.5.

FMD free zone where vaccination is practised

An FMD free zone where vaccination is practised can be established in either an FMD free country where vaccination is not practised or in a country of which parts are infected. In defining such zones the principles of Chapter 4.3. should be followed. Susceptible animals in the FMD free zone where vaccination is practised should be protected from neighbouring countries or zones if they are of a lesser animal health status by the application of animal health measures that effectively prevent the entry of the virus, taking into consideration physical or geographical barriers. These measures may include a protection zone.

To qualify for inclusion in the list of FMD free zones where vaccination is practised, a Member should:

1. have a record of regular and prompt animal disease reporting;
2. send a declaration to the OIE that within the proposed FMD free zone;
   a) there has been no outbreak of FMD for the past 2 years;
   b) no evidence of FMDV circulation has been found during the past 12 months;
3. supply documented evidence that:
   a) surveillance for FMD and FMDV infection circulation in accordance with Articles 8.5.42. to 8.5.48. is in operation;
   b) regulatory measures for the early detection, prevention and control of FMD have been implemented;
   c) routine vaccination is carried out for the purpose of the prevention of FMD;
   d) the vaccine used complies with the standards described in the Terrestrial Manual and is appropriate for the strains of virus currently circulating;
4. describe in detail and supply documented evidence that these are properly implemented and supervised:
   a) the boundaries of the proposed FMD free zone,
   b) the boundaries and measures of a protection zone if applicable,
   c) the system for preventing the entry of the virus (including the control of the movement of susceptible animals) into the proposed FMD free zone (in particular if the procedure described in Article 8.5.10. is implemented).

The proposed free zone will be included in the list of FMD free zones where vaccination is practised only after the submitted evidence has been accepted by the OIE. The information required in points 2, 3 and 4 b)-c) above should be re-submitted annually and changes in the epidemiological situation or other significant events including those relevant to points 3 b) and 4 b)-c) should be reported to the OIE according to the requirements in Chapter 1.1.

If a Member that has a zone which meets the requirements of a FMD free zone where vaccination is practised wishes to change the status of the zone to FMD free zone where vaccination is not practised, the status of this zone remains unchanged for a period of at least 12 months after vaccination has ceased. Evidence should also be provided showing that FMDV infection has not occurred in the said zone during that period.
FMD free compartment

A FMD free compartment can be established in either a FMD free country or zone or in an infected country or zone. In defining such a compartment the principles of Chapters 4.3. and 4.4. should be followed. Susceptible animals in the FMD free compartment should be separated from any other susceptible animals by the application of an effective biosecurity management system.

A Member wishing to establish a FMD free compartment should:

1. have a record of regular and prompt animal disease reporting and if not FMD free, have an official control programme and a surveillance system for FMD in place according to Articles 8.5.42. to 8.5.44. that allows an accurate knowledge of the prevalence of FMD in the country or zone;

2. declare for the FMD free compartment that:
   a) there has been no outbreak of FMD during the past 12 months;
   b) no evidence of FMDV infection has been found during the past 12 months;
   c) vaccination against FMD is prohibited;
   d) no animal vaccinated against FMD within the past 12 months is in the compartment;
   e) animals, semen and embryos should only enter the compartment in accordance with relevant Articles in this chapter;
   f) documented evidence shows that surveillance in accordance with Articles 8.5.42. to 8.5.48. is in operation for FMD and FMDV infection;
   g) an animal identification and traceability system in accordance with Chapters 4.1. and 4.2. is in place;

3. describe in detail the animal subpopulation in the compartment and the biosecurity plan for FMD and FMDV infection.

The compartment should be approved by the Veterinary Authority. The first approval should only be granted when no outbreak of FMD has occurred within the zone in which the compartment is situated, during the last 3 months.

FMD infected country or zone

For the purpose of this Chapter, an FMD infected country is a country that does not fulfil the requirements to qualify as either an FMD free country where vaccination is not practised or an FMD free country where vaccination is practised.

For the purpose of this Chapter, an FMD infected zone is a zone that does not fulfil the requirements to qualify as either an FMD free zone where vaccination is not practised or an FMD free zone where vaccination is practised.
Annex XVII (contd)

Article 8.5.7bis.

**OIE endorsed national FMD control programme**

Countries may apply for endorsement of their national FMD control programme when they have implemented measures that could potentially lead to OIE official recognition of FMD free status.

For a Member’s national FMD control programme to be endorsed by the OIE, the Member should:

1. have submitted documented evidence on the capacity of the veterinary services to control FMD; this evidence can be provided by countries following the OIE PVS pathway to identify gaps and the strategies to strengthen the veterinary services to sustainably control FMD;

2. submit documentation indicating that the national FMD control programme consistent with the recommendation of Chapter 8.5, is applicable to the entire territory or zone;

3. have a record of regular and prompt animal disease reporting according to the requirements in Chapter 1.1;

4. have submitted a dossier on the epidemiology of FMD in the country describing the following:
   a) the general epidemiology of FMD in the country highlighting the current knowledge and gaps,
   b) the measures to prevent introduction of infection from neighbouring countries;
   c) the prevailing livestock production systems and movement patterns of FMD susceptible animals and their products within and into the country;

5. have submitted a detailed plan on the approach to control and eventually eradicate FMD in the country or zone including:
   a) the timeline of the control programme,
   b) the performance indicators to assess the efficacy of the control measures implemented in the framework of the programme;

6. have submitted evidence that FMD surveillance, taking into account provisions in Chapter 1.4. of the Terrestrial Code and the provisions on surveillance of this Chapter, is in place;

7. have diagnostic capability and procedures which include regular submission of samples to a laboratory that carries out diagnosis and further characterisation of strains in accordance with the standards and methods described in the Terrestrial Manual;

8. where vaccination is practised as a part of national FMD control programme, provide legislation making vaccination compulsory on selected populations;

9. if applicable, provide detailed information on vaccination campaigns in particular on:
   a) target populations for vaccination,
   b) monitoring of vaccination coverage, including serological monitoring of population immunity,
   c) technical specification of the vaccines used and description of the licensing procedures in place,
   d) the proposed timeline for the transition to the use of vaccines, fully compliant with the standards and methods described in the Terrestrial Manual.
10. provide an Emergency Preparedness and Response Plan which is implemented in case of outbreaks.

The Member’s national programme will be included in the list of programmes endorsed by the OIE only after the submitted evidence has been accepted by the OIE. Retention on the list requires an annual update on the progress of the FMD control programme and information on significant changes concerning the points above. Changes in the epidemiological situation and other significant events should be reported to the OIE according to the requirements in Chapter 1.1.

The OIE may withdraw the endorsement of the national FMD control programme if there is evidence of:

11. a decreased capability of the veterinary services, or
12. an uncontrolled increase in incidence of FMD.

Article 8.5.8.

Establishment of a containment zone within an FMD free country or zone

In the event of limited outbreaks within an FMD free country or zone, including within a protection zone, with or without vaccination, a single containment zone, which includes all cases, can be established for the purpose of minimizing the impact on the entire country or zone. For this to be achieved and for the Member to take full advantage of this process, the Veterinary Authority should provide documented evidence as soon as possible to the OIE that:

1. the outbreaks are limited based on the following factors:
   a) immediately on suspicion, a rapid response including notification has been made;
   b) standstill of animal movements has been imposed, and effective controls on the movement of other commodities mentioned in this chapter are in place;
   c) epidemiological investigation (trace-back, trace-forward) has been completed;
   d) the infection has been confirmed;
   e) the primary outbreak has been identified and investigations on the likely source of the outbreak have been carried out;
   f) all cases have been shown to be epidemiologically linked;
   g) no new cases have been found in the containment zone within a minimum of two incubation periods as defined in Article 8.5.1. after the stamping-out of the last detected case is completed;
2. a stamping-out policy has been applied;
3. the susceptible animal population within the containment zones should be clearly identifiable as belonging to the containment zone;
4. increased passive and targeted surveillance in accordance with Articles 8.5.42. to 8.5.48. in the rest of the country or zone has been carried out and has not detected any evidence of infection;
5. animal health measures that effectively prevent the spread of the FMDV to the rest of the country or zone, taking into consideration physical and geographical barriers, are in place;
6. ongoing surveillance in the containment zone is in place.
Annex XVII (contd)

The free status of the areas outside the containment zone would be suspended pending the establishment of the containment zone. The free status of these areas could be reinstated irrespective of the provisions of Article 8.5.9., once the containment zone is clearly established, by complying with points 1 to 6 above. The containment zone should be managed in such a way that it can be demonstrated that commodities for international trade can be shown to have originated outside the containment zone.

The recovery of the FMD free status of the containment zone should follow the provisions of Article 8.5.9.

Article 8.5.9.

Recovery of free status

1. When an FMD outbreak or FMDV infection occurs in an FMD free country or zone where vaccination is not practised, one of the following waiting periods is required to regain the status of FMD free country or zone where vaccination is not practised:

   a) 3 months after the last case where a stamping-out policy and serological surveillance are applied in accordance with Articles 8.5.42. to 8.5.48.; or

   b) 3 months after the slaughter of all vaccinated animals where a stamping-out policy, emergency vaccination and serological surveillance are applied in accordance with Articles 8.5.42. to 8.5.48.; or

   c) 6 months after the last case or the last vaccination (according to the event that occurs the latest), where a stamping-out policy, emergency vaccination not followed by the slaughtering of all vaccinated animals, and serological surveillance are applied in accordance with Articles 8.5.42. to 8.5.48., provided that a serological survey based on the detection of antibodies to nonstructural proteins of FMDV demonstrates the absence of infection in the remaining vaccinated population.

   Where a stamping-out policy is not practised, the above waiting periods do not apply, and Article 8.5.2. or 8.5.4. applies.

2. When an FMD outbreak or FMDV infection occurs in an FMD free country or zone where vaccination is practised, one of the following waiting periods is required to regain the status of FMD free country or zone where vaccination is practised:

   a) 6 months after the last case where a stamping-out policy, emergency vaccination and serological surveillance in accordance with Articles 8.5.42. to 8.5.48. are applied, provided that the serological surveillance based on the detection of antibodies to nonstructural proteins of FMDV demonstrates the absence of virus circulation; or

   b) 18 months after the last case where a stamping-out policy is not applied, but emergency vaccination and serological surveillance in accordance with Articles 8.5.42. to 8.5.48. are applied, provided that the serological surveillance based on the detection of antibodies to nonstructural proteins of FMDV demonstrates the absence of virus circulation.

3. When a FMD outbreak or FMDV infection occurs in a FMD free compartment, Article 8.5.6. applies.

   Article 8.5.10.

Direct Transfer directly to slaughter of FMD susceptible animals from an infected zone for slaughter to in a free zone (where vaccination either is or is not practised) within a country

In order not to jeopardise the status of a free zone, FMD susceptible animals should only leave the infected zone if moved transported by mechanised transport directly to slaughter in the nearest designated abattoir under the following conditions:
Annex XVII (contd)

1. no FMD susceptible animal has been introduced into the establishment of origin and no animal in the establishment of origin has shown clinical signs of FMD for at least 30 days prior to movement;
2. the animals were kept in the establishment of origin for at least 3 months prior to movement;
3. FMD has not occurred within a 10-kilometre radius of the establishment of origin for at least 3 months prior to movement;
4. the animals should be transported under the supervision of the Veterinary Authority in a vehicle, which was cleansed and disinfected before loading, directly from the establishment of origin to the abattoir without coming into contact with other susceptible animals;
5. such an abattoir is not approved for the export of fresh meat during the time it is handling the meat of animals from the infected zone;
6. vehicles and the abattoir should be subjected to thorough cleansing and disinfection immediately after use.

The meat should be treated according to Article 8.5.25. or Article 8.5.26. Other products obtained from the animals and any products coming into contact with them should be considered infected, and treated in such a way as to destroy any residual virus in accordance with Articles 8.5.34. to 8.5.41.

Animals moved into a free zone for other purposes should be moved under the supervision of the Veterinary Authority and comply with the conditions in Article 8.5.14.

Article 8.5.11.

Transfer directly to slaughter of FMD susceptible animals from a containment zone to a free zone (where vaccination either is or is not practised) within a country

In order not to jeopardise the status of a free zone, FMD susceptible animals should only leave the containment zone if moved by mechanised transport directly to slaughter in the nearest designated abattoir under the following conditions:

1. the containment zone has been officially established according to the requirements in Article 8.5.8.;
2. the animals should be transported under the supervision of the Veterinary Authority in a vehicle, which was cleansed and disinfected before loading, directly from the establishment of origin to the abattoir without coming into contact with other susceptible animals;
3. such an abattoir is not approved for the export of fresh meat during the time it is handling the meat of animals from the containment zone;
4. vehicles and the abattoir should be subjected to thorough cleansing and disinfection immediately after use.

The meat should be treated according to point 2 of Article 8.5.25. or Article 8.5.26. Other products obtained from the animals and any products coming into contact with them should be treated in such a way as to destroy any residual virus in accordance with Articles 8.5.34. to 8.5.41.

Article 8.5.12.

Recommendations for importation from FMD free countries or zones where vaccination is not practised or FMD free compartments for FMD susceptible animals
Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of FMD on the day of shipment;
2. were kept since birth or for at least the past 3 months in a FMD free country or zone where vaccination is not practised or a FMD free compartment;
3. have not been vaccinated;
4. if transiting an infected zone, were not exposed to any source of FMDV infection during transportation to the place of shipment.

Article 8.5.13.

Recommendations for importation from FMD free countries or zones where vaccination is practised

for domestic ruminants and pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of FMD on the day of shipment;
2. were kept in an FMD free country or zone since birth or for at least the past 3 months; and
3. have not been vaccinated and were subjected, with negative results, to tests for antibodies against FMD virus, when destined to an FMD free country or zone where vaccination is not practised;
4. if transiting an infected zone, were not exposed to any source of FMDV infection during transportation to the place of shipment.

Article 8.5.14.

Recommendations for importation from FMD infected countries or zones

for domestic ruminants and pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of FMD on the day of shipment;
2. were kept in the establishment of origin since birth, or
   a) for the past 30 days, if a stamping-out policy is in force in the exporting country, or
   b) for the past 3 months, if a stamping-out policy is not in force in the exporting country,
   and that FMD has not occurred within a ten-kilometre radius of the establishment of origin for the relevant period as defined in points a) and b) above; and
3. were isolated in an establishment for the 30 days prior to shipment, and all animals in isolation were subjected to diagnostic tests (probang and serology) for evidence of FMDV infection with negative results at the end of that period, and that FMD did not occur within a ten-kilometre radius of the establishment during that period; or
4. were kept in a quarantine station for the 30 days prior to shipment, all animals in quarantine were subjected to diagnostic tests (probang and serology) for evidence of FMDV infection with negative results at the end of that period, and that FMD did not occur within a ten-kilometre radius of the quarantine station during that period;

5. were not exposed to any source of FMDV infection during their transportation from the quarantine station to the place of shipment.

Article 8.5.15.

Recommendations for importation from FMD free countries or zones where vaccination is not practised or FMD free compartments for fresh semen of domestic ruminants and pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen;
   b) were kept for at least 3 months prior to collection in a FMD free country or zone where vaccination is not practised or a FMD free compartment;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 8.5.16.

Recommendations for importation from FMD free countries or zones where vaccination is not practised or FMD free compartments for frozen semen of domestic ruminants and pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen and for the following 30 days;
   b) were kept for at least 3 months prior to collection in an FMD free country or zone where vaccination is not practised or a FMD free compartment;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 8.5.17.

Recommendations for importation from FMD free countries or zones where vaccination is practised for semen of domestic ruminants and pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen and for the following 30 days;
Annex XVII (contd)

b) were kept for at least 3 months prior to collection in a FMD free country or zone;

c) if destined to an FMD free country or zone where vaccination is not practised:
   i) have not been vaccinated and were subjected, not less than 21 days after collection of the semen, to tests for antibodies against FMD virus, with negative results; or
   ii) had been vaccinated at least twice, with the last vaccination not more than 12 and not less than one month prior to collection;

2. no other animal present in the artificial insemination centre has been vaccinated within the month prior to collection;

3. the semen:
   a) was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.;
   b) was stored in the country of origin for a period of at least one month following collection, and during this period no animal on the establishment where the donor animals were kept showed any sign of FMD.

Recommendations for importation from FMD infected countries or zones

for semen of domestic ruminants and pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen;
   b) were kept in an establishment where no animal had been added in the 30 days before collection, and that FMD has not occurred within 10 kilometres for the 30 days before and after collection;
   c) have not been vaccinated and were subjected, not less than 21 days after collection of the semen, to tests for antibodies against FMD virus, with negative results; or
   d) had been vaccinated at least twice, with the last vaccination not more than 12 and not less than one month prior to collection;

2. no other animal present in the artificial insemination centre has been vaccinated within the month prior to collection;

3. the semen:
   a) was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.;
   b) was subjected, with negative results, to a test for FMDV infection if the donor animal has been vaccinated within the 12 months prior to collection;
   c) was stored in the country of origin for a period of at least one month following collection, and during this period no animal on the establishment where the donor animals were kept showed any sign of FMD.
Annex XVII (contd)

Article 8.5.19.

Recommendations for the importation of in vivo derived embryos of cattle

Irrespective of the FMD status of the exporting country, zone or compartment, Veterinary Authorities should authorise without restriction on account of FMD the import or transit through their territory of in vivo derived embryos of cattle subject to the presentation of an international veterinary certificate attesting that the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.

Article 8.5.20.

Recommendations for importation from FMD free countries or zones where vaccination is not practised or FMD free compartments

for in vitro produced embryos of cattle

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) showed no clinical sign of FMD at the time of collection of the oocytes;
   b) were kept at the time of collection in a FMD free country or zone where vaccination is not practised or a FMD free compartment;

2. fertilisation was achieved with semen meeting the conditions referred to in Articles 8.5.15., 8.5.16., 8.5.17. or 8.5.18., as relevant;

3. the oocytes were collected, and the embryos were processed and stored in conformity with the provisions of Chapters 4.8. and 4.9., as relevant.

Article 8.5.21.

Recommendations for importation from FMD free countries or zones where vaccination is practised

for in vitro produced embryos of cattle

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) showed no clinical sign of FMD at the time of collection of the oocytes;
   b) were kept for at least 3 months prior to collection in a FMD free country or zone where vaccination is practised;
   c) if destined for an FMD free country or zone where vaccination is not practised or a FMD free compartment:
      i) have not been vaccinated and were subjected, with negative results, to tests for antibodies against FMD virus; or
      ii) had been vaccinated at least twice, with the last vaccination not less than one month and not more than 12 months prior to collection;
Annex XVII (contd)

2. no other animal present in the establishment has been vaccinated within the month prior to collection;

3. fertilization was achieved with semen meeting the conditions referred to in Articles 8.5.15., 8.5.16., 8.5.17. or 8.5.18., as relevant;

4. the oocytes were collected, and the embryos were processed and stored in conformity with the provisions of Chapters 4.8. and 4.9., as relevant.

Article 8.5.22.

Recommendations for importation from FMD free countries or zones where vaccination is or is not practised or from FMD free compartments

for fresh meat or meat products of FMD susceptible animals

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in the FMD free country or zone where vaccination is or is not practised, or in a FMD free compartment, or which have been imported in accordance with Article 8.5.12., Article 8.5.13. or Article 8.5.14.;

2. have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections for FMD with favourable results.

Article 8.5.23.

Recommendations for importation from FMD free countries or zones where vaccination is practised

for fresh meat of cattle and buffaloes (Bubalus bubalis) (excluding feet, head and viscera)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in the FMD free country or zone where vaccination is practised, or which have been imported in accordance with Article 8.5.12., Article 8.5.13. or Article 8.5.14.;

2. have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections for FMD with favourable results.

Article 8.5.24.

Recommendations for importation from FMD free countries or zones where vaccination is practised

for fresh meat or meat products of pigs and ruminants other than cattle and buffaloes

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in the FMD free country or zone where vaccination is practised, or which have been imported in accordance with Article 8.5.12., Article 8.5.13. or Article 8.5.14.;

2. have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections for FMD with favourable results.
Recommendations for importation from FMD infected countries or zones, where an official national FMD control programme exists, involving compulsory systematic vaccination of cattle, has been endorsed by the OIE for fresh meat of cattle and buffaloes (Bubalus bubalis) (excluding feet, head and viscera)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of meat:

1. comes from animals which:
   a) have remained in the exporting country for at least 3 months prior to slaughter;
   b) have remained, during this period, in a part of the country where cattle are regularly vaccinated against FMD and where official controls are in operation;
   c) have been vaccinated at least twice with the last vaccination not more than 12 months and not less than one month prior to slaughter;
   d) were kept for the past 30 days in an establishment, and that FMD has not occurred within a ten-kilometre radius of the establishment during that period;
   e) have been transported, in a vehicle which was cleansed and disinfected before the cattle were loaded, directly from the establishment of origin to the approved abattoir without coming into contact with other animals which do not fulfil the required conditions for export;
   f) have been slaughtered in an approved abattoir:
      i) which is officially designated for export;
      ii) in which no FMD has been detected during the period between the last disinfection carried out before slaughter and the shipment for export has been dispatched;
   g) have been subjected to ante-mortem and post-mortem inspections for FMD with favourable results within 24 hours before and after slaughter;

2. comes from deboned carcasses:
   a) from which the major lymphatic nodes have been removed;
   b) which, prior to deboning, have been submitted to maturation at a temperature above + 2°C for a minimum period of 24 hours following slaughter and in which the pH value was below 6.0 when tested in the middle of both the longissimus dorsi.

Recommendations for importation from FMD infected countries or zones for meat products of domestic ruminants and pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:
Annex XVII (contd)

1. the entire consignment of meat comes from animals which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections for FMD with favourable results;

2. the meat has been processed to ensure the destruction of the FMD virus in conformity with one of the procedures referred to in Article 8.5.34.;

3. the necessary precautions were taken after processing to avoid contact of the meat products with any potential source of FMD virus.

Article 8.5.27.

Recommendations for importation from FMD free countries or zones (where vaccination either is or is not practised) or FMD free compartments

for milk and milk products intended for human consumption and for products of animal origin (from FMD susceptible animals) intended for use in animal feeding or for agricultural or industrial use

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that these products come from animals which have been kept in a FMD free country, zone or compartment, or which have been imported in accordance with Article 8.5.12., Article 8.5.13. or Article 8.5.14.

Article 8.5.28.

Recommendations for importation from FMD infected countries or zones where an official control programme exists

for milk, cream, milk powder and milk products

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these products:
   a) originate from herds or flocks which were not infected or suspected of being infected with FMD at the time of milk collection;
   b) have been processed to ensure the destruction of the FMD virus in conformity with one of the procedures referred to in Article 8.5.38. and in Article 8.5.39.;

2. the necessary precautions were taken after processing to avoid contact of the products with any potential source of FMD virus.

Article 8.5.29.

Recommendations for importation from FMD infected countries

for blood and meat-meals (from domestic or wild ruminants and pigs)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the manufacturing method for these products included heating to a minimum core temperature of 70°C for at least 30 minutes.

Article 8.5.30.

Recommendations for importation from FMD infected countries

for wool, hair, bristles, raw hides and skins (from domestic or wild ruminants and pigs)
Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these products have been processed to ensure the destruction of the FMD virus in conformity with one of the procedures referred to in Articles 8.5.35., 8.5.36. and 8.5.37.;

2. the necessary precautions were taken after collection or processing to avoid contact of the products with any potential source of FMD virus.

Veterinary Authorities can authorise, without restriction, the import or transit through their territory of semi-processed hides and skins (limed hides, pickled pelts, and semi-processed leather - e.g. wet blue and crust leather), provided that these products have been submitted to the usual chemical and mechanical processes in use in the tanning industry.

Article 8.5.31.

Recommendations for importation from FMD infected countries or zones

for straw and forage

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that these commodities:

1. are free of grossly identifiable contamination with material of animal origin;

2. have been subjected to one of the following treatments, which, in the case of material sent in bales, has been shown to penetrate to the centre of the bale:

   a) either to the action of steam in a closed chamber such that the centre of the bales has reached a minimum temperature of 80°C for at least 10 minutes,

   b) or to the action of formalin fumes (formaldehyde gas) produced by its commercial solution at 35-40% in a chamber kept closed for at least 8 hours and at a minimum temperature of 19°C;

OR

3. have been kept in bond for at least 3 months (under study) before being released for export.

Article 8.5.32.

Recommendations for importation from FMD free countries or zones (where vaccination either is or is not practised)

for skins and trophies derived from FMD susceptible wild animals

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that these products are derived from animals that have been killed in such a country or zone, or which have been imported from a country or zone free of FMD (where vaccination either is or is not practised).

Article 8.5.33.

Recommendations for importation from FMD infected countries or zones

for skins and trophies derived from FMD susceptible wild animals
Annex XVII (contd)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that these products have been processed to ensure the destruction of the FMD virus in conformity with the procedures referred to in Article 8.5.40.

Article 8.5.34.

Procedures for the inactivation of the FMD virus in meat

For the inactivation of viruses present in meat, one of the following procedures should be used:

1. **Canning**
   - Meat is subjected to heat treatment in a hermetically sealed container to reach an internal core temperature of at least 70°C for a minimum of 30 minutes or to any equivalent treatment which has been demonstrated to inactivate the FMD virus.

2. **Thorough cooking**
   - Meat, previously deboned and defatted, shall be subjected to heating so that an internal temperature of 70°C or greater is maintained for a minimum of 30 minutes.

   After cooking, it shall be packed and handled in such a way that it cannot be exposed to a source of virus.

3. **Drying after salting**
   - When rigor mortis is complete, the meat must be deboned, salted with cooking salt (NaCl) and completely dried. It must not deteriorate at ambient temperature.

   ‘Drying’ is defined in terms of the ratio between water and protein which must not be greater than 2.25:1.

   Article 8.5.35.

Procedures for the inactivation of the FMD virus in wool and hair

For the inactivation of viruses present in wool and hair for industrial use, one of the following procedures should be used:

1. industrial washing, which consists of the immersion of the wool in a series of baths of water, soap and sodium hydroxide (soda) or potassium hydroxide (potash);

2. chemical depilation by means of slaked lime or sodium sulphide;

3. fumigation in formaldehyde in a hermetically sealed chamber for at least 24 hours. The most practical method is to place potassium permanganate in containers (which must NOT be made of plastic or polyethylene) and add commercial formalin; the amounts of formalin and potassium permanganate are respectively 53 ml and 35 g per cubic metre of the chamber;

4. industrial scouring which consists of the immersion of wool in a water-soluble detergent held at 60-70°C;

5. storage of wool at 18°C for 4 weeks, or 4°C for 4 months, or 37°C for 8 days.
Procedures for the inactivation of the FMD virus in bristles

For the inactivation of viruses present in bristles for industrial use, one of the following procedures should be used:

1. boiling for at least one hour;
2. immersion for at least 24 hours in a 1% solution of formaldehyde prepared from 30 ml commercial formalin per litre of water.

Procedures for the inactivation of the FMD virus in raw hides and skins

For the inactivation of viruses present in raw hides and skins for industrial use, the following procedure should be used: salting for at least 28 days in sea salt containing 2% sodium carbonate.

Procedures for the inactivation of the FMD virus in milk and cream for human consumption

For the inactivation of viruses present in milk and cream for human consumption, one of the following procedures should be used:

1. a sterilisation process applying a minimum temperature of 132°C for at least one second (ultra-high temperature [UHT]), or
2. if the milk has a pH less than 7.0, a sterilisation process applying a minimum temperature of 72°C for at least 15 seconds (high temperature - short time pasteurisation [HTST]), or
3. if the milk has a pH of 7.0 or over, the HTST process applied twice.

Procedures for the inactivation of the FMD virus in milk for animal consumption

For the inactivation of viruses present in milk for animal consumption, one of the following procedures should be used:

1. the HTST process applied twice;
2. HTST combined with another physical treatment, e.g. maintaining a pH 6 for at least one hour or additional heating to at least 72°C combined with dessication;
3. UHT combined with another physical treatment referred to in point 2 above.

Procedures for the inactivation of the FMD virus in skins and trophies from wild animals susceptible to the disease

For the inactivation of viruses present in skins and trophies from wild animals susceptible to FMD, one of the following procedures should be used prior to complete taxidermal treatment:
Annex XVII (contd)

1. boiling in water for an appropriate time so as to ensure that any matter other than bone, horns, hooves, claws, antlers or teeth is removed;

2. gamma irradiation at a dose of at least 20 kiloGray at room temperature (20°C or higher);

3. soaking, with agitation, in a 4% (w/v) solution of washing soda (sodium carbonate - Na2CO3) maintained at pH 11.5 or above for at least 48 hours;

4. soaking, with agitation, in a formic acid solution (100 kg salt [NaCl] and 12 kg formic acid per 1,000 litres water) maintained at below pH 3.0 for at least 48 hours; wetting and dressing agents may be added;

5. in the case of raw hides, salting for at least 28 days with sea salt containing 2% washing soda (sodium carbonate - Na2CO3).

Article 8.5.41.

Procedures for the inactivation of the FMD virus in casings of ruminants and pigs

For the inactivation of viruses present in casings of ruminants and pigs, the following procedures should be used:

salting for at least 30 days either with dry salt (NaCl) or with saturated brine (Aw < 0.80), or with phosphate salts/sodium chloride mixture, and kept at room temperature of about 20°C during this entire period.

Article 8.5.42.

Surveillance: introduction

Articles 8.5.42. to 8.5.48. define the principles and provide a guide for the surveillance of FMD in accordance with Chapter 1.4. applicable to Members seeking establishment of freedom from FMD, either with or without the use of vaccination. Guidance is provided for Members seeking reestablishment of freedom from FMD for the entire country or for a zone, either with or without vaccination, or a compartment, following an outbreak and for the maintenance of FMD status.

The impact and epidemiology of FMD differ widely in different regions of the world and therefore it is impossible to provide specific recommendations for all situations. Surveillance strategies employed for demonstrating freedom from FMD at an acceptable level of confidence will need to be adapted to the local situation. For example, the approach to proving freedom from FMD following an outbreak caused by a pig-adapted strain of FMD virus (FMDV) should differ significantly from an application designed to prove freedom from FMD for a country or zone where African buffaloes (Syncerus caffer) provide a potential reservoir of infection. It is incumbent upon the Member to submit a dossier to the OIE in support of its application that not only explains the epidemiology of FMD in the region concerned but also demonstrates how all the risk factors are managed. This should include provision of scientifically-based supporting data. There is therefore considerable latitude available to Members to provide a well-reasoned argument to prove that the absence of FMDV infection (in non-vaccinated populations) or circulation (in vaccinated populations) is assured at an acceptable level of confidence.

Surveillance for FMD should be in the form of a continuing programme designed to establish that the whole territory or part of it is free from FMDV infection/circulation.

For the purposes of this Chapter, virus circulation means transmission of FMDV as demonstrated by clinical signs, serological evidence or virus isolation.
Surveillance: general conditions and methods

1. A surveillance system in accordance with Chapter 1.4. should be under the responsibility of the Veterinary Authority. A procedure should be in place for the rapid collection and transport of samples from suspect cases of FMD to a laboratory for FMD diagnoses as described in the Terrestrial Manual.

2. The FMD surveillance programme should:
   a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers who have day-to-day contact with livestock, as well as diagnosticians, should report promptly any suspicion of FMD. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Authority. All suspect cases of FMD should be investigated immediately. Where suspicion cannot be resolved by epidemiological and clinical investigation, samples should be taken and submitted to a laboratory. This requires that sampling kits and other equipment are available for those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in FMD diagnosis and control;
   b) implement, when relevant, regular and frequent clinical inspection and serological testing of high-risk groups of animals, such as those adjacent to an FMD infected country or infected zone (for example, bordering a game park in which infected wildlife are present).

An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is FMDV. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. Applications for freedom from FMDV infection/circulation should, in consequence, provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement stand-still orders, etc.).

Surveillance strategies

1. Introduction

The target population for surveillance aimed at identifying disease and infection should cover all the susceptible species within the country, zone or compartment.

The design of surveillance programmes to prove the absence of FMDV infection/circulation needs to be carefully followed to avoid producing results that are either insufficiently reliable to be accepted by the OIE or international trading partners, or excessively costly and logistically complicated. The design of any surveillance programme, therefore, requires inputs from professionals competent and experienced in this field.

The strategy employed may be based on randomised sampling requiring surveillance consistent with demonstrating the absence of FMDV infection/circulation at an acceptable level of statistical confidence. The frequency of sampling should be dependent on the epidemiological situation. Targeted surveillance (e.g. based on the increased likelihood of infection in particular localities or species) may be an appropriate strategy. The Member should justify the surveillance strategy chosen as adequate to detect the presence of FMDV infection/circulation in accordance with Chapter 1.4. and the epidemiological situation. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clear clinical signs (e.g. cattle and pigs). If a Member wishes to apply for recognition of a specific zone within the country as being free from FMDV infection/circulation, the design of the survey and the basis for the sampling process would need to be aimed at the population within the zone.
For random surveys, the design of the sampling strategy will need to incorporate an epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect infection/circulation if it were to occur at a predetermined minimum rate. The sample size and expected disease prevalence determine the level of confidence in the results of the survey. The Member must justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Chapter 1.4. Selection of the design prevalence in particular clearly needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey design selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and production class of animals in the target population.

Irrespective of the testing system employed, surveillance design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following-up positives to ultimately determine with a high level of confidence, whether they are indicative of infection/circulation or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as herds which may be epidemiologically linked to it.

2. **Clinical surveillance**

Clinical surveillance aims at detecting clinical signs of FMD by close physical examination of susceptible animals. Whereas significant emphasis is placed on the diagnostic value of mass serological screening, surveillance based on clinical inspection should not be underrated. It may be able to provide a high level of confidence of detection of disease if a sufficiently large number of clinically susceptible animals is examined.

Clinical surveillance and laboratory testing should always be applied in series to clarify the status of FMD suspects detected by either of these complementary diagnostic approaches. Laboratory testing may confirm clinical suspicion, while clinical surveillance may contribute to confirmation of positive serology. Any sampling unit within which suspicious animals are detected should be classified as infected until contrary evidence is produced.

A number of issues must be considered in clinical surveillance for FMD. The often underestimated labour intensity and the logistical difficulties involved in conducting clinical examinations should not be underestimated and should be taken into account.

Identification of clinical cases is fundamental to FMD surveillance. Establishment of the molecular, antigenic and other biological characteristics of the causative virus, as well as its source, is dependent upon disclosure of such animals. It is essential that FMDV isolates are sent regularly to the regional reference laboratory for genetic and antigenic characterization.

3. **Virological surveillance**

Virological surveillance using tests described in the Terrestrial Manual should be conducted:

a) to monitor at risk populations;
b) to confirm clinically suspect cases;
c) to follow up positive serological results;
d) to test “normal” daily mortality, to ensure early detection of infection in the face of vaccination or in establishments epidemiologically linked to an outbreak.
4. Serological surveillance

Serological surveillance aims at detecting antibodies against FMDV. Positive FMDV antibody test results can have four possible causes:

a) natural infection with FMDV;

b) vaccination against FMD;

c) maternal antibodies derived from an immune dam (maternal antibodies in cattle are usually found only up to 6 months of age but in some individuals and in some species, maternal antibodies can be detected for considerably longer periods);

d) heterophile (cross) reactions.

It is important that serological tests, where applicable, contain antigens appropriate for detecting antibodies against viral variants (types, subtypes, lineages, topotypes, etc.) that have recently occurred in the region concerned. Where the probable identity of FMDVs is unknown or where exotic viruses are suspected to be present, tests able to detect representatives of all serotypes should be employed (e.g. tests based on nonstructural viral proteins – see below).

It may be possible to use serum collected for other survey purposes for FMD surveillance. However, the principles of survey design described in this Chapter and the requirement for a statistically valid survey for the presence of FMDV should not be compromised.

The discovery of clustering of seropositive reactions should be foreseen. It may reflect any of a series of events, including but not limited to the demographics of the population sampled, vaccinal exposure or the presence of field strain infection. As clustering may signal field strain infection, the investigation of all instances must be incorporated in the survey design. If vaccination cannot be excluded as the cause of positive serological reactions, diagnostic methods should be employed that detect the presence of antibodies to nonstructural proteins (NSPs) of FMDVs as described in the Terrestrial Manual.

The results of random or targeted serological surveys are important in providing reliable evidence that FMDV infection is not present in a country, zone or compartment. It is therefore essential that the survey be thoroughly documented.

Article 8.5.45.

Members applying for recognition of freedom from FMD for the whole country or a zone where vaccination is not practised: additional surveillance procedures

In addition to the general conditions described in the above-mentioned articles, a Member applying for recognition of FMD freedom for the country or a zone where vaccination is not practised should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and will be planned and implemented according to general conditions and methods in this Chapter, to demonstrate absence of FMDV infection, during the preceding 12 months in susceptible populations. This requires the support of a national or other laboratory able to undertake identification of FMDV infection through virus/antigen/genome detection and antibody tests described in the Terrestrial Manual.
Members applying for recognition of freedom from FMD for the whole country or a zone where vaccination is practised: additional surveillance procedures

In addition to the general conditions described in the above-mentioned articles, a Member applying for recognition of country or zone freedom from FMD with vaccination should show evidence of an effective surveillance programme planned and implemented according to general conditions and methods in this Chapter. Absence of clinical disease in the country or zone for the past 2 years should be demonstrated. Furthermore, surveillance should demonstrate that FMDV has not been circulating in any susceptible population during the past 12 months. This will require serological surveillance incorporating tests able to detect antibodies to NSPs as described in the Terrestrial Manual. Vaccination to prevent the transmission of FMDV may be part of a disease control programme. The level of herd immunity required to prevent transmission will depend on the size, composition (e.g. species) and density of the susceptible population. It is therefore impossible to be prescriptive. However, the aim should be for to vaccinate at least 80% of each vaccinated the susceptible population to be immunised. The vaccine must comply with the Terrestrial Manual. Based on the epidemiology of FMD in the country or zone, it may be that a decision is reached to vaccinate only certain species or other subsets of the total susceptible population. In that case, the rationale should be contained within the dossier accompanying the application to the OIE for recognition of status.

Evidence to show the effectiveness of the vaccination programme should be provided.

Members re-applying for recognition of freedom from FMD for the whole country or a zone where vaccination is either practised or not practised, following an outbreak: additional surveillance procedures

In addition to the general conditions described in the above-mentioned articles, a country re-applying for country or zone freedom from FMD where vaccination is practised or not practised should show evidence of an active surveillance programme for FMD as well as absence of FMDV infection/circulation. This will require serological surveillance incorporating, in the case of a country or a zone practising vaccination, tests able to detect antibodies to NSPs as described in the Terrestrial Manual.

Four strategies are recognised by the OIE in a programme to eradicate FMDV infection following an outbreak:

1. slaughter of all clinically affected and in-contact susceptible animals;
2. slaughter of all clinically affected and in-contact susceptible animals and vaccination of at-risk animals, with subsequent slaughter of vaccinated animals;
3. slaughter of all clinically affected and in-contact susceptible animals and vaccination of at-risk animals, without subsequent slaughter of vaccinated animals;
4. vaccination used without slaughter of affected animals or subsequent slaughter of vaccinated animals.

The time periods before which an application can be made for re-instatement of freedom from FMD depends on which of these alternatives is followed. The time periods are prescribed in Article 8.5.9.

In all circumstances, a Member re-applying for country or zone freedom from FMD with vaccination or without vaccination should report the results of an active surveillance programme implemented according to general conditions and methods in this Chapter.
The use and interpretation of serological tests (see Figure 1)

The recommended serological tests for FMD surveillance are described in the Terrestrial Manual.

*Animals* infected with FMDV produce antibodies to both the structural proteins (SP) and the nonstructural proteins (NSP) of the virus. Tests for SP antibodies to include SP-ELISAs and the virus neutralisation test (VNT). The SP tests are serotype specific and for optimal sensitivity should utilise an antigen or virus closely related to the field strain against which antibodies are being sought. Tests for NSP antibodies include NSP I-ELISA 3ABC and the electro-immunotransfer blotting technique (EITB) as recommended in the Terrestrial Manual or equivalent validated tests. In contrast to SP tests, NSP tests can detect antibodies to all serotypes of FMD virus. *Animals* vaccinated and subsequently infected with FMD virus develop antibodies to NSPs, but in some, the titre may be lower than that found in infected *animals* that have not been vaccinated. Both the NSP I-ELISA 3ABC and EITB tests have been extensively used in cattle. Validation in other species is ongoing. Vaccines used should comply with the standards of the Terrestrial Manual insofar as purity is concerned to avoid interference with NSP antibody testing.

Serological testing is a suitable tool for FMD surveillance. The choice of a serosurveillance system will depend on, amongst other things, the vaccination status of the country. A country, which is free from FMD without vaccination, may choose serosurveillance of high-risk subpopulations (e.g. based on geographical risk for exposure to FMDV). SP tests may be used in such situations for screening sera for evidence of FMDV infection/circulation if a particular virus of serious threat has been identified and is well characterised. In other cases, NSP testing is recommended in order to cover a broader range of strains and even serotypes. In both cases, serological testing can provide additional support to clinical surveillance. Regardless of whether SP or NSP tests are used in countries that do not vaccinate, a diagnostic follow-up protocol should be in place to resolve any presumptive positive serological test results.

In areas where *animals* have been vaccinated, SP antibody tests may be used to monitor the serological response to the vaccination. However, NSP antibody tests should be used to monitor for FMDV infection/circulation. NSP-ELISAs may be used for screening sera for evidence of infection/circulation irrespective of the vaccination status of the *animal*. All *herds* with seropositive reactors should be investigated. Epidemiological and supplementary laboratory investigation results should document the status of FMDV infection/circulation for each positive *herd*. Tests used for confirmation should be of high diagnostic specificity to eliminate as many false positive screening test reactors as possible. The diagnostic sensitivity of the confirmatory test should approach that of the screening test. The EITB or another OIE-accepted test should be used for confirmation.

Information should be provided on the protocols, reagents, performance characteristics and validation of all tests used.

1. **The follow-up procedure in case of positive test results if no vaccination is used in order to establish or re-establish FMD free status without vaccination**

Any positive test result (regardless of whether SP or NSP tests were used) should be followed up immediately using appropriate clinical, epidemiological, serological and, where possible, virological investigations of the reactor *animal* at hand, of susceptible *animals* of the same epidemiological unit and of susceptible *animals* that have been in contact or otherwise epidemiologically associated with the reactor *animal*. If the follow-up investigations provide no evidence for FMDV infection, the reactor *animal* shall be classified as FMD negative. In all other cases, including the absence of such follow-up investigations, the reactor *animal* should be classified as FMD positive.
2. The follow-up procedure in case of positive test results if vaccination is used in order to establish or re-establish FMD free status with vaccination

In case of vaccinated populations, one has to exclude that positive test results are indicative of virus circulation. To this end, the following procedure should be followed in the investigation of positive serological test results derived from surveillance conducted on FMD vaccinated populations.

The investigation should examine all evidence that might confirm or refute the hypothesis that the positive results to the serological tests employed in the initial survey were not due to virus circulation. All the epidemiological information should be substantiated, and the results should be collated in the final report.

It is suggested that in the primary sampling units where at least one animal reacts positive to the NSP test, the following strategy(ies) should be applied:

a. Following clinical examination, a second serum sample should be taken from the animals tested in the initial survey after an adequate interval of time has lapsed, on the condition that they are individually identified, accessible and have not been vaccinated during this period. The number of animals with antibodies against NSP in the population at the time of retest should be statistically either equal to or less than that observed in the initial test if virus is not circulating.

The animals sampled should remain in the holding pending test results and should be clearly identifiable. If the three conditions for retesting mentioned above cannot be met, a new serological survey should be carried out in the holding after an adequate period of time, repeating the application of the primary survey design and ensuring that all animals tested are individually identified. These animals should remain in the holding and should not be vaccinated, so that they can be retested after an adequate period of time.

b. Following clinical examination, serum samples should be collected from representative numbers of susceptible animals that were in physical contact with the primary sampling unit. The magnitude and prevalence of antibody reactivity observed should not differ in a statistically significant manner from that of the primary sample if virus is not circulating.

c. Following clinical examination, epidemiologically linked herds should be serologically tested and satisfactory results should be achieved if virus is not circulating.

d. Sentinel animals can also be used. These can be young, unvaccinated animals or animals in which maternally conferred immunity has lapsed and belonging to the same species resident within the positive initial sampling units. They should be serologically negative if virus is not circulating. If other susceptible, unvaccinated animals are present, they could act as sentinels to provide additional serological evidence.

Laboratory results should be examined in the context of the epidemiological situation. Corollary information needed to complement the serological survey and assess the possibility of viral circulation includes but is not limited to:

- characterization of the existing production systems;
- results of clinical surveillance of the suspects and their cohorts;
- quantification of vaccinations performed on the affected sites;
- sanitary protocol and history of the establishments with positive reactors;
- control of animal identification and movements;
- other parameters of regional significance in historic FMDV transmission.

The entire investigative process should be documented as standard operating procedure within the surveillance programme.
Fig. 1. Schematic representation of laboratory tests for determining evidence of FMDV infection through or following serological surveys

Key:

<table>
<thead>
<tr>
<th>Test</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>ELISA</td>
<td>Enzyme-linked immunosorbent assay</td>
</tr>
<tr>
<td>VNT</td>
<td>Virus neutralisation test</td>
</tr>
<tr>
<td>NSP</td>
<td>Nonstructural protein(s) of foot and mouth disease virus (FMDV)</td>
</tr>
<tr>
<td>3ABC</td>
<td>NSP antibody test</td>
</tr>
<tr>
<td>EITB</td>
<td>Electro-immuno transfer blotting technique (Western blot for NSP antibodies of FMDV)</td>
</tr>
<tr>
<td>SP</td>
<td>Structural protein test</td>
</tr>
<tr>
<td>S</td>
<td>No evidence of FMDV</td>
</tr>
</tbody>
</table>

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Questionnaire on foot and mouth disease

FMD FREE COUNTRY WHERE VACCINATION IS NOT PRACTISED

Report of a Member which applies for recognition of status, under Chapter 8.5. of the Terrestrial Animal Health Code (2010), as a FMD free country not practising vaccination

Please address concisely the following topics. National regulations laws and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. Introduction
   a) Geographical factors. Provide a general description of the country including physical, geographical and other factors that are relevant to FMD dissemination, countries sharing common borders and other countries that although may not be adjacent share a link for the potential introduction of disease. Provide a map identifying the factors above.
   b) Livestock industry. Provide a general description of the livestock industry in the country.

2. Veterinary system
   a) Legislation. Provide a list and summary of all relevant veterinary legislation in relation to FMD.
   b) Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the Terrestrial Code and 1.1.3. of the Terrestrial Manual and describe how the Veterinary Services supervise and control all FMD related activities. Provide maps and tables wherever possible.
   c) Role of farmers, industry and other relevant groups in FMD surveillance and control (include a description of training and awareness programmes on FMD).
   d) Role of private veterinary profession in FMD surveillance and control.

3. FMD eradication
   a) History. Provide a description of the FMD history in the country, date of first detection, origin of infection, date of eradication (date of last case), types and subtypes present.
   b) Strategy. Describe how FMD was controlled and eradicated (e.g. stamping-out, modified stamping-out, zoning), provide timeframe for eradication.
   c) Vaccines and vaccination. Was FMD vaccine ever used? If so, when was the last vaccination carried out? What species were vaccinated?
d) Legislation, organisation and implementation of the FMD eradication campaign. Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines exist and give a brief summary.

e) Animal identification and movement control. Are susceptible animals identified (individually or at a group level)? Provide a description of the methods of animal identification, herd registration and traceability. How are animal movements controlled in the country? Provide evidence on the effectiveness of animal identification and movement controls. Please provide information on pastoralism, transhumance and related paths of movement.

4. **FMD diagnosis**

Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3., and 2.1.5. of the *Terrestrial Manual* are applied. In particular, the following points should be addressed:

a) Is FMD laboratory diagnosis carried out in the country? If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to, the follow-up procedures and the time frame for obtaining results.

b) Provide an overview of the FMD approved laboratories, in particular to address the following points:

i) Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO, etc. that exist in, or planned for, the laboratory system.

ii) Give details of participation in inter-laboratory validation tests (ring tests).

iii) Is live virus handled?

iv) Biosecurity measures applied.

v) Details of the type of tests undertaken.

5. **FMD surveillance**

Provide documentary evidence that surveillance for FMD in the country complies with the provisions of Articles 8.5.4.2 to 8.5.4.8. of the *Terrestrial Code* and Chapter 2.1.5. of the *Terrestrial Manual*. In particular, the following points should be addressed:

a) Clinical suspicion. What are the criteria for raising a suspicion of FMD? What is the procedure to notify (by whom and to whom) and what penalties are involved for failure to report? Provide a summary table indicating, for the past two years, the number of suspect cases, the number of samples tested for FMDV, species, type of sample, testing method(s) and results (including differential diagnosis).

b) Serological surveillance. Are serological surveys conducted? If so, provide detailed information on the survey design (confidence level, sample size, stratification). How frequently are they conducted? Are wildlife susceptible species included in serological surveys? Provide a summary table indicating, for the past two years, the number of samples tested for FMDV, species, type of sample, testing method(s) and results (including differential diagnosis). Provide details on follow-up actions taken on all suspicious and positive results. Provide criteria for selection of populations for targeted surveillance and numbers of animals examined and samples tested. Provide details on the methods applied for monitoring the performance of the surveillance system including indicators.

c) Livestock demographics and economics. What is the susceptible animal population by species and production systems? How many herds, flocks, etc., of each susceptible species are in the country? How are they distributed (e.g. herd density, etc.)? Provide tables and maps as appropriate.

d) Wildlife demographics. What susceptible species are present in the country? Provide estimates of population sizes and geographic distribution. What are the measures in place to prevent contact between domestic and wildlife susceptible species?
Annex XVII (contd)

e) Slaughterhouses and markets. Where are the major livestock marketing or collection centres? What are the patterns of livestock movement within the country? How are the animals transported and handled during these transactions?

6. FMD prevention

a) Coordination with neighbouring countries. Are there any relevant factors about the adjacent countries or zones that should be taken into account (e.g. size, distance from adjacent border to affected herds or animals)? Describe coordination, collaboration and information sharing activities with neighbouring countries.

b) Are there controls in place for swill feeding to pigs containing animal products? If so provide information on the extent of the practice, and describe controls and surveillance measures.

bc) Import control procedures

From what countries or zones does the country authorize the import of susceptible animals or their products? What criteria are applied to approve such countries or zones? What controls are applied on entry of such animals and products, and subsequent internal movement? What import conditions and test procedures are required? Are imported animals of susceptible species required to undergo a quarantine or isolation period? If so, for how long and where? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports of susceptible animals and their products for the past two years, specifying country or zone of origin, species and volume.

i) Provide a map with the number and location of ports, airports and land crossings. Is the official service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the central Veterinary Services. Describe the communication systems between the central authorities and the border inspection posts, and between border inspection posts.

ii) Provide a description on the methods used for the safe disposal of waste from international traffic, who is responsible and provide a summary, for the past two years, of the quantity disposed of.

iii) Describe the regulations, procedures, type and frequency of checks at the point of entry into the country and/or their final destination, concerning the import and follow-up of the following:

- animals,
- genetic material (semen and embryos),
- animal products,
- veterinary medicinal products (i.e. biologics).

iv) Describe the action available under legislation, and actually taken, when an illegal import is detected. Provide information on detected illegal imports.

7. Control measures and contingency planning

a) Give details of any written guidelines, including contingency plans, available to the official services for dealing with suspected or confirmed outbreaks of FMD.

b) Is quarantine imposed on premises with suspicious cases, pending final diagnosis? What other procedures are followed regarding suspicious cases?

c) In the event of an FMD outbreak:
i) indicate the sampling and testing procedures used to identify and confirm presence of the causative agent;

ii) describe the actions taken to control the disease situation in and around any holdings found to be infected with FMD;

iii) indicate the control and/or eradication procedures (e.g. vaccination, stamping-out, partial slaughter/vaccination, etc.) that would be taken. Include details on antigen and vaccine banks;

iv) describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including any restrictions on restocking;

v) give details of any compensation payments made available to farmers, etc. when animals are slaughtered for disease control/eradication purposes and their prescribed timetable.

8. Compliance with the Terrestrial Code

a) In addition to the documentary evidence that the provisions of Article 8.5.2. are properly implemented and supervised, the Delegate of the country must submit a declaration indicating:

i) there has been no outbreak of FMD during the past 12 months;

ii) no evidence of FMDV infection has been found during the past 12 months;

iii) no vaccination against FMD has been carried out during the past 12 months,

b) and should confirm that since the cessation of vaccination no animals vaccinated against FMD have been imported.

9. Recovery of status

Countries applying for recovery of status should comply with the provisions of Article 8.5.9. of the Terrestrial Code and provide detailed information as specified in sections 3.a), 3.b), 3.c) and 5.b) of this questionnaire. Information in relation to other sections need only be supplied if relevant.

FMD FREE COUNTRY WHERE VACCINATION IS PRACTISED

Report of a Member which applies for recognition of status, under Chapter 8.5. of the Terrestrial Animal Health Code (2010), as a FMD free country practising vaccination

Please address concisely the following topics. National regulations laws and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. Introduction

a) Geographical factors. Provide a general description of the country including physical, geographical and other factors that are relevant to FMD dissemination, countries sharing common borders and other countries that although may not be adjacent share a link for the potential introduction of disease. Provide a map identifying the factors above.

b) Livestock industry. Provide a general description of the livestock industry in the country.
Annex XVII (contd)

2. Veterinary system

a) Legislation. Provide a list and summary of all relevant veterinary legislation in relation to FMD.

b) Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the Terrestrial Code and 1.1.3. of the Terrestrial Manual and describe how the Veterinary Services supervise and control all FMD related activities in the country and in the zone. Provide maps and tables wherever possible.

c) Role of farmers, industry and other relevant groups in FMD surveillance and control (include a description of training and awareness programmes on FMD).

d) Role of private veterinary profession in FMD surveillance and control.

3. FMD eradication

a) History. Provide a description of the FMD history in the country, provide date of first detection, origin of infection, date of eradication (date of last case), types and subtypes present.

b) Strategy. Describe how FMD was controlled and eradicated (e.g. stamping-out, modified stamping-out, zoning), provide timeframe for eradication.

c) Vaccines and vaccination. What type of vaccine is used? What species are vaccinated? Provide evidence that the vaccine used complies with Chapter 2.1.5. of the Terrestrial Manual. Describe the vaccination programme, including records kept, and provide evidence to show its effectiveness (e.g. vaccination coverage, serosurveillance, etc.).

d) Legislation, organisation and implementation of the FMD eradication campaign. Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines exist and give a brief summary.

e) Animal identification and movement control. Are susceptible animals identified (individually or at a group level)? Provide a description of the methods of animal identification, herd registration and traceability, including vaccination data. How are animal movements controlled in the country? Provide evidence on the effectiveness of animal identification and movement controls. Please provide information on pastoralism, transhumance and the related paths of movement.

4. FMD diagnosis

Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3. and 2.1.5. of the Terrestrial Manual are applied. In particular, the following points should be addressed:

a) Is FMD laboratory diagnosis carried out in the country? If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to and the follow-up procedures and the timeframe for obtaining results.

b) Provide an overview of the FMD approved laboratories, in particular to address the following points:

i) Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO, etc. that exist in, or planned for, the laboratory system.

ii) Give details of participation in inter-laboratory validation tests (ring tests).

iii) Is live virus handled?

iv) Biosecurity measures applied.
5. **FMD surveillance**

Provide documentary evidence that surveillance for FMD in the country complies with the provisions of Articles 8.5.42. to 8.5.48. of the Terrestrial Code and Chapter 2.1.5. of the Terrestrial Manual. In particular, the following points should be addressed:

a) Clinical suspicion. What are the criteria for raising a suspicion of FMD? What is the procedure to notify (by whom and to whom) and what penalties are involved for failure to report? Provide a summary table indicating, for the past two years, the number of suspect cases, the number of samples tested for FMDV, species, type of sample, testing method(s) and results (including differential diagnosis).

b) Surveillance. Are serological and virological surveys conducted, in particular applying the provisions of Article 8.5.46.? If so, provide detailed information on the survey design (confidence level, sample size, stratification). How frequently are they conducted? Are wildlife susceptible species included in serological surveys? Provide a summary table indicating, for the past two years, the number of samples tested for FMD and FMDV, species, type of sample, testing method(s) and results (including differential diagnosis). Provide details on follow-up actions taken on all suspicious and positive results. Provide criteria for selection of populations for targeted surveillance and numbers of animals examined and samples tested. Provide details on the methods applied for monitoring the performance of the surveillance system including indicators.

c) Livestock demographics and economics. What is the susceptible animal population by species and production systems? How many herds, flocks, etc., of each susceptible species are in the country? How are they distributed (e.g. herd density, etc.)? Provide tables and maps as appropriate.

d) Wildlife demographics. What susceptible species are present in the country? Provide estimates of population sizes and geographic distribution. What are the measures in place to prevent contact between domestic and wildlife susceptible species?

e) Slaughterhouses and markets. Where are the major livestock marketing or collection centres? What are the patterns of livestock movement within the country? How are the animals transported and handled during these transactions?

6. **FMD prevention**

a) Coordination with neighbouring countries. Are there any relevant factors about the adjacent countries or zones that should be taken into account (e.g. size, distance from adjacent border to affected herds or animals)? Describe coordination, collaboration and information sharing activities with neighbouring countries.

b) Import control procedures

From what countries or zones does the country authorize the import of susceptible animals or their products? What criteria are applied to approve such countries or zones? What controls are applied on entry of such animals and products, and subsequent internal movement? What import conditions and test procedures are required? Are imported animals of susceptible species required to undergo a quarantine or isolation period? If so, for how long and where? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports of susceptible animals and their products for the past two years, specifying country or zone of origin, species and volume.
Annex XVII (contd)

i) Provide a map with the number and location of ports, airports and land crossings. Is the official service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the central Veterinary Services. Describe the communication systems between the central authorities and the border inspection posts, and between border inspection posts.

ii) Provide a description on the methods used for the safe disposal of waste from international traffic, who is responsible and provide a summary, for the past two years, of the quantity disposed of.

iii) Describe the regulations, procedures, type and frequency of checks at the point of entry into the country and/or their final destination, concerning the import and follow-up of the following:

- animals,
- genetic material (semen and embryos),
- animal products,
- veterinary medicinal products (i.e. biologics).

iv) Describe the action available under legislation, and actually taken, when an illegal import is detected. Provide information on detected illegal imports.

7. Control measures and contingency planning

a) Give details of any written guidelines, including contingency plans, available to the official services for dealing with suspected or confirmed outbreaks of FMD.

b) Is quarantine imposed on premises with suspicious cases, pending final diagnosis? What other procedures are followed regarding suspicious cases?

c) In the event of an FMD outbreak:

i) indicate the sampling and testing procedures used to identify and confirm presence of the causative agent;

ii) describe the actions taken to control the disease situation in and around any holdings found to be infected with FMD;

iii) indicate the control and/or eradication procedures (e.g. vaccination, stamping-out, partial slaughter/vaccination, etc.) that would be taken. Include details on antigen and vaccine banks;

iv) describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including any restrictions on restocking;

v) give details of any compensation payments made available to farmers, etc. when animals are slaughtered for disease control/eradication purposes and their prescribed timetable.

8. Compliance with the Terrestrial Code

In addition to the documentary evidence that the provisions of Article 8.5.3. are properly implemented and supervised, the Delegate of the country must submit a declaration indicating that there has been no outbreak of FMD for the past 2 years and no evidence of FMDV circulation for the past 12 months, with documented evidence that:
Annex XVII (contd)

a) *surveillance* for FMD and FMDV circulation in accordance with Articles 8.5.42. to 8.5.48. is in operation, and that regulatory measures for the prevention and control of FMD have been implemented;

b) routine vaccination is carried out for the purpose of the prevention of FMD;

c) the vaccine used complies with the standards described in the *Terrestrial Manual*.

9. Recovery of status

Countries applying for recovery of status should comply with the provisions of Article 8.5.9. of the *Terrestrial Code* and provide detailed information as specified in sections 3.a), 3.b), 3.c) and 5.b) of this questionnaire. Information in relation to other sections need only be supplied if relevant.

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**FMD FREE ZONE WHERE VACCINATION IS NOT PRACTISED**

Report of a Member which applies for recognition of status, under Chapter 8.5. of the *Terrestrial Animal Health Code* (2010), as a FMD free zone not practising vaccination

Please address concisely the following topics. National regulations laws and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. **Introduction**

a) Geographical factors. Provide a general description of the country and the zone including physical, geographical and other factors that are relevant to FMD dissemination, countries or zones sharing common borders and other countries or zones that although may not be adjacent share a link for the potential introduction of disease. The boundaries of the zone must be clearly defined, including a protection zone if applied. Provide a digitalised, geo-referenced map with a precise text description of the geographical boundaries of the zone.

b) Livestock industry. Provide a general description of the livestock industry in the country and the zone.

2. **Veterinary system**

a) Legislation. Provide a list and summary of all relevant veterinary legislation in relation to FMD.

b) Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the *Terrestrial Code* and 1.1.3. of the *Terrestrial Manual* and describe how the Veterinary Services supervise and control all FMD related activities in the country and in the zone. Provide maps and tables wherever possible.

c) Role of farmers, industry and other relevant groups in FMD surveillance and control (include a description of training and awareness programmes on FMD).

d) Role of private veterinary profession in FMD surveillance and control.
Annex XVII (contd)

3. **FMD eradication**

   a) **History.** Provide a description of the FMD history in the country and zone, provide date of first detection, origin of infection, date of eradication in the zone (date of last case), types and subtypes present.

   b) **Strategy.** Describe how FMD was controlled and eradicated in the zone (e.g. stamping-out, modified stamping-out), provide timeframe for eradication.

   c) **Vaccines and vaccination.** If vaccination is used in the rest of the country, what type of vaccine is used? What species are vaccinated? Provide evidence that the vaccine used complies with Chapter 2.1.5. of the Terrestrial Manual. Describe the vaccination programme, including records kept, and provide evidence to show its effectiveness (e.g. vaccination coverage, serosurveillance, etc.).

   d) **Legislation, organisation and implementation of the FMD eradication campaign.** Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines exist and give a brief summary.

   e) **Animal identification and movement control.** Are susceptible animals identified (individually or at a group level)? Provide a description of the methods of animal identification, herd registration and traceability. How are animal movements controlled in and between zones of the same or different status, in particular if the provisions of the Terrestrial Code in Article 8.5.10. are applied? Provide evidence on the effectiveness of animal identification and movement controls. Please provide information on pastoralism, transhumance and the related paths of movement.

4. **FMD diagnosis**

   Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3. and 2.1.5. of the Terrestrial Manual are applied. In particular, the following points should be addressed:

   a) **Is FMD laboratory diagnosis carried out in the country?** If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to. Indicate the laboratory(ies) where samples originating from the zone are diagnosed, the follow-up procedures and the time frame for obtaining results.

   b) **Provide an overview of the FMD approved laboratories,** in particular to address the following points:

      a) **Procedures for the official accreditation of laboratories.** Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO, etc. that exist in, or planned for, the laboratory system.

      b) **Give details of participation in inter-laboratory validation tests (ring tests).**

      c) **Is live virus handled?**

      d) **Biosecurity measures applied.**

      e) **Details of the type of tests undertaken.**

5. **FMD surveillance**

   Provide documentary evidence that surveillance for FMD in the zone complies with the provisions of Articles 8.5.42. to 8.5.48. of the Terrestrial Code and Chapter 2.1.5. of the Terrestrial Manual. In particular, the following points should be addressed:
Annex XVII (contd)

a) Clinical suspicion. What are the criteria for raising a suspicion of FMD? What is the procedure to notify (by whom and to whom) and what penalties are involved for failure to report? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for FMDV, species, type of sample, testing method(s) and results (including differential diagnosis).

b) Serological surveillance. Are serological surveys conducted? If so, provide detailed information on the survey design (confidence level, sample size, stratification). How frequently are they conducted? Are wildlife susceptible species included in serological surveys? Provide a summary table indicating, for the past 2 years, the number of samples tested for FMDV, species, type of sample, testing method(s) and results (including differential diagnosis). Provide details on follow-up actions taken on all suspicious and positive results. Provide criteria for selection of populations for targeted surveillance and numbers of animals examined and samples tested. Provide details on the methods applied for monitoring the performance of the surveillance system including indicators.

c) Livestock demographics and economics. What is the susceptible animal population by species and production systems in the country and the zone? How many herds, flocks, etc., of each susceptible species are in the country? How are they distributed (e.g. herd density, etc.)? Provide tables and maps as appropriate.

d) Wildlife demographics. What susceptible species are present in the country and the zone? Provide estimates of population sizes and geographic distribution. What are the measures in place to prevent contact between domestic and wildlife susceptible species?

e) Slaughterhouses and markets. Where are the major livestock marketing or collection centres? What are the patterns of livestock movement within the country? How are the animals transported and handled during these transactions?

6. FMD prevention

a) Coordination with neighbouring countries. Are there any relevant factors about the adjacent countries and zones that should be taken into account (e.g. size, distance from adjacent border to affected herds or animals)? Describe coordination, collaboration and information sharing activities with neighbouring countries and zones.

If the FMD free zone without vaccination is situated in an FMD infected country or borders an infected country or zone, describe the animal health measures implemented to effectively prevent the introduction of the agent, taking into consideration physical or geographical barriers.

b) Import control procedures

From what countries or zones does the country authorize the import of susceptible animals or their products into a free zone? What criteria are applied to approve such countries or zones? What controls are applied on entry of such animals and products, and subsequent internal movement? What import conditions and test procedures are required? Are imported animals of susceptible species required to undergo a quarantine or isolation period? If so, for how long and where? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports of susceptible animals and their products for the past 2 years, specifying country or zone of origin, species and volume.

i) Provide a map with the number and location of ports, airports and land crossings. Is the official service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the central Veterinary Services. Describe the communication systems between the central authorities and the border inspection posts, and between border inspection posts.
Annex XVII (contd)

ii) Provide a description on the methods used for the safe disposal of waste from international traffic, who is responsible and provide a summary, for the past 2 years, of the quantity disposed of.

iii) Describe the regulations, procedures, type and frequency of checks at the point of entry into the country and/or their final destination, concerning the import and follow-up of the following:

- animals,
- genetic material (semen and embryos),
- animal products,
- veterinary medicinal products (i.e. biologics).

iv) Describe the action available under legislation, and actually taken, when an illegal import is detected. Provide information on detected illegal imports.

7. Control measures and contingency planning

a) Give details of any written guidelines, including contingency plans, available to the official services for dealing with suspected or confirmed outbreaks of FMD.

b) Is quarantine imposed on premises with suspicious cases, pending final diagnosis? What other procedures are followed regarding suspicious cases?

c) In the event of an FMD outbreak:

i) indicate the sampling and testing procedures used to identify and confirm presence of the causative agent;

ii) describe the actions taken to control the disease situation in and around any holdings found to be infected with FMD;

iii) indicate the control and/or eradication procedures (e.g. vaccination, stamping-out, partial slaughter/vaccination, etc.) that would be taken. Include details on antigen and vaccine banks;

iv) describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including any restrictions on restocking;

v) give details of any compensation payments made available to farmers, etc. when animals are slaughtered for disease control/eradication purposes and their prescribed timetable.

8. Compliance with the Terrestrial Code

In addition to the documentary evidence that the provisions of Article 8.5.4. are properly implemented and supervised, the Delegate of the country must submit a declaration indicating:

a) there has been no outbreak of FMD during the past 12 months;

b) no evidence of FMDV infection has been found during the past 12 months;

c) no vaccination against FMD has been carried out during the past 12 months;

d) no vaccinated animal has been introduced into the zone since the cessation of vaccination, except in accordance with Article 8.5.10.
9. Recovery of status

Countries applying for recovery of status should comply with the provisions of Article 8.5.9. of the Terrestrial Code and provide detailed information as specified in sections 3.a), 3.b), 3.c) and 5.b) of this questionnaire. Information in relation to other sections need only be supplied if relevant.

FMD FREE ZONE WHERE VACCINATION IS PRACTISED

Report of a Member which applies for recognition of status, under Chapter 8.5. of the Terrestrial Animal Health Code (2010), as a FMD free zone practising vaccination

Please address concisely the following topics. National regulations laws and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. Introduction
   a) Geographical factors. Provide a general description of the country and the zone including physical, geographical and other factors that are relevant to FMD dissemination, countries or zones sharing common borders and other countries or zones that although may not be adjacent share a link for the potential introduction of disease. The boundaries of the zone must be clearly defined, including a protection zone if applied. Provide a digitalised, geo-referenced map with a precise text description of the geographical boundaries of the zone.
   b) Livestock industry. Provide a general description of the livestock industry in the country and the zone.

2. Veterinary system
   a) Legislation. Provide a list and summary of all relevant veterinary legislation in relation to FMD.
   b) Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the Terrestrial Code and 1.1.3. of the Terrestrial Manual and describe how the Veterinary Services supervise and control all FMD related activities in the country and in the zone. Provide maps and tables wherever possible.
   c) Role of farmers, industry and other relevant groups in FMD surveillance and control (include a description of training and awareness programmes on FMD).
   d) Role of private veterinary profession in FMD surveillance and control.

3. FMD eradication
   a) History. Provide a description of the FMD history in the country and zone, provide date of first detection, origin of infection, date of eradication in the zone (date of last case), types and subtypes present.
   b) Strategy. Describe how FMD was controlled and eradicated in the zone (e.g. stamping-out, modified stamping-out), provide timeframe for eradication.
   c) Vaccines and vaccination. What type of vaccine is used? What species are vaccinated? Provide evidence that the vaccine used complies with Chapter 2.1.5. of the Terrestrial Manual. Describe the vaccination programme in the country and in the zone, including records kept, and provide evidence to show its effectiveness (e.g. vaccination coverage, serosurveillance, etc.).
Annex XVII (contd)

d) Legislation, organisation and implementation of the FMD eradication campaign. Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines exist and give a brief summary.

e) Animal identification and movement control. Are susceptible animals identified (individually or at a group level)? Provide a description of the methods of animal identification, herd registration and traceability, including vaccination data. How are animal movements controlled in and between zones of the same or different status, in particular if the provisions of the Terrestrial Code in Article 8.5.10. are applied? Provide evidence on the effectiveness of animal identification and movement controls. Please provide information on pastoralism, transhumance and the related paths of movement.

4. FMD diagnosis

Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3. and 2.1.5. of the Terrestrial Manual are applied. In particular, the following points should be addressed:

a) Is FMD laboratory diagnosis carried out in the country? If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to, the follow-up procedures and the time frame for obtaining results. Indicate the laboratory(ies) where samples originating from the zone are diagnosed.

b) Provide an overview of the FMD approved laboratories, in particular to address the following points.

i) Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO, etc. that exist in, or planned for, the laboratory system.

ii) Give details of participation in inter-laboratory validation tests (ring tests).

iii) Is live virus handled?

iv) Biosecurity measures applied.

v) Details of the type of tests undertaken.

5. FMD surveillance

Provide documentary evidence that surveillance for FMD in the zone complies with the provisions of Articles 8.5.42. to 8.5.48. of the Terrestrial Code and Chapter 2.1.5. of the Terrestrial Manual. In particular, the following points should be addressed:

a) Clinical suspicion. What are the criteria for raising a suspicion of FMD? What is the procedure to notify (by whom and to whom) and what penalties are involved for failure to report? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for FMDV, species, type of sample, testing method(s) and results (including differential diagnosis).

b) Surveillance. Are serological and virological surveys conducted, in particular applying the provisions of Article 8.5.46.? If so, provide detailed information on the survey design (confidence level, sample size, stratification). How frequently are they conducted? Are wildlife susceptible species included in serological surveys? Provide a summary table indicating, for the past 2 years, the number of samples tested for FMD and FMDV, species, type of sample, testing method(s) and results (including differential diagnosis). Provide details on follow-up actions taken on all suspicious and positive results. Provide criteria for selection of populations for targeted surveillance and numbers of animals examined and samples tested. Provide details on the methods applied for monitoring the performance of the surveillance system including indicators.
c) Livestock demographics and economics. What is the susceptible animal population by species and production systems in the country and the zone? How many herds, flocks, etc., of each susceptible species are in the country? How are they distributed (e.g. herd density, etc.)? Provide tables and maps as appropriate.

d) Wildlife demographics. What susceptible species are present in the country and in the zone? Provide estimates of population sizes and geographic distribution. What are the measures in place to prevent contact between domestic and wildlife susceptible species?

e) Slaughterhouses and markets. Where are the major livestock marketing or collection centres? What are the patterns of livestock movement within the country? How are the animals transported and handled during these transactions?

6. FMD prevention

a) Coordination with neighbouring countries. Are there any relevant factors about the adjacent countries and zones that should be taken into account (e.g. size, distance from adjacent border to affected herds or animals)? Describe coordination, collaboration and information sharing activities with neighbouring countries and zones.

If the FMD free zone with vaccination is situated in an FMD infected country or borders an infected country or zone, describe the animal health measures implemented to effectively prevent the introduction of the agent, taking into consideration physical or geographical barriers.

b) Import control procedures

From what countries or zones does the country authorize the import of susceptible animals or their products into a free zone? What criteria are applied to approve such countries or zones? What controls are applied on entry of such animals and products, and subsequent internal movement? What import conditions and test procedures are required? Are imported animals of susceptible species required to undergo a quarantine or isolation period? If so, for how long and where? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports of susceptible animals and their products for the past 2 years, specifying the country or zone of origin, the species and the volume.

i) Provide a map with the number and location of ports, airports and land crossings. Is the official service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the central Veterinary Services. Describe the communication systems between the central authorities and the border inspection posts, and between border inspection posts.

ii) Provide a description on the methods used for the safe disposal of waste from international traffic, who is responsible and provide a summary, for the past 2 years, of the quantity disposed of.

iii) Describe the regulations, procedures, type and frequency of checks at the point of entry into the country and/or their final destination, concerning the import and follow-up of the following:

- animals,
- genetic material (semen and embryos),
- animal products,
- veterinary medicinal products (i.e. biologics).
Annex XVII (contd)

iv) Describe the action available under legislation, and actually taken, when an illegal import is detected. Provide information on detected illegal imports.

7. Control measures and contingency planning

a) Give details of any written guidelines, including contingency plans, available to the official services for dealing with suspected or confirmed outbreaks of FMD.

b) Is quarantine imposed on premises with suspicious cases, pending final diagnosis? What other procedures are followed regarding suspicious cases?

c) In the event of an FMD outbreak:

i) indicate the sampling and testing procedures used to identify and confirm presence of the causative agent;

ii) describe the actions taken to control the disease situation in and around any holdings found to be infected with FMD;

iii) indicate the control and/or eradication procedures (e.g. vaccination, stamping-out, partial slaughter/vaccination, etc.) that would be taken. Include details on antigen and vaccine banks;

iv) describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including any restrictions on restocking;

v) give details of any compensation payments made available to farmers, etc. when animals are slaughtered for disease control/eradication purposes and their prescribed timetable.

8. Compliance with the Terrestrial Code

In addition to the documentary evidence that the provisions of Article 8.5.5. are properly implemented and supervised, the Delegate of the country must submit a declaration indicating:

a) that there has been no outbreak of FMD for the past 2 years,

b) no evidence of FMDV circulation for the past 12 months,

c) surveillance for FMD and FMDV circulation in accordance with Articles 8.5.42. to 8.5.48. is in operation.

9. Recovery of status

Countries applying for recovery of status should comply with the provisions of Article 8.5.9. of the Terrestrial Code and provide detailed information as specified in sections 3.a), 3.b), 3.c) and 5.b) of this questionnaire. Information in relation to other sections need only be supplied if relevant.

COUNTRY WITH AN OIE ENDORSED NATIONAL FMD CONTROL PROGRAMME

Report of a Member which applies for endorsement of status, under Chapter 8.5, of the Terrestrial Code (2010), as a Member with a endorsed national FMD control programme.

Please address concisely the following topics. National regulations laws and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.
1. Introduction
   
a) Provide a general description of geographical factors in the country and any zones, including physical, geographical and other factors that are relevant to FMD dissemination, countries or zones sharing common borders and other countries or zones that, although not adjacent, present a risk for the introduction of disease.

b) If the endorsed plan is gradually implemented to specific parts of the country, the boundaries of the zone(s) should be clearly defined, including the protection zone, if applied. Provide a digitalised, georeferenced map with a precise text description of the geographical boundaries of the zone(s).

c) Provide a general description of the livestock industry in the country and any zones.

2. Veterinary system
   
a) Legislation. Provide a list and summary of all relevant veterinary legislations in relation to the FMD control programme.

b) Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the Terrestrial Code and 1.1.3. of the Terrestrial Manual and describe how the veterinary services supervise and control all FMD related activities in the country and any zones. Provide maps and tables wherever possible.

c) Provide a description on the involvement and the participation of industry, producers, farmers, including subsistence and small scale producers, community animal health workers and the role of the private veterinary profession in FMD surveillance and control. Include a description of training and awareness programmes on FMD.

d) Provide information on any OIE PVS evaluation of the country and follow-up steps within the PVS Pathway.

3. FMD control
   
a) Provide a description of the FMD history in the country and any zones, including date of first detection, origin of infection, date of implementation of the control programme in the country and any zones, and types and subtypes of the FMD virus present.

b) Describe the general epidemiology of FMD in the country and the surrounding countries or zones highlighting the current knowledge and gaps.

c) Describe how FMD is controlled in the country or any zones. Submit a detailed plan on the measures to control and eventually eradicate FMD in the country. Include the timelines of the control programme and the performance indicators to assess the efficacy of the control measures and plan.

d) Provide a description of the legislation, organisation and implementation of the FMD control programme at the different levels. Indicate if detailed operational guidelines exist and give a brief summary. Describe the funding for the control programme and annual budgets for the duration of the control programme.
Annex XVII (contd)

e) Provide information on what types of vaccines are used and which species are vaccinated. Provide information on the licensing process of the vaccines used. Describe the vaccination programme in the country and in any zones, including records kept, and provide evidence to show its effectiveness (e.g. vaccination coverage, population immunity, etc.). Provide details on the studies carried out to determine the population immunity, indicating the study design, including threshold levels for within herd protective immunity and minimal herd level immunity. Provide details, if applicable, on a proposed timeline for the transition to the use of vaccines fully compliant with the standards and methods described in the Terrestrial Manual to enable demonstration of absence of virus circulation.

f) Provide a description of the methods of animal identification (at the individual or group level), herd registration and traceability; and how the movements of animals and products are assessed and controlled, including movement of infected animals to slaughter. Provide evidence on the effectiveness of animal identification and movement controls. Please provide information on pastoralism, transhumance and related paths of movement. Describe measures to prevent introduction of the virus from neighbouring countries or zones.

4. FMD surveillance

Provide documentary evidence on whether surveillance for FMD in the country complies with the provisions of Articles 8.5.40. to 8.5.46. of the Terrestrial Code and Chapter 2.1.5. of the Terrestrial Manual. In particular, the following points should be addressed:

a) Describe the criteria for raising a suspicion of FMD and the procedure to notify (by whom and to whom) and what penalties are involved for failure to report.

b) Describe how clinical surveillance is conducted, including which levels of the livestock production system are included in clinical surveillance (e.g. farms, markets, fairs, slaughterhouses, check points, etc.). Provide criteria for selection of populations for targeted surveillance and numbers of animals examined and samples tested in diagnostic laboratories. Provide details on the methods applied for monitoring the performance of the surveillance system including indicators. Explain whether serological and virological surveys are conducted and, if so, how frequently and for what purpose.

c) Provide a summary table indicating, for at least 2 consecutive years, the number of samples tested for FMD and FMDV, species, type of sample, testing method(s) and results (including differential diagnosis). Provide procedural details on follow-up actions taken on suspicious and positive results.

d) Provide information on livestock demographics and economics, including the susceptible animal population by species and production systems in the country and the zone. Identify how many herds, flocks, etc., of each susceptible species are in the country and how they are distributed (e.g., herd density, etc.). Provide tables and maps as appropriate.

e) Provide information on wildlife demographics, including which susceptible species are present in the country and any zones. Provide estimates of population sizes and geographic distribution. Identify whether susceptible wildlife are included in surveillance. Identify the measures in place to prevent contact between domestic and susceptible wildlife.

f) Identify the major livestock slaughter, marketing and collection centres. Provide information on the patterns of livestock movement within the country, including how animals are transported and handled during these transactions.

5. FMD laboratory diagnosis

Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3. and 2.1.5. of the Terrestrial Manual are applied. In particular, the following points should be addressed:
a) Is FMD laboratory diagnosis carried out in the country? If so, provide a list of laboratories approved by the competent authority to diagnose FMD. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to, the follow-up procedures and the time frame for obtaining results. If applicable, indicate the laboratory(ies) where samples originating from any zone are diagnosed. Is there regular submission of samples from the country or zone to a laboratory that carries out diagnosis and further characterisation of strains in accordance with the standards and methods described in the Terrestrial Manual?

b) Provide an overview of the FMD approved laboratories, in particular to address the following points.

i) Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO, etc. that exist in, or are planned for, the laboratory system.

ii) Give details on participation in inter-laboratory validation tests (ring tests).

iii) Is live virus handled?

iv) Biosecurity measures applied.

v) Details of the type of tests undertaken.

6. FMD prevention

Describe the procedures in place to prevent the introduction of FMD into the country. In particular, provide details on:

a) Coordination with neighbouring countries, trading partners and other countries within the same region. Identify relevant factors about the adjacent countries and zones that should be taken into account (e.g. size, distance from adjacent borders to affected herds or animals, surveillance carried in adjacent countries). Describe coordination, collaboration and information sharing activities with neighbouring countries and zones. Describe the measures implemented to effectively prevent the introduction of the agent, taking into consideration physical or geographical barriers. Describe the measures implemented to prevent the propagation of the agent within the country.

b) Provide information on countries or zones from which the country authorises the import of susceptible animals or their products into the country or zone. Describe the criteria applied to approve such countries or zones, the controls applied on entry of such animals and products, and subsequent internal movement. Describe the import conditions and test procedures required. Advise whether imported animals of susceptible species are required to undergo a quarantine or isolation period, and if so, the duration and location of quarantine. Advise whether import permits and health certificates are required. Describe any other procedures used. Provide summary statistics on imports of susceptible animals and their products for at least 2 consecutive years, specifying country or zone of origin, the species and the number or volume.

i) Provide a map with the number and location of ports, airports and land crossings. Advise whether the official service responsible for import controls is part of the official services, or if it is an independent body. If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the central veterinary services. Describe the communication systems between the central authorities and the border inspection posts, and between border inspection posts.

ii) Provide a description on the methods used for the safe disposal of waste from international traffic, who is responsible to supervise this and provide a summary, for the past 2 years, of the quantity disposed of.
Annex XVII (contd)

iii) Describe the regulations, procedures, type and frequency of checks at the point of entry into the country and/or their final destination, concerning the import and follow up of the following:
   - animals,
   - genetic material (semen and embryos),
   - animal products,
   - veterinary medicinal products (i.e. biologics).

iv) Describe the action available under legislation, and actually taken, when an illegal import is detected. Provide information on detected illegal imports, if available.

7. Control measures and emergency response

a) Give details of any written guidelines, including emergency response plans, available to the official services for dealing with suspected or confirmed outbreaks of FMD.

b) Advise whether quarantine is imposed on premises with suspicious cases, pending final diagnosis and any other procedures followed in respect of suspicious cases.

c) In the event of a FMD outbreak:

i) provide a detailed description of procedures that are followed in case of an outbreak including forward and backward tracing;

ii) indicate the sampling and testing procedures used to identify and confirm presence of the causative agent;

iii) describe the actions taken to control the disease situation in and around any holdings found to be infected with FMD;

iv) indicate the control and/or eradication procedures (e.g. vaccination, stamping-out, partial slaughter/vaccination, etc.) that would be taken;

v) describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including any restrictions on restocking;

vi) give details of any compensation payments made available to farmers, etc. when animals are slaughtered for disease control or eradication purposes and their prescribed timetable.

8. Recovery of status

Countries applying for recovery of the official endorsement of the national FMD control programme should provide updated information in compliance with the provisions of Article 8.5.7.bis of the Terrestrial Code.

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CHAPTER 8.10.

RABIES

Article 8.10.1.

General provisions

Rabies is a disease caused by any member of the Lyssavirus genus. All mammals including human are susceptible to infection. Carnivora and Chiroptera are the reservoirs for rabies.

For the purposes of the Terrestrial Code:

1. a case is any animal infected with the Rabies virus species;

2. the incubation period for rabies is variable, but will be considered less than 6 months, and the infective period for dogs, cats and ferrets is considered to start 10 days before the onset of the first apparent clinical signs.

The aim of this chapter is to mitigate the risk related to rabies for international trade and non-commercial movements of rabies susceptible species.

The most important species for international trade purposes are domestic carnivores (primarily dogs [Canis familiaris], cats [Felis catus] and ferrets [Mustela putorius furo]) and also include domestic livestock (equids, ruminants and suids).

Rabies can be suspected based on clinical signs or history of exposure to a rabid animal. Confirmation requires antigen detection or virus isolation. Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Members are encouraged to implement and maintain a programme for the management of stray dog populations consistent with Chapter 7.7.

Article 8.10.2.

Rabies free country

A country may be considered free from rabies when:

1. the disease is notifiable and any change in the epidemiological situation or relevant events should be reported in accordance with Chapter 1.1.;

2. an effective system of disease surveillance has been in operation for the last 2 years, with a minimum requirement being an on-going early detection programme to ensure investigation and reporting of rabies suspect animals;

3. regulatory measures for the prevention of rabies are implemented consistent with the recommendations in this chapter, including effective procedures for the importation of domestic dogs, cats and ferrets;

4. no case of indigenously acquired rabies virus infection has been confirmed during the past 2 years;
Annex XVIII (contd)

5. no imported case in reservoir species has been confirmed outside a quarantine station for the past 6 months;

6. an imported human case of rabies will not affect the rabies free status.

Members should implement and maintain a programme for the management of stray dog populations consistent with Chapter 7.7.

Article 8.10.3.

Country free from dog to dog transmission of rabies

A country may be considered free from dog to dog transmission of rabies when:

1. the disease is notifiable and any change in the epidemiological situation or relevant events are reported in accordance with Chapter 1.1.;

2. an effective system of disease surveillance has been in operation for the last 2 years, with a minimum requirement being an on-going early detection programme to ensure investigation and reporting of suspect animals;

3. regulatory measures for the prevention and control of rabies are implemented consistent with the recommendations in this chapter, including vaccination, identification and effective procedures for the importation of domestic dogs, cats and ferrets;

4. thorough epidemiological investigations have demonstrated no case of dog to dog transmission of rabies during the past 2 years.

Members should implement and maintain a programme for the management of stray dog populations consistent with Chapter 7.7.

Article 8.10.4.

Recommendations for importation from rabies free countries

for domestic mammals, and captive wild mammals

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of rabies the day prior to or on the day of shipment;

2. and either:
   a) were kept since birth or at least 6 months prior to shipment in the free country; or
   b) were imported in conformity with the regulations stipulated in Articles 8.10.7., 8.10.8., 8.10.9. or 8.10.10.

Article 8.10.5.

Recommendations for importation from rabies free countries

for wild mammals

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of rabies the day prior to or on the day of shipment;
2. and either:
   a) have been captured and remained in a rabies free country, at a sufficient distance, based on the biology of species, including home range, from any infected country. The distance should be defined according to the species exported and the reservoir species in the neighbouring infected countries; or
   b) were kept for the 6 months prior to shipment in a rabies free country.

Article 8.10.6

**Recommendations for importation of dogs from countries free from dog to dog transmission of rabies**

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the dogs:

1. were kept for at least the 6 months prior to shipment in a country free from dog to dog transmission of rabies;
2. were permanently identified (e.g., by a microchip or tattoo) and the identification number should be stated in the certificate;
3. received, prior to shipment, a valid anti-rabies vaccination, in accordance with the *Terrestrial Manual*, or revaccination if applicable, in accordance with the recommendations of the manufacturer;
4. showed no clinical sign of rabies the day prior to or on the day of shipment;

Article 8.10.7.

**Recommendations for importation of dogs, cats and ferrets from countries considered infected with rabies**

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the *animals*:

1. showed no clinical sign of rabies the day prior to or on the day of shipment;
   
   **AND EITHER:**

   2. were permanently identified (e.g., by a microchip or tattoo) and their identification number should be stated in the certificate; and
   3. received, prior to shipment, a valid anti-rabies vaccination in accordance with the *Terrestrial Manual*, or revaccination if applicable, in accordance with the recommendations of the manufacturer; and
   4. were subjected not less than 3 months and not more than 12 months prior to shipment to an antibody titration test as prescribed in the *Terrestrial Manual* with a positive result; 

   **OR**

   5. have not been vaccinated against rabies or do not meet all the conditions set out in points 2, 3 and 4 above; in such cases, the animals should be quarantined for 6 months prior to export.

Article 8.10.8.

**Recommendations for importation of domestic ruminants and suids from countries considered infected with rabies**
Annex XVIII (contd)

**Veterinary Authorities** should require the presentation of an *international veterinary certificate* attesting that the *animals* showed no clinical sign of rabies the day prior to or on the day of shipment.

Article 8.10.9.

**Recommendations for importation of domestic equids from countries considered infected with rabies**

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the *animals*:

1. showed no clinical sign of rabies the day prior to or on the day of shipment;

2. and either:
   
   a) were kept for the 6 months prior to shipment in an establishment where no contact with reservoir species was maintained and where no case of rabies was reported for at least 12 months prior to shipment; or
   
   b) were vaccinated as prescribed in the *Terrestrial Manual*.

Article 8.10.10.

**Recommendations for importation from countries considered infected with rabies**

for rodents and lagomorphs born and reared in a biosecure facility

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the *animals*:

1. showed no clinical sign of rabies on the day of shipment;

2. were kept since birth in a biosecure facility where no case of rabies was reported for at least 12 months prior to shipment.

Article 8.10.11.

**Recommendations for importation from countries considered infected with rabies**

for captive wild animals (other than non-human primates)

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the *animals*:

1. showed no clinical sign of rabies the day prior to or on the day of shipment;

2. were kept since birth, or for the 6 months prior to shipment, in an establishment where no contact with reservoir species and where no case of rabies was reported for at least 12 months prior to shipment.

Article 8.10.12.

**Recommendations for importation from countries considered infected with rabies**

for wild and feral animals (other than non-human primates and Chiroptera)

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the *animals*:

1. showed no clinical sign of rabies the day prior to or on the day of shipment;

2. were kept for the 6 months prior to shipment in an establishment where separation from wild *animals* and feral *animals* was maintained and where no case of rabies was reported for at least 12 months prior to shipment.
Recommendations for importation from countries considered infected with rabies

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

for captive non-human primates

1. the animals showed no clinical sign of rabies the day prior to or on the day of shipment;

2. quarantine measures were applied in accordance with Chapter 5.9. and Chapter 6.11.
CHAPTER 5.11.

RABIES

MODEL INTERNATIONAL VETERINARY CERTIFICATE FOR DOMESTIC DOGS (Canis familiaris), AND CATS (Felis catus) AND FERRETS (Mustela putorius furo) ORIGINATING FROM RABIES INFECTED COUNTRIES

I. OWNER

Name and address: ...........................................................................................................................
...........................................................................................................................................................
...............................................................................................................................................................
...............................................................................................................................................................

II. DESCRIPTION

Species of animal: ............................................................................................................................
Age or date of birth: ........................................................................................................................
Sex: ...................................................................................................................................................
Breed: ................................................................................................................................................
Colour: ................................................................................................................................................
Coat type and marking/Distinguishing marks: ..............................................................................
Identification number (tattoo or other permanent method of identification) (see note 1)

III. ADDITIONAL INFORMATION

Country of origin: ..............................................................................................................................
Countries visited............................................................................................................................
over the past 2 years ....................................................................................................................
as declared by the owner ............................................................................................................... (give dates) ..............................................................................................................................
IV. VACCINATION (Rabies)

I, the undersigned declare herewith that I have vaccinated the animal described in Part II against rabies as shown below. The animal was found to be healthy on the day of vaccination.

<table>
<thead>
<tr>
<th>Date of vaccination (dd/mm/yy)</th>
<th>Name of inactivated virus vaccine (see note 2)</th>
<th>1. Manufacturing laboratory</th>
<th>2. Batch number</th>
<th>3. Expiry date</th>
<th>Name (in capital letters) and signature of the veterinarian (see note 6)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>1..........................</td>
<td>2.....................</td>
<td>3..................</td>
<td></td>
</tr>
</tbody>
</table>

**PERIOD OF VALIDITY OF VACCINATION FOR INTERNATIONAL MOVEMENT (see note 3)**

<table>
<thead>
<tr>
<th>from (dd/mm/yy)</th>
<th>to (dd/mm/yy)</th>
<th>Name (in capital letters) and signature of the Official Veterinarian</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
V. SEROLOGICAL TESTING (Rabies)

I, the undersigned declare herewith that I have taken a blood sample from the animal described in Part II and have received the following result from the official diagnostic laboratory which has carried out the neutralising antibody titration test (see note 4).

<table>
<thead>
<tr>
<th>Date of sampling (dd/mm/yy)</th>
<th>Name and address of the official diagnostic laboratory</th>
<th>Result of the antibody titration test (in International Units [IU]/ml)</th>
<th>Name (in capital letters) and signature of the veterinarian (see note 6)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

PERIOD OF VALIDITY OF SEROLOGICAL TESTING FOR INTERNATIONAL MOVEMENT (see note 3)

<table>
<thead>
<tr>
<th>from (dd/mm/yy)</th>
<th>to (dd/mm/yy)</th>
</tr>
</thead>
</table>

<table>
<thead>
<tr>
<th>Name (in capital letters) and signature of the Official Veterinarian</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
</tr>
</tbody>
</table>
VI. CLINICAL EXAMINATION (Rabies)

I, the undersigned declare herewith that I have examined on the date indicated below the animal described in Part II and have found it to be clinically healthy (see note 5).

<table>
<thead>
<tr>
<th>Date (dd/mm/yy)</th>
<th>Name (in capital letters) and signature of the veterinarian (see note 6)</th>
<th>Name (in capital letters) and signature of the Official Veterinarian</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
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</tbody>
</table>
Annex XVIII (contd)

NOTE

1. The identification number stated in the certificate should be identical to that which can be found on the animal. When electronic identification is used, the type of microchip and the name of the manufacturer should be specified.

2. Only vaccines that comply with the recommendations of the Terrestrial Manual should be used. Inactivated virus vaccines are authorized for international movements of dogs and cats.

3. In the case of a primary vaccination or re-vaccination, the animal should have been vaccinated not less than 6 months and not more than 1 year prior to its introduction into the importing country; the vaccination should have been carried out when the animal was at least 3 months old.

   In the case of a booster vaccination, the animal should have been vaccinated not more than 1 year prior to its introduction into the importing country.

4. The animal should have been subjected not less than 3 months and not more than 24 months prior to its introduction into the importing country, to a neutralizing antibody titration test. It should be carried out by an official diagnostic laboratory approved by the Competent Authority of the exporting country. The animal's serum should contain at least 0.5 International Units (IU)/ml.

5. The clinical examination referred to in Part VI of the certificate must be carried out within 48 hours of shipment.

The Competent Authority of the importing country may require the placing of the animals which do not comply with any of the above-mentioned conditions in a quarantine station located on its territory; the conditions of stay in quarantine are laid down by the legislation of the importing country.

6. If the veterinarian whose name and signature appear on the certificate is not an official veterinarian, his signature must be authenticated in the relevant column by the signature and stamp of an official veterinarian. The expression 'Official Veterinarian' means a civil service veterinarian or a specially appointed veterinarian, as authorised by the Veterinary Authority of the country.

7. If so required, the certificate should be written in the language of the importing country. In such circumstances, it should also be written in a language understood by the certifying veterinarian.
General provisions

For the purposes of the *Terrestrial Code*, the *incubation period* for vesicular stomatitis (VS) shall be 21 days.

Standards for diagnostic tests are described in the *Terrestrial Manual*.

VS free country

A country may be considered free from VS when:

1. VS is notifiable in the country;
2. no clinical, epidemiological or other evidence of VS has been found during the past 2 years.

Trade in commodities

*Veterinary Authorities* of countries shall consider whether there is a risk with regard to VS in accepting importation or transit through their territory, from other countries, of ruminants, swine, Equidae, and their semen and embryos.

Recommendations for importation from VS free countries

for domestic cattle, sheep, goats, pigs and horses

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the animals:

1. showed no clinical sign of VS on the day of shipment;
2. were kept in a VS free country since birth or for at least the past 21 days.

Recommendations for importation from VS free countries

for wild bovine, ovine, caprine, porcine and equine animals and deer

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the animals:

1. showed no clinical sign of VS on the day of shipment;
2. come from a VS free country;
if the country of origin has a common border with a country considered infected with VS:

3. were kept in a quarantine station for the 30 days prior to shipment and were subjected to a diagnostic test for VS with negative results at least 21 days after the commencement of quarantine;

4. were protected from insect vectors during quarantine and transportation to the place of shipment.

Article 8.15.6.

Recommendations for importation from countries considered infected with VS for domestic cattle, sheep, goats, pigs and horses

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of VS on the day of shipment;

2. either:
   
   a) were kept, since birth or for the past 21 days, in an establishment where no case of VS was officially reported during that period; or

   b) were kept in a quarantine station for the 30 days prior to shipment and were subjected to a diagnostic test for VS with negative results at least 21 days after the commencement of quarantine;

3. were protected from insect vectors during quarantine and transportation to the place of shipment.

Article 8.15.7.

Recommendations for importation from countries considered infected with VS for wild bovine, ovine, caprine, porcine and equine animals and deer

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of VS on the day of shipment;

2. were kept in a quarantine station for the 30 days prior to shipment and were subjected to a diagnostic test for VS with negative results at least 21 days after the commencement of quarantine;

3. were protected from insect vectors during quarantine and transportation to the place of shipment.

Article 8.15.8.

Recommendations for importation from VS free countries or zones for in vivo derived embryos of ruminants, swine and horses

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females were kept in an establishment located in a VS free country or zone at the time of collection;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.
Article 8.15.9.

Recommendations for importation from countries or zones considered infected with VS

for in vivo derived embryos of ruminants, swine and horses

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females:

   a) were kept for the 21 days prior to, and during, collection in an establishment where no case of VS was reported during that period;

   b) were subjected to a diagnostic test for VS, with negative results, within the 21 days prior to embryo collection;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.
In each country, official health control of bee diseases should include:

a) an organisation for permanent health surveillance;

b) approval of breeding apiaries for export trade;

c) measures for cleaning, disinfection and disinfestation of apicultural equipment;

d) rules precisely stating the requirements for issuing an international veterinary certificate.

Organisation for permanent official sanitary surveillance of apiaries

Permanent official sanitary surveillance of apiaries should be under the authority of the Veterinary Authority and should be performed either by representatives of this Authority or by representatives of an approved organisation, with the possible assistance of bee-keepers specially trained to qualify as "health inspectors and advisers".

The official surveillance service thus established should be entrusted with the following tasks:

1. visit apiaries:
   a) annual visits during the most appropriate periods for the detection of diseases;
   b) unexpected visits to apiaries where breeding or transport operations are carried out for trade or transfer to other regions, or any other purpose whereby diseases could be spread, as well as to apiaries located in the vicinity;
   c) special visits for sanitary surveillance to sectors where breeding apiaries have been approved for export purposes;

2. collect the samples required for the diagnosis of contagious diseases and despatch them to an official laboratory; the results of laboratory examinations should be communicated within the shortest delay to the Veterinary Authority;

3. apply hygiene measures, comprising, in particular, treatment of colonies of bees, as well as disinfection of the equipment and possibly the destruction of affected or suspect colonies and of the contaminated equipment so as to ensure rapid eradication of any outbreak of a contagious disease.

Conditions for approval of breeding apiaries for export trade

The apiaries should:
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1. be situated in the centre of an area defined as follows and in which:
   a) no case of varroosis has been reported for at least the past 2 years within a radius of 50 kilometres;
   b) no case of any other contagious disease of bees included in this Terrestrial Code has been reported for at least the past 8 months within a radius of 5 kilometres;

2. have received, for at least the past 2 years, visits by a health inspector and adviser, carried out at least 3 times a year (in spring, during the breeding period and in autumn), for the systematic examination of at least 10% of the hives containing bees and of all the apicultural equipment, and for the collection of samples to be sent to an official laboratory.

Bee-keepers must should:

3. immediately notify the Veterinary Authority of any suspicion of a contagious disease of bees in the breeding apiary and in other apiaries in the vicinity;

4. not introduce into the apiary any bee (including larval stages) or apicultural material or product originating from another apiary unless health control has been previously performed by the Veterinary Authority;

5. apply special breeding and despatch techniques to ensure protection against any outside contamination, especially for the breeding and sending of queen-bees and accompanying bees and to enable retesting in the importing country;

6. collect at least every 30 days, during the breeding and despatch period, samples from breeding material, brood-combs, queen-bees and bees (including possibly separately raised accompanying bees), to be sent to an official laboratory.


Conditions for sanitation and disinfection of apicultural equipment

Veterinary Authorities of exporting countries are requested to regulate the use of products and means for sanitation and disinfection of apicultural equipment in their own country, taking into account the following recommendations.

1. Any apicultural equipment kept in an establishment which has been recognised as being affected with a contagious disease of bees shall be subjected to sanitary measures ensuring the elimination of pathogens.

2. In all cases, these measures comprise the initial cleaning and scraping of the equipment, followed by sanitation or disinfection depending on the disease concerned.

3. The kind of equipment (hives, small hives, combs, extractor, small equipment, appliances for handling or storage) shall also be taken into account in the choice of procedures to be applied.

4. Infected or contaminated equipment which cannot be subjected to the above-mentioned measures must should be destroyed, preferably by burning. Any equipment in bad condition, especially hives, as well as larvae in combs affected with varroosis, American foulbrood or European foulbrood, must should be destroyed by burning.

5. The products and means used for sanitation and disinfection shall be recognised as being effective by the Veterinary Authority. They shall be used in such a manner as to exclude any risk of contaminating the equipment which could eventually affect the health of bees or adulterate the products of the hive.

6. When these procedures are not performed, the products shall be kept away from the bees and any contact with apicultural equipment and products must should be prevented.
7. Waste water from the cleaning, sanitation and disinfection of apicultural equipment shall be kept away from the bees at all times and disposed of in a sewer or in an unused well.

Article 4.14.5.

Preparation of the international veterinary certificate for export

This Certificate covers hives containing bees, swarms, consignments of bees (worker bees or drones), queen bees (with accompanying bees), brood-combs, royal cells, etc.

This document shall be prepared in accordance with the model contained in Chapter 5.10.

— text deleted
CHAPTER 9.1.

ACARAPISOSIS OF HONEY BEES

Article 9.1.1.

General provisions

For the purposes of this Chapter, acarapisosis, acarine disease or tracheal mite infestation is a disease of the adult honey bee *Apis mellifera* L., and possibly of other *Apis* species (such as *Apis cerana*). It is caused by the Tarsonemid mite *Acarapis woodi* (Rennie). The mite is an internal obligate parasite of the respiratory system, living and reproducing mainly in the large prothoracic trachea of the bee. Early signs of infection normally go unnoticed, and only when infection is heavy does it become apparent; this is generally in the early spring. The infection spreads by direct contact from adult bee to adult bee, with newly emerged bees under 10 days old being the most susceptible. The mortality rate may range from moderate to high.

Standards for diagnostic tests are described in the Terrestrial Manual.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 9.1.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the acarapisosis status of the honey bee population of the exporting country or zone.

Article 9.1.2.

Trade in Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any acarapisosis related conditions, regardless of the acarapisosis status of the honey bee population of the exporting country or zone:

1. honey bee semen and honey bee venom;
2. used equipment associated with beekeeping;
3. extracted honey, pollen, propolis, royal jelly for human consumption, and processed beeswax, honey bee-collected pollen, propolis and royal jelly.

When authorising import or transit of other commodities listed in this Chapter, Veterinary Authorities should require the conditions prescribed in this Chapter relevant to the acarapisosis status of the honey bee population of the exporting country or zone.

Article 9.1.3.

Determination of the acarapisosis status of a country or zone/compartment

The acarapisosis status of a country or zone/compartment (under study) can only be determined after considering the following criteria:

1. a risk assessment has been conducted, identifying all potential factors for acarapisosis occurrence and their historic perspective;
2. acarapisosis should be notifiable in the whole country or zone/compartment (under study) and all clinical signs suggestive of acarapisosis should be subjected to field and laboratory investigations;
3. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of acarapisosis;

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4. the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees should have current knowledge of, and authority over, all domesticated apiaries in the whole country.

Article 9.1.4.

Country or zone/compartment (under study) free from acarapisosis

1. Historically free status

A country or zone/compartment (under study) may be considered free from acarapisosis after conducting a risk assessment as referred to in Article 9.1.3. but without formally applying a specific surveillance programme if the country or zone/compartment (under study) complies with the provisions of Chapter 1.4.

2. Free status as a result of an eradication programme

A country or zone/compartment (under study) which does not meet the conditions of point 1 above may be considered free from acarapisosis after conducting a risk assessment as referred to in Article 9.1.3. and when:

a) the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees has current knowledge of, and authority over, all domesticated apiaries existing in the country or zone/compartment (under study);

b) acarapisosis is notifiable in the whole country or zone/compartment (under study), and any clinical cases suggestive of acarapisosis are subjected to field and laboratory investigations;

c) for the 3 years following the last reported case of acarapisosis, annual surveys supervised by the Veterinary Authority, with negative results, have been carried out on a representative sample of apiaries in the country or zone/compartment (under study) to provide a confidence level of at least 95% of detecting acarapisosis if at least 1% of the apiaries were infected at a within-apiary prevalence rate of at least 5% of the hives; such surveys may be targeted towards apiaries, areas and seasons with a higher likelihood of disease;

d) to maintain free status, an annual survey supervised by the Veterinary Authority, with negative results, is carried out on a representative sample of apiaries in the country or zone/compartment (under study) to indicate that there has been no new cases; such surveys may be targeted towards areas with a higher likelihood of disease;

e) (under study) there is no self-sustaining feral population of A. mellifera or other possible host species in the country or zone/compartment (under study);

f) the importation of the commodities listed in this Chapter into the country or zone/compartment (under study) is carried out in conformity with the recommendations of this Chapter.

Article 9.1.5.

Recommendations for the importation of live queen honey bees, worker bees and drones with or without associated brood combs

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the bees come from a country or zone/compartment (under study) free from acarapisosis or the apiary meets the conditions prescribed in Article 4.14.3.
Recommendations for the importation of eggs, larvae and pupae of honey bees

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. were sourced from an officially free country or zone/compartment (under study); or
2. were examined by an official laboratory and declared free of all life stages of *A. woodi*; or
3. have originated from queens in a quarantine station and were examined microscopically and found free of all life stages of *A. woodi*.
CHAPTER 9.2.

AMERICAN FOULBROOD OF HONEY BEES

Article 9.2.1.

General provisions

For the purposes of this Chapter, American foulbrood is a disease of the larval and pupal stages of the honey bee *Apis mellifera* and other *Apis* spp., and occurs in most countries where such bees are kept. *Paenibacillus larvae*, the causative organism, is a bacterium that can produce over one billion spores in each infected larva. The spores are very long-living and extremely resistant to heat and chemical agents, and only the spores are capable of inducing the disease.

Combs of infected *apiaries* may show distinctive clinical signs which can allow the disease to be diagnosed in the field. However, subclinical infections are common and require laboratory diagnosis.

For the purposes of the *Terrestrial Code*, the incubation period for American foulbrood shall be 15 days (not including the wintering period which may vary according to country).

Standards for diagnostic tests are described in the *Terrestrial Manual*.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 9.2.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the American foulbrood status of the honey bee population of the *exporting country or zone*.

Article 9.2.2.

Trade in Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any American foulbrood related conditions, regardless of the American foulbrood status of the honey bee population of the *exporting country or zone*:

1. honey bee semen;
2. honey bee venom.

When authorising import or transit of other commodities listed in this Chapter, Veterinary Authorities should require the conditions prescribed in this Chapter relevant to the American foulbrood status of the honey bee population of the *exporting country or zone*.

Article 9.2.3.

Determination of the American foulbrood status of a country or zone/compartment

The American foulbrood status of a country or zone/compartment (under study) can only be determined after considering the following criteria:

1. a risk assessment has been conducted, identifying all potential factors for American foulbrood occurrence and their historic perspective;
2. American foulbrood should be notifiable in the whole country or zone/compartment (under study) and all clinical signs suggestive of American foulbrood should be subjected to field and/or laboratory investigations;
3. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of American foulbrood;

4. the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees should have current knowledge of, and authority over, all domesticated apiaries in the country.

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**Country or zone/compartment (under study) free from American foulbrood**

1. **Historically free status**

   A country or zone/compartment (under study) may be considered free from the disease after conducting a risk assessment as referred to in Article 9.2.3. but without formally applying a specific surveillance programme if the country or zone/compartment (under study) complies with the provisions of Chapter 1.4.

2. **Free status as a result of an eradication programme**

   A country or zone/compartment (under study) which does not meet the conditions of point 1 above may be considered free from American foulbrood after conducting a risk assessment as referred to in Article 9.2.3. and when:

   a) the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees has current knowledge of, and authority over, all domesticated apiaries existing in the country or zone/compartment (under study);

   b) American foulbrood is notifiable in the whole country or zone/compartment (under study), and any clinical cases suggestive of American foulbrood are subjected to field and/or laboratory investigations;

   c) for the 5 years following the last reported isolation of the American foulbrood agent, annual surveys supervised by the Veterinary Authority, with negative results, have been carried out on a representative sample of apiaries in the country or zone/compartment (under study) to provide a confidence level of at least 95% of detecting American foulbrood if at least 1% of the apiaries were infected at a within-apiary prevalence rate of at least 5% of the hives; such surveys may be targeted towards areas with the last reported isolation of the American foulbrood agent;

   d) to maintain free status, an annual survey supervised by the Veterinary Authority, with negative results, is carried out on a representative sample of hives in the country or zone/compartment (under study) to indicate that there has been no new isolations; such surveys may be targeted towards areas with a higher likelihood of isolation;

   e) (under study) there is no self-sustaining feral population of A. mellifera or other possible host species in the country or zone/compartment (under study);

   f) all equipment associated with previously infected apiaries has been sterilised or destroyed;

   g) the importation of the commodities listed in this Chapter into the country or zone/compartment (under study) is carried out in conformity with the recommendations of this Chapter.

**Recommendations for the importation of live queen honey bees, worker bees and drones with or without associated brood combs**

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the bees come from a country or zone/compartment (under study) officially free from American foulbrood or the apiary meets the conditions prescribed in Article 4.14.3.
Article 9.2.6.

Recommendations for the importation of eggs, larvae and pupae of honey bees

_Veterinary Authorities of importing countries_ should require the presentation of an _international veterinary certificate_ attesting that the products:

1. were sourced from a free country or _zone/compartment_ (under study); or
2. have been isolated from queens in a _quarantine station_, and all workers which accompanied the queen or a representative sample of eggs or larvae were examined for the presence of _P. larvae_ by bacterial culture or PCR in accordance with the _Terrestrial Manual._

Article 9.2.7.

Recommendations for the importation of used equipment associated with beekeeping

_Veterinary Authorities of importing countries_ should require the presentation of an _international veterinary certificate_ attesting that the equipment was sterilised under the supervision of the _Veterinary Authority_ by either immersion in 1% sodium hypochlorite for at least 30 minutes (suitable only for non-porous materials such as plastic and metal), gamma irradiation using a cobalt-60 source at a dose rate of 10 kGy, or processing to ensure the destruction of both bacillary and spore forms of _P. larvae_, in conformity with one of the procedures referred to in Chapter X.X. _recommended by the OIE_ (under study).

Article 9.2.8.

Recommendations for the importation of honey, honey bee-collected pollen, beeswax, propolis and royal jelly

_Veterinary Authorities of importing countries_ officially free from American foulbrood should require the presentation of an _international veterinary certificate_ attesting that the products:

1. were collected in a country or _zone/compartment_ (under study) free from American foulbrood; or
2. have been processed to ensure the destruction of both bacillary and spore forms of _P. larvae_, in conformity with one of the procedures referred to in Chapter X.X. _recommended by the OIE_ (under study).
CHAPTER 9.3.

EUROPEAN FOULBROOD OF HONEY BEES

Article 9.3.1.

General provisions

For the purposes of this Chapter, European foulbrood is a disease of the larval and pupal stages of the honey bee Apis mellifera and other Apis spp., and occurs in most countries where such bees are kept. The causative agent is the non-sporulating bacterium Melissococcus plutonius. Subclinical infections are common and require laboratory diagnosis. Infection remains enzootic because of mechanical contamination of the honeycombs. Recurrences of disease can therefore be expected in subsequent years.

For the purposes of the Terrestrial Code, the incubation period for European foulbrood shall be 15 days (not including the wintering period which may vary according to country).

Standards for diagnostic tests are described in the Terrestrial Manual.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 9.3.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the European foulbrood status of the honey bee population of the exporting country or zone.

Article 9.3.2.

Trade in Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any European foulbrood related conditions, regardless of the European foulbrood status of the honey bee population of the exporting country or zone:

1. honey bee semen;
2. honey bee venom.

When authorising import or transit of other commodities listed in this Chapter, Veterinary Authorities should require the conditions prescribed in this Chapter relevant to the European foulbrood status of the honey bee population of the exporting country or zone.

Article 9.3.3.

Determination of the European foulbrood status of a country or zone/compartment

The European foulbrood status of a country or zone/compartment (under study) can only be determined after considering the following criteria:

1. a risk assessment has been conducted, identifying all potential factors for European foulbrood occurrence and their historic perspective;
2. European foulbrood should be notifiable in the whole country or zone/compartment (under study) and all clinical signs suggestive of European foulbrood should be subjected to field and laboratory investigations;
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3. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of European foulbrood;

4. the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees should have current knowledge of, and authority over, all apiaries in the whole country.

Article 9.3.4.

Country or zone/compartment (under study) free from European foulbrood

1. Historically free status

A country or zone/compartment (under study) may be considered free from the disease after conducting a risk assessment as referred to in Article 9.3.3, but without formally applying a specific surveillance programme if the country or zone/compartment (under study) complies with the provisions of Chapter 1.4.

2. Free status as a result of an eradication programme

A country or zone/compartment (under study) which does not meet the conditions of point 1 above may be considered free from European foulbrood after conducting a risk assessment as referred to in Article 9.3.3, and when:

a) the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees has current knowledge of, and authority over, all domesticated apiaries existing in the country or zone/compartment (under study);

b) European foulbrood is notifiable in the whole country or zone/compartment (under study), and any clinical cases suggestive of European foulbrood are subjected to field and laboratory investigations;

c) for the 3 years following the last reported isolation of the European foulbrood agent, an annual survey supervised by the Veterinary Authority, with negative results, has been carried out on a representative sample of apiaries in the country or zone/compartment (under study) to provide a confidence level of at least 95% of detecting European foulbrood if at least 1% of the apiaries were infected at a within-apiary prevalence rate of at least 5% of the hives; such surveys may be targeted towards areas with the last reported isolation of the European foulbrood agent;

d) to maintain free status, an annual survey supervised by the Veterinary Authority, with negative results, is carried out on a representative sample of hives in the country or zone/compartment (under study) to indicate that there has been no new isolations; such surveys may be targeted towards areas with a higher likelihood of isolation;

e) (under study) there is no self-sustaining feral population of A. mellifera or other possible host species in the country or zone/compartment (under study);

f) the importation of the commodities listed in this Chapter into the country or zone/compartment (under study) is carried out in conformity with the recommendations of this Chapter.

Article 9.3.5.

Recommendations for the importation of live queen honey bees, worker bees and drones with or without associated brood combs

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the bees come from a country or zone/compartment (under study) free from European foulbrood or the apiary meets the conditions prescribed in Article 4.14.3.
Recommendations for the importation of eggs, larvae and pupae of honey bees

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. were sourced from a free country or zone/compartment (under study); or
2. have been isolated from queens in a quarantine station, and all workers which accompanied the queen or a representative sample of eggs or larvae were examined for the presence of *M. plutonius* by bacterial culture or PCR in accordance with the *Terrestrial Manual*.

Recommendations for the importation of used equipment associated with beekeeping

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the equipment was sterilised under the supervision of the Veterinary Authority by either immersion in 0.5% sodium hypochlorite for at least 20 minutes (suitable only for non-porous materials such as plastic and metal), gamma irradiation using a cobalt-60 source at a dose rate of 10 kGy, or processing to ensure the destruction of *M. plutonius*, in conformity with one of the procedures referred to in Chapter recommended by the OIE (under study).

Recommendations for the importation of honey, honey bee-collected pollen, beeswax, propolis and royal jelly

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. were collected in a country or zone/compartment (under study) free from European foulbrood; or
2. have been processed to ensure the destruction of *M. plutonius*, in conformity with one of the procedures referred to in Chapter recommended by the OIE (under study).
General provisions

For the purposes of this Chapter, small hive beetle (SHB) is an infestation of social bee colonies by the beetle *Aethina tumida*, which is a free-living predator and scavenger affecting populations of the honey bee *Apis mellifera* L. It can also invade bumble bee *Bombus terrestris* and stingless bee *Trigona carbonaria* colonies under experimental conditions, and although infestation has not been demonstrated in wild populations, *Bombus* spp. must also be considered to be susceptible to infestation.

The adult beetle is attracted to bee colonies to reproduce, although it can survive and reproduce independently in other natural environments, using other food sources, including certain types of fruit. Hence once it is established within a localised environment, it is extremely difficult to eradicate.

The life cycle of *A. tumida* begins with the adult beetle laying eggs within infested hives. These are usually laid in irregular masses in crevices or brood combs. After 2–6 days, the eggs hatch and the emerging larvae begin to feed voraciously on brood comb, bee eggs, pollen and honey within the hive. The SHB has a high reproductive potential. Each female can produce about 1,000 eggs in its 4–6 months of life. At maturation (approximately 10–29 days after hatching), the larvae exit the hive and burrow into soil around the hive entrance. Adult beetles emerge after an average of 3.4 weeks, although pupation can take between 8 and 60 days depending on temperature and moisture levels.

The life span of an adult beetle depends on environmental conditions such as temperature and humidity but, in practice, adult beetles can live for at least 6 months and, in favourable reproductive conditions, the female is capable of laying new egg batches every 5–12 weeks. The beetle is able to survive at least 2 weeks without food and 50 days on brood combs.

Early signs of infestation and reproduction in the debris may go unnoticed, but the growth of the beetle population is rapid, leading to high bee mortality in the hive. When the bees cannot prevent beetle mass reproduction on the combs, this leads to abandonment and/or collapse of the colony. Because *A. tumida* can be found and can thrive within the natural environment, and can fly up to 6–13 km from its nest site, it is capable of dispersing rapidly and directly invading new colonising hives. Dispersal of beetles includes following or accompanying swarms of bees. Spread of infestation does not require contact between adult bees. However, movement of adult bees, honeycomb and other apiculture products and used equipment associated with bee-keeping may all cause infestations to spread to previously unaffected colonies.

Standards for diagnostic tests are described in the *Terrestrial Manual*.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 9.4.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the *A. tumida* status of the honey bee and other social bee population of the exporting country or zone.

Article 9.4.2.

Trade in Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any small hive beetle infestation related conditions, regardless of the *A. tumida* status of the honey bee and bumble bee population of the exporting country or zone.
Annex XX (contd)

1. honey bee semen and honey bee venom;

2. packaged extracted honey for human consumption, refined or rendered beeswax, propolis and frozen or dried royal jelly.

When authorising import or transit of other commodities listed in this Chapter, Veterinary Authorities should require the conditions prescribed in this Chapter relevant to the A. tumida status of the honey bee and bumble bee population of the exporting country or zone.

Article 9.4.3.

Determination of the A. tumida status of a country or zone

The A. tumida status of a country or zone can only be determined after considering the following criteria:

1. A. tumida infestation should be notifiable in the whole country, and all signs suggestive of A. tumida infestation should be subjected to field and laboratory investigations;

2. on-going awareness and training programmes should be in place to encourage reporting of all cases suggestive of A. tumida infestation;

3. the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees should have current knowledge of, and authority over, all domesticated apiaries in the country.

Article 9.4.4.

Country or zone free from A. tumida

1. Historically free status

A country or zone may be considered free from the pest after conducting a risk assessment as referred to in Article 9.4.3. but without formally applying a specific surveillance programme if the country or zone complies with the provisions of Chapter 1.4.

2. Free status as a result of an eradication programme

A country or zone which does not meet the conditions of point 1 above may be considered free from A. tumida infestation after conducting a risk assessment as referred to in Article 9.4.3. and when:

a) the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees has current knowledge of, and authority over, all domesticated apiaries existing in the country or zone;

b) A. tumida infestation is notifiable in the whole country or zone, and any clinical cases suggestive of A. tumida infestation are subjected to field and laboratory investigations; a contingency plan is in place describing controls and inspection activities;

c) for the 5 years following the last reported case of A. tumida infestation, an annual survey supervised by the Veterinary Authority, with negative results, has been carried out on a representative sample of apiaries in the country or zone to provide a confidence level of at least 95% of detecting A. tumida infestation if at least 1% of the apiaries were infested at a within-apiary prevalence rate of at least 5% of the hives; such surveys may be targeted towards areas with a higher likelihood of infestation;

d) to maintain free status, an annual survey supervised by the Veterinary Authority, with negative results, is carried out on a representative sample of apiaries to indicate that there have been no new cases; such surveys may be targeted towards areas with a higher likelihood of infestation;
e) all equipment associated with previously infested apiaries has been destroyed, or cleaned and sterilised to ensure the destruction of *A. tumida* spp., in conformity with one of the procedures referred to in Chapter X.X. recommended by the OIE (under study);

f) the soil and undergrowth in the immediate vicinity of all infested apiaries has been treated with a soil drench or similar suitable treatment that is efficacious in destroying incubating *A. tumida* larvae and pupae;

g) the importation of the commodities listed in this Chapter into the country or zone is carried out, in conformity with the recommendations of this Chapter.

Article 9.4.5.

Recommendations for the importation of individual consignments containing a single live queen honey bee or queen bumble bee, accompanied by a small number of associated attendants (a maximum of 20 attendants per queen)

*Veterinary Authorities* of importing countries should require the presentation of an *international veterinary certificate* attesting that

1. the bees come from a country or zone officially free from *A. tumida* infestation.

OR

*Veterinary Authorities* of importing countries should require the presentation of an *international veterinary certificate* including an attestation from the *Veterinary Authority* of the exporting third country stating that

2. the bees come from hives or colonies which were inspected immediately prior to dispatch and show no signs or suspicion of the presence of *A. tumida* or its eggs, larvae or pupae; and

3. the bees come from an area of at least 100 km radius where no apiary has been subject to any restrictions associated with the occurrence of *A. tumida* for the previous 6 months; and

4. the bees and accompanying packaging presented for export have been thoroughly and individually inspected and do not contain *A. tumida* or its eggs, larvae or pupae; and

5. the consignment of bees is covered with fine mesh through which a live beetle cannot enter.

Article 9.4.6.

Recommendations for the importation of live worker bees, drone bees or bee colonies with or without associated brood combs or for live bumble bees

*Veterinary Authorities* of importing countries should require the presentation of an *international veterinary certificate* attesting that:

1. the bees come from a country or zone officially free from *A. tumida* infestation; and

2. the bees and accompanying packaging presented for export have been inspected and do not contain *A. tumida* or its eggs, larvae or pupae; and

3. the consignment of bees is covered with fine mesh through which a live beetle cannot enter.
Recommendations for the importation of eggs, larvae and pupae of honey bees or bumble bees

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. the products were sourced from a country or zone free from A. tumida infestation;

OR

2. the products have been bred and kept under a controlled environment within a recognised establishment which is supervised and controlled by the Veterinary Authority;

3. the establishment was inspected immediately prior to dispatch and all eggs, larvae and pupae show no clinical signs or suspicion of the presence of A. tumida or its eggs or larvae or pupae, and

4. the packaging material, containers, accompanying products and food are new and all precautions have been taken to prevent contamination with A. tumida or its eggs, larvae or pupae.

Recommendations for the importation of used equipment associated with beekeeping

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that:

1. the equipment:

   EITHER

   a) comes from a country or zone free from A. tumida infestation; and

   b) contains no live honey bees or bee brood;

   OR

   c) contains no live honey bees or bee brood; and

   d) has been thoroughly cleaned, and treated to ensure the destruction of A. tumida spp., in conformity with one of the procedures referred to in Chapter X.X. recommended by the OIE (under study); and

   AND

2. all precautions have been taken to prevent infestation/contamination.

Recommendations for the importation of honey-bee collected pollen and beeswax (in the form of honeycomb)

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that:
1. the products:

   EITHER

   a) comes from a country or zone free from *A. tumida* infestation; and

   b) contains no live honey bees or bee brood;

   OR

   c) contains no live honey bees or bee brood; and

   d) has been thoroughly cleaned, and treated to ensure the destruction of *A. tumida* spp., in conformity with one of the procedures referred to in Chapter X.X, recommended by the OIE (under study);

   AND

2. all precautions have been taken to prevent infestation/contamination.

   Article 9.4.10.

**Recommendations for the importation of comb honey**

*Veterinary Authorities* of importing countries should require the presentation of an *international veterinary certificate* attesting that the products:

1. comes from a country or zone free from *A. tumida* infestation; and

2. contains no live honey bees or bee brood;

OR

3. were subjected to a treatment at a temperature of -12°C or lower in the core of the product during at least 24 hours.
CHAPTER 9.5.

TROPILAEELAPS INFESTATION OF HONEY BEES

Article 9.5.1.

General provisions

For the purposes of this Chapter, Tropilaelaps infestation of the honey bee *Apis mellifera* L. is caused by the mites *Tropilaelaps clareae*, *T. koenigerum*, *T. thail* and *T. mercedesae*. The mite is an ectoparasite of brood of *Apis mellifera* L., *Apis laboriosa* and *Apis dorsata*, and cannot survive for periods of more than 14 days away from bee brood.

Early signs of infection normally go unnoticed, but the growth in the mite population is rapid leading to high hive mortality. The infection spreads by direct contact from adult bee to adult bee, and by the movement of infested bees and bee brood. The mite can also act as a vector for viruses of the honey bee.

Standards for diagnostic tests are described in the Terrestrial Manual.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 9.5.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the Tropilaelaps status of the honey bee population of the exporting country or zone.

Article 9.5.2.

Trade in Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any Tropilaelaps infestation related conditions, regardless of the Tropilaelaps status of the honey bee population of the exporting country or zone:

1. honey bee semen, honey bee eggs and honey bee venom;
2. extracted honey, pollen, propolis, royal jelly for human consumption and processed beeswax (not in the form of honeycomb).

When authorising import or transit of other commodities listed in this Chapter, Veterinary Authorities should require the conditions prescribed in this Chapter relevant to the Tropilaelaps status of the honey bee population of the exporting country or zone.

Article 9.5.3.

Determination of the Tropilaelaps status of a country or zone/compartment

The Tropilaelaps status of a country or zone/compartment (under study) can only be determined after considering the following criteria:

1. a risk assessment has been conducted, identifying all potential factors for Tropilaelaps occurrence and their historic perspective;
2. Tropilaelaps infestation should be notifiable in the whole country or zone/compartment (under study) and all clinical signs suggestive of Tropilaelaps infestation should be subjected to field and laboratory investigations;
3. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of Tropilaelaps infestation;
Annex XX (contd)

4. the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees should have current knowledge of, and authority over, all domesticated apiaries in the country.

Article 9.5.4.

Country or zone/compartment (under study) free from *Tropilaelaps* spp

1. **Historically free status**

A country or zone/compartment (under study) may be considered free from the disease after conducting a risk assessment as referred to in Article 9.5.3. but without formally applying a specific surveillance programme if the country or zone/compartment (under study) complies with the provisions of Chapter 1.4.

2. **Free status as a result of an eradication programme**

A country or zone/compartment (under study) which does not meet the conditions of point 1 above may be considered free from *Tropilaelaps* infestation after conducting a risk assessment as referred to in Article 9.5.3. and when:

a) the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees has current knowledge of, and authority over, all domesticated apiaries existing in the country or zone/compartment (under study);

b) *Tropilaelaps* infestation is notifiable in the whole country or zone/compartment (under study), and any clinical cases suggestive of *Tropilaelaps* infestation are subjected to field and laboratory investigations;

c) for the 3 years following the last reported case of *Tropilaelaps* infestation, an annual survey supervised by the Veterinary Authority, with negative results, have been carried out on a representative sample of apiaries in the country or zone/compartment (under study) to provide a confidence level of at least 95% of detecting *Tropilaelaps* infestation if at least 1% of the apiaries were infected at a within-apiary prevalence rate of at least 5% of the hives; such surveys may be targeted towards areas with a higher likelihood of infestation;

d) to maintain free status, an annual survey supervised by the Veterinary Authority, with negative results, is carried out on a representative sample of apiaries in the country or zone/compartment (under study) to indicate that there has been no new cases; such surveys may be targeted towards areas with a higher likelihood of disease;

e) (under study) there is no self-sustaining feral population of *A. mellifera, A. dorsata* or *A. laboriosa*, or other possible host species in the country or zone/compartment (under study);

f) the importation of the commodities listed in this Chapter into the country or zone/compartment (under study) is carried out, in conformity with the recommendations of this Chapter.

Article 9.5.5.

**Recommendations for the importation of live queen honey bees, worker bees and drones with associated brood combs**

*Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the bees come from a country or zone/compartment (under study) officially free from *Tropilaelaps* infestation or the apiary meets the conditions prescribed in Article 4.14.3.*
Recommendations for the importation of live queen honey bees, worker bees and drones without associated brood combs

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the bees have been held in isolation from brood and bees with access to brood, for a period of at least 21 days.

Article 9.5.7.

Recommendations for the importation of used equipment associated with beekeeping

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the equipment:

1. comes from a country or zone/compartment (under study) free from *Tropilaelaps* infestation; or

2. contains no live honey bees or bee brood and has been held away from contact with live honey bees for at least 21 days prior to shipment; or

3. has been treated to ensure the destruction of *Tropilaelaps* spp., in conformity with one of the procedures referred to in Chapter X.X. recommended by the OIE (under study).

Article 9.5.8.

Recommendations for the importation of honey-bee collected pollen, beeswax (in the form of honeycomb), comb honey and propolis

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. come from a country or zone/compartment (under study) free from *Tropilaelaps* infestation; or

2. contain no live honey bees or bee brood and has been held away from contact with live honey bees for at least 21 days prior to shipment; or

3. have been treated to ensure the destruction of *Tropilaelaps* spp., in conformity with one of the procedures referred to in Chapter X.X. recommended by the OIE (under study).
CHAPTER 9.6.

VARROOSIS OF HONEY BEES

Article 9.6.1.

General provisions

For the purposes of this Chapter, varroosis is a disease of the honey bee *Apis mellifera* L. It is caused by the Korea and Japan haplotypes of the mite *Varroa destructor*, the original hosts of which are the Korea and Japan haplotypes of *Apis cerana* (under study). The mite is an ectoparasite of adults and brood of *Apis mellifera* L. During its life cycle, sexual reproduction occurs inside the honey bee brood cells. Early signs of infection normally go unnoticed, and only when infection is heavy does it become apparent. The infection spreads by direct contact from adult bee to adult bee, and by the movement of infested bees and bee brood. The mite can also act as a vector for viruses of the honey bee.

The number of parasites steadily increases with increasing brood activity and the growth of the bee population, especially late in the season when clinical signs of infestation can first be recognised. The life span of an individual mite depends on temperature and humidity but, in practice, it can be said to last from some days to a few months.

Standards for diagnostic tests are described in the *Terrestrial Manual*.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 9.6.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the varroosis status of the honey bee population of the exporting country or zone.

Article 9.6.2.

Trade in Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any varroosis related conditions, regardless of the varroosis status of the honey bee population of the exporting country or zone:

1. honey bee semen, honey bee eggs and honey bee venom;
2. extracted honey, pollen, propolis, royal jelly for human consumption and processed beeswax (not in the form of honeycomb).

When authorising import or transit of other commodities listed in this Chapter, Veterinary Authorities should require the conditions prescribed in this Chapter relevant to the varroosis status of the honey bee population of the exporting country or zone.

Article 9.6.3.

Determination of the varroosis status of a country or zone/compartment

The varroosis status of a country or zone/compartment (under study) can only be determined after considering the following criteria:

1. a risk assessment has been conducted, identifying all potential factors for varroosis occurrence and their historic perspective;
Annex XX (contd)

2. varroosis should be notifiable in the whole country or zone/compartment (under study) and all clinical signs suggestive of varroosis should be subjected to field and laboratory investigations;

3. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of varroosis;

4. the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees should have current knowledge of, and authority over, all domesticated apiaries in the country.

Article 9.6.4.

Country or zone/compartment (under study) free from varroosis

1. Historically free status

A country or zone/compartment (under study) may be considered free from the disease after conducting a risk assessment as referred to in Article 9.6.3. but without formally applying a specific surveillance programme (historical freedom) if the country or zone/compartment (under study) complies with the provisions of Chapter 1.4.

2. Free status as a result of an eradication programme

A country or zone/compartment (under study) which does not meet the conditions of point 1 above may be considered free from varroosis after conducting a risk assessment as referred to in Article 9.6.3. and when:

a) the Veterinary Authority or other Competent Authority with responsibility for reporting and control of diseases of honey bees has current knowledge of, and authority over, all domesticated apiaries existing in the country or zone/compartment (under study);

b) varroosis is notifiable in the whole country or zone/compartment (under study), and any clinical cases suggestive of varroosis are subjected to field and laboratory investigations;

c) for the 3 years following the last reported case of varroosis, an annual survey supervised by the Veterinary Authority, with negative results, have been carried out on a representative sample of apiaries in the country or zone/compartment (under study) to provide a confidence level of at least 95% of detecting varroosis if at least 1% of the apiaries were infected at a within-apiary prevalence rate of at least 5% of the hives; such surveys may be targeted towards areas with a higher likelihood of disease;

d) to maintain free status, an annual survey supervised by the Veterinary Authority, with negative results, is carried out on a representative sample of apiaries in the country or zone/compartment (under study) to indicate that there has been no new cases; such surveys may be targeted towards areas with a higher likelihood of disease;

e) (under study) there is no self-sustaining feral population of A. mellifera, the Korea and Japan haplotypes of Apis cerana or other possible host species in the country or zone/compartment (under study);

f) the importation of the commodities listed in this Chapter into the country or zone/compartment (under study) is carried out in conformity with the recommendations of this Chapter.
Apiary free from varroosis

1. The apiary is located in a country or zone complying with the requirements in points 2. a) b) and f) of Article 9.6.4.;
2. the apiary should be situated in an area with a radius of 50 kilometres in which no case of varroosis has been reported for at least the past 2 years; and
3. the apiary meets the conditions prescribed in Article 4.14.3.

Recommendations for the importation of live queen honey bees, worker bees and drones with or without associated brood combs

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the bees come from a country or zone/compartment (under study) officially free from varroosis or the apiary meets the conditions prescribed in Article 9.6.4.bis.

Recommendations for the importation of larvae and pupae of honey bees

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:
1. were sourced from a free country or zone/compartment (under study); or
2. have originated from queens in a quarantine station and were inspected and found free of Varroa destructor.

(wait for member comments to modify larvae and pupae articles)

Recommendations for the importation of used equipment associated with beekeeping

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the equipment:
1. comes from a country or zone/compartment (under study) free from varroosis; or
2. contains no live honey bees or bee brood and has been held away from contact with live honey bees for at least 21 days prior to shipment; or
3. has been treated to ensure the destruction of Varroa destructor, in conformity with one of the procedures referred to in Chapter X.X., recommended by the OIE (under study).

Recommendations for the importation of honey-bee collected pollen, beeswax (in the form of honeycomb), comb honey and propolis

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:
Annex XX (contd)

1. come from a country or zone/compartment (under study) free from varroosis; or

2. contain no live honey bees or bee brood and has been held away from contact with live honey bees for at least 21 days prior to shipment; or

3. have been treated to ensure the destruction of *Varroa destructor*, in conformity with one of the procedures referred to in Chapter X.X. recommended by the OIE (under study).

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CHAPTER 10.4.

AVIAN INFLUENZA

Article 10.4.1.

General provisions

1. For the purposes of international trade, the Terrestrial Code, avian influenza in its notifiable form (NAI) is defined as an infection of poultry caused by any influenza A virus of the H5 or H7 subtypes or by any AI virus with an intravenous pathogenicity index (IVPI) greater than 1.2 (or as an alternative at least 75% mortality) as described below. NAI viruses can be divided into highly pathogenic notifiable avian influenza (HPNAI) and low pathogenicity notifiable avian influenza (LPNAI):

   a) HPNAI viruses have an IVPI in 6-week-old chickens greater than 1.2 or, as an alternative, cause at least 75% mortality in 4-to 8-week-old chickens infected intravenously. H5 and H7 viruses which do not have an IVPI of greater than 1.2 or cause less than 75% mortality in an intravenous lethality test should be sequenced to determine whether multiple basic amino acids are present at the cleavage site of the haemagglutinin molecule (HA0); if the amino acid motif is similar to that observed for other HPNAI isolates, the isolate being tested should be considered as HPNAI;

   b) LPNAI are all influenza A viruses of H5 and H7 subtype that are not HPNAI viruses.

2. Poultry is defined as ‘all domesticated birds, including backyard poultry, used for the production of meat or eggs for consumption, for the production of other commercial products, for restocking supplies of game, or for breeding these categories of birds, as well as fighting cocks used for any purpose’.

   Birds that are kept in captivity for any reason other than those reasons referred to in the preceding paragraph, including those that are kept for shows, races, exhibitions, competitions or for breeding or selling these categories of birds as well as pet birds, are not considered to be poultry.

3. For the purposes of the Terrestrial Code, the incubation period for NAI shall be 21 days.

4. For the purposes of international trade, this chapter deals not only with the occurrence of clinical signs caused by NAI virus, but also with the presence of infection with NAI virus in the absence of clinical signs.

4. For the purposes of international trade, a Member should not impose immediate bans on the trade in poultry commodities in response to a notification, according to Article 1.2.3. of the Terrestrial Code, of infection with HPAI and LPAI virus in birds other than poultry, including wild birds.

5. Antibodies to H5 or H7 subtype of NAI virus, which have been detected in poultry and are not a consequence of vaccination, have to be immediately investigated. In the case of isolated serological positive results, NAI infection may be ruled out on the basis of a thorough epidemiological and laboratory investigation that does not demonstrate further evidence of NAI infection.

6. The following defines the occurrence of infection with NAI virus:

   a) HPNAI virus has been isolated and identified as such or viral RNA specific for HPNAI has been detected in poultry or a product derived from poultry; or

   b) LPNAI virus has been isolated and identified as such or viral RNA specific for LPNAI has been detected in poultry or a product derived from poultry.
7. For the purposes of the Terrestrial Code, ‘NAI free establishment’ means an establishment in which the poultry have shown no evidence of NAI infection, based on surveillance in accordance with Articles 10.4.27. to 10.4.33.

For the purposes of the Terrestrial Code, the incubation period for NAI shall be 21 days.

8. Standards for diagnostic tests, including pathogenicity testing, are described in the Terrestrial Manual. Any vaccine used should comply with the standards described in the Terrestrial Manual.

9. A Member should not impose immediate bans on the trade in poultry commodities in response to a notification, according to Article 1.2.3. of the Terrestrial Code, of infection with HPAI and LPAI virus in birds other than poultry, including wild birds.

Article 10.4.2.

Determination of the NAI status of a country, zone or compartment

The NAI status of a country, a zone or a compartment can be determined on the basis of the following criteria:

1. NAI is notifiable in the whole country, an on-going NAI awareness programme is in place, and all notified suspect occurrences of NAI are subjected to field and, where applicable, laboratory investigations;

2. appropriate surveillance is in place to demonstrate the presence of infection in the absence of clinical signs in poultry, and the risk posed by birds other than poultry, this may be achieved through a NAI surveillance programme in accordance with Articles 10.4.27. to 10.4.33.;

3. consideration of all epidemiological factors for NAI occurrence and their historical perspective.

Article 10.4.3.

NAI free country, zone or compartment

A country, zone or compartment may be considered free from NAI when it has been shown that neither HPNAI nor LPNAI infection in poultry has been present in the country, zone or compartment for the past 12 months, based on surveillance in accordance with Articles 10.4.27. to 10.4.33.

If infection has occurred in poultry in a previously free country, zone or compartment, NAI free status can be regained:

1. In the case of HPNAI infections, 3 months after a stamping-out policy (including disinfection of all affected establishments) is applied, providing that surveillance in accordance with Articles 10.4.27. to 10.4.33. has been carried out during that three-month period.

2. In the case of LPNAI infections, poultry may be kept for slaughter for human consumption subject to conditions specified in Article 10.4.19. or a stamping-out policy may be applied; in either case, 3 months after the disinfection of all affected establishments, providing that surveillance in accordance with Articles 10.4.27. to 10.4.33. has been carried out during that three-month period.

Article 10.4.4.

HPNAI free country, zone or compartment

A country, zone or compartment may be considered free from HPNAI when:
1. it has been shown that HPNAI infection in poultry has not been present in the country, zone or compartment for the past 12 months, although its LPNAI status may be unknown; or

2. when, based on surveillance in accordance with Articles 10.4.27. to 10.4.33., it does not meet the criteria for freedom from NAI but any NAI virus detected has not been identified as HPNAI virus.

The surveillance may need to be adapted to parts of the country or existing zones or compartments depending on historical or geographical factors, industry structure, population data, or proximity to recent outbreaks.

If infection has occurred in poultry in a previously free country, zone or compartment, HPNAI free status can be regained 3 months after a stamping-out policy (including disinfection of all affected establishments) is applied, providing that surveillance in accordance with Articles 10.4.27. to 10.4.33. has been carried out during that three-month period.

Article 10.4.5.

**Recommendations for importation from a NAI free country, zone or compartment**

for live poultry (other than day-old poultry)

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that:

1. the poultry showed no clinical sign of NAI on the day of shipment;

2. the poultry were kept in a NAI free country, zone or compartment since they were hatched or for at least the past 21 days;

3. the poultry are transported in new or appropriately sanitized containers;

4. if the poultry have been vaccinated against NAI, it has been done in accordance with the provisions of the *Terrestrial Manual* and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.6.

**Recommendations for the importation of live birds other than poultry**

Regardless of the NAI status of the country of origin, *Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that:

1. on the day of shipment, the birds showed no clinical sign of infection with a virus which would be considered NAI in poultry;

2. the birds were kept in isolation approved by the *Veterinary Services* since they were hatched or for at least the 21 days prior to shipment and showed no clinical sign of infection with a virus which would be considered NAI in poultry during the isolation period;

3. a statistically valid sample of the birds, selected in accordance with the provisions of Article 10.4.29., was subjected to a diagnostic test within 14 days prior to shipment to demonstrate freedom from infection with a virus which would be considered NAI in poultry;

4. the birds are transported in new or appropriately sanitized containers;
Annex XXI (contd)

5. if the birds have been vaccinated against NAI, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.7.

Recommendations for importation from a NAI free country, zone or compartment

for day-old live poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the poultry were kept in a NAI free country, zone or compartment since they were hatched;
2. the poultry were derived from parent flocks which had been kept in a NAI free country, zone or compartment for at least 21 days prior to and at the time of the collection of the eggs;
3. the poultry are transported in new or appropriately sanitized containers;
4. if the poultry or the parent flocks have been vaccinated against NAI, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.8.

Recommendations for importation from a HPNAI free country, zone or compartment

for day-old live poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the poultry were kept in a HPNAI free country, zone or compartment since they were hatched;
2. the poultry were derived from parent flocks which had been kept in a NAI free establishment for at least 21 days prior to and at the time of the collection of the eggs;
3. the poultry are transported in new or appropriately sanitized containers;
4. if the poultry or the parent flocks have been vaccinated against NAI, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.9.

Recommendations for the importation of day-old live birds other than poultry

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. on the day of shipment, the birds showed no clinical signs of infection with a virus which would be considered NAI in poultry;
2. the birds were hatched and kept in isolation approved by the Veterinary Services;
3. the parent flock birds were subjected to a diagnostic test at the time of the collection of the eggs to demonstrate freedom from infection with NAIV;

4. the birds are transported in new or appropriately sanitized containers;

5. if the birds or parent flocks have been vaccinated against NAI, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.10.

Recommendations for importation from a NAI free country, zone or compartment

for hatching eggs of poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the eggs came from a NAI free country, zone or compartment;

2. the eggs were derived from parent flocks which had been kept in a NAI free country, zone or compartment for at least 21 days prior to and at the time of the collection of the eggs;

3. the eggs are transported in new or appropriately sanitized packaging materials;

4. if the parent flocks have been vaccinated against NAI, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.11.

Recommendations for importation from a HPNAI free country, zone or compartment

for hatching eggs of poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the eggs came from a HPNAI free country, zone or compartment;

2. the eggs were derived from parent flocks which had been kept in a NAI free establishment for at least 21 days prior to and at the time of the collection of the eggs;

3. the eggs have had their surfaces sanitized (in accordance with Chapter 6.4.);

4. the eggs are transported in new or appropriately sanitized packaging materials;

5. if the parent flocks have been vaccinated against NAI, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.12.

Recommendations for the importation of hatching eggs from birds other than poultry

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:
Annex XXI (contd)

1. the parent flock birds were subjected to a diagnostic test 7 days prior to and at the time of the collection of the eggs to demonstrate freedom from infection with NAIV;

2. the eggs have had their surfaces sanitized (in accordance with Chapter 6.4.);

3. the eggs are transported in new or appropriately sanitized packaging materials;

4. if the parent flocks have been vaccinated against NAI, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.4.13.

Recommendations for importation from a NAI free country, zone or compartment

for eggs for human consumption

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the eggs were produced and packed in a NAI free country, zone or compartment;

2. the eggs are transported in new or appropriately sanitized packaging materials.

Article 10.4.14.

Recommendations for importation from a HPNAI free country, zone or compartment

for eggs for human consumption

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the eggs were produced and packed in a HPNAI free country, zone or compartment;

2. the eggs have had their surfaces sanitized (in accordance with Chapter 6.4.);

3. the eggs are transported in new or appropriately sanitized packaging materials.

Article 10.4.15.

Recommendations for importation of egg products of poultry

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the commodity is derived from eggs which meet the requirements of Articles 10.4.13. or 10.4.14.; or

2. the commodity has been processed to ensure the destruction of NAI virus in accordance with Article 10.4.25.; AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

Article 10.4.16.

Recommendations for importation from a NAI free country, zone or compartment
for poultry semen

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the donor poultry:

1. showed no clinical sign of NAI on the day of semen collection;
2. were kept in a NAI free country, zone or compartment for at least the 21 days prior to and at the time of semen collection.

Article 10.4.17.

Recommendations for the importation from a HPNAI free country, zone or compartment

for poultry semen

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the donor poultry:

1. showed no clinical sign of HPNAI on the day of semen collection;
2. were kept in a HPNAI free country, zone or compartment for at least the 21 days prior to and at the time of semen collection.

Article 10.4.18.

Recommendations for the importation of semen of birds other than poultry

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the donor birds:

1. were kept in isolation approved by the Veterinary Services for at least the 21 days prior to semen collection;
2. showed no clinical sign of infection with a virus which would be considered NAI in poultry during the isolation period;
3. were tested within 14 days prior to semen collection and shown to be free of NAI infection.

Article 10.4.19.

Recommendations for importation from either a NAI or HPNAI free country, zone or compartment

for fresh meat of poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from poultry:

1. which have been kept in a country, zone or compartment free from HPNAI since they were hatched or for at least the past 21 days;
2. which have been slaughtered in an approved abattoir in a country, zone or compartment free from HPNAI and have been subjected to ante-mortem and post-mortem inspections in accordance with Chapter 6.2. and have been found free of any signs suggestive of NAI.
Recommendations for the importation of meat products of poultry

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the commodity is derived from fresh meat which meet the requirements of Article 10.4.19.; or

2. the commodity has been processed to ensure the destruction of NAI virus in accordance with Article 10.4.26.;

AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

Recommendations for the importation of products of poultry origin, other than feather meal and poultry meal, intended for use in animal feeding, or for agricultural or industrial use

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these commodities were processed in a NAI free country, zone or compartment from poultry which were kept in a NAI free country, zone or compartment from the time they were hatched until the time of slaughter or for at least the 21 days preceding slaughter; or

2. these commodities have been processed to ensure the destruction of NAI virus (under study);

AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

Recommendations for the importation of feathers and down of poultry

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these commodities originated from poultry as described in Article 10.4.19. and were processed in a NAI free country, zone or compartment, or

2. these commodities have been processed to ensure the destruction of NAI virus (under study);

AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

Recommendations for the importation of feathers and down of birds other than poultry

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:
1. these commodities have been processed to ensure the destruction of NAI virus (under study); and

2. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

Article 10.4.24.

Recommendations for the importation of feather meal and poultry meal

Regardless of the NAI status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these commodities were processed in a NAI free country, zone or compartment from poultry which were kept in a NAI free country, zone or compartment from the time they were hatched until the time of slaughter or for at least the 21 days preceding slaughter, or

2. these commodities have been processed either:
   a) with moist heat at a minimum temperature of 118°C for minimum of 40 minutes; or
   b) with a continuous hydrolysing process under at least 3.79 bar of pressure with steam at a minimum temperature of 122°C for a minimum of 15 minutes; or
   c) with an alternative rendering process that ensures that the internal temperature throughout the product reaches at least 74°C;

AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

Article 10.4.25.

Procedures for the inactivation of the AI virus in eggs and egg products

The following times for industry standard temperatures are suitable for the inactivation of AI virus present in eggs and egg products:

<table>
<thead>
<tr>
<th>Commodity</th>
<th>Core temperature (°C)</th>
<th>Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Whole egg</td>
<td>60</td>
<td>188 seconds</td>
</tr>
<tr>
<td>Whole egg blends</td>
<td>60</td>
<td>188 seconds</td>
</tr>
<tr>
<td>Whole egg blends</td>
<td>61.1</td>
<td>94 seconds</td>
</tr>
<tr>
<td>Liquid egg white</td>
<td>55.6</td>
<td>870 seconds</td>
</tr>
<tr>
<td>Liquid egg white</td>
<td>56.7</td>
<td>232 seconds</td>
</tr>
<tr>
<td>10% salted yolk</td>
<td>62.2</td>
<td>138 seconds</td>
</tr>
<tr>
<td>Dried egg white</td>
<td>67</td>
<td>20 hours</td>
</tr>
<tr>
<td>Dried egg white</td>
<td>54.4</td>
<td>513 hours</td>
</tr>
</tbody>
</table>
Annex XXI (contd)

The listed temperatures are indicative of a range that achieves a 7-log kill. Where scientifically documented, variances from these times and temperatures may also be suitable when they achieve the inactivation of the virus.

Article 10.4.26.

Procedures for the inactivation of the AI virus in meat

The following times for industry standard temperatures are suitable for the inactivation of AI virus present in meat.

<table>
<thead>
<tr>
<th>Core temperature (°C)</th>
<th>Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Poultry meat</td>
<td></td>
</tr>
<tr>
<td>60.0</td>
<td>507 seconds</td>
</tr>
<tr>
<td>65.0</td>
<td>42 seconds</td>
</tr>
<tr>
<td>70.0</td>
<td>3.5 seconds</td>
</tr>
<tr>
<td>73.9</td>
<td>0.51 seconds</td>
</tr>
</tbody>
</table>

The listed temperatures are indicative of a range that achieves a 7-log kill. Where scientifically documented, variances from these times and temperatures may also be suitable when they achieve the inactivation of the virus.

Article 10.4.27.

Surveillance: introduction

Articles 10.4.27. to 10.4.33. define the principles and provide a guide on the surveillance for NAI complementary to Chapter 1.4., applicable to Members seeking to determine their NAI status. This may be for the entire country, zone or compartment. Guidance for Members seeking free status following an outbreak and for the maintenance of NAI status is also provided.

The presence of avian influenza viruses in wild birds creates a particular problem. In essence, no Member can declare itself free from avian influenza (AI) in wild birds. However, the definition of NAI in this chapter refers to the infection in poultry only, and Articles 10.4.27. to 10.4.33. were developed under this definition.

The impact and epidemiology of NAI differ widely in different regions of the world and therefore it is impossible to provide specific recommendations for all situations. Surveillance strategies employed for demonstrating freedom from NAI at an acceptable level of confidence will need to be adapted to the local situation. Variables such as the frequency of contacts of poultry with wild birds, different biosecurity levels and production systems and the commingling of different susceptible species including domestic waterfowl require specific surveillance strategies to address each specific situation. It is incumbent upon the Member to provide scientific data that explains the epidemiology of NAI in the region concerned and also demonstrates how all the risk factors are managed. There is therefore considerable latitude available to Members to provide a well-reasoned argument to prove that absence of NAI virus (NAIV) infection is assured at an acceptable level of confidence.

Surveillance for NAI should be in the form of a continuing programme designed to establish that the country, zone or compartment, for which application is made, is free from NAIV infection.
Surveillance: general conditions and methods

1. A surveillance system in accordance with Chapter 1.4. should be under the responsibility of the Veterinary Authority. In particular:
   a) a formal and ongoing system for detecting and investigating outbreaks of disease or NAI infection should be in place;
   b) a procedure should be in place for the rapid collection and transport of samples from suspect cases of NAI to a laboratory for NAI diagnosis as described in the Terrestrial Manual;
   c) a system for recording, managing and analysing diagnostic and surveillance data should be in place.

2. The NAI surveillance programme should:
   a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers, who have day-to-day contact with poultry, as well as diagnosticians, should report promptly any suspicion of NAI to the Veterinary Authority. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Authority. All suspected cases of NAI should be investigated immediately. As suspicion cannot always be resolved by epidemiological and clinical investigation alone, samples should be taken and submitted to a laboratory for appropriate tests. This requires that sampling kits and other equipment are available for those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in NAI diagnosis and control. In cases where potential public health implications are suspected, notification to the appropriate public health authorities is essential;
   b) implement, when relevant, regular and frequent clinical inspection, serological and virological testing of high-risk groups of animals, such as those adjacent to a NAI infected country, zone or compartment, places where birds and poultry of different origins are mixed, such as live bird markets, poultry in close proximity to waterfowl or other potential sources of NAIV.

An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is NAIV. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. Applications for freedom from NAIV infection should, in consequence, provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement standstill orders, etc.).

Surveillance strategies

1. Introduction

The target population for surveillance aimed at identification of disease and infection should cover all the susceptible poultry species within the country, zone or compartment. Active and passive surveillance for NAI should be ongoing. The frequency of active surveillance should be at least every 6 months. Surveillance should be composed of random and targeted approaches using molecular, virological, serological and clinical methods.
The strategy employed may be based on randomised sampling requiring surveillance consistent with demonstrating the absence of NAIV infection at an acceptable level of confidence. Random surveillance is conducted using serological tests described in the Terrestrial Manual. Positive serological results should be followed up with molecular or virological methods.

Targeted surveillance (e.g. based on the increased likelihood of infection in particular localities or species) may be an appropriate strategy. Virological and serological methods should be used concurrently to define the NAI status of high risk populations.

A Member should justify the surveillance strategy chosen as adequate to detect the presence of NAIV infection in accordance with Chapter 1.4. and the prevailing epidemiological situation, including cases of HPAI detected in any birds. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clear clinical signs (e.g. chickens). Similarly, virological and serological testing could be targeted to species that may not show clinical signs (e.g. ducks).

If a Member wishes to declare freedom from NAIV infection in a specific zone or compartment, the design of the survey and the basis for the sampling process would need to be aimed at the population within the zone or compartment.

For random surveys, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect infection if it were to occur at a predetermined minimum rate. The sample size and expected disease prevalence determine the level of confidence in the results of the survey. The Member should justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Chapter 1.4. Selection of the design prevalence in particular clearly needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey approach selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and the different species in the target population.

Irrespective of the testing system employed, surveillance system design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of infection or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as flocks which may be epidemiologically linked to it.

The principles involved in surveillance for disease/infection are technically well defined. The design of surveillance programmes to prove the absence of NAIV infection/circulation needs to be carefully followed to avoid producing results that are either insufficiently reliable, or excessively costly and logistically complicated. The design of any surveillance programme, therefore, requires inputs from professionals competent and experienced in this field.

2. Clinical surveillance

Clinical surveillance aims at the detection of clinical signs of NAI at the flock level. Whereas significant emphasis is placed on the diagnostic value of mass serological screening, surveillance based on clinical inspection should not be underrated. Monitoring of production parameters, such as increased mortality, reduced feed and water consumption, presence of clinical signs of a respiratory disease or a drop in egg production, is important for the early detection of NAIV infection. In some cases, the only indication of LPNAIV infection may be a drop in feed consumption or egg production.
Clinical surveillance and laboratory testing should always be applied in series to clarify the status of NAI suspects detected by either of these complementary diagnostic approaches. Laboratory testing may confirm clinical suspicion, while clinical surveillance may contribute to confirmation of positive serology. Any sampling unit within which suspicious animals are detected should have restrictions imposed upon it until NAI infection is ruled out.

Identification of suspect flocks is vital to the identification of sources of NAIV and to enable the molecular, antigenic and other biological characteristics of the virus to be determined. It is essential that NAIV isolates are sent regularly to the regional Reference Laboratory for genetic and antigenic characterization.

3. Virological surveillance

Virological surveillance using tests described in the Terrestrial Manual should be conducted:

a) to monitor at risk populations;

b) to confirm clinically suspect cases;

c) to follow up positive serological results;

d) to test ‘normal’ daily mortality, to ensure early detection of infection in the face of vaccination or in establishments epidemiologically linked to an outbreak.

4. Serological surveillance

Serological surveillance aims at the detection of antibodies against NAIV. Positive NAIV antibody test results can have four possible causes:

a) natural infection with NAIV;

b) vaccination against NAI;

c) maternal antibodies derived from a vaccinated or infected parent flock are usually found in the yolk and can persist in progeny for up to 4 weeks;

d) false positive results due to the lack of specificity of the test.

It may be possible to use serum collected for other survey purposes for NAI surveillance. However, the principles of survey design described in these recommendations and the requirement for a statistically valid survey for the presence of NAIV should not be compromised.

The discovery of clusters of seropositive flocks may reflect any of a series of events, including but not limited to the demographics of the population sampled, vaccinal exposure or infection. As clustering may signal infection, the investigation of all instances should be incorporated in the survey design. Clustering of positive flocks is always epidemiologically significant and therefore should be investigated.

If vaccination cannot be excluded as the cause of positive serological reactions, diagnostic methods to differentiate antibodies due to infection or vaccination should be employed.

The results of random or targeted serological surveys are important in providing reliable evidence that no NAIV infection is present in a country, zone or compartment. It is therefore essential that the survey be thoroughly documented.
5. Virological and serological surveillance in vaccinated populations

The surveillance strategy is dependent on the type of vaccine used. The protection against AI is haemagglutinin subtype specific. Therefore, two broad vaccination strategies exist: 1) inactivated whole AI viruses, and 2) haemagglutinin expression-based vaccines.

In the case of vaccinated populations, the surveillance strategy should be based on virological and/or serological methods and clinical surveillance. It may be appropriate to use sentinel birds for this purpose. These birds should be unvaccinated, AI virus antibody free birds and clearly and permanently identified. Sentinel birds should be used only if no appropriate laboratory procedures are available. The interpretation of serological results in the presence of vaccination is described in Article 10.4.33.

Article 10.4.30.

Documentation of NAI or HPNAI free status

1. Members declaring freedom from NAI or HPNAI for the country, zone or compartment: additional surveillance procedures

In addition to the general conditions described in above mentioned articles, a Member declaring freedom from NAI or HPNAI for the entire country, or a zone or a compartment should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and should be planned and implemented according to general conditions and methods described in this Chapter, to demonstrate absence of NAIV or HPNAIV infection, during the preceding 12 months in susceptible poultry populations (vaccinated and non-vaccinated). This requires the support of a laboratory able to undertake identification of NAIV or HPNAIV infection through virus detection and antibody tests described in the Terrestrial Manual. This surveillance may be targeted to poultry population at specific risks linked to the types of production, possible direct or indirect contact with wild birds, multi-age flocks, local trade patterns including live bird markets, use of possibly contaminated surface water, and the presence of more than one species on the holding and poor biosecurity measures in place.

2. Additional requirements for countries, zones or compartments that practise vaccination

Vaccination to prevent the transmission of HPNAI virus may be part of a disease control programme. The level of flock immunity required to prevent transmission will depend on the flock size, composition (e.g. species) and density of the susceptible poultry population. It is therefore impossible to be prescriptive. The vaccine should also comply with the provisions stipulated for NAI vaccines in the Terrestrial Manual. Based on the epidemiology of NAI in the country, zone or compartment, it may be that a decision is reached to vaccinate only certain species or other poultry subpopulations.

In all vaccinated flocks there is a need to perform virological and serological tests to ensure the absence of virus circulation. The use of sentinel poultry may provide further confidence of the absence of virus circulation. The tests have to be repeated at least every 6 months or at shorter intervals according to the risk in the country, zone or compartment.

Evidence to show the effectiveness of the vaccination programme should also be provided.

Article 10.4.31.

Countries, zones or compartments declaring that they have regained freedom from NAI or HPNAI following an outbreak: additional surveillance procedures

In addition to the general conditions described in the above-mentioned articles, a Member declaring that it has regained country, zone or compartment freedom from NAI or HPNAI virus infection should show evidence of an active surveillance programme depending on the epidemiological circumstances of the outbreak to demonstrate the absence of the infection. This will require surveillance incorporating virus detection and antibody tests described in the Terrestrial Manual. The use of sentinel birds may facilitate the interpretation of surveillance results.
A Member declaring freedom of country, zone or compartment after an outbreak of NAI or HPNAI (with or without vaccination) should report the results of an active surveillance programme in which the NAI or HPNAI susceptible poultry population undergoes regular clinical examination and active surveillance planned and implemented according to the general conditions and methods described in these recommendations. The surveillance should at least give the confidence that can be given by a randomized representative sample of the populations at risk.

Article 10.4.32.

NAI free establishments within HPNAI free compartments: additional surveillance procedures

The declaration of NAI free establishments requires the demonstration of absence of NAIV infection. Birds in these establishments should be randomly tested using virus detection or isolation tests, and serological methods, following the general conditions of these recommendations. The frequency of testing should be based on the risk of infection and at a maximum interval of 21 days.

Article 10.4.33.

The use and interpretation of serological and virus detection tests

Poultry infected with NAI virus produce antibodies to haemagglutinin (HA), neuraminidase (NA), nonstructural proteins (NSPs), nucleoprotein/matrix (NP/M) and the polymerase complex proteins. Detection of antibodies against the polymerase complex proteins will not be covered in this chapter. Tests for NP/M antibodies include direct and blocking ELISA, and agar gel immunodiffusion (AGID) tests. Tests for antibodies against NA include the neuraminidase inhibition (NI), indirect fluorescent antibody and direct and blocking ELISA tests. For the HA, antibodies are detected in haemagglutination inhibition (HI), ELISA and neutralization (SN) tests. The HI test is reliable in avian species but not in mammals. The SN test can be used to detect subtype specific antibodies to the haemagglutinin and is the preferred test for mammals and some avian species. The AGID test is reliable for detection of NP/M antibodies in chickens and turkeys, but not in other avian species. As an alternative, blocking ELISA tests have been developed to detect NP/M antibodies in all avian species.

The HI and NI tests can be used to subtype AI viruses into 16 haemagglutinin and 9 neuraminidase subtypes. Such information is helpful for epidemiological investigations and in categorization of AI viruses.

Poultry can be vaccinated with a variety of AI vaccines including inactivated whole AI virus vaccines, and haemagglutinin expression-based vaccines. Antibodies to the haemagglutinin confer subtype specific protection. Various strategies can be used to differentiate vaccinated from infected birds including serosurveillance in unvaccinated sentinel birds or specific serological tests in the vaccinated birds.

AI virus infection of unvaccinated birds including sentinels is detected by antibodies to the NP/M, subtype specific HA or NA proteins, or NSP. Poultry vaccinated with inactivated whole AI vaccines containing an influenza virus of the same H sub-type but with a different neuraminidase may be tested for field exposure by applying serological tests directed to the detection of antibodies to the NA of the field virus. For example, birds vaccinated with H7N3 in the face of a H7N1 epidemic may be differentiated from infected birds (DIVA) by detection of subtype specific NA antibodies of the N1 protein of the field virus. Alternatively, in the absence of DIVA, inactivated vaccines may induce low titres of antibodies to NSP and the titre in infected birds would be markedly higher. Encouraging results have been obtained experimentally with this system, but it has not yet been validated in the field. In poultry vaccinated with haemagglutinin expression-based vaccines, antibodies are detected to the specific HA, but not any of the other AI viral proteins. Infection is evident by antibodies to the NP/M or NSP, or the specific NA protein of the field virus. Vaccines used should comply with the standards of the Terrestrial Manual.

All flocks with seropositive results should be investigated. Epidemiological and supplementary laboratory investigation results should document the status of NAI infection/circulation for each positive flock.
Annex XXI (contd)

A confirmatory test should have a higher specificity than the screening test and sensitivity at least equivalent than that of the screening test.

Information should be provided on the performance characteristics and validation of tests used.

1. **The follow-up procedure in case of positive test results if vaccination is used**

   In case of vaccinated populations, one has to exclude the likelihood that positive test results are indicative of virus circulation. To this end, the following procedure should be followed in the investigation of positive serological test results derived from surveillance conducted on NAI-vaccinated poultry. The investigation should examine all evidence that might confirm or refute the hypothesis that the positive results to the serological tests employed in the initial survey were not due to virus circulation. All the epidemiological information should be substantiated, and the results should be collated in the final report.

   Knowledge of the type of vaccine used is crucial in developing a serological based strategy to differentiate infected from vaccinated animals.

   a) Inactivated whole AI virus vaccines can use either homologous or heterologous neuraminidase subtypes between the vaccine and field strains. If poultry in the population have antibodies to NP/M and were vaccinated with inactivated whole AI virus vaccine, the following strategies should be applied:

      i) sentinel birds should remain NP/M antibody negative. If positive for NP/M antibodies, indicating AI virus infection, specific HI tests should be performed to identify H5 or H7 AI virus infection;

      ii) if vaccinated with inactivated whole AI virus vaccine containing homologous NA to field virus, the presence of antibodies to NSP could be indicative of infection. Sampling should be initiated to exclude the presence of NAIV by either virus isolation or detection of virus specific genomic material or proteins;

      iii) if vaccinated with inactivated whole AI virus vaccine containing heterologous NA to field virus, presence of antibodies to the field virus NA or NSP would be indicative of infection. Sampling should be initiated to exclude the presence of NAIV by either virus isolation or detection of virus specific genomic material or proteins.

   b) Haemagglutinin expression-based vaccines contain the HA protein or gene homologous to the HA of the field virus. Sentinel birds as described above can be used to detect AI infection. In vaccinated or sentinel birds, the presence of antibodies against NP/M, NSP or field virus NA is indicative of infection. Sampling should be initiated to exclude the presence of NAIV by either virus isolation or detection of virus specific genomic material or proteins.

2. **The follow-up procedure in case of positive test results indicative of infection for determination of infection due to HPNAI or LPNAI virus**

   The detection of antibodies indicative of a NAI virus infection as indicated in point a) above will result in the initiation of epidemiological and virological investigations to determine if the infections are due to HPNAI or LPNAI viruses.

   Virological testing should be initiated in all antibody-positive and at risk populations. The samples should be evaluated for the presence of AI virus, by virus isolation and identification, and/or detection of influenza A specific proteins or nucleic acids (Figure 2). Virus isolation is the gold standard for detecting infection by AI virus and the method is described in the Terrestrial Manual. All AI virus isolates should be tested to determine HA and NA subtypes, and in vitro tested in chickens and/or sequencing of HA proteolytic
cleavage site of H5 and H7 subtypes for determination of classification as HPNAI, LPNAI or LPAI (not notifiable) viruses. As an alternative, nucleic acid detection tests have been developed and validated; these tests have the sensitivity of virus isolation, but with the advantage of providing results within a few hours. Samples with detection of H5 and H7 HA subtypes by nucleic acid detection methods should either be submitted for virus isolation, identification, and in vivo testing in chickens, or sequencing of nucleic acids for determination of proteolytic cleavage site as HPNAI or LPNAI viruses. The antigen detection systems, because of low sensitivity, are best suited for screening clinical field cases for infection by Type A influenza virus looking for NP/M proteins. NP/M positive samples should be submitted for virus isolation, identification and pathogenicity determination.

Laboratory results should be examined in the context of the epidemiological situation. Corollary information needed to complement the serological survey and assess the possibility of viral circulation includes but is not limited to:

a) characterization of the existing production systems;
b) results of clinical surveillance of the suspects and their cohorts;
c) quantification of vaccinations performed on the affected sites;
d) sanitary protocol and history of the affected establishments;
e) control of animal identification and movements;
f) other parameters of regional significance in historic NAIV transmission.

The entire investigative process should be documented as standard operating procedure within the epidemiological surveillance programme.
Fig. 1. Schematic representation of laboratory tests for determining evidence of NAI infection through or following serological surveys

Key:
AGID  Agar gel immunodiffusion
DIVA  Differentiating infected from vaccinated animals
ELISA  Enzyme-linked immunosorbant assay
HA    Haemagglutinin
HI    Haemagglutination inhibition
NA    Neuraminidase
NP/M  Nucleoprotein and matrix protein
NSP   Nonstructural protein
S     No evidence of NAIV
**Fig. 2. Schematic representation of laboratory tests for determining evidence of NAI infection using virological methods**

The above diagrams indicate the tests which are recommended for use in the investigation of poultry flocks.

<table>
<thead>
<tr>
<th>Key:</th>
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<tbody>
<tr>
<td>AGID</td>
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<td>DIVA</td>
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<tr>
<td>ELISA</td>
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<td>HA</td>
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<td>NP/M</td>
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<td>NSP</td>
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CHAPTER 10.13.
NEWCASTLE DISEASE

Article 10.13.1.

General provisions

1. For the purposes of international trade, the Terrestrial Code, Newcastle disease (ND) is defined as an infection of poultry caused by a virus (NDV) of avian paramyxovirus serotype 1 (APMV-1) that meets one of the following criteria for virulence:

   a) the virus has an intracerebral pathogenicity index (ICPI) in day-old chicks (*Gallus gallus*) of 0.7 or greater; or

   b) multiple basic amino acids have been demonstrated in the virus (either directly or by deduction) at the C-terminus of the F2 protein and phenylalanine at residue 117, which is the N-terminus of the F1 protein. The term ‘multiple basic amino acids’ refers to at least three arginine or lysine residues between residues 113 and 116. Failure to demonstrate the characteristic pattern of amino acid residues as described above would require characterisation of the isolated virus by an ICPI test.

In this definition, amino acid residues are numbered from the N-terminus of the amino acid sequence deduced from the nucleotide sequence of the F0 gene, 113–116 corresponds to residues −4 to −1 from the cleavage site.’

2. Poultry is defined as ‘all domesticated birds, including backyard poultry, used for the production of meat or eggs for consumption, for the production of other commercial products, for restocking supplies of game, or for breeding these categories of birds, as well as fighting cocks used for any purpose’.

Birds that are kept in captivity for any reason other than those reasons referred to in the preceding paragraph, including those that are kept for shows, races, exhibitions, competitions, or for breeding or selling these categories of birds as well as pet birds, are not considered to be poultry.

3. For the purposes of the Terrestrial Code, the incubation period for ND shall be 21 days.

4. This chapter deals with NDV infection of poultry as defined in point 2 above, in the presence or absence of clinical signs. For the purposes of international trade, a Member should not impose immediate bans on the trade in poultry commodities in response to a notification, according to Article 1.2.3. of the Terrestrial Code, of infection with NDV in birds other than poultry, including wild birds.

5. The occurrence of infection with NDV is defined as the isolation and identification of NDV as such or the detection of viral RNA specific for NDV.

6. For the purposes of the Terrestrial Code, the incubation period for ND shall be 21 days.

7. Standards for diagnostic tests, including pathogenicity testing, are described in the Terrestrial Manual. When the use of ND vaccines is appropriate, those vaccines should comply with the standards described in the Terrestrial Manual.

8. A Member should not impose immediate bans on the trade in poultry commodities in response to a notification, according to Article 1.2.3. of the Terrestrial Code, of infection with NDV in birds other than poultry, including wild birds.
Determination of the ND status of a country, zone or compartment

The ND status of a country, a zone or a compartment can be determined on the basis of the following criteria:

1. ND is notifiable in the whole country, an on-going ND awareness programme is in place, and all notified suspect occurrences of ND are subjected to field and, where applicable, laboratory investigations;

2. appropriate surveillance is in place to demonstrate the presence of NDV infection in the absence of clinical signs in poultry, this may be achieved through an ND surveillance programme in accordance with Articles 10.13.22. to 10.13.26.;

3. consideration of all epidemiological factors for ND occurrence and their historical perspective.

ND free country, zone or compartment

A country, zone or compartment may be considered free from ND when it has been shown that NDV infection in poultry has not been present in the country, zone or compartment for the past 12 months, based on surveillance in accordance with Articles 10.13.22. to 10.13.26.

If infection has occurred in poultry in a previously free country, zone or compartment, ND free status can be regained three months after a stamping-out policy (including disinfection of all affected establishments) is applied, providing that surveillance in accordance with Articles 10.13.22. to 10.13.26. has been carried out during that three-month period.

Recommendations for importation from an ND free country, zone or compartment as defined in Article 10.13.3.

for live poultry (other than day-old poultry)

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the poultry showed no clinical sign suggestive of ND on the day of shipment;

2. the poultry were kept in an ND free country, zone or compartment since they were hatched or for at least the past 21 days;

3. the poultry are transported in new or appropriately sanitized containers;

4. if the poultry have been vaccinated against ND, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Recommendations for the importation of live birds other than poultry

Regardless of the ND status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the birds showed no clinical sign suggestive of infection by NDV on the day of shipment;
2. the birds were kept in isolation approved by the Veterinary Services since they were hatched or for at least the 21 days prior to shipment and showed no clinical sign of infection during the isolation period;

3. a statistically valid sample of the birds, selected in accordance with the provisions of Article 10.13.24., was subjected to a diagnostic test within 14 days prior to shipment to demonstrate freedom from infection with NDV;

4. the birds are transported in new or appropriately sanitized containers;

5. if the birds have been vaccinated against ND, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.13.6.

Recommendations for importation from an ND free country, zone or compartment

for day-old live poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the poultry were hatched and kept in an ND free country, zone or compartment since they were hatched;

2. the poultry were derived from parent flocks which had been kept in an ND free country, zone or compartment for at least 21 days prior to and at the time of the collection of the eggs;

3. the poultry are transported in new or appropriately sanitized containers;

4. if the poultry or parent flocks have been vaccinated against ND, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.

Article 10.13.7.

Recommendations for the importation of day-old live birds other than poultry

Regardless of the ND status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the birds showed no clinical sign suggestive of infection by NDV on the day of shipment;

2. the birds were hatched and kept in isolation approved by the Veterinary Services;

3. the parent flock birds were subjected to a diagnostic test at the time of the collection of the eggs to demonstrate freedom from infection with NDV;

4. the birds are transported in new or appropriately sanitized containers;

5. if the birds or parent flocks have been vaccinated against ND, it has been done in accordance with the provisions of the Terrestrial Manual and the nature of the vaccine used and the date of vaccination have been attached to the certificate.
Annex XXII (contd)

Article 10.13.8.

**Recommendations for importation from an ND free country, zone or compartment**

for hatching eggs of poultry

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that:

1. the eggs came from an ND free country, *zone or compartment*;
2. the eggs were derived from parent *flocks* which had been kept in an ND free country, *zone or compartment* for at least 21 days prior to and at the time of the collection of the eggs;
3. the eggs are transported in new or appropriately sanitized packaging materials;
4. if the parent *flocks* have been vaccinated against ND, it has been done in accordance with the provisions of the *Terrestrial Manual* and the nature of the vaccine used and the date of vaccination have been attached to the *certificate*.

Article 10.13.9.

**Recommendations for the importation of hatching eggs from birds other than poultry**

Regardless of the ND status of the country of origin, *Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that:

1. the parent *flock* birds were subjected to a diagnostic test 7 days prior to and at the time of the collection of the eggs to demonstrate freedom from infection with NDV;
2. the eggs have had their surfaces sanitized (in accordance with Chapter 6.4.);
3. the eggs are transported in new or appropriately sanitized packaging materials;
4. if the parent *flocks* have been vaccinated against ND, it has been done in accordance with the provisions of the *Terrestrial Manual* and the nature of the vaccine used and the date of vaccination have been attached to the *certificate*.

Article 10.13.10.

**Recommendations for importation from an ND free country, zone or compartment**

for eggs for human consumption

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that:

1. the eggs were produced and packed in an ND free country, *zone or compartment*;
2. the eggs are transported in new or appropriately sanitized packaging materials.

Article 10.13.11.

**Recommendations for importation of egg products of poultry**

Regardless of the ND status of the country of origin, *Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that:
1. the commodity is derived from eggs which meet the requirements of Article 10.13.10.; or
2. the commodity has been processed to ensure the destruction of NDV in accordance with Article 10.13.20.;

AND

3. the necessary precautions were taken to avoid contact of the egg products with any source of NDV.

Article 10.13.12.

Recommendations for importation from an ND free country, zone or compartment

for poultry semen

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the donor poultry:

1. showed no clinical sign suggestive of ND on the day of semen collection;
2. were kept in an ND free country, zone or compartment for at least the 21 days prior to and at the time of semen collection.

Article 10.13.13.

Recommendations for the importation of semen of birds other than poultry

Regardless of the ND status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the donor birds:

1. were kept in isolation approved by the Veterinary Services for at least the 21 days prior to and on the day of semen collection;
2. showed no clinical sign suggestive of infection with NDV during the isolation period and on the day of semen collection;
3. were subjected to a diagnostic test within 14 days prior to semen collection to demonstrate freedom from infection with NDV.


Recommendations for importation from an ND free country, zone or compartment

for fresh meat of poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from poultry:

1. which have been kept in an ND free country, zone or compartment since they were hatched or for at least the past 21 days;
2. which have been slaughtered in an approved abattoir in an ND free country, zone or compartment and have been subjected to ante-mortem and post-mortem inspections in accordance with Chapter 6.2. and have been found free of any sign suggestive of ND.
Recommendations for importation of meat products of poultry

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the commodity is derived from fresh meat which meet the requirements of Article 10.13.14.; or
2. the commodity has been processed to ensure the destruction of NDV in accordance with Article 10.13.21.;

AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NDV.

Recommendations for the importation of products of poultry origin, other than feather meal and poultry meal, intended for use in animal feeding, or for agricultural or industrial use

Regardless of the ND status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these commodities were processed in a ND free country, zone or compartment from poultry which were kept in a ND free country, zone or compartment from the time they were hatched until the time of slaughter or for at least the 21 days preceding slaughter; or
2. these commodities have been processed to ensure the destruction of NDV (under study);

AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NDV.

Recommendations for the importation of feathers and down of poultry

Regardless of the ND status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these commodities originated from poultry as described in Article 10.13.14. and were processed in a ND free country, zone or compartment; or
2. these commodities have been processed to ensure the destruction of NDV (under study);

AND

3. the necessary precautions were taken to avoid contact of the commodity with any source of NDV.

Recommendations for the importation of feathers and down of birds other than poultry

Regardless of the ND status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these commodities have been processed to ensure the destruction of NDV (under study); and
2. the necessary precautions were taken to avoid contact of the *commodity* with any source of NDV.

Article 10.13.19.

**Recommendations for the importation of feather meal and poultry meal**

Regardless of the ND status of the country of origin, *Veterinary Authorities* should require the presentation of an international veterinary certificate attesting that:

1. these *commodities* were processed in a ND free country, zone or compartment from poultry which were kept in a ND free country, zone or compartment from the time they were hatched until the time of *slaughter* or for at least the 21 days preceding *slaughter*; or

2. these *commodities* have been processed either:
   a) with moist heat at a minimum temperature of 118°C for minimum of 40 minutes; or
   b) with a continuous hydrolysing process under at least 3.79 bar of pressure with steam at a minimum temperature of 122°C for a minimum of 15 minutes; or
   c) with an alternative rendering process that ensures that the internal temperature throughout the product reaches at least 74°C for a minimum of 280 seconds;

AND

3. the necessary precautions were taken to avoid contact of the *commodity* with any source of ND virus.

Article 10.13.20.

**Procedures for the inactivation of the ND virus in eggs and egg products**

The following times and temperatures are suitable for the inactivation of ND virus present in eggs and egg products:

<table>
<thead>
<tr>
<th><em>Core temperature (°C)</em></th>
<th><em>Time</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>Whole egg</td>
<td>55</td>
</tr>
<tr>
<td>Whole egg</td>
<td>57</td>
</tr>
<tr>
<td>Whole egg</td>
<td>59</td>
</tr>
<tr>
<td>Liquid egg white</td>
<td>55</td>
</tr>
<tr>
<td>Liquid egg white</td>
<td>57</td>
</tr>
<tr>
<td>Liquid egg white</td>
<td>59</td>
</tr>
<tr>
<td>10% salted yolk</td>
<td>55</td>
</tr>
<tr>
<td>Dried egg white</td>
<td>57</td>
</tr>
</tbody>
</table>
Annex XXII (contd)

The listed temperatures are indicative of a range that achieves a 7-log kill. Where scientifically documented, variances from these times and temperatures may also be suitable when they achieve the inactivation of the virus.

Article 10.13.21.

Procedures for the inactivation of the ND virus in meat

The following times for industry standard temperatures are suitable for the inactivation of ND virus present in meat.

<table>
<thead>
<tr>
<th>Core temperature (°C)</th>
<th>Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Poultry meat</td>
<td></td>
</tr>
<tr>
<td>65.0</td>
<td>840 seconds</td>
</tr>
<tr>
<td>70.0</td>
<td>574 seconds</td>
</tr>
<tr>
<td>74.0</td>
<td>280 seconds</td>
</tr>
<tr>
<td>80.0</td>
<td>203 seconds</td>
</tr>
</tbody>
</table>

The listed temperatures are indicative of a range that achieves a 7-log kill. Where scientifically documented, variances from these times and temperatures may also be suitable when they achieve the inactivation of the virus.

Article 10.13.22.

Surveillance: introduction

Articles 10.13.22. to 10.13.26. define the principles and provide a guide on the surveillance for ND as defined in Article 10.13.1. and is complementary to Chapter 1.4. It is applicable to Members seeking to determine their ND status. This may be for the entire country, zone or compartment. Guidance for Members seeking free status following an outbreak and for the maintenance of ND status is also provided.

Surveillance for ND is complicated by the known occurrence of avian paramyxovirus serotype 1 (APMV-1) infections in many bird species, both domestic and wild, and the widespread utilization of ND vaccines in domestic poultry.

The impact and epidemiology of ND differ widely in different regions of the world and therefore it is not possible to provide specific recommendations for all situations. Therefore, surveillance strategies employed for demonstrating freedom from ND at an acceptable level of confidence will need to be adapted to the local situation. Variables such as the frequency of contacts of poultry with wild birds, different biosecurity levels, production systems and the commingling of different susceptible species require specific surveillance strategies to address each specific situation. It is incumbent upon the Member to provide scientific data that explains the epidemiology of ND in the region concerned and also demonstrates how all the risk factors are managed. There is, therefore, considerable latitude available to Members to provide a well-reasoned argument to prove freedom from NDV infection.
Surveillance for ND should be in the form of a continuing programme designed to establish that the country, zone or compartment, for which application is made, is free from NDV infection.

Article 10.13.23.

Surveillance: general conditions and methods

1. A surveillance system in accordance with Chapter 1.4. should be under the responsibility of the Veterinary Authority. In particular there should be in place:
   a) a formal and ongoing system for detecting and investigating outbreaks of disease or NDV infection;
   b) a procedure for the rapid collection and transport of samples from suspect cases of ND to a laboratory for ND diagnosis as described in the Terrestrial Manual;
   c) a system for recording, managing and analysing diagnostic and surveillance data.

2. The ND surveillance programme should:
   a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers, who have day-to-day contact with poultry, as well as diagnosticians, should report promptly any suspicion of ND to the Veterinary Authority. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Authority. All suspected cases of ND should be investigated immediately. As suspicion cannot be resolved by epidemiological and clinical investigation alone, samples should be taken and submitted to a laboratory for appropriate tests. This requires that sampling kits and other equipment are available to those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in ND diagnosis and control;
   b) implement, when relevant, regular and frequent clinical, virological and serological surveillance of high risk groups of poultry within the target population (e.g. those adjacent to an ND infected country, zone, compartment, places where birds and poultry of different origins are mixed, or other sources of NDV).

An effective surveillance system may identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is due to NDV infection. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. Applications for freedom from NDV infection should provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement standstill orders, etc.).

Article 10.13.24.

Surveillance strategies

1. Introduction

Any surveillance programme requires inputs from professionals competent and experienced in this field and should be thoroughly documented. The design of surveillance programmes to prove the absence of NDV infection / circulation needs to be carefully followed to avoid producing results that are either unreliable, or excessively costly and logistically complicated.
Annex XXII (contd)

If a Member wishes to declare freedom from NDV infection in a country, zone or compartment, the subpopulation used for the surveillance for the disease / infection should be representative of all poultry within the country, zone or compartment. Multiple surveillance methods should be used concurrently to accurately define the true ND status of poultry populations. Active and passive surveillance for ND should be ongoing with the frequency of active surveillance being appropriate to the disease situation in the country. Surveillance should be composed of random and/or targeted approaches, dependent on the local epidemiological situation and using clinical, virological and serological methods as described in the Terrestrial Manual. If alternative tests are used they should have been validated as fit-for-purpose in accordance with OIE standards. A Member should justify the surveillance strategy chosen as adequate to detect the presence of NDV infection in accordance with Chapter 1.4. and the prevailing epidemiological situation.

In surveys, the sample size selected for testing should be statistically justified to detect infection at a predetermined target prevalence. The sample size and expected prevalence determine the level of confidence in the results of the survey. The survey design and frequency of sampling should be dependent on the historical and current local epidemiological situation. The Member should justify the choice of survey design and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Chapter 1.4.

Targeted surveillance (e.g. based on the increased likelihood of infection in a population) may be an appropriate strategy.

It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clear clinical signs (e.g. unvaccinated chickens). Similarly, virological and serological testing could target species that may not show clinical signs (Article 10.13.2.) of ND and are not routinely vaccinated (e.g. ducks). Surveillance may also target poultry populations at specific risk, for example direct or indirect contact with wild birds, multi-age flocks, local trade patterns including live poultry markets, the presence of more than one species on the holding and poor biosecurity measures in place. In situations where wild birds have been shown to play a role in the local epidemiology of ND, surveillance of wild birds may be of value in alerting Veterinary Services to the possible exposure of poultry and, in particular, of free ranging poultry.

The sensitivity and specificity of the diagnostic tests are key factors in the choice of survey design, which should anticipate the occurrence of false positive and false negative reactions. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination / infection history and for the different species in the target population. If the characteristics of the testing system are known, the rate at which these false reactions are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of infection or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as flocks which may be epidemiologically linked to it.

The results of active and passive surveillance are important in providing reliable evidence that no NDV infection is present in a country, zone or compartment.

2. Clinical surveillance

Clinical surveillance aims to detect clinical signs suggestive of ND at the flock level and should not be underestimated as an early indication of infection. Monitoring of production parameters (e.g. a drop in feed or water consumption or egg production) is important for the early detection of NDV infection in some populations, as there may be no, or mild clinical signs, particularly if they are vaccinated. Any sampling unit within which suspicious animals are detected should be considered as infected until evidence to the contrary is produced. Identification of infected flocks is vital to the identification of sources of NDV.

A presumptive diagnosis of clinical ND in suspect infected populations should always be confirmed by virological testing in a laboratory. This will enable the molecular, antigenic and other biological characteristics of the virus to be determined.
It is desirable that NDV isolates are sent promptly to an OIE Reference Laboratory for archiving and further characterization if required.

3. **Virological surveillance**

Virological *surveillance* should be conducted using tests described in the *Terrestrial Manual* to:

a) monitor at risk populations;

b) confirm suspect clinical cases;

c) follow up positive serological results in unvaccinated populations or sentinel birds;

d) test ‘normal’ daily mortalities (if warranted by an increased risk e.g. *infection* in the face of vaccination or in establishments epidemiologically linked to an outbreak).

4. **Serological surveillance**

Where vaccination is carried out, serological *surveillance* is of limited value. Serological *surveillance* cannot be used to discriminate between NDV and other APMV-1. Test procedures and interpretations of results are as described in the *Terrestrial Manual*. Positive NDV antibody test results can have five possible causes:

a) natural infection with APMV-1;

b) vaccination against ND;

c) exposure to vaccine virus;

d) maternal antibodies derived from a vaccinated or infected parent flock are usually found in the yolk and can persist in progeny for up to 4 weeks;

e) non-specific test reactions.

It may be possible to use serum collected for other survey purposes for ND *surveillance*. However, the principles of survey design described in these recommendations and the requirement for a statistically valid survey for the presence of NDV should not be compromised.

Discovery of seropositive, unvaccinated flocks should be investigated further by conducting a thorough epidemiological investigation. Since seropositive results are not necessarily indicative of infection, virological methods should be used to confirm the presence of NDV in such populations. Until validated strategies and tools to differentiate vaccinated *animals* from those infected with field APMV-1 are available, serological tools should not be used to identify NDV infection in vaccinated populations.

5. **Use of sentinel poultry**

There are various applications of the use of sentinel *poultry* as a *surveillance* tool to detect virus circulation. They may be used to monitor vaccinated populations or species which are less susceptible to the development of clinical disease for the circulation of virus. Sentinel *poultry* should be immunologically naive and may be used in vaccinated flocks. In case of the use of sentinel *poultry*, the structure and organisation of the *poultry* sector, the type of vaccine used and local epidemiological factors will determine the type of production systems where sentinels should be placed, the frequency of placement and monitoring of the sentinels.
Annex XXII (contd)

Sentinel poultry should be in close contact with, but should be identified to be clearly differentiated from, the target population. Sentinel poultry should be observed regularly for evidence of clinical disease and any disease incidents investigated by prompt laboratory testing. The species to be used as sentinels should be proven to be highly susceptible to infection and ideally develop clear signs of clinical disease. Where the sentinel poultry do not necessarily develop overt clinical disease a programme of regular active testing by virological and serological tests should be used (the development of clinical disease may be dependent on the sentinel species used or use of live vaccine in the target population that may infect the sentinel poultry). The testing regime and the interpretation of the results will depend on the type of vaccine used in the target population. Sentinel birds should be used only if no appropriate laboratory procedures are available.

Article 10.13.25.

Documentation of ND free status: additional surveillance procedures

The requirements for a country, zone or compartment to declare freedom from ND are given in Article 10.13.3.

A Member declaring freedom of a country, zone or compartment (with or without vaccination) should report the results of a surveillance programme in which the ND susceptible poultry population undergoes regular surveillance planned and implemented according to the general conditions and methods described in these recommendations.

1. Members declaring freedom from ND for the country, zone or compartment

In addition to the general conditions described in the Terrestrial Code, a Member declaring freedom from ND for the entire country, or a zone or a compartment should provide evidence for the existence of an effective surveillance programme. The surveillance programme should be planned and implemented according to general conditions and methods described in this chapter to demonstrate absence of NDV infection in poultry during the preceding 12 months.

2. Additional requirements for countries, zones or compartments that practice vaccination

Vaccination against ND may be used as a component of a disease prevention and control programme. The vaccine used should comply with the provisions of the Terrestrial Manual.

In vaccinated populations there is a need to perform surveillance to ensure the absence of NDV circulation. The use of sentinel poultry may provide further confidence of the absence of virus circulation. The surveillance should be repeated at least every 6 months or at shorter intervals according to the risk in the country, zone or compartment, or evidence to show the effectiveness of the vaccination programme is regularly provided.


Countries, zones or compartments regaining freedom from ND following an outbreak: additional surveillance procedures

A Member regaining country, zone or compartment freedom from ND should show evidence of an active surveillance programme depending on the epidemiological circumstances of the outbreak to demonstrate the absence of the infection.

A Member declaring freedom of a country, zone or compartment after an outbreak of ND (with or without vaccination) should report the results of a surveillance programme in which the ND susceptible poultry population undergoes regular surveillance planned and implemented according to the general conditions and methods described in these recommendations.

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Annex XXIII

CHAPTER 1.6.

STATUS FOR OIE LISTED DISEASES:
PROCEDURES FOR SELF DECLARATION AND
FOR OFFICIAL RECOGNITION BY THE OIE

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Article 1.6.5.

Questionnaire on contagious bovine pleuropneumonia

CBPP FREE COUNTRY

Report of a Member which applies for recognition of status, under Chapter 11.8. of the Terrestrial Animal Health Code (2010), as a CBPP free country

Please address concisely the following topics. National regulations laws and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. Introduction
   a) Geographical factors. Provide a general description of the country including physical, geographical and other factors that are relevant to CBPP dissemination, countries sharing common borders and other countries that although may not be adjacent share a link for the potential introduction of disease. Provide a map identifying the factors above.
   b) Livestock industry. Provide a general description of the livestock industry in the country.

2. Veterinary system
   a) Legislation. Provide a list and summary of all relevant veterinary legislation in relation to CBPP.
   b) Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the Terrestrial Code and 1.1.3. of the Terrestrial Manual and describe how the Veterinary Services supervise and control all CBPP related activities. Provide maps and tables wherever possible.
   c) Role of farmers, industry and other relevant groups in CBPP surveillance and control (include a description of training and awareness programmes on CBPP).
   d) Role of private veterinary profession in CBPP surveillance and control.

3. CBPP eradication
   a) History. Provide a description of the CBPP history in the country, date of first detection, origin of infection, date of eradication (date of last case).
   b) Strategy. Describe how CBPP was controlled and eradicated (e.g. stamping-out, modified stamping-out, zoning), and provide timeframe for eradication.
Annex XXIII (contd)

c) Vaccines and vaccination. Was CBPP vaccine ever used? If so, when was the last vaccination carried out?

d) Legislation, organisation and implementation of the CBPP eradication campaign. Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines exist and give a brief summary.

e) Animal identification and movement control. Are susceptible animals identified (individually or at a group level)? Provide a description of the methods of animal identification, herd registration and traceability. How are animal movements controlled in the country? Provide evidence on the effectiveness of animal identification and movement controls. Please provide information on pastoralism, transhumance and the related paths of movement.

4. CBPP diagnosis

Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3. and 2.4.9. of the Terrestrial Manual are applied. In particular, the following points should be addressed:

a) Is CBPP laboratory diagnosis carried out in the country? If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to, the follow-up procedures and the time frame for obtaining results.

b) Provide an overview of the CBPP approved laboratories, in particular to address the following points:

i) Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO, etc. that exist in, or planned for, the laboratory system.

ii) Give details of participation in inter-laboratory validation tests (ring tests).

iii) Biosecurity measures applied.

iv) Details of the type of tests undertaken including procedures to isolate and identify \textit{M. mycoides subsp. mycoides SC} as opposed to \textit{M. mycoides subsp. mycoides LC}.

5. CBPP surveillance

Provide documentary evidence that surveillance for CBPP in the country complies with the provisions of Articles 11.8.12 to 11.8.17 of the Terrestrial Code and Chapter 2.4.9 of the Terrestrial Manual. In particular, the following points should be addressed:

a) Clinical surveillance. What are the criteria for raising a suspicion of CBPP? What is the procedure to notify (by whom and to whom) and what penalties are involved for failure to report? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for CBPP agent, species, type of sample, testing method(s) and results (including differential diagnosis).

b) Slaughterhouses, slaughter slabs, abattoirs. What are the criteria for raising a suspicion of CBPP lesion? What is the procedure to notify (by whom and to whom)? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for CBPP agent, species, type of sample, testing method(s) and results (including differential diagnosis).

c) Provide details on training programmes for personnel involved in clinical and slaughter facilities surveillance, and the approaches used to increase community involvement in CBPP surveillance programmes.
d) For countries where a significant proportion of *animals* are not slaughtered in controlled *abattoirs*, what are the alternative *surveillance* measures applied to detect CBPP (e.g. active clinical *surveillance* programmes, laboratory follow-up).

e) Livestock demographics and economics. What is the susceptible animal population by species and production systems? How many *herds* of each susceptible species are in the country? How are they distributed (e.g. *herd* density, etc.)? Provide tables and maps as appropriate.

f) Slaughterhouses and markets. Where are the major livestock marketing or collection centres? What are the patterns of livestock movement within the country? How are the *animals* transported and handled during these transactions?

g) Provide a description of the means employed during the 2 years preceding this application to rule out the presence of any *MmmSC* strain in the susceptible population. Provide criteria for selection of populations for targeted *surveillance* and numbers of *animals* examined and samples tested. Provide details on the methods applied for monitoring the performance of the *surveillance* system including indicators.

6. CBPP prevention

a) Coordination with neighbouring countries. Are there any relevant factors about the adjacent countries that should be taken into account (e.g. size, distance from adjacent border to affected *herds* or *animals*)? Describe coordination, collaboration and information sharing activities with neighbouring countries.

b) Import control procedures

From what countries or *zones* does the country authorize the import of susceptible *animals*? What criteria are applied to approve such countries or *zones*? What controls are applied on entry of such *animals*, and subsequent internal movement? What import conditions and test procedures are required? Are imported *animals* of susceptible species required to undergo a quarantine or isolation period? If so, for how long and where? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports of susceptible *animals* for the past 2 years, specifying country or *zone* of origin, species and volume.

i) Provide a map with the number and location of ports, airports and land crossings. Is the official service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the central *Veterinary Services*. Describe the communication systems between the central authorities and the border inspection posts, and between border inspection posts.

ii) Describe the regulations, procedures, type and frequency of checks at the point of entry into the country and/or their final destination, concerning the import and follow-up of the following:

- *animals*,

- *semen, embryos and oocytes*,

- veterinary medicinal products (i.e. biologics).

iii) Describe the action available under legislation, and actually taken, when an illegal import is detected. Provide information on detected illegal imports.
7. **Control measures and contingency planning**

   a) Give details of any written guidelines, including contingency plans, available to the official services for dealing with suspected or confirmed outbreaks of CBPP.

   b) Is quarantine imposed on premises with suspicious cases, pending final diagnosis? What other procedures are followed regarding suspicious cases?

   c) In the event of a CBPP outbreak:

      i) indicate the sampling and testing procedures used to identify and confirm presence of the causative agent;

      ii) describe the actions taken to control the disease situation in and around any holdings found to be infected with CBPP;

      iii) indicate the control and/or eradication procedures (e.g. vaccination, stamping-out, partial slaughter/vaccination, etc.) that would be taken;

      iv) describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including any restrictions on restocking;

      v) give details of any compensation payments made available to farmers, etc. when animals are slaughtered for disease control/eradication purposes and their prescribed timetable.

8. **Compliance with the Terrestrial Code**

   In addition to the documentary evidence that the provisions of Article 11.8.3. are properly implemented and supervised, the Delegate of the country must submit a declaration indicating:

   a) no clinical CBPP has been detected for at least 2 years;

   b) no CBPP vaccines have been used for at least 2 years in any susceptible species;

   c) the country operates both clinical surveillance and disease reporting systems for CBPP adequate to detect clinical disease if it were present;

   d) all clinical and pathological evidence suggestive of CBPP is investigated by field and laboratory methods (including serological assessment) to refute a possible diagnosis of CBPP;

   e) there are effective measures in force to prevent the re-introduction of the disease.

9. **Recovery of status**

   Countries applying for recovery of status should comply with the provisions of Article 11.8.4. of the Terrestrial Code and provide detailed information as specified in sections 3.a), 3.b), 3.c), 5.b), 5.c) and 5.d) of this questionnaire. Information in relation to other sections need only be supplied if relevant.
CBPP FREE ZONE

Report of a Member which applies for recognition of status, under Chapter 11.8. of the Terrestrial Animal Health Code (2010), as a CBPP free zone

Please address concisely the following topics. National regulations laws and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. Introduction
   a) Geographical factors. Provide a general description of the country including physical, geographical and other factors that are relevant to CBPP dissemination, countries sharing common borders and other countries that although may not be adjacent share a link for the potential introduction of disease. Provide a map identifying the factors above. The boundaries of the zone must be clearly defined. Provide a digitalised, geo-referenced map with a precise text description of the geographical boundaries of the zone.
   
   b) Livestock industry. Provide a general description of the livestock industry in the country.

2. Veterinary system
   a) Legislation. Provide a list and summary of all relevant veterinary legislation in relation to CBPP.
   
   b) Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the Terrestrial Code and 1.1.3. of the Terrestrial Manual and describe how the Veterinary Services supervise and control all CBPP related activities. Provide maps and tables wherever possible.
   
   c) Role of farmers, industry and other relevant groups in CBPP surveillance and control (include a description of training and awareness programmes on CBPP).
   
   d) Role of private veterinary profession in CBPP surveillance and control.

3. CBPP eradication
   a) History. Provide a description of the CBPP history in the zone, date of first detection, origin of infection, date of eradication (date of last case).
   
   b) Strategy. Describe how CBPP was controlled and eradicated in the zone (e.g. stamping-out, modified stamping-out, zoning) and provide timeframe for eradication.
   
   c) Vaccines and vaccination. Was CBPP vaccine ever used? In the entire country? If vaccination was used, when was the last vaccination carried out? Where in the country?
   
   d) Legislation, organisation and implementation of the CBPP eradication campaign. Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines exist and give a brief summary.
   
   e) Animal identification and movement control. Are susceptible animals identified (individually or at a group level)? Provide a description of the methods of animal identification, herd registration and traceability. How are animal movements controlled in the zone? Provide evidence on the effectiveness of animal identification and movement controls. Please provide information on pastoralism, transhumance and the related paths of movement.
Annex XXIII (contd)

4. CBPP diagnosis

Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3. and 2.4.9. of the Terrestrial Manual are applied. In particular, the following points should be addressed:

a) Is CBPP laboratory diagnosis carried out in the country? If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to, the follow-up procedures and the time frame for obtaining results.

b) Provide an overview of the CBPP approved laboratories, in particular to address the following points:

i) Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO, etc. that exist in, or planned for, the laboratory system.

ii) Give details of participation in inter-laboratory validation tests (ring tests).

iii) Biosecurity measures applied.

iv) Details of the type of tests undertaken including procedures to isolate and identify M. mycoides subsp. mycoides SC as opposed to M. mycoides subsp. mycoides LC.

5. CBPP surveillance

Provide documentary evidence that surveillance for CBPP in the country complies with the provisions of Articles 11.8.12. to 11.8.17. of the Terrestrial Code and Chapter 2.4.9. of the Terrestrial Manual. In particular, the following points should be addressed:

a) Clinical surveillance. What are the criteria for raising a suspicion of CBPP? What is the procedure to notify (by whom and to whom) and what penalties are involved for failure to report? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for CBPP agent, species, type of sample, testing method(s) and results (including differential diagnosis).

b) Slaughterhouses, slaughter slabs, abattoirs. What are the criteria for raising a suspicion of CBPP lesion? What is the procedure to notify (by whom and to whom)? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for CBPP agent, species, type of sample, testing method(s) and results (including differential diagnosis).

c) Provide details on training programmes for personnel involved in clinical and slaughter facilities surveillance, and the approaches used to increase community involvement in CBPP surveillance programmes.

d) For countries where a significant proportion of animals in the zone are not slaughtered in controlled abattoirs, what are the alternative surveillance measures applied to detect CBPP (e.g. active clinical surveillance programme, laboratory follow-up).

e) Livestock demographics and economics. What is the susceptible animal population by species and production systems? How many herds of each susceptible species are in the zone? How are they distributed (e.g. herd density, etc.)? Provide tables and maps as appropriate.

f) Slaughterhouses and markets. Where are the major livestock marketing or collection centres? What are the patterns of livestock movement within the country and the zone? How are the animals transported and handled during these transactions?
Annex XXIII (contd)

6. CBPP prevention

a) Coordination with neighbouring countries and zones. Are there any relevant factors about the adjacent countries and zones that should be taken into account (e.g. size, distance from adjacent border to affected herds or animals)? Describe coordination, collaboration and information sharing activities with neighbouring countries and zones. If the CBPP free zone is situated in a CBPP infected country or borders an infected country or zone, describe the animal health measures implemented to effectively prevent the introduction of the agent, taking into consideration physical or geographical barriers.

b) Import control procedures

From what countries or zones does the country authorize the import of susceptible animals? What criteria are applied to approve such countries or zones? What controls are applied on entry of such animals, and subsequent internal movement? What import conditions and test procedures are required? Are imported animals of susceptible species required to undergo a quarantine or isolation period? If so, for how long and where? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports of susceptible animals for the past 2 years, specifying country or zone of origin, species and volume.

i) Provide a map with the number and location of ports, airports and land crossings. Is the official service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the central Veterinary Services. Describe the communication systems between the central authorities and the border inspection posts, and between border inspection posts.

ii) Describe the regulations, procedures, type and frequency of checks at the point of entry into the zone and/or their final destination, concerning the import and follow-up of the following:

- animals,
- veterinary medicinal products (i.e. biologics).

iii) Describe the action available under legislation, and actually taken, when an illegal import is detected. Provide information on detected illegal imports.

7. Control measures and contingency planning

a) Give details of any written guidelines, including contingency plans, available to the official services for dealing with suspected or confirmed outbreaks of CBPP.

b) Is quarantine imposed on premises with suspicious cases, pending final diagnosis? What other procedures are followed regarding suspicious cases?

c) In the event of a CBPP outbreak:

i) indicate the sampling and testing procedures used to identify and confirm presence of the causative agent;
Annex XXIII (contd)

ii) describe the actions taken to control the disease situation in and around any holdings found to be infected with CBPP;

iii) indicate the control and/or eradication procedures (e.g. vaccination, stamping-out, partial slaughter/vaccination, etc.) that would be taken;

iv) describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including any restrictions on restocking;

v) give details of any compensation payments made available to farmers, etc. when animals are slaughtered for disease control/eradication purposes.

8. Compliance with the Terrestrial Code

In addition to the documentary evidence that the provisions of Article 11.8.3. are properly implemented and supervised, the Delegate of the country must submit a declaration indicating that in the zone:

a) no clinical CBPP has been detected for at least 2 years;

b) no CBPP vaccines have been used for at least 2 years in any susceptible species;

c) the country operates both clinical surveillance and disease reporting systems for CBPP adequate to detect clinical disease if it were present in the zone;

d) all clinical and pathological suggestive of CBPP is investigated by field and laboratory methods (including serological assessment) to refute a possible diagnosis of CBPP;

e) there are effective measures in force to prevent the re-introduction of the disease.

9. Recovery of status

Countries applying for recovery of status should comply with the provisions of Article 11.8.4. of the Terrestrial Code and provide detailed information as specified in sections 3.a), 3.b), 3.c), 5.b), 5.c) and 5.d) of this questionnaire. Information in relation to other sections need only be supplied if relevant.

1. Accounts of the ages for eruption of the incisor teeth vary markedly and are clearly dependent on species, breed, nutritional status and nature of the feed. Therefore, for the purposes of serosurveillance, it should be noted that a) cattle having only one pair of erupted permanent central incisor teeth are aged between 21 and 36 months (Asian buffalos 24-48 months) and b) cattle having only two pairs of erupted permanent central incisor teeth are aged between 30 and 48 months (Asian buffalos 48-60 months).
CHAPTER 11.12.

LUMPY SKIN DISEASE
(cause by group III virus, type Neethling)

Article 11.12.1.

General provisions

For the purposes of the Terrestrial Code, the incubation period for lumpy skin disease (LSD) shall be 28 days.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

When authorising import or transit of the commodities covered in the chapter, with the exception of those listed in Article 11.12.1.bis, Veterinary Authorities should require the conditions prescribed in this chapter relevant to the LSD status of the cattle population of the exporting country.

Article 11.12.1.bis

Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any LSD related conditions regardless of the LSD status of the cattle population of the exporting country or zone:

1. milk and milk products;
2. meat and meat products.

Article 11.12.2.

LSD free country

A country may be considered free from LSD when:

1. LSD is notifiable in the country;
2. no case of LSD has been confirmed for at least the past 3 years;
3. no vaccination against LSD has been performed for at least 3 years;
4. commodities have been imported in accordance with this chapter.

Article 11.12.3.

Trade in commodities

Veterinary Authorities of LSD free countries may prohibit importation or transit through their territory, from countries considered infected with LSD, of the following commodities:

1. domestic and wild animals of the bovine species;
2. semen of animals of the bovine species.
Annex XXIV (contd)

Article 11.12.4.

Recommendations for importation from LSD free countries

for domestic cattle

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:
1. showed no clinical sign of LSD on the day of shipment;
2. come from an LSD free country.

Article 11.12.5.

Recommendations for importation from LSD free countries

for wild cattle

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:
1. showed no clinical sign of LSD on the day of shipment;
2. come from an LSD free country;
if the country of origin has a common border with a country considered infected with LSD:
3. were kept in a quarantine station for the 28 days prior to shipment.

Article 11.12.6.

Recommendations for importation from countries considered infected with LSD

for domestic cattle

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:
1. showed no clinical sign of LSD on the day of shipment;
2. either:
   a. were not vaccinated against LSD during the 30 days and were tested negative using tests according to the Terrestrial Manual within 14 days prior to shipment; or
   b. were vaccinated against LSD not more than 3 months between 30 days and 90 days prior to shipment;
   AND
   4. either:
      a. were kept since birth, or for the past 28 days, in an establishment where no case of LSD was officially reported during that period; or
      b. were kept in a quarantine station for the 28 days prior to shipment.

Article 11.12.7.

Recommendations for importation from countries considered infected with LSD

for wild cattle
Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of LSD on the day of shipment;

2. were kept in a quarantine station for the 28 days prior to shipment.

Article 11.12.8.

**Recommendations for importation from LSD free countries**

for semen of cattle

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a. showed no clinical sign of LSD on the day of collection of the semen;
   b. were kept for at least 28 days prior to collection in an LSD free country;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 11.12.9.

**Recommendations for importation from countries considered infected with LSD**

for semen of cattle

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a. showed no clinical sign of LSD on the day of collection of the semen and for the following 28 days;
   b. were kept in the exporting country for the 28 days prior to collection, in an establishment or artificial insemination centre where no case of LSD was officially reported during that period, and that the establishment or artificial insemination centre was not situated in an LSD infected zone.

c. and either:

   i) were vaccinated against LSD between 28 days and 90 days before semen collection and thereafter vaccinated annually; or

   ii) were tested with negative results using a serum neutralisation test (SNT) or an indirect enzyme-linked immunosorbent assay (ELISA) for LSD on the day of first semen collection or up to 90 days after last collection; or

   iii) showed stable seropositivity (not more than a two-fold rise in titre) on paired samples (tested side by side) to indirect ELISA or SNT carried out in quarantine, 28-60 days apart, with the first sample taken on the day of first semen collection;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.
Annex XXIV (contd)

Article 11.12.10.

**Recommendations for importation from LSD free countries**

**for embryos/oocytes of cattle**

Veterinary Authorities should require the presentation of an *international veterinary certificate* attesting that:

1. the donor *animals* showed no clinical sign of LSD on the day of collection of the embryos/oocytes; and

2. the embryos/oocytes were collected, processed and stored in conformity with the provisions of Chapters 4.7., 4.8. and 4.9., as relevant.

Article 11.12.11.

**Recommendations for importation from countries considered infected with LSD**

**for embryos/oocytes of cattle**

Veterinary Authorities should require the presentation of an *international veterinary certificate* attesting that:

1. the donor *animals*:
   a. were kept in an *establishment* where no *case* of LSD has been reported during the 28 days prior to collection; and
   b. showed no clinical sign of LSD on the day of collection;
   c. and either:
      i. were vaccinated against LSD between 30 28 days and 90 days before first embryo/oocyte collection and thereafter vaccinated annually; or
      ii. were tested with negative results using a serum neutralisation test (SNT) or an indirect enzyme-linked immunosorbent assay (ELISA) for LSD on the day of embryo/oocyte collection or up to 90 days after last collection according to the *Terrestrial Manual*; or
      iii. showed serostability (stable seropositivity (not more than a two-fold rise in titre) on paired samples (to indirect ELISA tests, tested side by side, carried out in isolation) to indirect ELISA or SNT carried out in quarantine, 14 28–60 days apart with one of the samples taken on the day of embryo/oocyte collection of the embryos/oocytes;

2. the embryos/oocytes were collected, processed and stored in conformity with the provisions of Chapters 4.7., 4.8. and 4.9., as relevant.

Article 11.12.12.

**Recommendations for importation from LSD free countries**

**for products of animal origin (from cattle) intended for agricultural or industrial use**

Veterinary Authorities should require the presentation of an *international veterinary certificate* attesting that these products come from *animals* which have been kept in an LSD free country since birth or for at least the past 28 days.
Recommendations for importation from countries considered infected with LSD

for products of animal origin (from cattle) intended for agricultural or industrial use

_Veterinary Authorities_ should require the presentation of an _international veterinary certificate_ attesting that these products have been processed to ensure the destruction of the LSD virus.


Recommendations for importation from countries considered infected with LSD

for raw hides of cattle

_Veterinary Authorities_ should require the presentation of an _international veterinary certificate_ attesting that these products were stored for at least 40 days before shipment.

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Annex XXV

CHAPTER 12.1.

AFRICAN HORSE SICKNESS

Article 12.1.1.

General provisions

For the purposes of the Terrestrial Code, the infective period for African horse sickness virus (AHSV) shall be 40 days for domestic horses. Although critical information is lacking for some species, this chapter applies to all equidae.

All countries or zones neighbouring adjacent to, or considered to be at risk from, a country or zone not having free status should determine their AHSV status from an ongoing surveillance programme. Throughout the chapter, surveillance is in all cases understood as being conducted as described in Chapter 1.4. Article 12.11.1. to 12.11.3.

The following defines a case of AHS:

1. AHSV has been isolated and identified from an equid or a product derived from that equid; or

2. viral antigen or viral RNA specific to one or more of the serotypes of AHSV has been identified in samples from one or more equids showing clinical signs consistent with AHS, or epidemiologically linked to a suspected or confirmed case; or

3. serological evidence of active infection with AHSV by detection of seroconversion with production of antibodies to structural or nonstructural proteins of AHSV that are not a consequence of vaccination have been identified in one or more equids that either show clinical signs consistent with AHS, or epidemiologically linked to a suspected or confirmed case.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 12.1.2.

AHSV free country or zone

1. A country or zone may be considered free from AHSV when African horse sickness (AHS) is notifiable in the whole country, systematic vaccination is prohibited, importation of equidae and their semen, oocytes or embryos are carried out in accordance with this chapter, and either:

   a) historical freedom as described in Chapter 1.4. has demonstrated no evidence of AHSV in the country or zone, or

   b) the country or zone has not reported any case of AHS for at least 2 years and is not adjacent to a country or zone not having a free status; or

   c) a surveillance programme has demonstrated no evidence of AHSV in the country or zone for at least 12 months and includes a complete season of vector activity; or

   d) the country or zone has not reported any case of AHS for at least 40 days and a surveillance programme has demonstrated no evidence of Culicoides likely to be competent AHSV vectors for at least 2 years in the country or zone.
Annex XXV (contd)

22. A AHS free country or zone adjacent to an infected country or infected zone should include a zone in which surveillance is conducted in accordance with Articles 12.1.11. to 12.1.13. Animals within this zone should be subjected to continuing surveillance. The boundaries of this zone should be clearly defined, and should take account of geographical and epidemiological factors that are relevant to AHS transmission.

23. An AHSV free country or zone will not lose its free status through the importation of vaccinated or seropositive equidae and their semen, oocytes or embryos from infected countries or infected zones, provided these imports are carried out in accordance with this chapter.

4. To qualify for inclusion in the existing list of AHSV free countries or zones, a Member should:
   a) have a record of regular and prompt animal disease reporting;
   b) send a declaration to the OIE stating:
      i) the section under paragraph 1 on the base of which the application is made;
      ii) no systematic vaccination against AHS has been carried out during the past 12 months in the country or zone;
      iii) equidae are imported in accordance with paragraph 3 above;
   c) supply documented evidence that:
      i) surveillance for both AHS and AHSV infection in accordance with Articles 12.1.11. to 12.1.13 is in operation;
      ii) regulatory measures for the early detection, prevention and control of AHS have been implemented.

The Member will be included in the list only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information in points 4b(ii) and iii) and 4c) above be re-submitted annually and changes in the epidemiological situation or other significant events should be reported to the OIE according to the requirements in Chapter 1.1., and in particular, formally state that:

1. there has been no outbreak of AHS during the past 12 months in the country or zone;

2. no evidence of AHSV infection has been found during the past 12 months in the country or zone.

Article 12.1.3.

AHSV seasonally free zone

1. An AHSV seasonally free zone is a part of an infected country or an infected zone in which for part of a year, ongoing surveillance and monitoring consistently demonstrated neither evidence of AHSV transmission nor the evidence of the presence of adult Culicoides likely to be competent AHSV vectors.

2. For the application of Articles 12.1.6., 12.1.8. and 12.1.9., the seasonally free period is:
   a) taken to commence the day following the last evidence of AHSV transmission and of the cessation of activity of adult Culicoides likely to be competent AHSV vectors as demonstrated by an ongoing surveillance programme, and
b) taken to conclude either:

i) at least 40 days before the earliest date that historical data show AHSV activity has recommenced; or

ii) immediately when current climatic data or data from a surveillance and monitoring programme indicate an earlier resurgence of activity of adult Culicoides likely to be competent AHSV vectors.

3. An AHSV seasonally free zone will not lose its free status through the importation of vaccinated or seropositive equidae and their semen, oocytes or embryos from infected countries or infected zones, provided these imports are carried out in accordance with this chapter.

**Article 12.1.4.**

**AHSV infected country or zone**

For the purpose of this chapter, an AHSV infected country or infected zone is one that does not fulfil the requirements to qualify as either AHSV free country or zone or AHSV seasonally free zone in which the conditions of Article 12.1.2. or Article 12.1.3. do not apply.

**Article 12.1.4bis.**

**Establishment of a containment zone within an AHS free country or zone**

In the event of limited outbreaks within an AHS free country or zone, including within a protection zone, a single containment zone, which includes all cases, can be established for the purpose of minimizing the impact on the entire country or zone. For this to be achieved, the Veterinary Authority should provide documented evidence that:

1. the outbreaks are limited based on the following factors:
   
   a) immediately on suspicion, a rapid response including notification has been made;
   
   b) standstill of movements of equidae has been imposed, and effective controls on the movement of equidae and their products mentioned in this chapter are in place;
   
   c) epidemiological investigation (trace-back, trace-forward) has been completed;
   
   d) the infection has been confirmed;
   
   e) the primary outbreak and likely source of the outbreak has been identified;
   
   f) all cases have been shown to be epidemiologically linked;
   
   g) no new cases have been found in the containment zone within a minimum of two infectious periods as defined in Article 12.1.1.;

2. the equidae within the containment zone should be clearly identifiable as belonging to the containment zone;

3. increased passive and targeted surveillance in accordance with Articles 12.1.11. to 12.1.13. has increased in the rest of the country or zone and has not detected any evidence of infection;

4. animal health measures that effectively prevent the spread of the AHS to the rest of the country or zone, taking into consideration the establishment of a protection zone within the containment zone, the seasonal vector conditions and existing physical, geographical and ecological barriers;

5. ongoing surveillance is in place in the containment zone.
Annex XXV (contd)

The free status of the areas outside the containment zone is suspended pending the establishment of the containment zone in accordance with points 1 to 5 above. The free status of the areas outside the containment zone could be reinstated irrespective of the provisions of Article 12.1.4tris, once the containment zone is recognised by OIE.

The recovery of the AHS free status of the containment zone should follow the provisions of Article 12.1.4tris.

Article 12.1.4tris

Recovery of free status

When an AHS outbreak occurs in an AHS free country or zone, the following waiting period is required to regain the status of AHS free country or zone:

1. 12 months after the last case and completion of the emergency vaccination and where surveillance, applied in accordance with Articles 12.1.11. to 12.1.13., has shown no evidence of AHSV infection; or
2. the conditions of Article 12.1.2 apply.

Article 12.1.5.

Recommendations for importation from AHSV free countries that are neither neighbouring nor considered to be at risk from an AHSV infected country or infected zone for equidae

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of AHS on the day of shipment;
2. have not been vaccinated against AHS within the last 40 days;
3. were kept in an AHSV free country or zone since birth or for at least 40 days prior to shipment;
4. either:
   a) did not transit through an infected country or infected zone during transportation to the place of shipment; or
   b) were protected from attacks by Culicoides at all times when transiting through an infected country or infected zone.

Article 12.1.6.

Recommendations for importation from AHSV free countries or free zones or from AHSV seasonally free zones (during the seasonally free period) that are neighbouring or are considered to be at risk from an AHSV infected country or infected zone for equidae

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical signs of AHS on the day of shipment;
2. have not been vaccinated against AHS within the last 40 days;
3. and either:
Annex XXV (contd)

a. were kept in an AHSV free country, free zone or seasonally free zone during the seasonally free period since birth or for at least 40 days prior to shipment; or

4b. in a country or zone considered to be at risk, were held in quarantine isolation for at least 40 days prior to shipment and protected at all times from attacks by Culicoides; and

a. for a period at least 28 days and a serological test according to the Terrestrial Manual to detect antibodies to the AHSV group, was carried out with a negative result on a blood sample collected at least 28 days after introduction into the quarantine station; or

b. for a period at least 40 days and serological tests according to the Terrestrial Manual to detect antibodies against AHSV were carried out with no significant increase in antibody titre on blood samples collected on two occasions, with an interval of not less than 21 days, the first sample being collected at least 7 days after introduction into the quarantine station; or

c. for a period at least 14 days and an agent identification test according to the Terrestrial Manual were was carried out with a negative result on a blood sample collected on two occasions with an interval of not less than 14 days between collection, the first sample being collected at least 7 days after introduction into the quarantine station;

4d. were protected from attacks by Culicoides at all times during transportation (including to and at the place of shipment) when transiting through an infected zone.

Article 12.1.7.

Recommendations for importation from AHSV infected countries or zones for equidae

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of AHS on the day of shipment;

2. have not been vaccinated against AHS within the last 40 days;

3. were held continuously during the quarantine period of at least 40 days, in a vector proof protected quarantine station and protected at all times from attacks by Culicoides; and

a) for a period at least 28 days and a serological test according to the Terrestrial Manual to detect antibodies to the AHSV group, was carried out with a negative result on a blood sample collected at least 28 days after introduction into the quarantine station; or

b) for a period at least 40 days and serological tests according to the Terrestrial Manual to detect antibodies against AHSV were carried out with no significant increase in antibody titre on blood samples collected on two occasions, with an interval of not less than 21 days, the first sample being collected at least 7 days after introduction into the quarantine station; or

c) for a period at least 14 days and an agent identification test according to the Terrestrial Manual were was carried out with a negative result on a blood sample collected on two occasions with an interval of not less than 14 days between collection, the first sample being collected at least 7 days after introduction into the quarantine station;

4. were protected from attacks by Culicoides at all times during transportation (including transportation to and at the place of shipment).
Annex XXV (contd)

Article 12.1.8.

Recommendations for the importation of equid semen

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the donor animals:

1. showed no clinical sign of AHS on the day of collection of the semen and for the following 40 days;

2. had not been immunised against AHS with a live attenuated vaccine within 40 days prior to the day of collection;

3. were either:

   a) kept in an AHSV free country or free zone or from an AHSV seasonally free zone (during the seasonally free period) for at least 40 days before commencement of, and during collection of the semen, or

   b) kept in an AHSV free vector-proof protected artificial insemination centre throughout the collection period, and subjected to either:

      i) a serological test according to the Terrestrial Manual to detect antibody to the AHSV group, carried out with a negative result on a blood sample collected at least 28 days and not more than 90 days after the last collection of semen; or

      ii) agent identification tests according to the Terrestrial Manual carried out with negative results on blood samples collected at commencement and conclusion of, and at least every 7 days, during semen collection for this consignment.

Article 12.1.9.

Recommendations for the importation of in vivo derived equid embryos/oocytes

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:

   a) showed no clinical sign of AHS on the day of collection of the embryos/oocytes and for the following 40 days;

   b) had not been immunised against AHS with a live attenuated vaccine within 40 days prior to the day of collection;

   c) were either:

      i) kept in an AHSV free country or free zone or from an AHSV seasonally free zone (during the seasonally free period) for at least 40 days before commencement of, and during collection of the embryos/oocytes, or
ii) kept in an AHSV free vector-proof protected collection centre throughout the collection period, and subjected to either:

- a serological test according to the *Terrestrial Manual* to detect antibody to the AHSV group carried out with a negative result on a blood sample collected at least 28 days and not more than 90 days after the last collection of embryos/oocytes; or

- agent identification tests according to the *Terrestrial Manual* carried out with negative results on blood samples collected at commencement and conclusion of, and at least every 7 days during embryos/oocytes collection for this consignment;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapter 4.7. or Chapter 4.9., as relevant;

3. semen used to fertilize the oocytes, complies at least with the requirements in Article 12.1.8.

Article 12.1.10.

**Protecting animals from *Culicoides* attack**

1. **Vector-protected establishment or facility**

   The means of protection of the *establishment* or facility should at least comprise the following:

   a) double-door entry-exit system;

   b) openings of the building are vector screened with mesh of appropriate aperture size (under study) impregnated regularly with an approved insecticide according to manufacturers’ instruction;

   c) vector surveillance and control within and around the building;

   d) measures to limit breeding sites for vectors in vicinity of the *establishment* or facility;

   e) Standard Operating Procedure, including description of back-up and alarm systems, for operation of the *establishment* or facility and transport of horses to the place of loading.

2. **During transportation**

   When transporting equines through AHSV infected countries or AHSV infected zones, *Veterinary Authorities* should require strategies to protect animals from attacks by *Culicoides* during transport, taking into account the local ecology of the vector.

   a) **Transport by road**

      Potential risk management strategies include a combination of:

      1. treating animals with chemical repellents prior to and during transportation, in sanitized *vehicles* treated with appropriate residual contact insecticide;

      2. loading, transporting and unloading animals at times of low vector activity (i.e. bright sunshine and low temperature);

      3. ensuring *vehicles* do not stop en route during dawn or dusk, or overnight, unless the animals are held behind insect proof netting;
Annex XXV (contd)

4iv. darkening the interior of the vehicle, for example by covering the roof and/or sides of vehicles with shade cloth;

5v. monitoring for vectors at common stopping and offloading points to gain information on seasonal variations;

6vi. using historical, ongoing and/or AHS modelling information to identify low risk ports and transport routes.

b) Transport by air

Prior to loading the equids, the crates, containers or jetstalls are sprayed with an insecticide approved in the country of dispatch.

Crates, containers or jet stalls in which equidae are being transported and the cargo hold of the aircraft must be sprayed with an approved insecticide just after the doors to the aircraft are closed and prior to takeoff.

In addition, during any stop over in countries or zones not free of AHS, prior to the opening of any aircraft door and until all doors are closed prior to takeoff, netting of appropriate aperture size (under study) impregnated with an approved insecticide must be placed over all crates, containers or jetstalls.

Surveillance: introduction

Articles 12.1.11. to 12.1.13. define the principles and provide a guide on the surveillance for AHS, complementary to Chapters 1.4. and 1.5., applicable to Members seeking to determine their AHSV status. This may be for the entire country or zone. Guidance for Members seeking free status following an outbreak and for the maintenance of AHS status is also provided.

AHS is a vector-borne infection transmitted by a limited number of species of Culicoides insects. Unlike the related bluetongue virus, AHSV is so far geographically restricted to sub Saharan Africa with periodic excursions into North Africa, southwest Europe, the Middle East and adjacent regions of Asia. An important component of AHSV epidemiology is vectorial capacity which provides a measure of disease risk that incorporates vector competence, abundance, seasonal incidence, biting rates, survival rates and the extrinsic incubation period. However, methods and tools for measuring some of these vector factors remain to be developed, particularly in a field context.

According to this chapter, a Member demonstrating freedom from AHSV infection for the entire country or a zone should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and should be planned and implemented according to general conditions and methods described in this chapter. This requires the support of a laboratory able to undertake identification of AHSV infection through the virus detection and antibody tests described in the Terrestrial Manual.

Susceptible captive wild, feral and wild equid populations should be included in the surveillance programme.

For the purposes of surveillance, a case refers to an equid infected with AHSV.

The purpose of surveillance is to determine if a country or zone is free from AHSV or if a zone is seasonally free from AHSV. Surveillance deals not only with the occurrence of clinical signs caused by AHSV, but also with evidence of infection with AHSV in the absence of clinical signs.
The following defines the occurrence of AHSV infection:

1. AHSV has been isolated and identified as such from an equid or a product derived from that equid, or

2. Viral antigen or viral RNA specific to one or more of the serotypes of AHSV has been identified in samples from one or more equids showing clinical signs consistent with AHS, or epidemiologically linked to a confirmed or suspected case, or

3. Serological evidence of active infection with AHSV by detection of seroconversion with production of antibodies to structural or non-structural proteins of AHSV that are not a consequence of vaccination have been identified in one or more equids that either show clinical signs consistent with AHS, or epidemiologically linked to a suspected case.

Article 12.1.12.

Surveillance: general conditions and methods

1. A surveillance system should be under the responsibility of the Veterinary Authority. In particular the following should be in place:

   a) A formal and ongoing system for detecting and investigating outbreaks of disease;

   b) A procedure for the rapid collection and transport of samples from suspect cases of AHS to a laboratory for AHS diagnosis as described in the Terrestrial Manual;

   c) A system for recording, managing and analysing diagnostic, epidemiological and surveillance data.

2. The AHS surveillance programme should:

   a) in a country/zone, free or seasonally free, include an early warning system for reporting suspicious cases. Persons who have regular contact with equids, as well as diagnosticians, should report promptly any suspicion of AHS to the Veterinary Authority. An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is AHS. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. All suspected cases of AHS should be investigated immediately and samples should be taken and submitted to a laboratory. This requires that sampling kits and other equipment are available for those responsible for surveillance;

   b) conduct random or targeted serological and virological surveillance appropriate to the infection status of the country or zone in accordance with Chapter 1.4.

Article 12.1.13.

Surveillance strategies

The target population for surveillance aimed at identification of disease and/or infection should cover susceptible equids within the country or zone. Active and passive surveillance for AHSV infection should be ongoing. Surveillance should be composed of random or targeted approaches using virological, serological and clinical methods appropriate for the infection status of the country or zone.

A Member should justify the surveillance strategy chosen as appropriate to detect the presence of AHSV infection in accordance with Chapter 1.4 and the prevailing epidemiological situation. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clinical signs (e.g. horses). Similarly, virological and serological testing may be targeted to species that rarely show clinical signs (e.g. donkeys).
Annex XXV (contd)

In vaccinated populations serological and virological surveillance is necessary to detect the AHSV types circulating to ensure that all circulating types are included in the vaccination programme.

If a Member wishes to declare freedom from AHSV infection in a specific zone, the design of the surveillance strategy would need to be aimed at the population within the zone.

For random surveys, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect infection if it were to occur at a predetermined minimum rate. The sample size, expected prevalence and diagnostic sensitivity of the tests determine the level of confidence in the results of the survey. The Member must justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Chapter 1.4. Selection of the design prevalence, in particular, needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey approach selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and the different species in the target population.

Irrespective of the testing system employed, surveillance system design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of infection or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as those which may be epidemiologically linked to it.

The principles for surveillance for disease/infection are technically well defined. Surveillance programmes to prove the absence of AHSV infection/circulation, need to be carefully designed to avoid producing results that are either insufficiently reliable to be accepted by international trading partners, or excessively costly and logistically complicated. The design of any surveillance programme, therefore, requires inputs from professionals competent and experienced in this field.

1. Clinical surveillance

Clinical surveillance aims at the detection of clinical signs of AHS in equids particularly during a newly introduced infection. In horses, clinical signs may include pyrexia, oedema, hyperaemia of mucosal membranes and dyspnoea.

AHS suspects detected by clinical surveillance should always be confirmed by laboratory testing.

2. Serological surveillance

Serological surveillance of equid populations is an important tool to confirm absence of AHSV transmission in a country or zone. The species tested should reflect the local epidemiology of AHSV infection, and the equine species available. Management variables that may reduce the likelihood of infection, such as the use of insecticides and animal housing, should be taken into account when selecting equids to be included in the surveillance system.

Samples should be examined for antibodies against AHSV using tests prescribed in the Terrestrial Manual. Positive AHSV antibody tests results can have four possible causes:

a) natural infection with AHSV;

b) vaccination against AHSV;
c) maternal antibodies;

d) positive results due to the lack of specificity of the test.

It may be possible to use sera collected for other purposes for AHSV surveillance. However, the principles of survey design described in these recommendations and the requirements for a statistically valid survey for the presence of AHSV infection should not be compromised.

The results of random or targeted serological surveys are important in providing reliable evidence that no AHSV infection is present in a country or zone. It is, therefore, essential that the survey is thoroughly documented. It is critical to interpret the results in light of the movement history of the animals being sampled.

Serological surveillance in a free zone should target those areas that are at highest risk of AHSV transmission, based on the results of previous surveillance and other information. This will usually be towards the boundaries of the free zone. In view of the epidemiology of AHSV, either random or targeted sampling is suitable to select herds and/or animals for testing.

Serological surveillance in a free country or zone should be carried out over an appropriate distance from the border with an infected country or infected zone, based upon geography, climate, history of infection and other relevant factors. The surveillance should be carried out over a distance of at least 100 kilometres from the border with that country or zone, but a lesser distance could be acceptable if there are relevant ecological or geographical features likely to interrupt the transmission of AHSV. An AHSV free country or zone may be protected from an adjacent infected country or infected zone by a protection zone.

Serological surveillance in infected zones will identify changes in the boundary of the zone, and can also be used to identify the AHSV types circulating. In view of the epidemiology of AHSV infection, either random or targeted sampling is suitable.

3. **Virological surveillance**

   Isolation and genetic analysis of AHSV from a proportion of infected animals is beneficial in terms of providing information on serotype and genetic characteristics of the viruses concerned.

   Virological surveillance using tests described in the *Terrestrial Manual* can be conducted:

   a) to identify virus circulation in at risk populations;

   b) to confirm clinically suspect cases;

   c) to follow up positive serological results;

   d) to better characterize the genotype of circulating virus in a country or zone.

4. **Sentinel animals**

   Sentinel animals are a form of targeted surveillance with a prospective study design. They comprise groups of unexposed equids that are not vaccinated and are managed at fixed locations and observed and sampled regularly to detect new AHSV infections.

   The primary purpose of a sentinel equid programme is to detect AHSV infections occurring at a particular place, for instance sentinel groups may be located on the boundaries of infected zones to detect changes in distribution of AHSV. In addition, sentinel equid programmes allow the timing and dynamics of infections to be observed.
A sentinel equid programme should use animals of known source and history of exposure, control management variables such as use of insecticides and animal housing (depending on the epidemiology of AHSV in the area under consideration), and be flexible in its design in terms of sampling frequency and choice of tests.

Care is necessary in choosing the sites for the sentinel groups. The aim is to maximise the chance of detecting AHSV activity at the geographical location for which the sentinel site acts as a sampling point. The effect of secondary factors that may influence events at each location, such as climate, may also be analysed. To avoid confounding factors sentinel groups should comprise animals selected to be of similar age and susceptibility to AHSV infection. The only feature distinguishing groups of sentinels should be their geographical location. Sera from sentinel animal programmes should be stored methodically in a serum bank to allow retrospective studies to be conducted in the event of new serotypes being isolated.

The frequency of sampling should reflect the equid species used and the reason for choosing the sampling site. In endemic areas virus isolation will allow monitoring of the serotypes and genotypes of AHSV circulating during each time period. The borders between infected and non infected areas can be defined by serological detection of infection. Monthly sampling intervals are frequently used. Sentinels in declared free zones add to confidence that AHSV infections are not occurring unobserved. Here sampling prior to and after the possible period of transmission is sufficient.

Definitive information on AHSV circulating in a country or zone is provided by isolation and identification of the viruses. If virus isolation is required sentinels should be sampled at sufficiently frequent intervals to ensure that some samples are collected during the period of viraemia.

5. **Vector surveillance**

AHSV is transmitted between equine hosts by species of *Culicoides* which vary across the world. It is therefore important to be able to identify potential vector species accurately although many such species are closely related and difficult to differentiate with certainty.

The main purpose of vector surveillance is to define high, medium and low-risk areas and local details of seasonality by determining the various species present in an area, their respective seasonal occurrence, and abundance. Vector surveillance has particular relevance to potential areas of spread. Long term surveillance can also be used to assess vector abatement measures.

The most effective way of gathering this information should take account of the biology and behavioural characteristics of the local vector species of *Culicoides* and may include the use of Onderstepoort-type light traps or similar, operated from dusk to dawn in locations adjacent to equids.

Vector surveillance should be based on scientific sampling techniques. The choice of the number and types of traps to be used in vector surveillance and the frequency of their use should take into account the size and ecological characteristics of the area to be surveyed.

The operation of vector surveillance sites at the same locations as sentinel animals is advisable.

The use of a vector surveillance system to detect the presence of circulating virus is not recommended as a routine procedure as the typically low vector infection rates mean that such detections can be rare. Other surveillance strategies are preferred to detect virus circulation.
CHAPTER 1.6.

STATUS FOR OIE LISTED DISEASES:
PROCEDURES FOR SELF DECLARATION AND
FOR OFFICIAL RECOGNITION BY THE OIE

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Article 1.6.6.

Questionnaire on African horse sickness

AHS FREE COUNTRY

Report of a Member which applies for recognition of status, under Chapter 12.1. of the Terrestrial Animal Health Code (2010), as a AHS free country

Please address concisely the following topics. National legislation, regulations and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. Introduction

   a. Geographical factors. Provide a general description of the country including physical, geographical and other factors that are relevant to AHS introduction. Provide a map identifying the factors above.
   
        b. Equine sector. Provide a general description of the equine sector and their relative economic importance in the country. Outline significant changes observed (if relevant documents are available, please attach).

               i. Sport and race horses
               ii. Breeding stock equidae

               iii. Working and production equidae (including horses for slaughter)
               iv. Leisure equidae
               v. Captive wild, wild and feral equidae

2. Description of equid population

   a. Demographics of domestic equidae. What is the equidae population by species within the various sectors? Provide a description of the methods of animal identification, holding and individual animal registration systems if in place. How are they distributed (e.g. density, etc.)? Provide tables and maps as appropriate.

   b. Wildlife demographics. What captive wild, wild or feral equidae are present in the country? Provide estimates of population sizes and geographic distribution. What are the measures in place to prevent contact between domestic and captive wild, wild or feral equidae?
Annex XXV (contd)

3. **Veterinary system**
   
a. Legislation. Provide a list and summary of all relevant veterinary legislation in relation to AHS.

b. Veterinary Services. Provide documentation on the compliance of the *Veterinary Service* of the country with the provisions of Chapters 3.1. and 3.2. of the *Terrestrial Code* and 1.1.3. of the *Terrestrial Manual* and describe how the *Veterinary Services* supervise and control all AHS related activities. Provide maps and tables wherever possible.

c. Role of farmers, keepers, industry, regulatory bodies, and other relevant groups in AHS surveillance and control (include a description of training and awareness programmes on AHS).

d. Role of private veterinary profession in AHS surveillance and control.

4. **AHS eradication**
   
a. History. Provide a description of the AHS history in the country if applicable, date of first detection, origin of *infection*, date of eradication (date of last *case*), and serotypes present.

b. Strategy. Describe how AHS was controlled and eradicated (e.g. isolation of cases, *stamping-out policy*, zoning), provide time frame for eradication.

c. Vaccines and vaccination. What type of vaccine was used? What equine species were vaccinated? Were vaccinated animals marked or was vaccination recorded in a unique identification document?

d. Legislation, organisation and implementation of the AHS eradication campaign. Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines were used and give a brief summary.

e. Animal identification. Are equidae identified (individually or at a group level)?

f. Movements of equidae. How are movements of equidae controlled in the country? Provide evidence on the effectiveness of equidae identification and movement controls. Please provide information on pastoralism, transhumance and related movements.

g. Leisure and competition movements of equidae. How are movements of competition and leisure equidae controlled in the country. Please provide information on systems including any use of registration. Provide information on any events that include international movements of equidae.

h. Describe the market systems for equidae, in particular, if markets require the international movement of equidae.

5. **AHS diagnosis**

Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3., and 2.5.1. of the *Terrestrial Manual* are applied. In particular, the following points should be addressed:

a. Is AHS laboratory diagnosis carried out in the country? If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to, the follow-up procedures and the time frame for obtaining results.

b. Provide an overview of the AHS approved laboratories, in particular to address the following points:
   
i. Details on the types of tests undertaken.
ii. Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO that exist in, or planned for, the laboratory system.

iii. Give details of participation in inter-laboratory validation tests (ring tests).

iv. Describe biosecurity measures applied, particularly in the case where live virus is handled.

6. **AHS surveillance**

Provide documentary evidence that *surveillance* for AHS in the country complies with the provisions of Articles 12.1.11. to 12.1.13. of the *Terrestrial Code*, and Chapter 2.5.1. of the *Terrestrial Manual*. In particular, the following points should be addressed:

a. Clinical suspicion. What are the criteria for raising a suspicion of AHS? What is the procedure to notify (by whom and to whom), is there a compensation system in place and what penalties are involved for failure to report? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for AHS, species, type of sample, testing method(s) and results (including differential diagnosis).

b. Surveillance. Are the following undertaken?

i. Serological surveillance

ii. Virological surveillance

iii. Sentinel animals

iv. Vector surveillance

If so, provide detailed information on the survey designs. How frequently are they conducted? Which were the equine species included? Are wildlife species included? Provide a summary table indicating detailed results, for at least the past 2 years. Provide details on follow-up actions taken on all suspicious and positive results. Provide criteria for selection of populations for targeted surveillance and numbers of equidae examined and samples tested. Provide details on the methods selected and applied for monitoring the performance of the surveillance system.

7. **AHS prevention**

a. Coordination with neighbouring countries. Are there any relevant factors about the adjacent countries or zones that have been taken into account (e.g. size, distance from adjacent border to infected equidae)? Describe coordination, collaboration and information sharing activities with neighbouring countries.

If the AHS free country borders an infected country or zone, describe the animal health measures implemented to effectively prevent the introduction of the agent and/or vectors, taking into consideration the seasonal vector conditions and existing physical, geographical and ecological barriers.

b. Import control procedures

From what countries or zones does the country authorize the import of equidae or their products? What criteria are applied to approve such countries or zones? What controls are applied on entry of such equidae and products, and subsequent internal movement? What import conditions (e.g. quarantine) and test procedures are required? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports, temporary admissions or re-entry of equidae and their products for at least the past 2 years, specifying country or zone of origin and volume.
Annex XXV (contd)

i. Provide a map with the number and location of ports, airports and land crossings. Is the service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the Competent Authority. Describe the communication systems between the Competent Authority and the border inspection posts, and between border inspection posts.

ii. Describe the regulations, procedures, type and frequency of checks at the point of entry into the country and/or their final destination, concerning the import and follow-up of the following:

- Equidae,
- genetic material (semen, ova and embryos of the equine species),
- equine derived (by-)products and biological.

iii. Describe the action available under legislation, and actually taken, when an illegal introduction is detected. Provide information on detected illegal introduction.

8. Control measures and contingency planning

a. Give details of any written guidelines, contingency plans (including information on vaccine banks) available to the Competent Authority for dealing with suspected or confirmed cases of AHS.

b. In the event of a suspected or confirmed AHS outbreak:

   i. is quarantine imposed on premises with suspicious cases, pending final diagnosis?

   ii. are movement restrictions applied on suspicion?

   iii. describe the sampling and testing procedures used to identify and confirm presence of the causative agent;

   iv. describe the actions taken to control the disease situation in and around any holdings found to be infected with AHS;

   v. describe the control and/or eradication procedures (e.g. vaccination, modified stamping-out);

   vi. describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including conditions for restocking;

   vii. give details of any compensation made available when equidae are killed, for disease control/eradication purposes.

9. Compliance with the Terrestrial Code

a. In addition to the documentary evidence that the provisions of Article 12.1.2 are properly implemented and supervised, the Delegate of the country must submit a declaration stating:

   i. The section under paragraph 1 (of Article 12.1.2.) on the base of which the application is made;

   ii. there has been no outbreak of AHS during the past 12 months;

   iii. no systematic vaccination against AHS has been carried out during the past 12 months;

b. and that vaccinated equidae were imported in accordance with Chapter 12.1.
10. **Recovery of status**

Countries applying for recovery of status should comply with the provisions of Article 12.1.2. of the *Terrestrial Code* and provide detailed information as specified in sections 4(a), b), c and 6, and highlight any measures introduced to prevent a recurrence of the infection under section 7 of this questionnaire. Information in relation to other sections need only be supplied if relevant.

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**AHS FREE ZONE**

Report of a Member which applies for recognition of status, under Chapter 12.1. of the *Terrestrial Animal Health Code* (2010), as a AHS free zone

Please address concisely the following topics. National legislation, regulations and Veterinary Administration directives may be referred to and annexed as appropriate in one of the OIE official languages.

1. **Introduction**

   a. Geographical factors. Provide a general description of the country and the zone including physical, geographical and other factors that are relevant to AHS introduction. Provide a map identifying the factors above. The boundaries of the zone must be clearly defined, including a protection zone, if applied. Provide a digitalised, geo-referenced map with a precise text description of the geographical boundaries of the zone (and of the protection zone) established in accordance with Chapter 4.3.

   b. Equine sector. Provide a general description of the equine sector and their relative economic importance in the country and the zone. Outline significant changes observed (if relevant documents are available, please attach).

      i. Sport and race horses

      ii. Breeding stock equidae

      iii. Working and production equidae (including horses for slaughter)

      iv. Leisure equidae

      v. Captive wild, wild and feral equidae

2. **Description of equidae population**

   a. Demographics of domestic equidae. What is the equidae population by species within the various sectors in the country and the zone? Provide a description of the methods of animal identification, holding and individual animal registration systems in the country and the zone if in place. How are they distributed (e.g. density, etc.)? Provide tables and maps as appropriate.

   b. Wildlife demographics. What captive wild, wild or feral equidae are present in the country and the zone? Provide estimates of population sizes and geographic distribution. What are the measures in place to prevent contact between domestic and captive wild, wild or feral equidae?
Annex XXV (contd)

3. Veterinary system
   a. Legislation. Provide a list and summary of all relevant veterinary legislation in relation to AHS.
   b. Veterinary Services. Provide documentation on the compliance of the Veterinary Service of the country with the provisions of Chapters 3.1. and 3.2. of the Terrestrial Code and 1.1.3. of the Terrestrial Manual and describe how the Veterinary Services supervise and control all AHS related activities in the country and in the zone. Provide maps and tables wherever possible.
   c. Role of farmers, keepers, industry, regulatory bodies, and other relevant groups in AHS surveillance and control (include a description of training and awareness programmes on AHS).
   d. Role of private veterinary profession in AHS surveillance and control.

4. AHS eradication
   a. History. Provide a description of the AHS history in the country and zone, if applicable, date of first detection, origin of infection, date of eradication in the zone (date of last case), and serotypes present.
   b. Strategy. Describe how AHS was controlled and eradicated in the zone (e.g. isolation of cases, stamping-out policy, zoning), provide time frame for eradication.
   c. Vaccines and vaccination. What type of vaccine was used in the zone and the rest of the country? What equine species were vaccinated? Were vaccinated animals marked or was vaccination recorded in a unique identification document?
   d. Legislation, organisation and implementation of the AHS eradication campaign. Provide a description of the organizational structure at the different levels. Indicate if detailed operational guidelines were used and give a brief summary.
   e. Animal identification. Are equidae identified (individually or at a group level)?
   f. Movements of equidae. How are movements of equidae controlled in, and between zones of the country? Provide evidence on the effectiveness of equidae identification and movement controls in the zone. Please provide information on pastoralism, transhumance and related paths of movements.
   g. Leisure and competition movements of equidae. How are movements of competition and leisure equidae controlled in the country and the zones. Please provide information on systems including any use of registration. Provide information on any events that include international movements of equidae.
   h. Describe the market systems for equidae in the country and the zones, in particular, if markets require the international movement of equidae.

5. AHS diagnosis
   Provide documentary evidence that the provisions in Chapters 1.1.2., 1.1.3., and 2.5.1. of the Terrestrial Manual are applied in the country and the zone. In particular, the following points should be addressed:
   a. Is AHS laboratory diagnosis carried out in the country and the zone? If so, provide a list of approved laboratories. If not, provide the name(s) of and the arrangements with the laboratory(ies) samples are sent to, the follow-up procedures and the time frame for obtaining results. Indicate the laboratory (ies) where samples originating from the zone are diagnosed.
   b. Provide an overview of the AHS approved laboratories, in particular to address the following points:
Annex XXV (contd)

i. Details on the types of tests undertaken.

ii. Procedures for the official accreditation of laboratories. Give details of internal quality management systems, e.g. Good Laboratory Practice, ISO that exist in, or planned for, the laboratory system.

iii. Give details of participation in inter-laboratory validation tests (ring tests).

iv. Describe biosecurity measures applied, particularly in the case where live virus is handled.

6. AHS surveillance

Provide documentary evidence that surveillance for AHS in the zone complies with the provisions of Articles 12.1.11. to 12.1.13. of the Terrestrial Code, and Chapter 2.5.1. of the Terrestrial Manual. In particular, the following points should be addressed:

a. Clinical suspicion. What are the criteria for raising a suspicion of AHS? What is the procedure to notify (by whom and to whom), is there a compensation system in place and what penalties are involved for failure to report? Provide a summary table indicating, for the past 2 years, the number of suspect cases, the number of samples tested for AHS, species, type of sample, testing method(s) and results (including differential diagnosis) from the zone.

b. Surveillance. Are the following undertaken?

i. Serological surveillance

ii. Virological surveillance

iii. Sentinel animals

iv. Vector surveillance

If so, provide detailed information on the survey designs. How frequently are they conducted? Which were the equine species included? Are wildlife species included? Provide a summary table indicating detailed results, for at least the past 2 years. Provide details on follow-up actions taken on all suspicious and positive results. Provide criteria for selection of populations for targeted surveillance and numbers of equidae examined and samples tested. Provide details on the methods selected and applied for monitoring the performance of the surveillance system.

7. AHS prevention

a. Coordination with neighbouring countries. Are there any relevant factors about the adjacent countries and/or zones that have been taken into account (e.g. size, distance from adjacent border to infected equidae)? Describe coordination, collaboration and information sharing activities with neighbouring countries and zones.

If the AHS free zone is established in an AHS infected country or borders an infected country or zones, describe the animal health measures implemented to effectively prevent the introduction of the agent and/or vectors, taking into consideration the seasonal vector conditions and existing physical, geographical and ecological barriers.
Annex XXV (contd)

b. Import control procedures. From what countries or zones does the country authorize the import of equidae or their products into the free zone? What criteria are applied to approve such countries or zones? What controls are applied on entry of such equidae and products, and subsequent internal movement? What import conditions (e.g. quarantine) and test procedures are required? Are import permits and health certificates required? What other procedures are used? Provide summary statistics of imports, temporary admissions or re-entry of equidae and their products to the free zone for at least the past 2 years, specifying country or zone of origin and volume.

i. Provide a map with the number and location of ports, airports and land crossings in the zone. Is the service responsible for import controls part of the official services, or is it an independent body? If it is an independent body, describe its management structure, staffing levels and resources, and its accountability to the Competent Authority. Describe the communication systems between the Competent Authority and the border inspection posts, and between border inspection posts.

ii. Describe the regulations, procedures, type and frequency of checks at the points of entry into the zone and/or their final destination, concerning the import and follow-up of the following:
   - Equidae,
   - genetic material (semen, ova and embryos of the equine species),
   - equine derived (by-)products and biologicals,

iii. Describe the action available under legislation, and actually taken, when an illegal introduction into the zone is detected. Provide information on detected illegal introductions into the zone.

8. Control measures and contingency planning

a. Give details of any written guidelines, contingency plans (including information on vaccine banks) available to the Competent Authority for dealing with suspected or confirmed cases of AHS in the country and the zone (including the protection zone if applicable).

b. In the event of a suspected or confirmed AHS outbreak in the zone:

i. is quarantine imposed on premises with suspicious cases, pending final diagnosis?

ii. are movement restrictions applied on suspicion?

iii. describe the sampling and testing procedures used to identify and confirm presence of the causative agent;

iv. describe the actions taken to control the disease situation in and around any holdings found to be infected with AHS;

v. describe the control and/or eradication procedures (e.g. vaccination, modified stamping-out);

vi. describe the procedures used to confirm that an outbreak has been successfully controlled/eradicated, including conditions for restocking;

vii. give details of any compensation made available when equidae are killed, for disease control/eradication purposes.
9. **Compliance with the Terrestrial Code**

   a. In addition to the documentary evidence that the provisions of Article 12.1.2 are properly implemented and supervised, the Delegate of the country must submit a declaration stating:
      
      i. The section under paragraph 1 (of Article 12.1.2.) on the base of which the application is made
      
      ii. there has been no outbreak of AHS during the past 12 months in the zone;
      
      iii. no systematic vaccination against AHS has been carried out during the past 12 months in the zone;

   b. and that vaccinated equidae were imported into the zone in accordance with Chapter 12.1.

10. **Recovery of status**

    Countries applying for recovery of status should comply with the provisions of Article 12.1.2. of the *Terrestrial Code* and provide detailed information as specified in sections 4 (a), (b), (c) and 6 and highlight any measures introduced to prevent a recurrence of the infection under Section 7 of this questionnaire.
CHAPTER 12.6.

EQUINE INFLUENZA

Article 12.6.1.

General provisions

For the purposes of the Terrestrial Code, equine influenza (EI) is defined as an infection of domestic horses, donkeys and mules.

For the purposes of international trade, this chapter deals not only with the occurrence of clinical signs caused by equine influenza virus (EIV), but also with the presence of infection with EIV in the absence of clinical signs.

For the purposes of this chapter, isolation is defined as ‘the separation of domestic equids from domestic equids of a different equine influenza health status, utilising appropriate biosecurity measures, with the purpose of preventing the transmission of infection’.

For the purposes of the Terrestrial Code, the infective period for EI 21 days.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

When authorising import or transit of the commodities listed in this chapter, with the exception of those listed in Article 12.7.2., Veterinary Authorities should require the conditions prescribed in this chapter relevant to the EI status of the equine population of the exporting country, zone or compartment.

Article 12.6.2.

Safe commodities

When authorising import or transit of the following commodities, Veterinary Authorities should not require any EIV related conditions, regardless of the EI status of the equine population of the exporting country, zone or compartment:

1. semen;

2. in vivo derived equine embryos collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant (under study).

Article 12.6.3.

Determination of the EI status of a country, a zone or a compartment

The EI status of a country, a zone or a compartment can be determined on the basis of the following criteria:

1. the outcome of a risk assessment identifying all potential factors for EI occurrence and their historic perspective;

2. whether EI is notifiable in the whole country, an on-going EI awareness programme is in place, and all notified suspect occurrences of EI are subjected to field and, where applicable, laboratory investigations;

3. appropriate surveillance is in place to demonstrate the presence of infection in the absence of clinical signs in domestic equids.
Annex XXV (contd)

Article 12.6.4.

EI free country, zone or compartment

A country, zone or compartment may be considered free from EI provided the disease is notifiable in the whole country and it shows evidence, through an effective surveillance programme, planned and implemented according to the general principles in Chapter 1.4., that no case of EI occurred in the past 2 years.

The surveillance may need to be adapted to parts of the country, zone or compartment depending on historical or geographical factors, industry structure, population data, movements of equids within and into the country, zone or compartment, wild equid populations or proximity to recent outbreaks.

A country, zone or compartment seeking freedom from EI, in which vaccination is practised, should also demonstrate that EIV has not been circulating in the population of domestic and wild equids during the past 12 months, through surveillance, in accordance with Chapter 1.4.

In a country in which vaccination is not practised, surveillance may be conducted using serological testing alone. In countries where vaccination is practised, the surveillance should include agent identification methods described in the Terrestrial Manual for evidence of infection.

A country, zone or compartment seeking freedom from EI should apply appropriate movement controls to minimise the risk of introduction of EIV in accordance with this chapter.

If an outbreak of clinical EI occurs in a previously free country, zone or compartment, free status can be regained 12 months after the last clinical case, providing that surveillance for evidence of infection has been carried out during that twelve-month period in accordance with Chapter 1.4.

Article 12.6.5.

Recommendations for the importation of domestic equids for immediate slaughter

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the domestic equids showed no clinical sign of EI on the day of shipment.

Article 12.6.6.

Recommendations for the importation of domestic equids for unrestricted movement

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the domestic equids:

1. came from an EI free country, zone or compartment in which they had been resident for at least 21 days; in the case of a vaccinated domestic equid, information on its vaccination status should be included in the veterinary certificate;

OR

2. came from a country, zone or compartment not known to be free from EI, were subjected to pre-export isolation for 21 days and showed no clinical sign of EI during isolation nor on the day of shipment; and

3. were immunised according to the manufacturer’s instructions with a vaccine complying with the standards described in the Terrestrial Manual between 21 and 90 days before shipment either with a primary course or a booster; information on their vaccination status should be included in the veterinary certificate.
For additional security, countries that are free of EI or undertaking an eradication programme may also request that the domestic equids were tested negative for EIV by an agent identification test for EI described in the *Terrestrial Manual* conducted on samples collected on two occasions at 7 to 14 days and less than 5 days before shipment.

**Article 12.6.7.**

**Recommendations for the importation of domestic equids which will be kept in isolation (see Article 12.6.1.)**

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the domestic equids:

1. came from an EI free country, zone or compartment in which they had been resident for at least 21 days; in the case of a vaccinated domestic equid, information on its vaccination status should be included in the veterinary certificate;

OR

2. showed no clinical sign of EI in any premises in which the domestic equids had been resident for the 21 days prior to shipment nor on the day of shipment; and

3. were immunised according to the manufacturer’s instructions with a vaccine complying with the standards described in the *Terrestrial Manual*; information on their vaccination status should be included in the veterinary certificate.

**Article 12.6.8.**

**Recommendations for the importation of fresh meat of equids**

*Veterinary Authorities* should require the presentation of an *international veterinary certificate* attesting that the *fresh meat* came from equids which had been subjected to ante-mortem and post-mortem inspections as described in Chapter 6.2.
CHAPTER 12.9.

EQUINE VIRAL ARTERITIS

Article 12.9.1.

General provisions

The infective period for equine viral arteritis (EVA) shall be 28 days for all categories of equine except sexually mature stallion where the infective period may be for the life of the animal. Because the infective period may be extended in the case of virus shedding in semen, the status of seropositive stallions should be checked to ensure that they do not shed virus in their semen.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 12.9.2.

Recommendations for the importation of uncastrated male equines

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the animals showed no clinical sign of EVA on the day of shipment and during the 28 days prior to shipment and met one of the following requirements:

1. were isolated for the 28 days prior to shipment and were subjected, to a test for EVA, as prescribed in the Terrestrial Manual, carried out on a single blood sample collected during the 21 days prior to shipment with negative result; or

2. were subjected between 6 and 9 months of age to a test for EVA, as prescribed in the Terrestrial Manual, carried out on two blood samples collected at least 14 days apart with stable or decreasing titre, immediately vaccinated for EVA and regularly revaccinated according to the manufacturer's instructions; or

3. met the following requirements:
   a) were isolated; and
   b) not earlier than 7 days of commencing isolation were tested, with negative results, with subjected to a test for EVA as prescribed in the Terrestrial Manual on a blood sample with negative results; and
   c) were then immediately vaccinated; and
   d) were kept separated from other equidae for 21 days following vaccination; and
   e) were revaccinated regularly according to the manufacturer's instructions; or

4. have been subjected to a test for EVA, as prescribed in the Terrestrial Manual, carried out on a blood sample with positive results and then: either
   a) were subsequently test mated to two mares within 6 months prior to shipment which were subjected to two tests for EVA as prescribed in the Terrestrial Manual with negative results on blood samples collected at the time of test mating and again 28 days after the mating; or
   b) were subjected to a test for equine arteritis virus as prescribed in the Terrestrial Manual with negative results, carried out on semen collected during the 6 months prior to shipment;
Annex XXV (contd)

c) were subjected to a test for equine arteritis virus as prescribed in the *Terrestrial Manual* with negative results, carried out on semen collected within 6 months after the blood sample was tested, then immediately vaccinated, and revaccinated regularly.

Article 12.9.3.

**Recommendations for the importation of equines other than uncastrated males**

*Veterinary Authorities* of importing countries should require the presentation of an *international veterinary certificate* attesting that the *animals* showed no clinical sign of EVA on the day of shipment and were kept in an *establishment* where no *animals* have shown any signs of EVA for the 28 days prior to shipment; and

**EITHER**

1. were kept in an *establishment* where no *animals* have shown any signs of EVA for the 28 days prior to shipment; and

   a) were subjected to a test for EVA, as prescribed in the *Terrestrial Manual*, carried out on blood samples collected on two occasions at least 14 days apart within 28 days prior to shipment, which demonstrated stable or declining antibody titres; or

   b) were regularly vaccinated according to the manufacturer’s instructions;

   OR

2. were isolated for the 28 days prior to shipment and during this period the *animals* showed no signs of EVA.

Article 12.9.4.

**Recommendations for the importation of semen**

*Veterinary Authorities* of importing countries should require the presentation of an *international veterinary certificate* attesting that the animal donors were kept for the 28 days prior to semen collection in an *establishment* where no equine has shown any clinical sign of EVA during that period and showed no clinical sign of EVA on the day of semen collection; and

1. were subjected between 6 and 9 months of age to a test for EVA as prescribed in the *Terrestrial Manual* on two blood samples collected at least 14 days apart with a stable or decreasing titre, immediately vaccinated for EVA and regularly revaccinated according to the manufacturer’s instructions; or

2. were isolated and not earlier than 7 days of commencing isolation were subjected to a test for EVA as prescribed in the *Terrestrial Manual* on a blood sample with negative results, immediately vaccinated for EVA, kept for 21 days following vaccination separated from other equidae and regularly revaccinated according to the manufacturer’s instructions; or

3. were subjected to a test for EVA as prescribed in the *Terrestrial Manual* on a blood sample with negative results within 14 days prior to semen collection, and had been separated from other equidae not of an equivalent EVA status for 14 days prior to blood sampling from the time of the taking of the blood sample until the end of semen collection; or

4. have been subjected to a test for EVA as prescribed in the *Terrestrial Manual* carried out on a blood sample with positive results and then: either
a) were subsequently test mated to two mares within 6 months prior to semen collection, which were subjected to two tests for EVA as prescribed in the *Terrestrial Manual* with negative results on blood samples collected at the time of test mating and again 28 days after the test mating, or

b) were subjected to a test for equine arteritis virus as prescribed in the *Terrestrial Manual* with negative results, carried out on semen collected within 6 months prior to collection of the semen to be exported; or

c) were subjected to a test for equine arteritis virus as prescribed in the *Terrestrial Manual* with negative results, carried out on semen collected within 6 months after the blood sample was tested, then immediately vaccinated, and revaccinated regularly; or

5. were, for frozen semen, subjected with negative results either:

a) to a test for EVA as prescribed in the *Terrestrial Manual* carried out on a blood sample taken not earlier than 14 days and not later than 12 months after the collection of the semen for export; or

b) to a test for equine arteritis virus as prescribed in the *Terrestrial Manual* carried out on an aliquot of the semen collected immediately prior to processing or on an aliquot of semen collected within 14 to 30 days after the first collection of the semen to be exported.

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CHAPTER 14.5.

CHLAMYDOPHILA ABORTUS INFECTION

ENZOOTIC ABORTION OF EWES
(OVINE CHLAMYDIOSIS)

Article 14.5.1.

General provisions

For the purposes of the Terrestrial Code, enzootic abortion of ewes (EAE), also known as ovine chlamydiosis or ovine enzootic abortion, is an infection of domestic sheep and goats by the bacterium Chlamyphila abortus.

For the purposes of the Terrestrial Code, the following information should be considered with regard to the incubation period for enzootic abortion of ewes (EAE). Susceptible animals become infected through ingestion of infectious materials. In lambs and non-pregnant ewes, the infection remains latent until conception. Ewes exposed to infection late in pregnancy may not exhibit signs of infection until the subsequent pregnancy. Countries should take account of these risk factors.

Standards for diagnostic tests are described in the Terrestrial Manual.

Article 14.5.2.

Recommendations for the importation of sheep and/or goats for breeding

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the animals:

1. have remained since birth, or for the previous 2 years, in establishments where no EAE has been diagnosed during the past 2 years;
2. showed no clinical sign of EAE on the day of shipment;
3. were subjected to a diagnostic test for EAE with negative results within the 30 days prior to shipment.

Article 14.5.3.

Sheep flocks and/or goat herds free from EAE infection

To qualify as free from EAE infection, a sheep flock or goat herd shall satisfy the following requirements:

1. it is under official veterinary surveillance;
2. all sheep and goats showed no clinical evidence of EAE infection during the past 2 years;
3. a statistically valid number of sheep and goats over 6 months of age were subjected to a diagnostic test for EAE with negative results within the past 6 months;
4. all sheep or goats are permanently identified;
5. no sheep or goat has been added to the flock or herd since 30 days prior to the flock or herd test referred to in point 3 above unless:

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a) either the additions were isolated from other members of the flock or herd in the establishment of origin for a minimum period of 30 days and then were subjected to a diagnostic test for EAE with negative results, before entry into the new flock or herd; or

b) they originated from an establishment of equal health status.

Article 14.5.4.

Recommendations for the importation of semen of sheep

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) have been kept in establishments or artificial insemination centres free from EAE during the past 2 years, and have not been in contact with animals of a lower health status;
   b) were subjected to a diagnostic test for EAE with negative results 2 to 3 weeks after collection of the semen;

2. an aliquot of the semen to be exported was shown to be free of *Chlamydia psittaci* *Chlamydophila abortus*, by culture techniques.

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General provisions and safe commodities
Scrapie is a neurodegenerative disease of sheep and goats. The main mode of transmission is from mother to offspring immediately after birth and to other susceptible neonates exposed to the birth fluids and tissues of an infected animal. Transmission occurs at a much lower frequency to adults exposed to the birth fluids and tissues of an infected animal. A variation in genetic susceptibility of sheep has been recognised. The incubation period of the disease is variable; however, it is usually measured in years. The duration in incubation period can be influenced by a number of factors including host genetics and strain of agent.

Scrapie is not considered to pose a risk to human health. The recommendations in this chapter are intended to manage the animal health risks associated with the presence of the scrapie agent in sheep and goats. The chapter does not cover so-called ‘atypical’ scrapie which because this condition is clinically, pathologically, biochemically and epidemiologically unrelated to ‘classical’ scrapie, may not be contagious and may, in fact, be a spontaneous degenerative condition of older sheep.

1. When authorising import or transit of the following commodities derived from sheep or goats and any products made from these commodities and containing no other tissues from sheep or goats, Veterinary Authorities should not require any scrapie-related conditions, regardless of the scrapie risk status of the sheep and goat populations of the exporting country, zone or compartment:

   a) in vivo derived sheep embryos handled in accordance with Chapter 4.7. of this Terrestrial Code;

   b) meat (excluding materials as referred to in Article 14.9.12.);

   c) hides and skins;

   d) gelatine;

   e) collagen prepared from hides or skins;

   f) tallow (maximum level of insoluble impurities of 0.15% in weight) and derivatives made from this tallow;

   g) dicalcium phosphate (with no trace of protein or fat);

   h) wool or fibre.

2. When authorising import or transit of other commodities listed in this chapter, Veterinary Authorities should require the conditions prescribed in this chapter relevant to the scrapie risk status of the sheep and goat populations of the exporting country, zone or compartment.

Standards for diagnostic tests are described in the Terrestrial Manual.

Determination of the scrapie status of the sheep and goat populations of a country, zone, compartment or establishment
Annex XXVII (contd)

The scrapie status of the sheep and goat populations of a country, zone, compartment or establishment should be determined on the basis of the following criteria:

1. the outcome of a risk assessment identifying all potential factors for scrapie occurrence and their historic perspective, in particular the:
   a) importation or introduction of sheep and goats or their semen, in vivo derived goat embryos or in vitro processed sheep and goat embryos/oocytes potentially infected with scrapie;
   b) extent of knowledge of the population structure and husbandry practices of sheep and goats;
   c) feeding practices, including consumption of meat-and-bone meal or greaves derived from ruminants;
   d) importation of milk and milk products of sheep or goats origin intended for use in feeding of sheep and goats;

2. an on-going awareness programme for veterinarians, farmers, and workers involved in transportation, marketing and slaughter of sheep and goats to facilitate recognition and encourage reporting of all animals with clinical signs compatible with scrapie;

3. a surveillance and monitoring system including the following:
   a) official veterinary surveillance, reporting and regulatory control in accordance with the provisions of Chapter 1.4.;
   b) a Veterinary Authority with current knowledge of, and authority over, all establishments which contain sheep and goats in the whole country;
   c) compulsory notification and clinical investigation of sheep and goats showing clinical signs compatible with scrapie;
   d) examination, in accordance with the Terrestrial Manual, in a laboratory of appropriate material from sheep and goats older than 18 months displaying clinical signs compatible with scrapie;
   e) maintenance of records including the number and results of all investigations for at least 7 years.

Scrapie free country or zone

Countries or zones may be considered free from scrapie if within the said territory:

1. a risk assessment, as described in point 1 of Article 14.9.2., has been conducted, and it has been demonstrated that appropriate measures are currently in place and have been taken for the relevant period of time to manage any risk identified and points 2 and 3 have been complied with for the preceding 7 years;

   AND

2. one of the following conditions should be met:
   a) the country or the zone have demonstrated historical freedom as follows:
      i) scrapie has been notifiable for at least 25 years; and
ii) a formal programme of targeted surveillance and monitoring, which includes testing of sheep and goats displaying clinical signs compatible with scrapie and those over 18 months of age slaughtered, culled or found dead on farm, can be documented as having been in place for at least 10 years; and

iii) appropriate measures to prevent scrapie introduction can be documented as having been in place for at least 25 years; and

- either scrapie has never been reported; or
- no case of scrapie has been reported for at least 25 years;

b) for at least 7 years, sheep and goats displaying clinical signs compatible with scrapie have been tested. Also a sufficient number of sheep and goats over 18 months of age, representative of slaughtered, culled or found dead on farm, have been tested annually, to provide a 95% level of confidence of detecting scrapie if it is present in that population at a prevalence rate exceeding 0.1% and no case of scrapie has been reported during this period; or

c) all establishments containing sheep or goats have been accredited free as described in Article 14.9.5.;

AND

3. the feeding to sheep and goats of meat-and-bone meal or greaves of ruminant origin has been banned and effectively enforced in the whole country for at least 7 years;

AND

4. introductions of sheep and goats or their semen, in vivo derived goat embryos or in vitro processed sheep and goat embryos/oocytes from countries or zones not free from scrapie are carried out in accordance with Articles 14.9.6., 14.9.7., 14.9.8. or 14.9.9., as relevant.

Article 14.9.4.

**Compartment free from scrapie**

To qualify as a compartment free from scrapie, all sheep and goats in a compartment should be certified by the Veterinary Authority as satisfying the following requirements:

1. all establishments within the compartment are free from scrapie according to Article 14.9.5.;

2. all establishments within the compartment are managed under a common biosecurity plan protecting them from introduction of scrapie, and the compartment has been approved by the Veterinary Authority in accordance with Chapters 4.3. and 4.4.;

3. introductions of sheep and goats are allowed only from accredited free establishments or free countries;

4. introductions of in vivo derived goat embryos and in vitro processed sheep and goat embryos/oocytes are allowed either from accredited free establishments or in accordance with Article 14.9.9.;

5. sheep and goat semen should be introduced into the compartment in accordance with Article 14.9.8.;

6. sheep and goats in the compartment should have no direct or indirect contact, including shared grazing, with sheep or goats from establishments not within the compartment.
Annex XXVII (contd)

Scrapie free establishment

To qualify as free from scrapie, an establishment of sheep and goats should satisfy the following requirements:

1. in the country or zone where the establishment is situated, the following conditions are fulfilled:
   a) the disease is compulsorily notifiable;
   b) an awareness, surveillance and monitoring system as referred to in Article 14.9.2. is in place;
   c) affected sheep and goats are killed and completely destroyed;
   d) the feeding to sheep and goats of meat-and-bone meal or greaves of ruminant origin has been banned and effectively enforced in the whole country for at least 7 years;
   e) an official accreditation scheme is in operation under the supervision of the Veterinary Authority, including the measures described in point 2 below;

2. in the establishment the following conditions have been complied with for at least 7 years:
   a) sheep and goats are permanently identified and records maintained, to enable trace back to their establishment of birth;
   b) records of movements of sheep and goats in and out of the establishment are maintained;
   c) introductions of sheep and goats are allowed only from free establishments or establishment at an equal or higher stage in the process of accreditation;
   d) introduction of in vivo derived goat embryos and in vitro processed sheep and goat embryos /oocytes should comply with Article 14.9.9.;
   e) sheep and goat semen should be introduced into the establishment in accordance with Article 14.9.8.;
   f) an Official Veterinarian inspects sheep and goats in the establishments and audits the records at least once a year;
   g) no case of scrapie has been reported;
   h) sheep and goats of the establishments should have no direct or indirect contact, including shared grazing, with sheep or goats from establishments of a lower status;
   i) all culled sheep and goats over 18 months of age are inspected by an Official Veterinarian, and a proportion of those exhibiting wasting signs and all those exhibiting neurological signs are tested in a laboratory for scrapie. The selection of the sheep and goats to be tested should be made by the Official Veterinarian. Sheep and goats over 18 months of age that have died or have been killed for reasons other than routine slaughter should also be tested (including ‘fallen’ stock and those sent for emergency slaughter).

Recommendations for importation from countries or zones not considered free from scrapie

for sheep and goats for breeding or rearing
Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals come from an establishment free from scrapie as described in Article 14.9.5.

Article 14.9.7.

Recommendations for importation from countries or zones not considered free from scrapie for sheep and goats for slaughter

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. in the country or zone:
   a) the disease is compulsorily notifiable;
   b) an awareness, surveillance and monitoring system as referred to in Article 14.9.2 is in place;
   c) affected sheep and goats are killed and completely destroyed;
2. the sheep and goats selected for export showed no clinical sign of scrapie on the day of shipment.

Article 14.9.8.

Recommendations for importation from countries or zones not considered free from scrapie for semen of sheep and goats

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) are permanently identified to enable trace back to their establishment of origin;
   b) showed no clinical sign of scrapie at the time of semen collection;
2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 14.9.9.

Recommendations for importation from countries or zones not considered free from scrapie for in vivo derived goat embryos and in vitro processed sheep and goat embryos/oocytes

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. in the country or zone:
   a) the disease is compulsorily notifiable;
   b) an awareness, surveillance and monitoring system as referred to in Article 14.9.2 is in place;
   c) affected sheep and goats are killed and completely destroyed;
   d) the feeding to sheep and goats of meat-and-bone meal or greaves of ruminant origin has been banned and effectively enforced in the whole country;
2. the donor animals either have been kept since birth in a free establishment, or meet the following conditions:
   a) are permanently identified to enable trace back to their establishment of origin;
   b) have been kept since birth in establishments in which no case of scrapie had been confirmed during their residency;
   c) showed no clinical sign of scrapie at the time of embryo/oocyte collection;
3. the embryos/oocytes were collected, processed and stored in conformity with the provisions of Chapters 4.7., 4.8. and 4.9., as relevant.

Recommendations for importation from countries or zones not considered free from scrapie

for milk and milk products of sheep or goat origin intended for use in feeding of sheep and goats

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the milk and milk products come from scrapie free establishments.

Recommendations on meat-and-bone meal

Meat-and-bone meal containing any sheep or goat protein, or any feedstuffs containing that type of meat-and-bone meal, which originate from countries not considered free of scrapie should not be traded between countries for ruminant feeding.

Recommendations for importation from countries or zones not considered free from scrapie

for skulls including brains, ganglia and eyes, vertebral column including ganglia and spinal cord, tonsils, thymus, spleen, intestine, adrenal gland, pancreas, or liver, and protein products derived therefrom, from sheep and goats

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. these commodities should not be traded for use in ruminant feeds;
2. for purposes other than ruminant feeding, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:
   a) in the country or zone:
      i) the disease is compulsorily notifiable;
      ii) an awareness, surveillance and monitoring system as referred to in Article 14.9.2. is in place;
      iii) affected sheep and goats are killed and completely destroyed;
   b) the materials come from sheep and goats that showed no clinical sign of scrapie on the day of slaughter.

Recommendations for the importation of ovine and caprine materials destined for the preparation of biologicals

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products originate from sheep and goats born and raised in a scrapie free country, zone or establishment.

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CHAPTER 15.2.

CLASSICAL SWINE FEVER

Article 15.2.1.

General provisions

The pig is the only natural host for classical swine fever (CSF) virus. The definition of pig includes all varieties of Sus scrofa, both domestic and wild. For the purposes of this chapter, a distinction is made between domestic pig and wild pig (including feral pigs) populations.

For the purposes of international trade, the Terrestrial Code, classical swine fever (CSF) is defined as an infection of domestic pigs.

Domestic pig is defined as ‘all domesticated pigs, permanently captive or farmed free range, used for the production of meat for consumption, for the production of other commercial products or for breeding these categories of pigs.

Pigs exposed to CSF virus prenatally may be persistently infected throughout life and may have an incubation period of several months before showing signs of disease. Pigs exposed postnatally have an incubation period of 2-14 days, and are usually infective between post-infection days 5 and 14, but up to 3 months in cases of chronic infections.

For the purposes of international trade, a Member should not impose trade bans in response to a notification of infection with classical swine fever virus in wild pigs according to Article 1.2.3. of the Terrestrial Code after the Member confirms that Article 15.2.2. is appropriately implemented.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

A Member should not impose trade bans in response to a notification of infection with classical swine fever virus in wild pigs according to Article 1.2.3. of the Terrestrial Code after the Member confirms that Article 15.2.2. is appropriately implemented.

Article 15.2.2.

Determination of the CSF status of a country, zone or compartment

The CSF status of a country, zone or compartment can only be determined after considering the following criteria in domestic and wild pigs, as applicable:

1. CSF should be notifiable in the whole territory, and all clinical signs suggestive of CSF should be subjected to appropriate field and/or laboratory investigations;

2. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of CSF;

3. the Veterinary Authority should have current knowledge of, and authority over, all domestic pigs in the country, zone or compartment;

4. the Veterinary Authority should have current knowledge about the population and habitat of wild pigs in the country or zone,
Annex XXVIII (contd)

5. for domestic pigs, appropriate surveillance, capable of detecting the presence of infection even in the absence of clinical signs, and the risk posed by wild pigs, is in place; this may be achieved through a surveillance programme in accordance with Articles 15.2.23. to 15.2.28.;

6. for wild pigs, if present in the country or zone, a surveillance programme is in place according to Article 15.2.28., taking into account the presence of natural and artificial boundaries, the ecology of the wild pig population, and an assessment of the risks of disease spread.

7. Based on the assessed risk of spread within the wild pig population, and according to Article 15.2.26., the domestic pig population should be separated from the wild pig population by appropriate biosecurity measures to prevent transmission of CSF from wild to domestic pigs.

Article 15.2.3.

CSF free country, zone or compartment

A country, zone or compartment may be considered free from CSF when surveillance in accordance with Articles 15.2.23. to 15.2.28. has been in place for at least 12 months, and when:

1. there has been no outbreak of CSF in domestic pigs during the past 12 months;

2. no evidence of CSFV infection has been found in domestic pigs during the past 12 months;

3. no vaccination against CSF has been carried out in domestic pigs during the past 12 months unless there are means, validated to OIE standards (Chapter 2.8.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs;

4. imported domestic pigs comply with the requirements in Article 15.2.5. or Article 15.2.6.

Article 15.2.4.

Recovery of free status

Should a CSF outbreak occur in a free country, zone or compartment, the free status may be restored where surveillance in accordance with Articles 15.2.23. to 15.2.28. has been carried out with negative results either:

1. 3 months after the last case where a stamping-out policy without vaccination is practised;

OR

2. where a stamping-out policy with emergency vaccination is practised:
   a) 3 months after the last case and the slaughter of all vaccinated animals, or
   b) 3 months after the last case without the slaughter of vaccinated animals where there are means, validated to OIE standards (Chapter 2.8.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs;

OR

3. where a stamping-out policy is not practised, the provisions of Article 15.2.3. should be followed.

Article 15.2.5.

Recommendations for importation from countries, zones or compartments free of CSF
Annex XXVIII (contd)

for domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CSF on the day of shipment;
2. were kept in a country, zone or compartment free of CSF since birth or for at least the past 3 months;
3. have not been vaccinated against CSF, nor are they the progeny of vaccinated sows, unless there are means, validated to OIE standards (Chapter 2.8.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs.

Article 15.2.6.

Recommendations for importation from CSF infected countries or zones

for domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CSF on the day of shipment;
2. were kept since birth or for the past 3 months in a CSF free compartment;
3. have not been vaccinated against CSF nor are they the progeny of vaccinated sows, unless there are means, validated to OIE standards (Chapter 2.8.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs.

Article 15.2.7.

Recommendations for the importation of wild pigs

Regardless of the CSF status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CSF on the day of shipment;
2. were kept in a quarantine station for 40 days prior to shipment, and were subjected to a virological test and a serological test performed at least 21 days after entry into the quarantine station, with negative results;
3. have not been vaccinated against CSF, unless there are means, validated to OIE standards (Chapter 2.8.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs.

Article 15.2.8.

Recommendations for importation of semen of domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in a country, zone or compartment free of CSF since birth or for at least 3 months prior to collection;
Annex XXVIII (contd)

b) showed no clinical sign of CSF on the day of collection of the semen;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 15.2.9.

Recommendations for importation from CSF infected countries or zones for semen of domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in a compartment free of CSF since birth or for at least 3 months prior to collection;
   b) showed no clinical sign of CSF on the day of collection of the semen and for the following 40 days;
   c) met one of the following conditions:
      i) have not been vaccinated against CSF and were subjected to a serological test performed at least 21 days after collection, with negative results; or
      ii) have been vaccinated against CSF and were subjected to a serological test in accordance with the Terrestrial Manual performed at least 21 days after collection and it has been conclusively demonstrated that any antibody is due to the vaccine; or
      iii) have been vaccinated against CSF and were subjected to a virological test performed in accordance with the Terrestrial Manual on a sample taken on the day of collection and it has been conclusively demonstrated that the boar is negative for virus genome;

2. the semen was collected, processed and stored in conformity with the provisions of Chapters 4.5. and 4.6.

Article 15.2.10.

Recommendations for importation from countries, zones or compartments free of CSF for in vivo derived embryos of domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:

1. the donor females showed no clinical sign of CSF on the day of collection of the embryos;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.

Article 15.2.11.

Recommendations for importation from CSF infected countries or zones for in vivo derived embryos of domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that:
Annex XXVIII (contd)

1. the donor females:
   a) were kept in a compartment free of CSF since birth or for at least 3 months prior to collection;
   b) showed no clinical sign of CSF on the day of collection of the embryos and for the following 40 days;
   c) and either:
      i) have not been vaccinated against CSF and were subjected, with negative results, to a serological test performed at least 21 days after collection; or
      ii) have been vaccinated against CSF and were subjected to a serological test performed at least 21 days after collection and it has been conclusively demonstrated by means, validated to OIE standards (Chapter 2.8.3. of the Terrestrial Manual), that any antibody is due to the vaccine;

2. the embryos were collected, processed and stored in conformity with the provisions of Chapters 4.7. and 4.9., as relevant.

Article 15.2.12.

Recommendations for importation from countries, zones or compartments free of CSF

for fresh meat of domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from animals which:

1. have been kept in a country, zone or compartment free of CSF, or which have been imported in accordance with Article 15.2.5. or Article 15.2.6.;

2. have been slaughtered in an approved abattoir, have been subjected to ante-mortem and post-mortem inspections in accordance with Chapter 6.2. and have been found free of any sign suggestive of CSF.

Article 15.2.13.

Recommendations for the importation of fresh meat of wild pigs

Regardless of the CSF status of the country of origin, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from animals:

1. the entire consignment of fresh meat comes from animals which have been subjected to a post-mortem inspection in accordance with Chapter 6.2. in an approved examination centre, and have been found free of any sign suggestive of CSF;

2. where the CSF-free status of the wild pig population cannot be assured, the entire consignment of meat comes from animals from each of which a sample has been collected and has been subjected to a virological test and a serological test for CSF, with negative results.
Annex XXVIII (contd)

Article 15.2.14.

Recommendations for the importation of meat and meat products of pigs, or for products of animal origin (from fresh meat of pigs) intended for use in animal feeding, for agricultural or industrial use, or for pharmaceutical or surgical use

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. have been prepared:
   a) exclusively from fresh meat meeting the conditions laid down in Article 15.2.12.;
   b) in a processing establishment:
      i) approved by the Veterinary Authority for export purposes;
      ii) processing only meat meeting the conditions laid down in Article 15.2.12.;

OR

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the CSF virus in conformity with one of the procedures referred to in Article 15.2.21. and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.

Article 15.2.15.

Recommendations for the importation of products of animal origin (from pigs, but not derived from fresh meat) intended for use in animal feeding

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. originated from domestic pigs in a CSF free country, zone or compartment and have been prepared in a processing establishment approved by the Veterinary Authority for export purposes; or

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the CSF virus in accordance with Article 15.2.20. and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.

Article 15.2.16.

Recommendations for the importation of products of animal origin (from pigs, but not derived from fresh meat) intended for agricultural or industrial use

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. originated from domestic pigs in a CSF free country, zone or compartment and have been prepared in a processing establishment approved by the Veterinary Authority for export purposes; or

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the CSF virus (under study) and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.
Recommendations for the importation of bristles

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. originated from domestic pigs in a CSF free country, zone or compartment and have been prepared in a processing establishment approved by the Veterinary Authority for export purposes; or

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the CSF virus (under study) and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.

Article 15.2.17.

Recommendations for the importation of litter and manure

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. originated from domestic pigs in a CSF free country, zone or compartment and have been prepared in a processing establishment approved by the Veterinary Authority for export purposes; or

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the CSF virus (under study) and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.

Article 15.2.18.

Recommendations for the importation of skins and trophies

Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

1. originated from domestic pigs in a CSF free country, zone or compartment and have been prepared in a processing establishment approved by the Veterinary Authority for export purposes; or

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the CSF virus in conformity with one of the procedures referred to in Article 15.2.22, and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.

Article 15.2.19.

Procedures for the inactivation of the CSF virus in swill

For the inactivation of CSF viruses likely to be present in swill, one of the following procedures should be used:

1. the swill should be maintained at a temperature of at least 90°C for at least 60 minutes, with continuous stirring; or

2. the swill should be maintained at a temperature of at least 121°C for at least 10 minutes at an absolute pressure of 3 bar.
Annex XXVIII (contd)

Article 15.2.21.

Procedures for the inactivation of the CSF virus in meat

For the inactivation of viruses present in meat, one of the following procedures should be used:

1. **Heat treatment**

   Meat shall be subjected to one of the following treatments:
   
   a) heat treatment in a hermetically sealed container with a Fo value of 3.00 or more;
   
   b) heat treatment at a minimum temperature of 70°C, which should be reached throughout the meat.

2. **Natural fermentation and maturation**

   The meat should be subjected to a treatment consisting of natural fermentation and maturation having the following characteristics:
   
   a) an aw value of not more than 0.93, or
   
   b) a pH value of not more than 6.0.

   Hams should be subjected to a natural fermentation and maturation process for at least 190 days and loins for 140 days.

3. **Dry cured pork meat**

   a) Italian style hams with bone-in should be cured with salt and dried for a minimum of 313 days.
   
   b) Spanish style pork meat with bone-in should be cured with salt and dried for a minimum of 252 days for Iberian hams, 140 days for Iberian shoulders, 126 days for Iberian loin, and 140 days for Serrano hams.

Article 15.2.22.

Procedures for the inactivation of the CSF virus in skins and trophies

For the inactivation of CSF viruses likely to be present in skins and trophies, one of the following procedures should be used:

1. boiling in water for an appropriate time so as to ensure that any matter other than bone, tusks or teeth is removed;

2. gamma irradiation at a dose of at least 20 kiloGray at room temperature (20°C or higher);

3. soaking, with agitation, in a 4% (w/v) solution of washing soda (sodium carbonate - Na2CO3) maintained at pH 11.5 or above for at least 48 hours;

4. soaking, with agitation, in a formic acid solution (100 kg salt [NaCl] and 12 kg formic acid per 1,000 litres water) maintained at below pH 3.0 for at least 48 hours; wetting and dressing agents may be added;

5. in the case of raw hides, salting for at least 28 days with sea salt containing 2% washing soda (sodium carbonate - Na2CO3).
Annex XXVIII (contd)

Article 15.2.23.

Surveillance: introduction

Articles 15.2.23. to 15.2.28. define the principles and provide a guide on the surveillance for CSF, complementary to Chapter 1.4., applicable to Members seeking to determine their CSF status. This may be for the entire country, or a zone or a compartment. Guidance for Members seeking free status following an outbreak and for the maintenance of CSF status is also provided.

The impact and epidemiology of CSF differ widely in different regions of the world, and it is, therefore, impossible to provide specific recommendations for all situations. The surveillance strategies employed for demonstrating freedom from CSF at an acceptable level of confidence will need to be adapted to the local situation. For example, the approach should be tailored in order to prove freedom from CSF for a country or zone where wild pigs provide a potential reservoir of infection, or where CSF is present in adjacent countries. The method should examine the epidemiology of CSF in the region concerned and adapt to the specific risk factors encountered. This should include provision of scientifically based supporting data. There is, therefore, latitude available to Members to provide a well-reasoned argument to prove that absence of classical swine fever virus (CSFV) infection is assured at an acceptable level of confidence.

Surveillance for CSF should be in the form of a continuing programme designed to establish that a population in a country, zone or compartment is free from CSFV infection or to detect the introduction of CSFV into a population already recognized as free. Consideration should be given to the specific characteristics of CSF epidemiology which include: the role of swill feeding and the impact of different production systems on disease spread, the role of semen in transmission of the virus, the lack of pathognomonic gross lesions and clinical signs, the frequency of clinically inapparent infections, the occurrence of persistent and chronic infections, and the genotypic, antigenic, and virulence variability exhibited by different strains of CSFV. Serological cross-reactivity with other pestiviruses has to be taken into consideration when interpreting data from serological surveys. A common route by which ruminant pestiviruses can infect pigs is the use of vaccines contaminated with bovine viral diarrhoea virus (BVDV).

For the purposes of this chapter, virus infection means presence of CSFV as demonstrated directly by virus isolation, the detection of virus antigen or virus nucleic acid, or indirectly by seroconversion which is not the result of vaccination.

Article 15.2.24.

Surveillance: general conditions and methods

1. A surveillance system in accordance with Chapter 1.4. should be under the responsibility of the Veterinary Authority. A procedure should be in place for the rapid collection and transport of samples to an accredited laboratory as described in the Terrestrial Manual.

   a) a formal and ongoing system for detecting and investigating outbreaks of disease or CSFV infection should be in place;

   b) a procedure should be in place for the rapid collection and transport of samples from suspect cases of CSF to a laboratory for CSF diagnosis as described in the Terrestrial Manual;

   c) a system for recording, managing and analysing diagnostic and surveillance data should be in place.
Annex XXVIII (contd)

2. The CSF surveillance programme should:

a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers, who have day-to-day contact with livestock, as well as diagnosticians, should report promptly any suspicion of CSF to the Veterinary Authority. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Authority. Since many strains of CSFV do not induce pathognomonic gross lesions or clinical signs, cases in which CSF cannot be ruled out should be immediately investigated employing clinical, pathological, and laboratory diagnosis. This requires that sampling kits and other equipment are available to those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in CSF diagnosis, epidemiological evaluation, and control;

b) implement, when relevant, regular and frequent clinical inspections and serological testing of high-risk groups of animals (for example, where swill feeding is practised), or those adjacent to a CSF infected country or zone (for example, bordering areas where infected wild pigs are present).

An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is CSFV. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot, therefore, be reliably predicted. Recognitions for freedom from CSFV infection should, as a consequence, provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement standstill orders, etc.).

Article 15.2.25.

Surveillance strategies

1. Introduction

There are two basic strategies that can be employed for CSF surveillance depending on the purpose of the Member for seeking recognition of freedom from CSF. In countries free of CSF, surveillance programmes should be designed to detect the presence introduction of CSFV infection into domestic or wild swine. The optimal strategy to meet this objective is most often targeted surveillance.

The population covered by surveillance aimed at detecting disease and infection should include domestic and wild pig populations within the country or zone to be recognised as free from CSFV infection. Such surveillance may involve opportunistic testing of samples submitted for other purposes, but a more efficient and effective strategy is one which includes targeted surveillance.

Although surveillance may involve opportunistic testing of samples submitted for other purposes, the optimal strategy to meet this objective is usually targeted surveillance. Surveillance is targeted to aimed at the domestic and wild pig population which presents the highest risk of infection (for example, swill fed farms, pigs reared outdoors, specific wild pig sub-populations or farms in proximity to infected wild pigs). Each Member will need to identify its individual risk factors. Targeted surveillance may include randomized sampling in selected high risk populations, based on the risk factors present. These may include: temporal and spatial distribution of past outbreaks, pig movements and demographics, etc.

For reasons of cost, the longevity of antibody levels, as well as the existence of clinically inapparent infections and difficulties associated with differential diagnosis of other diseases, serology is often the most effective and efficient surveillance methodology. In some circumstances, which will be discussed later, clinical and virological surveillance may also have value.
The surveillance strategy chosen by the Member should be justified as adequate to detect the presence of CSFV infection in accordance with Chapter 1.4. and the epidemiological situation. Cumulative survey results in combination with the results of passive surveillance, over time, will increase the level of confidence in the surveillance strategy. If a Member wishes to apply for recognition by other Members of a specific zone within the country as being free from CSFV infection, the design of the surveillance strategy and the basis for any sampling process would need to be aimed at the population within the zone.

When applying randomized sampling, either at the level of the entire population or within targeted sub-populations, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalences for the selected populations. The sample size selected for testing will need to be large enough to detect infection if it were to occur at a predetermined minimum rate. The sample size and expected disease prevalence determine the level of confidence in the results of the survey. The choice of design prevalence and confidence level should be justified based on the objectives of surveillance and the epidemiological situation, in accordance with Chapter 1.4. Selection of the design prevalence in particular, clearly needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey design approach selected, the sensitivity and specificity of the diagnostic tests in the target populations employed should be considered factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and production class of animals in the target population.

Irrespective of the testing system employed, the surveillance system design should anticipate the occurrence of false positive reactions. This is especially true of the serological diagnosis of CSF because of the recognized cross-reactivity with ruminant pestiviruses. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether or not they are indicative of CSFV infection. This should involve confirmatory and differential tests for pestiviruses, as well as further investigations concerning the original sampling unit as well as animals which may be epidemiologically linked.

2. Clinical and virological surveillance

Beyond their role in targeted surveillance, clinical and virological surveillance for CSF has two aims: a) to shorten the period between introduction of CSF virus into a disease free country or zone and its detection, and b) to confirm that no unnoticed outbreaks have occurred.

In the past, the value of clinical identification of cases was the cornerstone of early detection of CSF. However, emergence of surveillance alone is limited due to the low virulence of some strains of CSF, as well as the emergence of new diseases - such as post-weaning multisystemic wasting syndrome and porcine dermatitis and nephropathy syndrome - have made such reliance less effective, and, in countries where such diseases are common, can add significant risk of masking the presence of CSF which can mask the presence of CSF. Therefore, clinical surveillance should be supplemented, as appropriate, by serological and virological surveillance.

The spectrum of disease signs and gross pathology seen in CSF infections, along with the plethora of other agents that can mimic CSF, renders the value of clinical examination alone somewhat inefficient as a surveillance tool. These factors, along with the compounding effects of concurrent infections and diseases caused by ruminant pestiviruses, dictate the need for laboratory testing in order to clarify the status of CSF suspects detected by clinical monitoring.
Nevertheless, clinical and pathological signs presentation should not be ignored as a tool are useful for early detection; in particular, any cases where clinical signs or lesions consistent with CSF are accompanied by high morbidity and/or mortality should be investigated without delay. In CSFV infections involving low virulence strains, high mortality may only be seen in young animals and adult animals may not show clinical sign. Otherwise close physical examination of susceptible animals is useful as a selection criteria for CSF surveillance, particularly in diagnostic laboratories or slaughter establishments or when applied to high risk populations such as swill feeding operations.

The difficulties in detecting chronic disease, manifested by non-specific clinical signs and delayed seroconversion and seronegativity, in persistently infected piglets, both of which may be clinically normal, maker virological investigation essential. As part of a herd investigation, such animals are likely to be in a minority and would not confound a diagnosis based on serology. Individually or as part of recently mixed batches, such animals may, however, escape detection by this method. A holistic approach to investigation, taking note of herd history, pig, personnel and vehicle movements and disease status in neighbouring zones or countries, can also assist in targeting surveillance in order to increase efficiency and enhance the likelihood of early detection.

The labour intensive nature of clinical, pathological and virological investigations, along with the smaller ‘window of opportunity’ inherent in virus, rather than antibody detection, has, in the past, resulted in greater emphasis being placed on mass serological screening as the best method for surveillance. However, surveillance based on clinical and pathological inspection and virological testing should not be underrated. If targeted at high risk groups in particular, it provides an opportunity for early detection that can considerably reduce the subsequent spread of disease. Herds predominated by adult animals, such as nucleus herds and artificial insemination studs, are particularly useful groups to monitor, since infection by low virulence viruses in such groups may be clinically inapparent, yet the degree of spread may be high.

Clinical and virological monitoring may also provide a high level of confidence of rapid detection of disease if a sufficiently large number of clinically susceptible animals is examined. In particular, molecular detection methods are increasingly able to offer the possibility of such large-scale screening for the presence of virus, at reasonable cost.

Wild pigs and, in particular, those with a wholly free-living existence, rarely present the opportunity for clinical observation, but should form part of any surveillance scheme and should, ideally, be monitored for virus as well as antibody.

3. Virological surveillance

Virological surveillance should be conducted using tests described in the Terrestrial Manual:

a) to monitor at risk populations;

b) to confirm clinically suspect cases;

c) to follow up positive serological results;

d) to test abnormal daily mortality, to ensure early detection of infection.

Molecular detection methods can be applied to large-scale screening for the presence of virus. If targeted at high risk groups, they provide an opportunity for early detection that can considerably reduce the subsequent spread of disease. Epidemiological understanding of the pathways of spread of CSFV can be greatly enhanced by molecular analyses of viruses in endemic areas and those involved in outbreaks in disease free areas.
Vaccine design and diagnostic methodologies, and in particular methods of virus detection, are increasingly reliant on up-to-date knowledge of the molecular, antigenic and other biological characteristics of viruses currently circulating and causing disease. Furthermore, epidemiological understanding of the pathways of spread of CSFV can be greatly enhanced by molecular analyses of viruses in endemic areas and those involved in outbreaks in disease-free areas. It is therefore essential that CSFV isolates are sent regularly to the regional OIE Reference Laboratory for genetic and antigenic characterization.

34. Serological surveillance

Serological surveillance aims at detecting antibodies against CSFV. Positive CSFV antibody test results can have five possible causes:

a. natural infection with CSF;
b. legal or illegal vaccination against CSF;
c. maternal antibodies derived from an immune sow (maternal antibodies) are usually found only up to 4.5 months of age, but, in some individuals, maternal antibodies can be detected for considerably longer periods;
d. cross-reactions with other pestiviruses;
e. non-specific reactors.

The infection of pigs with other pestiviruses may complicate a surveillance strategy based on serology. Antibodies to bovine viral diarrhoea virus (BVDV) and Border disease virus (BDV) can give positive results in serological tests for CSF, due to common antigens. Such samples will require differential tests to confirm their identity. Although persistently infected immunotolerant pigs are themselves seronegative, they continuously shed virus, so the prevalence of antibodies at the herd level will be high.

CSFV may lead to persistently infected, sero-negative young animals, which continuously shed virus. CSFV infection may also lead to chronically infected pigs which may have undetectable or fluctuating antibody levels. Even though serological methods will not detect these animals, such animals are likely to be in a minority and would not confound a diagnosis based on serology as part of a herd investigation.

It may be possible to use sera collected for other survey purposes for CSF surveillance. However, the principles of survey design described in this chapter and the requirement for statistical validity should not be compromised.

The discovery of clustering of seropositive reactions should be foreseen. It may reflect any of a series of events, including but not limited to the demographics of the population sampled, vaccinal exposure or the presence of infection by field strains or other pestiviruses. Because clustering may signal field strain infection, the investigation of all instances should be incorporated in the survey design. Clustering of positive animals is always epidemiologically significant and therefore should be investigated.

In countries or zones that are moving towards freedom, serosurveillance can provide valuable information on the disease status and efficacy of any control programme. In countries, zones or compartments that are heading towards freedom from CSF and have recently discontinued the use of vaccination, targeted serosurveillance of young, unvaccinated animals will indicate whether newly circulating virus is present, although the presence of maternal antibody will also need to be considered. Maternal antibodies are usually found up to four and a half months of age and can interfere with the interpretation of serological results. If conventional attenuated vaccine is currently being used or has been used in the recent past, serology aimed at detecting the presence of field virus will likewise need to be targeted at unvaccinated animals and after the disappearance of maternal antibody. General usage in such situations may also be used to assess levels of vaccine coverage.
Marker vaccines also exist which, when used in conjunction with accompanying DIVA tests as described in the Terrestrial Manual dedicated serological tests, may allow discrimination between vaccinal antibody and that induced by field natural infection. Such tools, described in the Terrestrial Manual, will need to be fully validated. They do not confer the same degree of protection as that provided by conventional vaccines, particularly with respect to preventing transplacental infections. Furthermore, however, the interpretation of serosurveillance results using DIVA techniques is only meaningful on a herd level. Such differentiation requires cautious interpretation on a herd basis.

The results of random or targeted serological surveys are important in providing reliable evidence that no CSFV infection is present in a country or zone. It is therefore essential that the survey be thoroughly documented.

The free status should be reviewed whenever evidence emerges to indicate that changes which may alter the underlying assumption of continuing freedom, has occurred. Such changes include but are not limited to:

f. an emergence or an increase in the prevalence of CSF in countries or zones from which live pigs or products are imported;

g. an increase in the volume of imports or a change in their country or zone of origin;

h. an increase in the prevalence of CSF in the domestic or wild pigs of adjacent countries or zones;

i. an increased entry from, or exposure to, infected wild pig populations of adjacent countries or zones.

Article 15.2.26.

Countries, zones or compartments declaring freedom from CSF: additional surveillance procedures

1. Country or zone free of CSF

In addition to the general conditions described above, a Member seeking recognition of CSF freedom for the country or a zone, whether or not vaccination had been practiced, should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances in and around the country or zone and will be planned and implemented according to the general conditions and methods described in this chapter. The objective is to demonstrate the absence of CSFV infection in domestic pigs and ascertain the infection status in wild pig populations, as described in Article 15.2.28. This requires the support of a national or other laboratory able to undertake identification of CSFV infection through virus detection and serological tests described in the Terrestrial Manual.

2. Compartment free of CSF

The objective of surveillance is to demonstrate the absence of CSFV infection in the compartment. The provisions of Chapter 4.3. and 4.4. should be followed. The frequency and intensity of surveillance should be defined and adapted to the prevailing epidemiological situation in the country or zone. Any deterioration in the epidemiological situation should trigger a review of the biosecurity measures and an intensification of surveillance. The effective separation of the two subpopulations should be demonstrated. To this end, a biosecurity plan that includes but is not limited to the following provisions should be implemented:

a. proper containment of domestic pigs;

b. control of movement of vehicles with cleaning and disinfection as appropriate;

c. control of personnel entering into the establishments and awareness of risk of fomite spread;
d. prohibition of introduction to the establishments of wild caught animals and their products;

e. record of animal movements into and out of establishments;

f. information and training programmes for farmers, processors, veterinarians, etc.

The biosecurity plan implemented also requires internal and external monitoring by the Veterinary Authority. This monitoring should include:

g. periodic clinical and serological monitoring of herds in the country or zone, and adjacent wild pig populations following these recommendations;

h. herd registration;

i. official accreditation of biosecurity plans;

j. periodic monitoring and review.

Monitoring the CSF status of wild and domestic pig populations outside the compartment will be of value in assessing the degree of risk they pose to the CSF free compartment. The design of a monitoring system should follow the provision described in this Chapter and in Chapter 1.4, dependent on several factors such as the size and distribution of the population, the organisation of the Veterinary Services and resources available. The occurrence of CSF in wild and domestic pigs may vary considerably among countries. Surveillance design should be epidemiologically based, and the Member should justify its choice of design prevalence and level of confidence based on Chapter 1.4.

The geographic distribution and approximate size of wild pig populations need to be assessed as a prerequisite for designing a monitoring system. Sources of information may include government wildlife authorities, wildlife conservation organisations, hunter associations and other available sources. The objective of a surveillance programme when the disease is already known to exist should be to determine the geographic distribution and the extent of the infection.

Recovery of free status: additional surveillance procedures

In addition to the general conditions described in the above mentioned articles, a Member seeking reestablishment of country or zone freedom from CSF should show evidence of an active surveillance programme to demonstrate absence of CSFV infection.

Populations under this surveillance programme should include:

a. establishments in the proximity of the outbreak;

b. establishments epidemiologically linked to the outbreak;

c. animals used to re-populate affected establishments and any establishments where contiguous culling is carried out;

d. wild pig populations in the area of the outbreak.

In all circumstances, a Member seeking reestablishment of country or zone freedom from CSF with vaccination or without vaccination should report the results of an active and a passive surveillance programme, in which the pig population should undergo regular clinical, pathological, virological, and/or serological examination, planned and implemented according to the general conditions and methods described in these recommendations. The surveillance should be based on a statistically representative sample of the populations at risk. To regain CSF free status, the surveillance approach should provide at least the same level of confidence as demonstrated during the previous declaration of freedom.
Annex XXVIII (contd)

Article 15.2.28.

Surveillance for CSFV infection in wild pigs

1. The objective of a surveillance programme is to determine the CSFV infection status of wild pigs, as well as the geographic distribution and prevalence, if present. While the same principles apply, surveillance in wild pigs presents challenges beyond those encountered in domestic populations in each of the following areas:
   a) determination of the distribution, size and movement patterns associated with the wild pig population;
   b) assessment of the possible presence of CSF within the population;
   c) determination of the practicability of establishing a zone.

2. The design of a monitoring system for wild pigs is dependent on several factors such as the organisation of the Veterinary Services and resources available. The geographic distribution and approximate size of wild pig populations need to be assessed as a prerequisite for designing a monitoring system. Sources of information to aid in the design of a monitoring system may include wildlife conservation organisations, hunter associations and other available sources, etc. The objective of a surveillance programme is to determine if a given disease is present, and if so, at what prevalence.

3. Estimates of wild pig populations can be made using advanced a variety of methods (e.g. including radio tracking, linear transect method, capture/recapture) or estimates based on the number of animals hunted traditional methods based on the number of animals that can be hunted to allow for natural restocking (hunting bags).

4. For implementation of the monitoring programme, it will be necessary to define the limits of the territory over which wild pigs range in order to delineate the epidemiological units within the monitoring programme. It is often difficult to define epidemiological units for wild animals. The most practical approach is based on natural and artificial barriers.

5. The monitoring programme should also include animals found dead, road kills, animals showing abnormal behaviour or exhibiting gross lesions during dressing.

6. There may be situations where a more targeted surveillance programme can provide additional assurance. The criteria to define high risk areas for targeted surveillance include:
   a. areas with past history of CSF;
   b. sub-regions with large populations of wild pigs;
   c. border regions with CSF affected countries or zones;
   d. interface between wild and domestic pig populations;
   e. picnic and camping areas;
   f. farms with free-ranging pigs;
   g. garbage dumps;
   h. other risk areas determined by the Veterinary Authority such as garbage dumps and picnic and camping areas.
Chapter 8.15.164.

Swine Vesicular Disease

Article 8.15.164.1.

The pig is the only natural host for swine vesicular disease (SVD) virus. The definition of pig includes all varieties of Sus scrofa, both domestic and wild.

For the purposes of the Terrestrial Code, the incubation period for swine vesicular disease (SVD) shall be 28 days.

For the purposes of this Chapter, the Terrestrial Code, SVD is defined as an infection of susceptible animals include domestic and wild pigs.

Domestic pig is defined as all domesticated pigs, permanently captive or farmed free range, used for the production of meat for consumption, for the production of other commercial products or for breeding these categories of pigs.

For the purposes of the Terrestrial Code, the incubation period for SVD shall be 28 days.

For the purposes of this Chapter, a case includes an animal infected with SVD virus (SVDV).

For the purposes of international trade, this chapter deals not only with the occurrence of clinical signs caused by SVDV virus (SVDV), but also with the presence of infection with SVDV in the absence of clinical signs. For the purposes of this Chapter, virus The following defines the occurrence of infection means presence of with SVDV as demonstrated by:

1. virus isolation, or detection of virus antigen or virus nucleic acid, or
2. seroconversion, or
3. clinical signs associated with serological evidence, or
4. clinical signs or serological evidence associated with an epidemiological link.

Standards for diagnostic tests are described in the Terrestrial Manual.

A Member should not impose trade bans in response to a notification of infection with SVDV in wild pigs according to Article 1.2.3. of the Terrestrial Code.

Article 15.4.1bis.

Determination of the SVD status of a country, zone or compartment

The SVD status of a country, zone or compartment can only be determined after considering the following criteria, as applicable:

1. SVD should be notifiable in the whole territory, and all clinical signs suggestive of SVD should be subjected to appropriate field and/or laboratory investigations;
2. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of SVD;
3. the Veterinary Authority should have current knowledge of, and authority over, all domestic pigs in the country, zone or compartment.
Annex XXIX (contd)

4. The Veterinary Authority should have current knowledge about the population and habitat of wild pigs in the country or zone.

5. For domestic pigs, appropriate surveillance, capable of detecting the presence of infection even in the absence of clinical signs, is in place; this may be achieved through a surveillance programme in accordance with Articles 15.4.14. to 15.4.19.

Article 8.15.46.2.

SVD free country, zone or compartment

Susceptible animals in the SVD free country or zone or compartment should be separated from neighbouring infected countries or zones by animal health measures (biosecurity measures, which may include a buffer zone) that effectively prevent the entry of the virus, or by physical barriers.

The SVD status of a country, zone or compartment can only be determined by applying surveillance recommendations described Chapter 1.4. according to two possibilities:

1. Historically free status

A country or zone may be considered free from the disease without formally applying a pathogen specific surveillance programme if the provisions of Article 1.4.6. are complied with.

2. Free status as a result of a specific surveillance programme

A country, zone or compartment which does not meet the conditions of point 1 above may be considered free from SVD when:

a) surveillance for both SVD and SVDV infection in accordance with Articles 15.4.14. to 15.4.19 and Chapter 1.4. has been in place for at least 3 years;

b) no outbreak of SVD and no evidence of SVDV circulation has been found during the past 3 years;

c) regulatory measures for the prevention and control of SVD have been implemented, including the control of the movement of susceptible animals pigs and other relevant measures for preventing the entry of the virus.

If a stamping out policy was applied in respect of the most recent outbreak, the requirement of 3 years in points a) and b) above is shortened to 12 months.

Article 8.15.46.3.

SVD infected country or zone

An SVD infected country or zone is a country or zone one that does not fulfill the requirements to be considered as free.

Article 8.15.46.4.

Establishment of a containment zone within an SVD free country or SVD free zone

In the event of a limited outbreaks within an SVD free country or SVD free zone, a single containment zone, which includes all cases, can be established for the purpose of minimizing the impact on the entire country or zone. For this to be achieved, the Veterinary Authority should be able to provide documented evidence that:
1. the outbreak is limited based on the following factors:
   a) immediately on suspicion, a rapid response including notification has been made;
   b) standstill of animal pig movements has been imposed, and effective controls on the movement of other commodities mentioned in this chapter are in place;
   c) the infection has been confirmed;
   d) epidemiological investigation (trace-back, trace-forward) has been carried out;
   e) the primary outbreak has been identified and investigations of the likely source of the outbreak have been identified carried out;
   f) all cases have been shown to be epidemiologically linked;

2. surveillance in accordance with Articles 15.4.14 -15.4.19 and Chapter 1.4. is in place and demonstrates that there are no undetected cases in the containment zone;

3. a stamping-out policy has been applied;

4. the pig population within the containment zones should be clearly identifiable as belonging to the containment zone;

5. increased passive and targeted surveillance in accordance with Articles 15.4.14 -15.4.19 and Chapter 1.4. in the rest of the country or zone has been carried out and has not detected any evidence of infection;

6. measures to prevent spread of the infection from the containment zone to the rest of the country or zone, are in place.

The free status of the area outside the containment zone would be suspended pending the establishment of the containment zone. The suspension of free status of this area could be lifted irrespective of the provisions of Article 815.164.5., once the containment zone is clearly established, by complying with points 1 to 6 above.

The recovery of the SVD free status of the containment zone should follow the provisions of Article 815.164.5.

When importing from containment zones, provisions of Articles 815.164.6., 815.164.98., 815.164.140., 15.4.12 and 815.164.13., concerning the importation from countries or zones considered infected with SVD, should be applied.

Article 815.164.5.

Recovery of free status

When an SVD outbreak or SVDV infection occurs in an SVD free country or zone, one of the following waiting periods is required to regain the status of SVD free country or zone.

1. 2 months after the stamping-out of the last case, where a containment zone and serological surveillance have been applied in accordance with this chapter and Chapter 1.4.; or

2. 12 months after the stamping-out of the last case, where the conditions for the establishment of a containment zone are not fulfilled, a stamping-out policy and serological surveillance have been applied in accordance with this chapter and Chapter 1.4.
Annex XXIX (contd)

Where both a stamping-out policy and serological surveillance in accordance with this chapter X.X. have not been practiced, the above waiting periods do not apply, and Article 815.164.2. applies.

Direct Transfer of pigs from an infected zone for directly to slaughter of SVD susceptible animals from an infected zone to a free zone within a country.

In order not to jeopardise the status of a free zone, pigs SVD susceptible animals should only leave the an infected zone if moved by mechanised transported directly to slaughter in to the nearest designated abattoir, located in the buffer zone (if established), directly to slaughter under the following conditions:

In the absence of an abattoir in the buffer zone, or in the absence of a buffer zone, live SVD susceptible animals can be transported to the nearest abattoir in a free zone directly to slaughter only under the following conditions:

1. no SVD susceptible animal pig has been introduced into the establishment of origin and no animal pig in the establishment of origin has shown clinical signs of SVD for at least 60 days prior to movement;
2. a representative sample of animals of pigs in the herd of origin, including all animals pigs to be moved for slaughter has been serologically tested with negative findings;
3. the animals pigs were kept in the establishment of origin for at least 2 months prior to movement;
4. SVD has not occurred within a 1 kilometre radius of the establishment of origin for at least 2 months prior to movement;
5. the animals pigs must should be transported under the supervision of the Veterinary Authority in a vehicle, which was cleansed and disinfected before loading, directly from the establishment of origin to the abattoir without coming into contact with other susceptible animals pigs;
6. such an abattoir is not approved for the export of fresh meat during the time it is handling the meat of animals pigs from the infected zone and, to be re-approved, must should apply disinfection able to that will destroy any residual infectivity;
7. vehicles and the abattoir must should be subjected to thorough cleansing and disinfection able to that will destroy any residual infectivity immediately after use.

All products obtained from the animals pigs and any products coming into contact with them must should be identified and traded only on domestic market.

Animals Pigs moved into a free zone for other purposes must should be moved under the supervision of the Veterinary Authority and comply with the conditions in Article 815.164.98.

Recommendations for importation from SVD free countries, zones or compartment for domestic pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the animals pigs:

1. showed no clinical sign of SVD on the day of shipment;
2. were kept in an SVD free country, zone or compartment since birth or for at least the past 60 days.

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Article 8.16.8.

Recommendations for importation from SVD free countries or zones for wild pigs

_Veterinary Authorities_ should require the presentation of an _international veterinary certificate_ attesting that the animals:

1. showed no clinical sign of SVD on the day of shipment;
2. came from an SVD free country or zone;
3. if the country or zone of origin has a common border with a country or zone considered infected with SVD:
   3. were kept in a _quarantine station_ for the 60 days prior to shipment and were subjected to a prescribed serological test for SVD with negative results during that period.

Article 8.15.144, 145, 146, 147.

Recommendations for importation from countries or zones considered infected with SVD for domestic and wild pigs

_Veterinary Authorities_ should require the presentation of an _international veterinary certificate_ attesting that the animals:

1. showed no clinical sign of SVD on the day of shipment;
2. were kept in a _quarantine station_ for the 60 days prior to shipment and were subjected to a prescribed serological test for SVD with negative findings at the end of that period.

Article 8.15.144, 145, 146, 147.

Recommendations for importation from SVD free countries or zones or compartments for semen of pigs

_Veterinary Authorities_ should require the presentation of an _international veterinary certificate_ attesting that:

1. the donor animals:
   a) showed no clinical sign of SVD on the day of collection of the semen;
   b) were kept in an SVD free country or zone or compartment for not less than 60 days prior to collection;
2. the semen was collected, processed and stored in conformity with the provisions of Chapter 4.6.

Article 8.15.144, 146, 147.

Recommendations for importation from countries or zones considered infected with SVD for semen of pigs

_Veterinary Authorities_ should require the presentation of an _international veterinary certificate_ attesting that:

1. the donor animals showed no clinical sign of SVD on the day of collection of the semen and were subjected to a prescribed serological test for SVD with negative findings;
Annex XXIX (contd)

2. the donor animals pigs were kept in the exporting country or zone for the 60 days prior to collection, in an establishment or artificial insemination centre where no case of SVD was officially reported during that period, and that the establishment or artificial insemination centre was not situated within one km from an outbreak occurring in the last 60 days;

3. a representative sample of animals pigs of in the herd of origin has been serologically tested with negative findings;

4. the semen was collected, processed and stored in conformity with the provisions of Chapter 4.6.

Recommendations for importation from SVD free countries, zones or compartments

for fresh meat of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals:

1. which have been kept in an SVD free country, zone or compartment since birth or for at least the past 60 days;

2. which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections for SVD with favourable outcome.

All the necessary measures have been taken to avoid cross contamination.

Recommendations for importation from SVD infected countries, zones or compartments

for meat products of pigs

Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the entire consignment of meat products have been processed in an establishment approved by the Veterinary Authority so as to ensure the destruction of the SVD virus by either:

1. heat treatment in a hermetically sealed container with an F0 value of 3.00 or more, or

2. heat treatment at a minimum temperature of 70 °C, which must be reached throughout the meat, or

3. heat treatment in a hermetically sealed container to at least 60 °C for a minimum of 4 hours, during which time the core temperature must be at least 70 °C for 30 minutes, or

4. natural fermentation and maturation of not less than nine months, resulting in the following characteristics: $A_w$ value of not more than 0.93 or a pH value of not more than 6.0, and

5. all the necessary measures have been taken to avoid cross contamination.

Recommendations for the importation of meat products of pigs (either domestic or wild), or for products of animal pig origin (from fresh meat of pigs) intended for use in animal feeding, for agricultural or industrial use, or for pharmaceutical or surgical use, or for trophies derived from wild pigs.
Veterinary Authorities of importing countries should require the presentation of an international veterinary certificate attesting that the products:

4. have been prepared:
   a1. exclusively from fresh meat meeting the conditions laid down in Article 8.15.16.12, as relevant; or
   a2. from meat products meeting the conditions laid down in Article 15.4.12;
   b) in a processing establishment:
      i) approved by the Veterinary Authority for export purposes;
      ii) processing only meat meeting the conditions laid down in Article 8.16.12, as relevant;
   OR

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the SVD virus.

Article 15.4.14.

Surveillance: introduction

The Articles 15.4.14 – 15.4.19 define the principles and provides a guide for the surveillance of SVD complementary to Chapter 1.4., applicable to Members seeking to determine their SVD status for the whole country or a zone, or a compartment. Guidance on surveillance for countries seeking re-establishment of freedom from SVD for the whole country or a zone, or a compartment following an outbreak, as well as for demonstrating the maintenance of SVD free status is also provided.

Consideration should be given to the known characteristics of SVD epidemiology, which include the impact of different production systems on disease spread, the lack of pathognomonic gross lesions and clinical signs, and the frequency of clinically inapparent infection. Serological cross-reactivity with other agents has to be taken into consideration when interpreting data from serological surveys.

Clinically, SVD may be indistinguishable from foot and mouth disease (FMD), and this is its main importance. And since any vesicular condition in pigs may be FMD, it is therefore essential that cases of SVD be distinguished urgently from FMD by laboratory investigation.

Article 15.4.15.

Surveillance: general conditions and methods

1. A surveillance system in accordance with Chapter 1.4. should be under the control of the Veterinary Authority:
   a) a formal and ongoing system for detecting and investigating outbreaks of disease or SVDV infection should be in place;
   b) a procedure should be in place for the rapid collection and transport of samples from suspect cases of SVD to a laboratory for SVD diagnosis as described in the Terrestrial Manual;
   c) a system for recording, managing and analysing diagnostic and surveillance data should be in place.
The SVD surveillance programme should:

a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers, who have day-to-day contact with livestock, as well as diagnosticians, should report promptly any suspect case of SVD. All suspected cases of SVD should be investigated immediately. Where suspicion cannot be resolved by epidemiological and clinical investigation, samples should be taken and submitted to an approved laboratory. This requires that sampling kits and other equipment are available for those responsible for the surveillance. Personnel responsible for the surveillance should be able to call for assistance from a team with expertise in vesicular diseases diagnosis and control.

b) implement when relevant, regular and frequent clinical inspection and serological testing of high-risk groups of animals (risks linked to the types of production cycle, local trade pattern, holding with poor bio-security measures, possible direct or indirect contact with other pigs).

An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is SVD. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot, therefore, be reliably predicted. Recognition for freedom from SVD infection should, as a consequence, provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were submitted during the investigation (quarantine, movement stand-still orders, etc.).

**Article 15.4.16.**

**Surveillance strategies**

1. **Introduction**

The population targeted by surveillance programs aimed at identifying disease and infection should include domestic pig populations within the country or zone or compartment to be recognised as free from SVD.

Given the existence of clinically inapparent infection and difficulties associated with clinical diagnosis of SVD, serology is often the most effective and efficient surveillance methodology. In some circumstances, which will be discussed later, clinical and virological surveillance may also have a value.

2. **Clinical surveillance**

SVD can be sub-clinical, mild or severe depending on the strain of virus involved, the route and dose of infection, and the husbandry condition under which the pigs are kept.

Clinically, SVD is indistinguishable from FMD and, when a vesicular condition is seen in pigs, it must be assumed to be FMD until investigated by laboratory tests and proven otherwise.
Nevertheless, SVD caused by mild strains may remain unobserved, and in this case the value of clinical examination alone is insufficient as a surveillance tool; in this case serology is often the most effective and efficient surveillance methodology.

Clinical surveillance and laboratory testing should always be applied in series to clarify the status of suspected cases detected by either of these complementary diagnostic approaches. Laboratory testing may confirm clinical suspects, while clinical surveillance may contribute to confirmation of positive serology. Any sampling unit within which suspicious animals are detected should be classified as infected until contrary evidence is produced.

Identification of suspected cases is vital to identify the sources of SVDV. It is essential that SVDV isolates are sent regularly to a Reference Laboratory to enable the determination of the molecular, antigenic and other biological characteristics of the virus.

3. Virological surveillance

Virological surveillance using tests described in the Terrestrial Manual should be conducted:

a) to monitor an at risk population;
b) to confirm clinically suspected cases;
c) to follow up positive serological results.

The most suitable samples for virological testing are vesicular lesion materials from clinically affected pigs and faeces from pigs without lesions.

4. Serological surveillance

Serological surveillance aims at the detection of antibodies against SVD. Positive SVD antibody test results can have three possible causes:

a) natural infection with SVD;
b) maternal antibodies derived from immune sows (no published data exist so far on the duration of maternal passive immunity against SVD);
c) non-specific reactors.

The use and interpretation of serological tests

Any positive test result should be followed up immediately using appropriate clinical, epidemiological, serological and virological investigations of the reactor animals at hand, and of susceptible animals of the same epidemiological unit and those that have been in contact or otherwise epidemiologically associated with the reactor animals. If the follow-up investigations provide no evidence for SVDV active infection, the reactor animal shall be classified as non SVD infected. In all the other cases, including the absence of such follow-up investigations, the reactor animals should be classified as SVD positive.
It is suggested that in the primary sampling units where at least one animal reacts positive to the screening test, the following strategy should be applied (Figure 1):

1. In case of positive results to the screening test (ELISA), all positive sera from the herd should be tested using the Virus Neutralization (VN) test. If there are pigs that test serologically positive by VN test, the positive sample may be tested to identify the isotype of antibody (IgM or IgG).

2. The positive herd should undergo clinical examination with collection of samples for virological testing (vesicular lesions and/or faces). In the presence of symptoms compatible with SVD and/or detection of virus, the herd is to be considered infected.

3. Identification of the isotype of antibody present in positive sera can be helpful in the evaluation of the epidemiological meaning of results, as sera from recently infected pigs usually contain specific IgM alone, subsequently both IgM and IgG, and later exclusively IgG. Therefore, in the sero-positive herd:
   a) The clinical examination and virological testing of sero-positive animals and animals in contact should be targeted to the IgM positive animals and to those living in their proximity, rather than to the IgG positives.
   b) The presence of IgG positives exclusively may indicate a low likelihood of SVDV circulation.
   c) The presence of a single reactor, containing exclusively IgM also on re-testing, without increase of VN titre, in the absence of symptoms and seroconversion in animals in contact, is usually due to non-specific reaction.

4. In the case of seroreactor herds without clinical signs or positive virological findings, after an adequate interval of time has lapsed (at least 7 days), following clinical examination, a second serum sample should be collected from the positive animals and also from a representative number of pigs in contact with the positives in the primary sampling. These samples are tested using ELISA and VN test and antibody titres at the time of retest should be equal to or lesser than those observed in the initial test if virus is not circulating.

5. In case of the detection of an outbreak, an epidemiological investigation has to be performed and a representative sample of animals in all epidemiologically linked herds should be serologically tested.

Possible alternative strategies may be adopted, but in this case the country should justify the procedure chosen as adequate to detect the presence of SVDV infection. Possible shortcomings in the sensitivity of alternative diagnostic strategies should be addressed by appropriate changes in the surveillance design and in the sample size.

Fig 1: Should confirm that SVD virus could be demonstrated in samples from pigs on seroreactor herds before declaring an outbreak, even if clinical signs suggestive of SVD were found.

Article 15.4.18.

Countries, zones or compartments declaring freedom from SVD: Additional surveillance procedures

1. Country or zone free of SVD

In addition to the general conditions described in this chapter, a Member declaring freedom from SVD for the entire country or a zone should provide evidence for the existence of an effective surveillance programme. The strategy and the design of the surveillance programme will depend on the prevailing epidemiological circumstances. It will be planned and implemented to demonstrate the absence of SVDV infection in susceptible populations, during the preceding 3 years, according to general conditions and methods described in this chapter. This requires the support of a national or other laboratory able to undertake identification of SVDV infection through virus detection and antibody tests described in the Terrestrial Manual.
This surveillance may be targeted to a pig population at specific risks linked to the types of production, local trade patterns, holdings with poor biosecurity measures in place.

2. Compartment free of SVD

The objective of surveillance is to demonstrate the absence of SVDV infection in the compartment. The provisions of Chapters 4.3. and 4.4. should be followed. The frequency and intensity of surveillance should be defined and adapted to the prevailing epidemiological situation in the country or zone. Any deterioration in the epidemiological situation should trigger a review of the biosecurity measures and an intensification of surveillance.

Article 15.4.19

Recovery of status: additional surveillance procedures

In addition to the general conditions described in this chapter, a country, zone or compartment regaining freedom from SVDV infection should show evidence of an active surveillance programme aimed to demonstrate the absence of the infection.

The population under this surveillance programme should include:

a) in the establishments in the area of the outbreak;

b) in the establishments epidemiologically linked to the outbreak;

c) used to re-populate affected establishments.

This will require surveillance incorporating virus detection and antibody tests described in the Terrestrial Manual.

In all circumstances, a Member self-declaring freedom of a country, zone or compartment after an outbreak, should report the results of an active surveillance programme in which pigs undergo regular active surveillance, planned and implemented according to the general conditions and methods described in this chapter.
Figure 1. Use and interpretation of serological tests

Sampled herd

Screening test (ELISA)

- Positive result
  - VN
    - Positive result
      - STOP
    - Negative result
      - Negative herd

- Negative result
  - STOP

- Positive result
  - Isotype identification
    - Isotypes suggestive of recent infection
      - See text for details
      - Outbreak
      - STOP
    - Isotypes not suggestive of recent infection
      - See text for details
      - Outbreak
      - ELISA+VN
        - No seroconversion
          - No outbreak
        - Seroconversion
          - Outbreak
  - Clinical exam of positive source and in contact after 7 days
    - No lesions or symptoms
      - STOP
    - Lesions or symptoms
      - Outbreak

- Negative herd

Epidemiological investigation, trace-back, trace forth and testing of all herds in contact

STOP
CHAPTER 3.4
COMMUNICATION

Article 3.4.1.

General considerations
In general communication entails the exchange of information between various individual, institutional and public audiences for purposes of informing, guiding and motivating action. The application of the science and technique of communication involves modulating messages according to situations, objectives and target audiences.

The recognition of communication as a discipline of the Veterinary Services and its incorporation within it is critical for their operations. The integration of veterinary and communication expertises is essential for effective communication.

Communication should be an integral part of all the activities of the Veterinary Services including animal health (surveillance, early detection and rapid response, prevention and control), animal welfare and veterinary public health (food safety, zoonoses) and veterinary medicine.

Objectives of this chapter on communication for the Veterinary Services are to provide guidance for the development of a communication system, strategic and operational communication plans and elements to assess their quality.

Article 3.4.2.

Principles of communication
1. Veterinary Services should have the authority and capability to communicate on matters within their mandate.

2. Veterinary and communication expertises should be combined.

3. Communication should be targeted and follow the fundamental criteria of transparency, consistency, timeliness, balance, accuracy, honesty and empathy and respect the fundamental principles of quality of Veterinary Services (article 3.1.2).

4. Communication should be a continuous process.

5. Veterinary Services should be responsible for planning, implementing, monitoring, evaluating and revising their strategic and operational communication plans.

Article 3.4.3.

Definitions
Communication means the discipline of informing, influencing, guiding and motivating individual, institutional and public audiences, preferably ideally on the basis of interactive exchanges, about any issue falling under the mandate of the OIE and under the competence of the Veterinary Services.
Annex XXX (contd)

Crisis

means a time situation of great danger, threat, difficulty or uncertainty when problems related to any issues falling under the mandate of the OIE and the competence of the Veterinary Services require immediate action.

Crisis Communication

means the process of communicating information of potentially incomplete nature within time constraints, in the event of a crisis, that allows an individual, affected and/or interested parties, an entire community or the general public to make best possible decisions and be informed of and/or accept policy decisions and rationale behind policy decisions during a crisis.

Outbreak communication

means the process of communicating in the event of an outbreak. Outbreak communication includes notification.

Article 3.4.4.

Communication system

In addition to the Principles for Communication the following critical elements should be used in conjunction with Chapter 3.1., when planning, implementing and assessing a communication system.

Critical elements

1. Authority and organizational structure

   a) Legislation providing authority to Veterinary Services under the responsibility of the CVO to communicate on matters within their mandate

   b) Identified and accessible official contact points for communication

   c) Organizational chart indicating direct link to the CVO through chain of command (e.g. dedicated communication unit, communication officer)

2. Human resources

   a) Job descriptions of communication personnel identifying roles and responsibilities

   b) Sufficient number of qualified personnel with knowledge, skills, attitude and abilities relevant to communication

   c) Continuous training and education on communication provided to communication personnel

3. Financial and material resources

   a) Clearly identified budget for communication that provides adequate funding

   b) Provision and/or access to appropriate material resources in order to carry out roles and responsibilities: suitable premise/accommodation that is adequately equipped with sufficient office and technical equipment, including information technology and access to the Internet
4. Management of the communication system
   
a) Roles and responsibilities of the communication unit
   
   i) Report to the CVO
   
   ii) Engaged in decision-making process
   
   iii) Responsible for the planning, implementation and evaluation of the strategic and operational plans for communication and relevant standard operating procedures
   
   iv) Function as contact point on communication issues for the *Veterinary Services*
   
   v) Provide guidance and expertise on communication issues to the *Veterinary Services*
   
   vi) Provide and coordinate continuous education on communication for the *Veterinary Services*
   
b) Strategic plan for communication
   
   A well-designed strategic plan for communication should support the *Veterinary Services* strategic plan and have management support and commitment. The strategic plan for communication should address all high level organization-wide communication objectives. The plan should be a long-term plan.

   A strategic plan for communication should be monitored, periodically reviewed and should identify measurable performance objectives and techniques to assess.

   The strategic plan for communication should consider the different types of communication: routine communication, risk communication, outbreak communication and crisis communication.

   The key outcomes in effectively implementing a strategic plan for communication are increased knowledge and awareness of issues by the public and stakeholders, higher understanding of the role of the Veterinary Services, higher visibility of and improved trust and credibility in the Veterinary Services. These will enhance understanding and/or acceptance of policy decisions and subsequent change of perception, attitude and/or behaviour.

   c) Operational plans for communication
   
   Operational plans for communication should be based on the assessment of specific issues and should identify specific objectives and target audiences such as staff, partners, stakeholders, media and the general public.

   Each operational plan for communication should consist of a well-planned series of activities using different techniques, tools, messages and channels to achieve intended objectives and utilizing available resources within a specific timeframe.