Annex I

MEETING OF THE OIE
TERRESTRIAL ANIMAL HEALTH STANDARDS COMMISSION

Agenda

1. Welcome - President of the Commission
2. Meeting with Director General
3. Joint meetings of commissions
   • Meeting of the Code Commission and the Scientific Commissions
   • Meeting of the Code Commission and the Biological Standards Commission
4. Examination of Member Countries’ comments and work of relevant Expert Groups

Item 1 General definitions
   a) General definitions (Chapter 1.1.1.)
   b) Status report on incorporation of new definitions (Veterinary authority, etc.)
   c) New definition proposed by Permanent Animal Welfare Working Group (PAWWG)

Item 2 Model Certificates
   a) General obligations (Chapter 1.2.1.)
   b) Notes for Guidance on Veterinary Certificates for International Trade in Live Animals, Hatching Eggs and Products of Animal Origin (Appendix X.X.X)
   c) Other horizontal chapters

Item 3 Evaluation of Veterinary Services
   a) Chapter 1.3.3., Chapter 1.3.4.
   b) Report of the ad hoc Group on the Evaluation of Veterinary Services
   c) OIE Tool for the Evaluation of Performance of Veterinary Services (OIE PVS Tool)

Item 4 Zoning and compartmentalisation
   a) Chapter 1.3.5.
   b) General guidelines on the application of compartmentalisation (Appendix X.X.X)
   c) Checklist on the practical application of compartmentalisation for avian influenza and Newcastle disease
   d) Use of the compartmentalisation concept for vector borne diseases

Item 5 Rabies (Chapter 2.2.5.)

Item 6 Foot and mouth disease
   a) Chapter 2.2.10
   b) Guidelines on Surveillance for foot and mouth disease (Appendix 3.8.7)
   c) Virus inactivation procedures (Appendix 3.6.2)
Annex I (contd)

Item 7 Rinderpest
   a) Chapter 2.2.12.
   b) Guidelines on surveillance for rinderpest (Appendix 3.8.2.)

Item 8 Contagious bovine pleuropneumonia (Chapter 2.3.15. and Appendix 3.8.3.)

Item 9 General guidelines on animal health surveillance (Appendix 3.8.1.)

Item 10 Bluetongue
   a) Chapter 2.2.13.
   b) Guidelines on surveillance for bluetongue (Appendix 3.8.10.)

Item 11 Bovine brucellosis (Chapter 2.3.1.)

Item 12 Bovine tuberculosis (Chapter 2.3.3.)

Item 13 Bovine spongiform encephalopathy
   a) Chapter 2.3.13.
   b) Risk assessment recommendations (Appendix 3.8.5.)

Item 14 Equine influenza (Chapter 2.5.5.)

Item 15 Equine diseases (other than equine influenza and AHS)
   a) Equine rhinopneumonitis (Chapter 2.5.7.)
   b) Equine viral arteritis (Chapter 2.5.10.)

Item 16 African horse sickness
   a) Chapter 2.5.14.
   b) Guidelines on surveillance for African horse sickness (Appendix 3.8.X.)

Item 17 African swine fever (Chapter 2.6.6.)

Item 18 Classical swine fever
   a) Chapter 2.6.7.
   b) Guidelines on surveillance for classical swine fever (Appendix 3.8.8.)

Item 19 Avian influenza
   a) Chapter 2.7.12.
   b) Guidelines on the inactivation of avian influenza virus (Appendix 3.6.5.)
   c) Guidelines on surveillance for avian influenza (Appendix 3.8.9.)

Item 20 Newcastle disease
   a) Chapter 2.7.13.
   b) Guidelines on surveillance for Newcastle disease (Appendix 3.8.X.)
   c) Guidelines on the inactivation of the Newcastle disease virus

Item 21 West Nile fever (Chapter 2.X.XX.)

Item 22 Draft guidelines on the design and implementation of identification system to achieve animal traceability

Item 23 Guidelines on the control of hazards of animal health and public health importance in animal feed

Item 24 Guidelines on the detection, control and prevention of Salmonella Enteritidis and S. Typhimurium in poultry producing eggs for human consumption (Appendix 3.10.2.)
Annex I (contd)

Item 25  Animal welfare

  a)  Guidelines on the transport of animals by sea and land

  b)  *Ad hoc* Group on slaughter and humane killing

  c)  Draft guidelines on dog population control

  d)  Update on 2nd OIE Global Conference on Animal Welfare 2008

  e)  Update from PAWWG meeting 5-7 September 2007

      - animal production systems

      - control of dog populations

      - laboratory animals

      - wildlife harvest

Item 26  Infectious bursal disease (IBD)

Item 27  Small hive beetle

Item 28  Leptospirosis (Chapter 2.2.4.)

Item 29  Paratuberculosis (Chapter 2.2.6.)

5.  Other issues

Item 30  Commodity based measures in the *Code*

Item 31  Division of the *Code* into two volumes

Item 32  Report of OIE/FAO *ad hoc* Group meeting on Guide to Good Farming Practice

Item 33  Other documents

  a)  The Role of the Veterinary Services in Food Safety

  b)  Report of the OIE *ad hoc* Group on the Notification of Terrestrial Animal Diseases / Pathogenic Agents

Item 34  Future work programme

Item 35  Others
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OIE Terrestrial Animal Health Standards Commission/September 2007
CHAPTER 1.1.1. GENERAL DEFINITIONS

Article 1.1.1.1.

For the purposes of the Terrestrial Code:

**Animal welfare**
means the state of animal as regards its attempts to cope with its environment and includes both the extent of failure to cope and the ease or difficulty in coping.

**Approved abattoir**
means premises used for the slaughter of animals for human consumption or animal feeding and approved by the Veterinary Authority for export purposes.

**Area of direct transit**
means a special area established in a transit country, approved by the relevant Veterinary Authority and placed under its immediate control, where animals stay for a short time pending further transport to their final destination.

**Breeding birds**
means birds kept for the purpose of producing hatching eggs.

**Collecting centre**
means a premise or a place where animals for breeding or rearing or animals for slaughter from different establishments or markets are collected together.

**Commodity**
means live animals, products of animal origin, animal genetic material, intended for human consumption, for animal feeding, for pharmaceutical or surgical use or for agricultural or industrial use, semen, embryos/ova, biological products and pathological material.

**Flock of birds**
means any group of birds continuously housed in one building or part of a building separated from other parts of that building by a solid partition and having its own ventilation system, or, in the case of free range birds, any group of birds having common access to one or more buildings or More than one flock of birds may exist in one establishment.

**Flock**
means a number of animals of one kind kept together under human control or a congregation of gregarious wild animals. For the purposes of the Terrestrial Code, a flock is usually regarded as an epidemiological unit.

**Herd**
means a number of animals of one kind kept together under human control or a congregation of gregarious wild animals. For the purposes of the Terrestrial Code, a herd is usually regarded as an epidemiological unit.

**Infected country**
means a country in which the absence of the disease under consideration has not been demonstrated by the requirements specified in the Terrestrial Code being met.
Annex III (contd)

Infection
means the presence of the pathogenic agent in the host entry and development or multiplication of an infectious agent in the body of men or animals.

Laying birds
means birds kept for the purpose of producing eggs not intended for hatching.

Monitoring
means the continuous investigation of intermittent performance and analysis of routine measurements, aimed at changes in the environment or health status of a given population or subpopulation, and its environment, to detect changes in the prevalence of a disease or characteristics of a pathogenic agent.

Official veterinary control
means that the Veterinary Services knows the location of the animals and the identity of their owner or responsible keeper and is able to apply appropriate animal health measures, as required.

Quarantine station
means a facility where animals are maintained in isolation with no direct or indirect contact with other animals, to prevent the transmission of specified pathogen(s) while the animals are undergoing observation for a specified length of time and, if appropriate, testing and treatment.

Risk
means the likelihood of the occurrence and the likely magnitude of the consequences of an adverse event to animal or human health in the importing country during a specified time period, as a result of a hazard.

Risk assessment
means the evaluation of the likelihood and the biological and economic consequences of entry, establishment, or spread of a pathogenic agent within the territory of an importing country.

Sanitary measure
means any measure applied, such as those described in various chapters of the Terrestrial Code, designed to protect animal or human health or life within the territory of the Member Country from risks arising from the entry, establishment or spread of a hazard. Note: A detailed definition of sanitary measure may be found in the Agreement on the Application of Sanitary and Phytosanitary Measures of the World Trade Organization.

Surveillance
means the investigation of a given population or subpopulation to detect the presence of a pathogenic agent or disease; the frequency and type of surveillance will be determined by the epidemiology of the pathogenic agent or disease, and the desired outputs the systematic ongoing collection, collation, and analysis of data and the timely dissemination of information to those who need to know so that action can be taken.

Veterinary Services
means the governmental and non-governmental organisations that implement animal health and welfare measures and other standards and guidelines in the Terrestrial Code in the country. The Veterinary Services are under the overall control and direction of the Veterinary Authority. Private sector organisations or veterinarians are normally accredited or approved to deliver functions by the Veterinary Authority.
CHAPTER 1.2.1.

GENERAL OBLIGATIONS

Article 1.2.1.1.

Safety of international trade in animals and animal products depends on a combination of factors which should be taken into account to ensure unimpeded trade, without incurring unacceptable risks to human and animal health.

Because of differences between countries in their animal health situations, various options are offered by the Terrestrial Code. The animal health situation in the exporting country, in the transit country or countries and in the importing country should be considered before determining the requirements which have to be met for trade. To maximise harmonisation of the sanitary aspects of international trade, Veterinary Authorities of Members Countries should base their import requirements on the OIE standards, and guidelines and recommendations.

These requirements should be included in the model certificates approved by the OIE which form are included in Part 4 of the Terrestrial Code.

Certification requirements should be exact and concise, and should clearly convey the wishes of the importing country. For this purpose, prior consultation between Veterinary Authorities of importing and exporting countries is useful and may be necessary. It enables the setting out of the exact requirements so that the signing veterinarian can, if necessary, be given a note of guidance explaining the understanding between the Veterinary Authorities involved.

When Members officials of a Veterinary Authority wish to visit another country for matters of professional interest to the Veterinary Authority of the other country, the latter should be informed.

Article 1.2.1.2.

Responsibilities of the importing country

1. The import requirements included in the international veterinary certificate should assure that commodities introduced into the importing country comply with the national level of protection that it has chosen for animal and human health. Importing countries should restrict their requirements to those justified for such level of protection. If these are stricter than the OIE standards, they should be based on an import risk analysis.

2. The international veterinary certificate should not include requirements for the exclusion of pathogens or animal diseases which are present within the territory of in the importing country and are not subject to any official control programme. The requirements applying to pathogens or disease subject to official control programmes in a country or zone should not provide a higher level of protection on imports than that provided for the same pathogens or disease by the measures applied within that country or zone. The measures imposed on imports to manage the risks posed by a specific pathogen or disease should not require a higher level of protection than that provided by measures applied as part of the official control programme operating within the importing country.
3. The international veterinary certificate should not include requirements for disease agents, measures against pathogens, or diseases which are not OIE listed, unless the importing country has identified the disease agent as presenting a significant risk for that country, after conducting a scientifically based import risk analysis according to the guidelines in Section 1.3 demonstrated through import risk analysis carried out in accordance with Section 1.3, that the pathogen or disease poses a significant risk to the importing country.

4. The transmission by the Veterinary Authority of certificates or the communication of import requirements to persons other than the Veterinary Authority of another country, necessitates that copies of these documents are also sent to the Veterinary Authority. This important procedure avoids delays and difficulties which may arise between traders and Veterinary Authorities when the authenticity of the certificates or permits is not established.

This information is usually the responsibility of Veterinary Authorities. However, it can be issued by private sector veterinarians at the place of origin of the animals when this practice is the subject of appropriate approval and authentication by the Veterinary Authority.

Article 1.2.1.3.

Responsibilities of the exporting country

1. An exporting country should, on request, be prepared to supply the following information to importing countries on request:

   a) information on the animal health situation and national animal health information systems to determine whether that country is free or has free zones of listed diseases, including the regulations and procedures in force to maintain its free status;

   b) regular and prompt information on the occurrence of transmissible notifiable diseases;

   c) details of the country's ability to apply measures to control and prevent the relevant listed diseases;

   d) information on the structure of the Veterinary Services and the authority which they exercise;

   e) technical information, particularly on biological tests and vaccines applied in all or part of the national territory.

2. Veterinary Authorities of exporting countries should:

   a) have official procedures for authorisation of certifying veterinarians, defining their functions and duties as well as conditions covering possible suspension and termination of the appointment;

   b) ensure that the relevant instructions and training are provided to certifying veterinarians;

   c) monitor the activities of the certifying veterinarians to verify their integrity and impartiality.

3. The Head of the Veterinary Service of the exporting country is ultimately accountable for veterinary certification used in international trade.
Article 1.2.1.4.

Responsibilities in case of an incident occurring after related to importation

1. International trade involves a continuing ethical responsibility. Therefore, if within the recognised incubation periods of the various diseases subsequent to an export taking place, the Veterinary Authority becomes aware of the appearance or reappearance of a disease which has been specifically included in the international veterinary certificate, there is an obligation for the Administration to notify the importing country, so that the imported stock may be inspected or tested and appropriate action be taken to limit the spread of the disease should it have been inadvertently introduced.

2. Equally, if a disease condition appears in imported stock within a time period after importation consistent with the recognised incubation period of the disease, the Veterinary Authority of the exporting country should be informed so as to enable an investigation to be made, since this may be the first available information on the occurrence of the disease in a previously free herd. The Veterinary Authority of the importing country should be informed of the result of the investigation since the source of infection may not be in the exporting country.

3. In case of suspicion, on reasonable grounds, that an official certificate may be fraudulent, the Veterinary Authority of the importing country and exporting country should conduct an investigation. Consideration should also be given to notifying any third country(ies) that may have been implicated. All associated consignments should be kept under official control, pending the outcome of the investigation. The Veterinary Authorities of all countries involved should fully cooperate with the investigation. If the certificate is found to be fraudulent, every effort should be made to identify those responsible so that appropriate action can be taken according to the relevant legislation.
CHAPTER 1.3.2.

GUIDELINES FOR IMPORT RISK ANALYSIS

Article 1.3.2.1.

Introduction

An import risk analysis begins with a description of the commodity proposed for import and the likely annual quantity of trade. It must be recognised that whilst an accurate estimate of the anticipated quantity of trade is desirable to incorporate into the risk estimate, it may not be readily available, particularly where such trade is new.

Hazard identification is an essential step which must be conducted before the risk assessment.

The risk assessment process consists of four interrelated steps. These steps clarify the stages of the risk assessment, describing them in terms of the events necessary for the identified potential risk(s) to occur, and facilitate understanding and evaluation of the outputs. The product is the risk assessment report which is used in risk communication and risk management.

The relationships between risk assessment and risk management processes are outlined in Figure 1.

Fig 1. The relationship between risk assessment and risk management processes
Annex IV (contd)

Article 1.3.2.2.

Hazard identification

The hazard identification involves identifying the pathogenic agents which could potentially produce adverse consequences associated with the importation of a commodity.

The potential hazards identified would be those appropriate to the species being imported, or from which the commodity is derived, and which may be present in the exporting country. It is then necessary to identify whether each potential hazard is already present in the importing country, and whether it is a notifiable disease or is subject to control or eradication in that country and to ensure that import measures are not more trade restrictive than those applied within the country.

Hazard identification is a categorisation step, identifying biological agents dichotomously as potential hazards or not. The risk assessment may be concluded if hazard identification fails to identify potential hazards associated with the importation.

The evaluation of the Veterinary Services, surveillance and control programmes and zoning and compartmentalisation systems are important inputs for assessing the likelihood of hazards being present in the animal population of the exporting country.

An importing country may decide to permit the importation using the appropriate sanitary standards recommended in the Terrestrial Code, thus eliminating the need for a risk assessment.

Article 1.3.2.3.

Principles of risk assessment

1. Risk assessment should be flexible to deal with the complexity of real life situations. No single method is applicable in all cases. Risk assessment must be able to accommodate the variety of animal commodities, the multiple hazards that may be identified with an importation and the specificity of each disease, detection and surveillance systems, exposure scenarios and types and amounts of data and information.

2. Both qualitative risk assessment and quantitative risk assessment methods are valid.

3. The risk assessment should be based on the best available information that is in accord with current scientific thinking. The assessment should be well-documented and supported with references to the scientific literature and other sources, including expert opinion.

4. Consistency in risk assessment methods should be encouraged and transparency is essential in order to ensure fairness and rationality, consistency in decision making and ease of understanding by all the interested parties.

5. Risk assessments should document the uncertainties, the assumptions made, and the effect of these on the final risk estimate.

6. Risk increases with increasing volume of commodity imported.

7. The risk assessment should be amenable to updating when additional information becomes available.
Risk assessment steps

1. **Release assessment**

   Release assessment consists of describing the biological pathway(s) necessary for an importation activity to 'release' (that is, introduce) pathogenic agents into a particular environment, and estimating the probability of that complete process occurring, either qualitatively (in words) or quantitatively (as a numerical estimate). The release assessment describes the probability of the 'release' of each of the potential hazards (the pathogenic agents) under each specified set of conditions with respect to amounts and timing, and how these might change as a result of various actions, events or measures. Examples of the kind of inputs that may be required in the release assessment are:

   a) Biological factors
      - species, age and breed of animals
      - agent predilection sites
      - vaccination, testing, treatment and quarantine.

   b) Country factors
      - incidence/prevalence
      - evaluation of Veterinary Services, surveillance and control programmes and zoning systems of the exporting country.

   c) Commodity factors
      - quantity of commodity to be imported
      - ease of contamination
      - effect of processing
      - effect of storage and transport.

   If the release assessment demonstrates no significant risk, the risk assessment conclude.

2. **Exposure assessment**

   Exposure assessment consists of describing the biological pathway(s) necessary for exposure of animals and humans in the importing country to the hazards (in this case the pathogenic agents) released from a given risk source, and estimating the probability of the exposure(s) occurring, either qualitatively (in words) or quantitatively (as a numerical estimate).
The probability of exposure to the identified hazards is estimated for specified exposure conditions with respect to amounts, timing, frequency, duration of exposure, routes of exposure (e.g. ingestion, inhalation, or insect bite), and the number, species and other characteristics of the animal and human populations exposed. Examples of the kind of inputs that may be required in the exposure assessment are:

a) Biological factors
   - properties of the agent.

b) Country factors
   - presence of potential vectors
   - human and animal demographics
   - customs and cultural practices
   - geographical and environmental characteristics.

c) Commodity factors
   - quantity of commodity to be imported
   - intended use of the imported animals or products
   - disposal practices.

If the exposure assessment demonstrates no significant risk, the risk assessment may conclude at this step.

3. Consequence assessment

Consequence assessment consists of describing the relationship between specified exposures to a biological agent and the consequences of those exposures. A causal process must exist by which exposures produce adverse health or environmental consequences, which may in turn lead to socio-economic consequences. The consequence assessment describes the potential consequences of a given exposure and estimates the probability of them occurring. This estimate may be either qualitative (in words) or quantitative (a numerical estimate).

Examples of consequences include:

a) Direct consequences
   - animal infection, disease, and production losses
   - public health consequences.

b) Indirect consequences
   - surveillance and control costs
- compensation costs
- potential trade losses
- adverse consequences to the environment.

4. Risk estimation

Risk estimation consists of integrating the results from the release assessment, exposure assessment, and consequence assessment to produce overall measures of risks associated with the hazards identified at the outset. Thus risk estimation takes into account the whole of the risk pathway from hazard identified to unwanted outcome.

For a quantitative assessment, the final outputs may include:

- estimated numbers of herds, flocks, animals or people likely to experience health impacts of various degrees of severity over time;
- probability distributions, confidence intervals, and other means for expressing the uncertainties in these estimates;
- portrayal of the variance of all model inputs;
- a sensitivity analysis to rank the inputs as to their contribution to the variance of the risk estimation output;
- analysis of the dependence and correlation between model inputs.

Article 1.3.2.5.

Principles of risk management

1. Risk assessment is the process of deciding upon and implementing measures to achieve the Member Country's appropriate level of protection, whilst at the same time ensuring that negative effects on trade are minimised. The objective is to manage risk appropriately to ensure that a balance is achieved between a country's desire to minimise the likelihood or frequency of disease incursions and their consequences and its desire to import commodities and fulfil its obligations under international trade agreements.

2. The international standards of the OIE are the preferred choice of sanitary measures for risk management. The application of these sanitary measures should be in accordance with the intentions in the standards.

Article 1.3.2.6.

Risk management components

1. Risk evaluation - the process of comparing the risk estimated in the risk assessment with the Member Country's appropriate level of protection.
Annex IV (contd)

2. Option evaluation - the process of identifying, evaluating the efficacy and feasibility of, and selecting measures in order to reduce the risk associated with an importation in line with the Member Country's appropriate level of protection. The efficacy is the degree to which an option reduces the likelihood and/or magnitude of adverse health and economic consequences. Evaluating the efficacy of the options selected is an iterative process that involves their incorporation into the risk assessment and then comparing the resulting level of risk with that considered acceptable. The evaluation for feasibility normally focuses on technical, operational and economic factors affecting the implementation of the risk management options.

3. Implementation - the process of following through with the risk management decision and ensuring that the risk management measures are in place.

4. Monitoring and review - the ongoing process by which the risk management measures are continuously audited to ensure that they are achieving the results intended.

Article 1.3.2.7.

Principles of risk communication

1. Risk communication is the process by which information and opinions regarding hazards and risks are gathered from potentially affected and interested parties during a risk analysis, and by which the results of the risk assessment and proposed risk management measures are communicated to the decision-makers and interested parties in the importing and exporting countries. It is a multidimensional and iterative process and should ideally begin at the start of the risk analysis process and continue throughout.

2. A risk communication strategy should be put in place at the start of each risk analysis.

3. The communication of the risk should be an open, interactive, iterative and transparent exchange of information that may continue after the decision on importation.

4. The principal participants in risk communication include the authorities in the exporting country and other stakeholders such as domestic and foreign industry groups, domestic livestock producers and consumer groups.

5. The assumptions and uncertainty in the model, model inputs and the risk estimates of the risk assessment should be communicated.

6. Peer review is a component of risk communication in order to obtain scientific critique and to ensure that the data, information, methods and assumptions are the best available.
CHAPTER 1.4.1.

ANIMAL HEALTH MEASURES APPLICABLE BEFORE AND AT DEPARTURE

Article 1.4.1.1.

1. Countries should only authorise the exportation from their territory of animals for breeding, rearing or slaughter which are correctly identified and which meet the requirements of the importing country.

2. Biological tests and/or vaccinations required by the importing country should be carried out in accordance with the recommendations in the Terrestrial Code and Terrestrial Manual, as well as disinfection and disinfestation procedures.

3. Observation of the animals before leaving the country may be carried out either in the establishment where they were reared, or in a quarantine station. When they have been found to be clinically healthy and free from diseases listed by the OIE by an Official Veterinarian during the period of observation, the animals should be transported to the place of shipment in specially constructed vehicles, previously cleansed and disinfected. This must be done without delay and without the animals coming into contact with other susceptible animals, unless these animals have animal health guarantees similar to those of the transported animals.

4. The transportation of the animals for breeding or rearing or animals for slaughter from the establishment of origin to the point of departure from the exporting country shall be carried out in conformity with the conditions agreed between the importing country and exporting country.

Article 1.4.1.2.

Countries should only undertake the export from its territory of:

a) semen,

b) embryos/ova,

c) hatching eggs,

from artificial insemination centres, collection centres or farms which meet the requirements of the importing country.

Article 1.4.1.3.

Countries exporting animals, semen, embryos/ova or hatching eggs should inform the country of destination and where necessary the transit countries if, after exportation, a disease listed by the OIE occurs within the incubation period of that particular disease in the establishment of origin, or in an animal which was in a collecting centre where animals for breeding or rearing or animals for slaughter from different establishments or markets are collected together, or in a market, at the same time as the exported animals.
Annex IV (contd)

Article 1.4.1.4.

Before the departure of animals, semen, embryos/ova, hatching eggs and brood-combs of bees, an Official Veterinarian should, within the 24 hours prior to shipment, provide an international veterinary certificate conforming with the models approved by the OIE (as shown in Part 4 of the Terrestrial Code) and worded in the languages agreed upon between the exporting country and the importing country, and, where necessary, with the transit countries.

Article 1.4.1.5.

1. Before the departure of an animal or a consignment of animals on an international journey, the Veterinary Authority of the port, airport or district in which the border post is situated may, if it is considered necessary, carry out a clinical examination of the animal or consignment. The time and place of the examination shall be arranged taking into account customs and other formalities and in such a way as not to impede or delay departure.

2. The Veterinary Authority referred to in point 1 above shall take necessary measures to:

   a) prevent the shipment of animals affected or suspected of being affected with any disease listed by the OIE or with any other infectious disease;

   b) avoid entry into the vehicle of possible vectors or causal agents of infection.

Article 1.4.1.6.

1. Countries should only authorise the export from their territory of meat and products of animal origin intended for human consumption, which are fit for human consumption. They must be accompanied by an international veterinary certificate conforming with the models approved by the OIE (as shown in Part 4. of the Terrestrial Code). These must be worded in the languages agreed upon between the exporting country and the importing country, and, where necessary, with the transit countries.

2. Products of animal origin intended for use in animal feeding, or for pharmaceutical or surgical or agricultural or industrial use, should be accompanied by an international veterinary certificate conforming with the models approved by the OIE (as shown in Part 4. of the Terrestrial Code).
CHAPTER 1.4.3.

BORDER POSTS AND QUARANTINE STATIONS IN THE IMPORTING COUNTRY

Article 1.4.3.1.

1. Countries and their Veterinary Authorities shall, wherever possible, take the necessary action to ensure that the border posts and quarantine stations in their territory shall be provided with an adequate organisation and sufficient equipment for the application of the measures recommended in the Terrestrial Code.

2. Each border post and quarantine station shall be provided with facilities for the feeding and watering of animals.

Article 1.4.3.2.

When justified by the amount of international trade and by the epidemiological situation, border posts and quarantine stations shall be provided with a Veterinary Service comprising personnel, equipment and premises as the case may be and, in particular, means for:

a) making clinical examinations and obtaining specimens of material for diagnostic purposes from live animals or carcasses of animals affected or suspected of being affected by an epizootic disease, and obtaining specimens of animal products suspected of contamination;

b) detecting and isolating animals affected by or suspected of being affected by an epizootic disease;

c) carrying out disinfection and possibly disinfestation of vehicles used to transport animals and animal products.

In addition to this, each port and international airport should ideally be provided with equipment for the sterilisation or incineration of swill or any other material dangerous to animal health.

Article 1.4.3.3.

When required for the transit of commodities in international trade, airports shall be provided, as soon as possible, with areas of direct transit. These must, however, comply with the conditions required by Veterinary Authorities, especially to prevent the risk of introducing diseases transmitted by insects.

Article 1.4.3.4.

Each Veterinary Authority, when requested, shall make available for the Central Bureau and any interested country on request:

a) a list of border posts, quarantine stations, approved abattoirs and storage depots in its territory which are approved for international trade.
Annex IV (contd)

b) the period of time required for notice to be given for the application of the arrangements contained in point 2 of Articles 1.4.4.1. to 1.4.4.4.;

c) a list of airports in its territory which are provided with an area of direct transit, approved by the relevant Veterinary Authority and placed under its immediate control, where animals stay for a short time pending further transport to their final destination.
CHAPTER 1.3.5.

ZONING AND COMPARTMENTALISATION

Article 1.3.5.1.

Introduction

For the purposes of the Terrestrial Code, ‘zoning’ and ‘regionalisation’ have the same meaning.

Given the difficulty of establishing and maintaining a disease free status for an entire country, especially for diseases the entry of which is difficult to control through measures at national boundaries, there may be benefits to a Member in establishing and maintaining a subpopulation with a distinct health status within its territory. Subpopulations may be separated by natural or artificial geographical barriers or, in certain situations, by the application of appropriate management practices.

Zoning and compartmentalisation are procedures implemented by a country under the provisions of this chapter with a view to defining subpopulations of distinct health status within its territory for the purpose of disease control and/or international trade. While zoning applies to an animal subpopulation defined primarily on a geographical basis (using natural, artificial or legal boundaries), compartmentalisation applies to an animal subpopulation defined primarily by management and husbandry practices related to biosecurity. In practice, spatial considerations and good management including biosecurity plans play important roles in the application of both concepts.

A particular application of the concept of zoning is the establishment of a containment zone. In the event of a limited outbreak of a specified disease within an otherwise free country or zone, a single containment zone, which includes all cases, can be established for the purpose of minimizing the impact on the entire country or zone.

This chapter is to assist OIE Members wishing to establish and maintain different subpopulations within their territory using the principles of compartmentalisation and zoning. These principles should be applied in accordance with the measures recommended in the relevant disease chapter(s). This chapter also outlines a process through which trading partners may recognise such subpopulations. This process is best implemented by trading partners through establishing parameters and gaining agreement on the necessary measures prior to disease outbreaks.

Before trade in animals or their products may occur, an importing country needs to be satisfied that its animal health status will be appropriately protected. In most cases, the import regulations developed will rely in part on judgements made about the effectiveness of sanitary procedures undertaken by the exporting country, both at its borders and within its territory.

As well as contributing to the safety of international trade, zoning and compartmentalisation may assist disease control or eradication within Members’ countries. Zoning may encourage the more efficient use of resources within certain parts of a country and compartmentalisation may allow the functional separation of a subpopulation from other domestic or wild animals through biosecurity measures, which a zone (through geographical separation) would not achieve. Following a disease outbreak, the use of compartmentalization may allow a Member to take advantage of epidemiological links among subpopulations or common practices relating to biosecurity, despite diverse geographical locations, to facilitate disease control and/or the continuation of trade.
Annex V (contd)

Zoning and compartmentalisation cannot be applied to all diseases but separate requirements will be developed for each disease for which the application of zoning or compartmentalisation is considered appropriate.

To regain free status following a disease outbreak in a zone or compartment, Members should follow the recommendations in the relevant disease chapter in the Terrestrial Code.

Article 1.3.5.2.

**General considerations**

The Veterinary Services of an exporting country which is establishing a zone or compartment within its territory for international trade purposes should clearly define the subpopulation in accordance with the recommendations in the relevant chapters in the Terrestrial Code, including those on surveillance, and the identification and traceability of live animals. The Veterinary Services of an exporting country should be able to explain to the Veterinary Services of an importing country the basis for its claim of a distinct animal health status for the zone or compartment in such terms.

The procedures used to establish and maintain the distinct animal health status of a zone or compartment should be appropriate to the particular circumstances, and will depend on the epidemiology of the disease, environmental factors and applicable biosecurity measures.

The authority, organisation and infrastructure of the Veterinary Services, including laboratories, must be clearly documented in accordance with the chapter on the evaluation of Veterinary Services of the Terrestrial Code, to provide confidence in the integrity of the zone or compartment. The final authority of the zone or compartment, for the purposes of domestic and international trade, lies with the Veterinary Authority.

In the context of maintaining the animal health status of a population, references to ‘import’, ‘importation’ and ‘imported animals/products’ found in the Terrestrial Code apply both to importation into a country and to the movement of animals and their products into zones and compartments. Such movements should be the subject of appropriate measures to preserve the health status of the zone or compartment.

The exporting country should be able to demonstrate, through detailed documentation provided to the importing country, that it has implemented the recommendations in the Terrestrial Code for establishing and maintaining such a zone or compartment.

An importing country should recognise the existence of this zone or compartment when the appropriate measures recommended in the Terrestrial Code are applied and the Veterinary Authority of the exporting country certifies that this is the case.

The exporting country should conduct an assessment of the resources needed and available to establish and maintain a zone or compartment for international trade purposes. These include the human and financial resources, and the technical capability of the Veterinary Services (and of the relevant industry, in the case of a compartment) including disease surveillance and diagnosis.

Biosecurity and surveillance are essential components of zoning and compartmentalisation, and the arrangements should be developed through cooperation of industry and Veterinary Services.

Industry’s responsibilities include the application of biosecurity measures, documenting and recording movements of animals, quality assurance schemes, monitoring the efficacy of the measures, documenting corrective actions, conducting surveillance, rapid reporting and maintenance of records in a readily accessible form.
The Veterinary Services should provide movement certification, and carry out documented periodic inspections of facilities, biosecurity measures, records and surveillance procedures. Veterinary Services should conduct or audit surveillance, reporting and laboratory diagnostic examinations.

Article 1.3.5.3.

Principles for defining a zone or compartment, including containment zone

In conjunction with the above considerations, the following principles should apply when Members define a zone or a compartment.

1. The extent of a zone and its geographical limits should be established by the Veterinary Authority on the basis of natural, artificial and/or legal boundaries, and made public through official channels.

2. Establishment of a containment zone should be based on a rapid response including appropriate standstill of movement of animals and commodities upon notification of suspicion of the specified disease and the demonstration that the outbreak is contained within this zone through epidemiological investigation (trace-back, trace-forward) after confirmation of infection. The primary outbreak and likely source of the outbreak should be identified and all cases shown to be epidemiologically linked. For the effective establishment of a containment zone, it is necessary to demonstrate that there have been no new cases in the containment zone within a minimum of two incubation periods from the last detected case.

A stamping-out policy or another effective control strategy should be applied and the susceptible animal population within the containment zones should be clearly identifiable as belonging to the containment zone. Increased passive and targeted surveillance in accordance with Appendix 3.8.7. in the rest of the country or zone should be carried out and has not detected any evidence of infection. Measures to prevent spread of the infection from the containment zone to the rest of the country or zone including ongoing surveillance in the containment zone should be in place.

The free status of the areas outside the containment zone would be suspended pending the establishment of the containment zone. The suspension of free status of these areas could be lifted, once the containment zone is clearly established, irrespective of the provisions of the disease specific chapter.

The recovery of the free status of the containment zone should follow the provisions of the disease specific chapter.

3. The factors defining a compartment should be established by the Veterinary Authority on the basis of relevant criteria such as management and husbandry practices related to biosecurity, and made public through official channels.

34. Animals and herds belonging to such subpopulations need to be recognisable as such through a clear epidemiological separation from other animals and all things presenting a disease risk. For a zone or compartment, the Veterinary Authority should document in detail the measures taken to ensure the identification of the subpopulation and the establishment and maintenance of its animal health status through a biosecurity plan. The measures used to establish and maintain the distinct animal health status of a zone or compartment should be appropriate to the particular circumstances, and will depend on the epidemiology of the disease, environmental factors, the health status of animals in adjacent areas, applicable biosecurity measures (including movement controls, use of natural and artificial boundaries, the spatial separation of animals, and commercial management and husbandry practices), and surveillance.
Annex V (contd)

Annex V (contd)

45. Relevant animals within the zone or compartment should be identified in such a way that their history can be audited. Depending on the system of production, identification may be done at the herd, flock lot or individual animal level. Relevant animal movements into and out of the zone or compartment should be well documented, controlled and supervised. The existence of a valid animal identification system is a prerequisite to assess the integrity of the zone or compartment.

56. For a compartment, the biosecurity plan should describe the partnership between the relevant industry and the Veterinary Authority, and their respective responsibilities. It should also describe the routine operating procedures to provide clear evidence that the surveillance conducted, the live animal identification and traceability system, and the management practices are adequate to meet the definition of the compartment. In addition to information on animal movement controls, the plan should include herd or flock production records, feed sources, surveillance results, birth and death records, visitor logbook, morbidity and mortality history, medications, vaccinations, documentation of training of relevant personnel and any other criteria necessary for evaluation of risk mitigation. The information required may vary according to the species and disease(s) under consideration. The biosecurity plan should also describe how the measures will be audited to ensure that the risks are regularly re-assessed and the measures adjusted accordingly.

Article 1.3.5.4.

Sequence of steps to be taken in establishing a zone/compartment and having it recognised for international trade purposes

There is no single sequence of steps which should be followed in establishing a zone or a compartment. The steps that the Veterinary Services of the importing country and the exporting country choose and implement will generally depend on the circumstances existing within the countries and at their borders, and their trading history. The recommended steps are:

1. For zoning

   a) The exporting country identifies a geographical area within its territory, which it considers to contain an animal subpopulation with a distinct health status with respect to a specific disease/specific diseases, based on surveillance.

   b) The exporting country describes in the biosecurity plan for the zone the measures which are being, or will be, applied to distinguish such an area epidemiologically from other parts of its territory, in accordance with the recommendations in the Terrestrial Code.

   c) The exporting country provides:

      i) the above information to the importing country, with an explanation of why the area can be treated as an epidemiologically separate zone for international trade purposes;

      ii) access to enable the procedures or systems that establish the zone to be examined and evaluated upon request by the importing country.

   d) The importing country determines whether it accepts such an area as a zone for the importation of animals and animal products, taking into account:
i) an evaluation of the exporting country's veterinary Services;

ii) the result of a risk assessment based on the information provided by the exporting country and its own research;

iii) its own animal health situation with respect to the disease(s) concerned; and

iv) other relevant OIE standards.

e) The importing country notifies the exporting country of its determination and the underlying reasons, within a reasonable period of time, being:

i) recognition of the zone; or

ii) request for further information; or

iii) rejection of the area as a zone for international trade purposes.

f) An attempt should be made to resolve any differences over recognition of the zone, either in the interim or finally, by using an agreed mechanism to reach consensus such as the OIE in-house procedure for settlement of disputes (Article 1.3.1.3.)

g) The Veterinary Authorities of the importing and exporting countries should enter into a formal agreement recognizing the zone.

2. For compartmentalisation

a) Based on discussions with the relevant industry, the exporting country identifies within its territory a compartment of one or more establishments or other premises which operates under common management practices related to biosecurity and which contains an identifiable animal subpopulation with a distinct health status with respect to a specific disease/specific diseases; the exporting country describes how this status is maintained through a partnership between the relevant industry and the Veterinary Authority of the exporting country.

b) The exporting country examines the compartment’s biosecurity plan and confirms through an audit that:

i) the compartment is epidemiologically closed throughout its routine operating procedures as a result of effective implementation of its biosecurity plan; and

ii) the surveillance and monitoring programme in place is appropriate to verify the status of such establishment(s) with respect to such disease(s).

c) The exporting country describes the compartment, in accordance with the recommendations in the Terrestrial Code.
Annex V (contd)

d) The exporting country provides:

i) the above information to the importing country, with an explanation of why such an establishment(s) can be treated as an epidemiologically separate compartment for international trade purposes; and

ii) access to enable the procedures or systems that establish the compartment to be examined and evaluated upon request by the importing country.

e) The importing country determines whether it accepts such establishment(s) as a compartment for the importation of animals and animal products, taking into account:

i) an evaluation of the exporting country’s Veterinary Services;

ii) the result of a risk assessment based on the information provided by the exporting country and its own research;

iii) its own animal health situation with respect to the disease(s) concerned; and

iv) other relevant OIE standards.

f) The importing country notifies the exporting country of its determination and the underlying reasons, within a reasonable period of time, being:

i) recognition of the compartment; or

ii) request for further information; or

iii) rejection of such establishment(s) as a compartment for international trade purposes.

g) An attempt should be made to resolve any differences over recognition of the compartment, either in the interim or finally, by using an agreed mechanism to reach consensus such as the OIE in-house procedure for settlement of disputes (Article 1.3.1.3.).

h) The Veterinary Authorities of the importing and exporting countries should enter into a formal agreement recognizing the compartment.

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APPENDIX 3.x.x.

GENERAL GUIDELINES FOR THE APPLICATION OF COMPARTMENTALISATION

Article 3.x.x.1.

Introduction and objectives

The guidelines in this appendix provide a structured framework for the application and recognition of compartments within countries or zones, based on the provisions of Chapter 1.3.5. with the objective to facilitate trade in animals and products of animal origin and as a tool for disease management.

Establishing and maintaining a disease-free status for an entire country may be difficult, especially in the case of diseases that can easily cross international boundaries. For many diseases, OIE Member Countries have traditionally applied the concept of zoning to establish and maintain an animal subpopulation with a different animal health status within national boundaries.

Chapter 1.1.1. defines a compartment as “one or more establishments under a common biosecurity management system containing an animal subpopulation with a distinct health status with respect to a specific disease or specific diseases for which required surveillance, control and biosecurity measures have been applied for the purpose of international trade.”

The essential difference between zoning and compartmentalisation is that the recognition of zones is based on geographical boundaries whereas the recognition of compartments is based on management practices and biosecurity. However, spatial considerations and good management practices play a role in the application of both concepts.

Compartmentalisation is not a new concept for Veterinary Services; in fact, it has been applied for a long time in many disease control programmes that are based on the concept of disease-free herds/flocks.

The fundamental requirement for compartmentalisation is the implementation of management and biosecurity measures to create a functional separation of establishments and allows the Veterinary Services to make a clear epidemiological differentiation to be made between subpopulations of differing health status.

For example, a confinement operation for poultry or swine operation in an infected country or zone might have biosecurity measures and management practices that result in negligible risk from diseases or agents. The concept of a compartment extends the application of a ‘risk boundary’ beyond that of a geographical interface and considers all epidemiological factors that can help to create an effective disease-specific separation between subpopulations.

In disease-free countries or zones, compartments preferably should be defined prior to the occurrence of a disease outbreak. In the event of an outbreak or in endemic infected countries or zones, compartmentalisation may be used to facilitate trade.

For the purpose of international trade, compartments must be under the direct control and responsibility of the Veterinary Administration Authority in the country. For the purposes of this appendix, compliance by the Members with Chapters 1.1.2. and 1.3.3. are an essential prerequisite.
Principles for defining a compartment

A compartment may be established with respect of a specific disease or diseases. A compartment must be clearly defined, indicating the location of all its components including establishments, as well as related functional units (such as feed mills, slaughterhouses, rendering plants, etc.), their interrelationships and their contribution to an epidemiological separation between the animals in a compartment and subpopulations with a different health status. The definition of compartment may revolve around disease specific epidemiological factors, animal production systems, biosecurity practices and similar functional demarcations.

Separation of a compartment from potential sources of infection

The management of a compartment must provide to the veterinary administration documented evidence on the following:

a) Physical or spatial factors that affect the status of biosecurity in a compartment

While a compartment is primarily based on management and biosecurity measures, a review of geographical factors is needed to ensure that the functional boundary provides adequate separation of a compartment from adjacent animal populations with a different health status. The following factors should be taken into consideration in conjunction with biosecurity measures and, in some instances, may alter the degree of confidence achieved by general biosecurity and surveillance measures:

i) disease status in adjacent areas and in areas epidemiologically linked to the compartment;

ii) location, disease status and biosecurity of the nearest epidemiological units or other epidemiologically relevant premises. Consideration should be given to the distance and physical separation from:

- flocks or herds with a different health status in close proximity to the compartment, including wildlife and migratory routes;
- slaughterhouses, rendering plants or feed mills;
- markets, fairs, agricultural shows, sporting events, zoos, circuses and other points of animal concentration.

b) Infrastructural factors

Structural aspects of the establishments within a compartment contribute to the effectiveness of its biosecurity. Consideration should be given to:

i) fencing or other effective means of physical separation;

ii) facilities for people entry including access control, changing area and showers;

iii) vehicle access including washing and disinfection procedures;
iv) unloading and loading facilities;

v) isolation facilities for introduced animals;

vi) infrastructure to store feed and veterinary products;

vii) disposal of carcasses, manure and waste;

viii) water supply;

ix) physical measures to prevent exposure to living mechanical or biological vectors such as insects, rodents and wild birds;

datail; x) air supply;

xi) feed supply/source.

More detailed recommendations for certain establishments can be found in Sections 3.2., 3.3. and 3.4. of the Terrestrial Code.

c) Biosecurity plan

The integrity of the compartment relies on effective biosecurity. The management of the compartment should develop, implement and monitor a comprehensive biosecurity plan.

The biosecurity plan should describe in detail:

i) potential pathways for introduction and spread into the compartment of the agents for which the compartment was defined, including animal movements, rodents, fauna, aerosols, arthropods, vehicles, people, biological products, equipment, fomites, feed, waterways, drainage or other means. Consideration should also be given to the survivability of the agent in the environment;

ii) the critical control points for each pathway;

iii) measures to mitigate exposure for each critical control point;

iv) standard operating procedures including:

   - implementation, maintenance, monitoring of the measures;

   - application of corrective actions;

   - verification of the process;

   - record keeping;

v) contingency plan in the event of a change in the level of exposure;

vi) reporting procedures to the Veterinary Administration Authority;

vii) the programme for educating and training workers to ensure that all persons involved are knowledgeable and informed on biosecurity principles and practices;

viii) the surveillance programme in place.
In any case, sufficient evidence should be submitted to assess the efficacy of the biosecurity plan in accordance with the level of risk for each identified pathway. The biosecurity risk of all operations of the compartment should be regularly re-assessed. Based on the outcome of the assessment, concrete and documented mitigation steps should be taken to reduce the likelihood of introduction of the disease agent into the compartment.

d) **Traceability system**

A prerequisite for assessing the integrity of a compartment is the existence of a valid traceability system. All animals within a compartment should be individually identified and registered in such a way that their history and movements can be documented and audited. In cases where individual identification may not be feasible, such as broilers and day-old chicks, the Veterinary Administration Authority should provide sufficient assurance of traceability.

All animal movements into and out of the compartment should be certified by the Veterinary Administration Authority and recorded at the compartment level. Movements within the compartment need not be certified but should be recorded at the compartment level.

**Article 3.x.x.4.**

**Documentation of factors critical to the definition of a compartment**

Documentation must provide clear evidence that the biosecurity, surveillance, traceability and management practices defined for a compartment are effectively and consistently applied. In addition to animal movement information, the necessary documentation should include herd or flock production records, feed sources, laboratory tests, birth and death records, the visitor logbook, morbidity history, medication and vaccination records, biosecurity plans, training documentation and any other criteria necessary for the evaluation of disease exclusion.

The historical status of a compartment for the disease(s) for which it was defined should be documented and demonstrate compliance with the requirements for freedom in the relevant Terrestrial Code chapter.

In addition, a compartment seeking recognition should submit to the Veterinary Administration Authority a baseline animal health report indicating the presence or absence of OIE listed diseases. This report should be regularly updated to reflect the current animal health situation of the compartment.

Vaccination records including the type of vaccine and frequency of administration must be available to enable interpretation of surveillance data.

The time period for which all records should be kept may vary according to the species and disease(s) for which the compartment was defined.

All relevant information must be recorded in a transparent manner and be easily accessible so as to be auditable by the Veterinary Administration Authority.

**Article 3.x.x.5.**

**Surveillance for the agent or disease**

The surveillance system should comply with Appendix 3.8.1. on General Guidelines for Surveillance and the specific guidelines for surveillance for the disease(s) for which the compartment was defined, if available.
If there is an increased risk of exposure to the agent for which the compartment has been defined, the detection level of the internal surveillance and the level of biosecurity should be raised.

a) Internal surveillance

Surveillance should involve the collection and analysis of disease/infection data such so that the Veterinary Authority can certify that the animals in all the establishments comply with the defined status of that compartment. A surveillance system that is able to ensure early detection in the event that the agent enters an establishment is essential. Depending on the disease(s) for which the compartment was defined, different surveillance strategies may be applied to achieve the desired confidence in disease freedom.

b) External surveillance

The biosecurity measures applied in a compartment must be appropriate to the level of exposure of the compartment. External surveillance will help identify a significant change in the level of exposure for the identified pathways for disease introduction into the compartment.

An appropriate combination of active and passive surveillance is necessary to achieve the goals described above. Based on the recommendations of Appendix 3.8.1., targeted surveillance based on an assessment of risk factors may be the most efficient surveillance approach. Targeted surveillance should in particular include epidemiological units in close proximity to the compartment or those that have a potential epidemiological link with it.

Article 3.x.x.6.

Diagnostic capabilities and procedures

Officially-designated laboratory facilities complying with the OIE standards for quality assurance, as defined in Chapter I.1.2. of the Terrestrial Manual, should be available for sample testing. All laboratory tests and procedures should comply with the recommendations of the Terrestrial Manual for the specific disease. Each laboratory that conducts testing should have systematic procedures in place for rapid reporting of disease results to the Veterinary Administration. Where appropriate, results should be confirmed by an OIE Reference Laboratory.

Article 3.x.x.7.

Emergency response and notification

Early detection, diagnosis and notification of disease are critical to minimise the consequences of outbreaks.

In case of a suspicion or an occurrence of any OIE listed disease not present according to the baseline animal health report referred to in Article 3.x.x.4., the management of the compartment should notify the Veterinary Administration, as this may indicate a breach in the biosecurity measures. The Veterinary Administration should immediately suspend export certification and should notify the importing countries to reassess the biosecurity of the compartment and if a significant breach is detected, export certification should be suspended. Trade may only be resumed after the compartment has adopted the necessary measures to re-establish the biosecurity level and the Veterinary Administration re-approves the compartment for trade.
In the event of suspicion of occurrence of the disease for which the compartment was defined, export certification should be immediately suspended. If confirmed, the status of the compartment should be immediately revoked and importing countries should be notified following the provisions of Chapter 1.1.2.

Positive findings of the disease(s) for which the compartment has been defined, should be immediately notified following the provisions of Chapter 1.1.2.

Article 3.x.x.8.

Supervision and control of a compartment

The authority, organisation, and infrastructure of the Veterinary Services, including laboratories, must be clearly documented in accordance with the chapter on the evaluation of Veterinary Services of the OIE Terrestrial Code, to provide confidence in the integrity of the compartment.

The Veterinary Administration Authority has the final authority in granting, suspending and revoking the status of a compartment. The Veterinary Administration Authority should continuously supervise compliance with all the requirements critical to the maintenance of the compartment status described in this appendix and ensure that all the information is readily accessible to the importing countries. Any significant change should be notified to the importing country.
CHAPTER 2.2.5.

RABIES

Article 2.2.5.1.
For the purposes of the Terrestrial Code, the incubation period for rabies shall be 6 months, and the infective period in domestic carnivores starts 15 days before the onset of the first clinical signs and ends when the animal dies.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.2.5.2.

Rabies free country

A country may be considered free from rabies when:
1. the disease is notifiable;
2. an effective system of disease surveillance is in operation;
3. all regulatory measures for the prevention and control of rabies have been implemented including effective importation procedures;
4. no case of indigenously acquired rabies infection has been confirmed in man or any animal species during the past 2 years; however, this status would not be affected by the isolation of an Australian or European Bat Lyssavirus (EBL1 or EBL2);
5. no imported case in carnivores has been confirmed outside a quarantine station for the past 6 months.

Article 2.2.5.3.

When importing from rabies free countries, Veterinary Authorities should require:

for domestic mammals, and wild mammals reared under confined conditions
the presentation of an international veterinary certificate attesting that the animals:
1. showed no clinical sign of rabies on the day of shipment;
2. were kept since birth or for the 6 months prior to shipment in a rabies free country or were imported in conformity with the regulations stipulated in Articles 2.2.5.5., 2.2.5.6. or 2.2.5.7.

Article 2.2.5.4.

When importing from rabies free countries, Veterinary Authorities should require:

for wild mammals not reared under confined conditions
the presentation of an international veterinary certificate attesting that the animals:
Annex VI (contd)

1. showed no clinical sign of rabies on the day of shipment;
2. have been captured in a rabies free country, at a sufficient distance from any infected country. The distance should be defined according to the species exported and the reservoir species in the infected country.

**Article 2.2.5.5.**

When importing from countries considered infected with rabies, Veterinary Authorities should require:

for dogs and cats

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of rabies within 48 hours of shipment;

**AND EITHER**

2. were vaccinated against rabies:
   a) not less than 6 months and not more than one year prior to shipment in the case of a primary vaccination, which should have been carried out when the animals were at least 3 months old;
   b) not more than one year prior to shipment in the case of a booster vaccination;
   c) with an inactivated virus vaccine or with a recombinant vaccine expressing the rabies virus glycoprotein and were identified by a permanent mark (including a microchip) before the vaccination (their identification number shall be stated in the certificate);

3. were subjected not less than 3 months and not more than 24 months prior to shipment to an antibody test as prescribed in the Terrestrial Manual with a positive result equivalent to at least 0.5 IU/ml;

**OR**

4. have not been vaccinated against rabies or do not meet all the conditions set out in points 1, 2 and 3 above; in such cases, the importing country may require the placing of the animals in a quarantine station located on its territory, in conformity with the conditions stipulated in its animal health legislation.

**Article 2.2.5.6.**

When importing from countries considered infected with rabies, Veterinary Authorities should require:

for domestic ruminants, equines and pigs

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of rabies on the day of shipment;
2. was maintained and where no case of rabies was reported for at least 12 months prior to shipment.

**Article 2.2.5.7.**

When importing from countries considered infected with rabies, Veterinary Authorities should require:
for laboratory reared rodents and lagomorphs, and lagomorphs or wild mammals (other than non-human primates) reared under confined conditions

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of rabies on the day of shipment;

2. were kept since birth, or for the 12 months prior to shipment, in an establishment where no case of rabies was reported for at least 12 months prior to shipment.

Article 2.2.5.8.

When importing from countries considered infected with rabies, Veterinary Authorities should require:

for wild mammals not belonging to the orders of primates or carnivores and not reared under confined conditions

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of rabies on the day of shipment;

2. were kept in a quarantine station for the 6 months prior to shipment.

Article 2.2.5.9.

When importing from countries considered infected with rabies, Veterinary Authorities should require:

for frozen semen of dogs

the presentation of an international veterinary certificate attesting that the donor animals showed no clinical sign of rabies during the 15 days following collection of the semen.
CHAPTER 2.2.10.

FOOT AND MOUTH DISEASE

Article 2.2.10.1.

For the purposes of this Terrestrial Code, the incubation period for foot and mouth disease (FMD) shall be 14 days.

For the purposes of this Chapter, ruminants include animals of the family of Camelidae.

For the purposes of this Chapter, a case includes an animal infected with FMD virus (FMDV).

For the purposes of international trade, this Chapter deals not only with the occurrence of clinical signs caused by FMDV, but also with the presence of infection with FMDV in the absence of clinical signs.

The following defines the occurrence of FMDV infection:

1. FMDV has been isolated and identified as such from an animal or a product derived from that animal; or
2. viral antigen or viral ribonucleic acid (RNA) specific to one or more of the serotypes of FMDV has been identified in samples from one or more animals, whether showing clinical signs consistent with FMD or not, or epidemiologically linked to a confirmed or suspected outbreak of FMD, or giving cause for suspicion of previous association or contact with FMDV; or
3. antibodies to structural or nonstructural proteins of FMDV that are not a consequence of vaccination, have been identified in one or more animals showing clinical signs consistent with FMD, or epidemiologically linked to a confirmed or suspected outbreak of FMD, or giving cause for suspicion of previous association or contact with FMDV.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.2.10.2.

FMD free country where vaccination is not practised

Susceptible animals in the FMD free country should be separated from neighbouring infected countries by a buffer zone, or physical or geographical barriers, and animal health measures that effectively prevent the entry of the virus should be implemented.

To qualify for inclusion in the existing list of FMD free countries where vaccination is not practised, a country should:

1. have a record of regular and prompt animal disease reporting;
2. send a declaration to the OIE stating that:
   a) there has been no outbreak of FMD during the past 12 months;
   b) no evidence of FMDV infection has been found during the past 12 months;
Appendix VII (contd)

c) no vaccination against FMD has been carried out during the past 12 months;

d) no vaccinated animal has been introduced since the cessation of vaccination;

3. supply documented evidence that:

a) surveillance for both FMD and FMDV infection in accordance with Appendix 3.8.7. is in operation;

b) regulatory measures for the prevention and control of FMD have been implemented.

The country will be included in the list only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information in points 2 and 3a) above be re-submitted annually and changes in the epidemiological situation or other significant events should be reported promptly to the OIE.

Article 2.2.10.3.

FMD free country where vaccination is practised

Susceptible animals in the FMD free country where vaccination is practised should be separated from neighbouring infected countries by a buffer zone or by physical/geographical barriers, and animal health measures that effectively prevent the entry of the virus should be implemented.

To qualify for inclusion in the list of FMD free countries where vaccination is practised, a country should:

1. have a record of regular and prompt animal disease reporting;

2. send a declaration to the OIE that there has been no outbreak of FMD for the past 2 years and no evidence of FMDV circulation for the past 12 months, with documented evidence that:

a) surveillance for FMD and FMDV circulation in accordance with Appendix 3.8.7. is in operation and that regulatory measures for the prevention and control of FMD have been implemented;

b) routine vaccination is carried out for the purpose of the prevention of FMD;

c) the vaccine used complies with the standards described in the Terrestrial Manual.

The country will be included in the list only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information in point 2 above be re-submitted annually and changes in the epidemiological situation or other significant events should be reported promptly to the OIE.

If an FMD free country where vaccination is practised wishes to change its status to FMD free country where vaccination is not practised, the country should wait for 12 months after vaccination has ceased then notify the OIE and provide evidence showing that FMDV circulation has not occurred during that period.
FMD free zone where vaccination is not practised

An FMD free zone where vaccination is not practised can be established in either an FMD free country where vaccination is practised or in a country of which parts are infected. In defining such zones the principles of Chapter 1.3.5. should be followed. Susceptible animals in the FMD free zone should be separated by a buffer zone or by physical/geographical barriers from the rest of the country and from neighbouring countries if they are of a different health status, and animal health measures that effectively prevent the entry of the virus should be implemented.

A country in which an FMD free zone where vaccination is not practised is to be established should:

1. have a record of regular and prompt animal disease reporting;

2. send a declaration to the OIE stating that it wishes to establish an FMD free zone where vaccination is not practised and that within the proposed FMD free zone:
   a) there has been no outbreak of FMD during the past 12 months;
   b) no evidence of FMDV infection has been found during the past 12 months;
   c) no vaccination against FMD has been carried out during the past 12 months;
   d) no vaccinated animal has been introduced since the cessation of vaccination, except in accordance with Article 2.2.10.9.;
   e) documented evidence shows that surveillance in accordance with Appendix 3.8.7. is in operation for both FMD and FMDV infection;

3. describe in detail:
   a) regulatory measures for the prevention and control of both FMD and FMDV infection,
   b) the boundaries of the proposed FMD free zone and, if applicable, the buffer zone or physical or geographical barriers,
   c) the system for preventing the entry of the virus (including the control of the movement of susceptible animals) into the proposed FMDV free zone (in particular if the procedure described in Article 2.2.10.9. is implemented),

and supply documented evidence that these are properly implemented and supervised.

The proposed free zone will be included in the list of FMD free zones where vaccination is not practiced only after the submitted evidence has been accepted by the OIE.

The information required in points 2 and 3c) above should be re-submitted annually as well as any relevant changes in the epidemiological situation or other significant events including those relevant to under points 3a) and 3b) should be reported promptly to the OIE.
Appendix VII (contd)

Article 2.2.10.5.

FMD free zone where vaccination is practised

An FMD free zone where vaccination is practised can be established in either an FMD free country where vaccination is not practised or in a country of which parts are infected. In defining such zones the principles of Chapter 1.3.5. should be followed. Susceptible animals in the FMD free zone where vaccination is practised should be separated by a buffer zone or by physical/geographical barriers from the rest of the country and from neighbouring countries if they are of a different health status, and animal health measures that effectively prevent the entry of the virus should be implemented.

A country in which an FMD free zone where vaccination is practised is to be established should:

1. have a record of regular and prompt animal disease reporting;

2. send a declaration to the OIE that it wishes to establish an FMD free zone where vaccination is practised and that within the proposed FMD free zone:
   a) there has been no outbreak of FMD for the past 2 years;
   b) no evidence of FMDV circulation for the past 12 months;
   c) documented evidence shows that surveillance in accordance with Appendix 3.8.7. is in operation for FMD and FMDV circulation;

3. supply documented evidence that the vaccine used complies with the standards described in the Terrestrial Manual;

4. describe in detail:
   a) regulatory measures for the prevention and control of both FMD and FMDV circulation,
   b) the boundaries of the proposed FMD free zone where vaccination is practised and, if applicable, the buffer zone or physical or geographical barriers,
   c) the system for preventing the entry of the virus into the proposed FMD free zone (in particular if the procedure described in Article 2.2.10.9. is implemented),

and supply evidence that these are properly implemented and supervised.

The proposed free zone will be included in the list of FMD free zones where vaccination is practised only after the submitted evidence has been accepted by the OIE. The information required in points 2, 3 and 4c) above should be re-submitted annually as well as any relevant changes in the epidemiological situation or other significant events including those relevant to under points 4a) and 4b) should be reported promptly to the OIE.

If a country that has an FMD free zone where vaccination is practised wishes to change the status of the zone to FMD free zone where vaccination is not practised, a waiting period of 12 months after vaccination has ceased is required and evidence must be provided showing that FMDV infection has not occurred in the said zone during that period.
Article 2.2.10.6.

FMD infected country or zone

An FMD infected country is a country that does not fulfil the requirements to qualify as either an FMD free country where vaccination is not practised or an FMD free country where vaccination is practised.

An FMD infected zone is a zone that does not fulfil the requirements to qualify as either an FMD free zone where vaccination is not practised or an FMD free zone where vaccination is practised.

Article 2.2.10.7.

Establishment of a containment zone within an FMD free country or zone

In the event of a limited outbreak within an FMD free country or zone with or without vaccination, a single containment zone, which includes all cases, can be established for the purpose of minimizing the impact on the entire country or zone. For this to be achieved, the Veterinary Authority should provide documented evidence that:

1. the outbreak is limited based on the following factors:
   a) immediately on suspicion, a rapid response including notification has been made;
   b) standstill of animal movements has been imposed, and effective controls on the movement of other commodities mentioned in this chapter are in place;
   c) epidemiological investigation (trace-back, trace-forward) has been completed;
   d) the infection has been confirmed;
   e) the primary and likely source of the outbreak has been identified;
   f) all cases have been shown to be epidemiologically linked;
   g) no new cases have been found in the containment zone within a minimum of two incubation periods as defined in Article 2.2.10.1. from the last detected case;

2. surveillance in accordance with Appendix 3.8.7. demonstrates that there are no undetected cases in the containment zone;

3. a stamping-out policy or another effective control strategy has been applied;

4. the susceptible animal population within the containment zones should be clearly identifiable as belonging to the containment zone;

5. increased passive and targeted surveillance in accordance with Appendix 3.8.7. in the rest of the country or zone has been carried out and has not detected any evidence of infection;

6. measures to prevent spread of the infection from the containment zone to the rest of the country or zone, including ongoing surveillance in the containment zone, are in place.

The free status of the areas outside the containment zone would be suspended pending the establishment of the containment zone. The suspension of free status of these areas could be lifted irrespective of the provisions of Article 2.2.10.8., once the containment zone is clearly established, by complying with points 1 to 5 above.
Annex VII (contd)

The recovery of the FMD free status of the containment zone should follow the provisions of Article 2.2.10.8.

Article 2.2.10.8.

Recovery of free status

1. When an FMD outbreak or FMDV infection occurs in an FMD free country or zone where vaccination is not practised, one of the following waiting periods is required to regain the status of FMD free country or zone where vaccination is not practised:

   a) 3 months after the last case where a stamping-out policy and serological surveillance are applied in accordance with Appendix 3.8.7.; or

   b) 3 months after the slaughter of all vaccinated animals where a stamping-out policy, emergency vaccination and serological surveillance are applied in accordance with Appendix 3.8.7.; or

   c) 6 months after the last case or the last vaccination (according to the event that occurs the latest), where a stamping-out policy, emergency vaccination not followed by the slaughtering of all vaccinated animals, and serological surveillance are applied in accordance with Appendix 3.8.7., provided that a serological survey based on the detection of antibodies to nonstructural proteins of FMDV demonstrates the absence of infection in the remaining vaccinated population.

   Where a stamping-out policy is not practised, the above waiting periods do not apply but either Article 2.2.10.2. or Article 2.2.10.4. applies.

2. When an FMD outbreak or FMDV infection occurs in an FMD free country or zone where vaccination is practised, one of the following waiting periods is required to regain the status of FMD free country or zone where vaccination is practised:

   a) 6 months after the last case where a stamping-out policy, emergency vaccination and serological surveillance in accordance with Appendix 3.8.7. are applied, provided that the serological surveillance based on the detection of antibodies to nonstructural proteins of FMDV demonstrates the absence of virus circulation; or

   b) 18 months after the last case where a stamping-out policy is not applied, but emergency vaccination and serological surveillance in accordance with Appendix 3.8.7. are applied, provided that the serological surveillance based on the detection of antibodies to nonstructural proteins of FMDV demonstrates the absence of virus circulation.

Article 2.2.10.9.

Transfer directly to slaughter of FMD susceptible animals from an infected zone to a free zone within a country

FMD susceptible animals should only leave the infected zone if moved by mechanised transport to the nearest designated abattoir located in the buffer zone directly to slaughter.

In the absence of an abattoir in the buffer zone, live FMD susceptible animals can be transported to the nearest abattoir in a free zone directly to slaughter only under the following conditions:

1. no FMD susceptible animal has been introduced into the establishment of origin and no animal in the establishment of origin has shown clinical signs of FMD for at least 30 days prior to movement;
2. the animals were kept in the establishment of origin for at least 3 months prior to movement;

3. FMD has not occurred within a 10-kilometre radius of the establishment of origin for at least 3 months prior to movement;

4. the animals must be transported under the supervision of the Veterinary Authority in a vehicle, which was cleansed and disinfected before loading, directly from the establishment of origin to the abattoir without coming into contact with other susceptible animals;

5. such an abattoir is not approved for the export of fresh meat during the time it is handling the meat of animals from the infected zone;

6. vehicles and the abattoir must be subjected to thorough cleansing and disinfection immediately after use.

All products obtained from the animals and any products coming into contact with them must be considered infected and treated in such a way as to destroy any residual virus in accordance with Appendix 3.6.2.

Animals moved into a free zone for other purposes must be moved under the supervision of the Veterinary Authority and comply with the conditions in Article 2.2.10.12.

Article 2.2.10.10.

When importing from FMD free countries where vaccination is not practised or FMD free zones where vaccination is not practised, Veterinary Authorities should require:

for FMD susceptible animals

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of FMD on the day of shipment;

2. were kept in an FMD free country or zone where vaccination is not practised since birth or for at least the past 3 months;

3. have not been vaccinated.

Article 2.2.10.11.

When importing from FMD free countries where vaccination is practised or from FMD free zones where vaccination is practised, Veterinary Authorities should require:

for domestic ruminants and pigs

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of FMD on the day of shipment;

2. were kept in an FMD free country or zone since birth or for at least the past 3 months; and

3. have not been vaccinated and were subjected, with negative results, to tests for antibodies against FMD virus, when destined to an FMD free country or zone where vaccination is not practised.
Article 2.2.10.12.

When importing from FMD infected countries or zones, Veterinary Authorities should require:

for domestic ruminants and pigs

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of FMD on the day of shipment;

2. were kept in the establishment of origin since birth, or
   a) for the past 30 days if a stamping-out policy is in force in the exporting country, or
   b) for the past 3 months if a stamping-out policy is not in force in the exporting country, and that FMD has not occurred within a ten-kilometre radius of the establishment of origin for the relevant period as defined in points a) and b) above; and

3. were isolated in an establishment for the 30 days prior to shipment, that all animals in isolation were subjected to diagnostic tests (probang and serology) for evidence of FMDV infection with negative results at the end of that period, and that FMD did not occur within a ten-kilometre radius of the establishment during that period; or

4. were kept in a quarantine station for the 30 days prior to shipment, that all animals in quarantine were subjected to diagnostic tests (probang and serology) for evidence of FMDV infection with negative results at the end of that period, and that FMD did not occur within a ten-kilometre radius of the quarantine station during that period;

5. were not exposed to any source of FMD infection during their transportation from the quarantine station to the place of shipment.

Article 2.2.10.13.

When importing from FMD free countries where vaccination is not practised or FMD free zones where vaccination is not practised, Veterinary Authorities should require:

for fresh semen of domestic ruminants and pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen;
   b) were kept in an FMD free country or zone where vaccination is not practised for at least 3 months prior to collection;

2. the semen was collected, processed and stored in conformity with the provisions of Appendix 3.2.1. or Appendix 3.2.2., as relevant.

Article 2.2.10.14.

When importing from FMD free countries where vaccination is not practised or FMD free zones where vaccination is not practised, Veterinary Authorities should require:
for frozen semen of domestic ruminants and pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen and for the following 30 days;
   b) were kept in an FMD free country or zone where vaccination is not practised for at least 3 months prior to collection;

2. the semen was collected, processed and stored in conformity with the provisions of Appendix 3.2.1. or Appendix 3.2.2., as relevant.

Article 2.2.10.15.

When importing from FMD free countries where vaccination is practised or from FMD free zones where vaccination is practised, Veterinary Authorities should require:

for semen of domestic ruminants and pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen and for the following 30 days;
   b) were kept in a country or zone free from FMD for at least 3 months prior to collection;
   c) if destined to an FMD free country or zone where vaccination is not practised:
      i) have not been vaccinated and were subjected, not less than 21 days after collection of the semen, to tests for antibodies against FMD virus, with negative results; or
      ii) had been vaccinated at least twice, with the last vaccination not more than 12 and not less than one month prior to collection;

2. no other animal present in the artificial insemination centre has been vaccinated within the month prior to collection;

3. the semen:
   a) was collected, processed and stored in conformity with the provisions of Appendix 3.2.1. or Appendix 3.2.2., as relevant;
   b) was stored in the country of origin for a period of at least one month following collection, and during this period no animal on the establishment where the donor animals were kept showed any sign of FMD.

Article 2.2.10.16.

When importing from FMD infected countries or zones, Veterinary Authorities should require:
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for semen of domestic ruminants and pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of FMD on the day of collection of the semen;
   b) were kept in an establishment where no animal had been added in the 30 days before collection, and that FMD has not occurred within 10 kilometres for the 30 days before and after collection;
   c) have not been vaccinated and were subjected, not less than 21 days after collection of the semen, to tests for antibodies against FMD virus, with negative results; or
   d) had been vaccinated at least twice, with the last vaccination not more than 12 and not less than one month prior to collection;

2. no other animal present in the artificial insemination centre has been vaccinated within the month prior to collection;

3. the semen:
   a) was collected, processed and stored in conformity with the provisions of Appendix 3.2.1. or Appendix 3.2.2., as relevant;
   b) was subjected, with negative results, to a test for FMDV infection if the donor animal has been vaccinated within the 12 months prior to collection;
   c) was stored in the country of origin for a period of at least one month following collection, and during this period no animal on the establishment where the donor animals were kept showed any sign of FMD.

Article 2.2.10.17.

Irrespective of the FMD status of the exporting country or zone, Veterinary Authorities should authorise without restriction on account of FMD the import or transit through their territory of in vivo derived embryos of cattle subject to the presentation of an international veterinary certificate attesting that the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1. or Appendix 3.3.3., as relevant.

Article 2.2.10.18.

When importing from FMD free countries where vaccination is not practised or FMD free zones where vaccination is not practised, Veterinary Authorities should require:

for in vitro produced embryos of cattle

the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) showed no clinical sign of FMD at the time of collection of the oocytes;
b) were kept in a country or zone free from FMD at the time of collection;

2. fertilisation was achieved with semen meeting the conditions referred to in Articles 2.2.10.13., 2.2.10.14., 2.2.10.15. or 2.2.10.16., as relevant;

3. the oocytes were collected, and the embryos were processed and stored in conformity with the provisions of Appendix 3.3.2. or Appendix 3.3.3., as relevant.

Article 2.2.10.19.

When importing from FMD free countries where vaccination is practised or from FMD free zones where vaccination is practised, Veterinary Authorities should require:

for in vitro produced embryos of cattle

the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) showed no clinical sign of FMD at the time of collection of the oocytes;
   b) were kept in a country or zone free from FMD for at least 3 months prior to collection;
   c) if destined for an FMD free country or zone where vaccination is not practised:
      i) have not been vaccinated and were subjected, with negative results, to tests for antibodies against FMD virus, or
      ii) had been vaccinated at least twice, with the last vaccination not less than one month and not more than 12 months prior to collection;

2. no other animal present in the establishment has been vaccinated within the month prior to collection;

3. fertilisation was achieved with semen meeting the conditions referred to in Articles 2.2.10.13., 2.2.10.14., 2.2.10.15. or 2.2.10.16., as relevant;

4. the oocytes were collected, and the embryos were processed and stored in conformity with the provisions of Appendix 3.3.2. or Appendix 3.3.3., as relevant.

Article 2.2.10.20.

When importing from FMD free countries where vaccination is not practised or FMD free zones where vaccination is not practised, Veterinary Authorities should require:

for fresh meat of FMD susceptible animals

the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in the FMD free country or zone where vaccination is not practised since birth, or which have been imported in accordance with Article 2.2.10.10., Article 2.2.10.11. or Article 2.2.10.12.;
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2. have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections to rule out the presence of FMD with favourable results.

Article 2.2.10.21.

When importing from FMD free countries where vaccination is practised or from FMD free zones where vaccination is practised, Veterinary Authorities should require:

for fresh meat of cattle and buffalo (Bubalus bubalis) (excluding feet, head and viscera) the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in the FMD free country or zone where vaccination is practised since birth, or which have been imported in accordance with Article 2.2.10.10., Article 2.2.10.11. or Article 2.2.10.12.;

2. have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections to rule out the presence of FMD with favourable results.

Article 2.2.10.22.

When importing from FMD free countries where vaccination is practised or from FMD free zones where vaccination is practised, Veterinary Authorities should require:

for fresh meat or meat products of pigs and ruminants other than cattle and buffalo the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in the FMD free country or zone where vaccination is practised since birth, or which have been imported in accordance with Article 2.2.10.10., Article 2.2.10.11. or Article 2.2.10.12.;

2. have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections to rule out the presence of FMD with favourable results.

Article 2.2.10.23.

When importing from FMD infected countries or zones, where an official control programme exists, involving compulsory systematic vaccination of cattle, Veterinary Authorities should require:

for fresh meat of cattle and buffalo (Bubalus bubalis) (excluding feet, head and viscera) the presentation of an international veterinary certificate attesting that the entire consignment of meat:

1. comes from animals which:
   a) have remained in the exporting country for at least 3 months prior to slaughter;
   b) have remained, during this period, in a part of the country where cattle are regularly vaccinated against FMD and where official controls are in operation;
   c) have been vaccinated at least twice with the last vaccination not more than 12 months and not less than one month prior to slaughter;
Annex VII (contd)

d) were kept for the past 30 days in an establishment, and that FMD has not occurred within a ten-kilometre radius of the establishment during that period;

e) have been transported in a vehicle, which was cleansed and disinfected before the cattle were loaded, directly from the establishment of origin to the approved abattoir without coming into contact with other animals which do not fulfil the required conditions for export;

f) have been slaughtered in an approved abattoir:

i) which is officially designated for export;

ii) in which no FMD has been detected during the period between the last disinfection carried out before slaughter and the shipment for export has been dispatched;

g) have been subjected to ante-mortem and post-mortem inspections to rule out the presence of FMD with favourable results within 24 hours before and after slaughter;

2. comes from deboned carcasses:

a) from which the major lymphatic nodes have been removed;

b) which, prior to deboning, have been submitted to maturation at a temperature above +2°C for a minimum period of 24 hours following slaughter and in which the pH value was below 6.0 when tested in the middle of both the longissimus dorsi.

Article 2.2.10.24.

When importing from FMD infected countries or zones, Veterinary Authorities should require:

for meat products of domestic ruminants and pigs

the presentation of an international veterinary certificate attesting that:

1. the entire consignment of meat comes from animals which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections to rule out the presence of FMD with favourable results;

2. the meat has been processed to ensure the destruction of the FMD virus in conformity with one of the procedures referred to in Article 3.6.2.1.;

3. the necessary precautions were taken after processing to avoid contact of the meat products with any potential source of FMD virus.

Article 2.2.10.25.

When importing from FMD free countries or zones (where vaccination either is or is not practised), Veterinary Authorities should require:

for milk and milk products intended for human consumption and for products of animal origin (from FMD susceptible animals) intended for use in animal feeding or for agricultural or industrial use

the presentation of an international veterinary certificate attesting that these products come from animals which have been kept in the country or zone since birth, or which have been imported in accordance with Article 2.2.10.10, Article 2.2.10.11. or Article 2.2.10.12.
Annex VII (contd)

Article 2.2.10.26.

When importing from FMD infected countries or zones where an official control programme exists, Veterinary Authorities should require:

for milk, cream, milk powder and milk products

the presentation of an international veterinary certificate attesting that:

1. these products:
   a) originate from herds or flocks which were not infected or suspected of being infected with FMD at the time of milk collection;
   b) have been processed to ensure the destruction of the FMD virus in conformity with one of the procedures referred to in Article 3.6.2.5. and in Article 3.6.2.6.;

2. the necessary precautions were taken after processing to avoid contact of the products with any potential source of FMD virus.

Article 2.2.10.27.

When importing from FMD infected countries, Veterinary Authorities should require:

for blood and meat-meals (from domestic or wild ruminants and pigs)

the presentation of an international veterinary certificate attesting that the manufacturing method for these products included heating to a minimum internal temperature of 70°C for at least 30 minutes.

Article 2.2.10.28.

When importing from FMD infected countries, Veterinary Authorities should require:

for wool, hair, bristles, raw hides and skins (from domestic or wild ruminants and pigs)

the presentation of an international veterinary certificate attesting that:

1. these products have been processed to ensure the destruction of the FMD virus in conformity with one of the procedures referred to in Article 3.6.2.2., Article 3.6.2.3. and Article 3.6.2.4.;

2. the necessary precautions were taken after collection or processing to avoid contact of the products with any potential source of FMD virus.

Veterinary Authorities can authorise, without restriction, the import or transit through their territory of semi-processed hides and skins (limed hides, pickled pelts, and semi-processed leather - e.g. wet blue and crust leather -), provided that these products have been submitted to the usual chemical and mechanical processes in use in the tanning industry.

Article 2.2.10.29.

When importing from FMD infected countries or zones, Veterinary Authorities should require:

for straw and forage
Annex VII (contd)

the presentation of an international veterinary certificate attesting that these commodities:

1. are free of grossly identifiable contamination with material of animal origin;

2. have been subjected to one of the following treatments, which, in the case of material sent in bales, has been shown to penetrate to the centre of the bale:
   
   a) either to the action of steam in a closed chamber such that the centre of the bales has reached a minimum temperature of 80°C for at least 10 minutes,
   
   b) or to the action of formalin fumes (formaldehyde gas) produced by its commercial solution at 35-40% in a chamber kept closed for at least 8 hours and at a minimum temperature of 19°C;

   OR

3. have been kept in bond for at least 3 months (under study) before being released for export.

Article 2.2.10.30.

When importing from FMD free countries or zones (where vaccination either is or is not practised), Veterinary Authorities should require:

for skins and trophies derived from FMD susceptible wild animals

the presentation of an international veterinary certificate attesting that these products are derived from animals that have been killed in such a country or zone, or which have been imported from a country or zone free of FMD (where vaccination either is or is not practised).

Article 2.2.10.31.

When importing from FMD infected countries or zones, Veterinary Authorities should require:

for skins and trophies derived from FMD susceptible wild animals

the presentation of an international veterinary certificate attesting that these products have been processed to ensure the destruction of the FMD virus in conformity with the procedures referred to in Article 3.6.2.7.

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APPENDIX 3.8.7.

GUIDELINES FOR ON THE SURVEILLANCE OF FOR FOOT AND MOUTH DISEASE

Article 3.8.7.1.

**Introduction**

This Appendix defines the principles and provides a guide for the surveillance of foot and mouth disease (FMD) in accordance with Appendix 3.8.1. applicable to countries seeking recognition from the OIE for freedom from FMD, either with or without the use of vaccination. This may be for the entire country or a zone within the country. Guidance for countries seeking reestablishment of freedom from FMD for the whole country or a zone within the country, either with or without vaccination, following an outbreak, as well as guidelines for the maintenance of FMD status are provided. These guidelines are intended to expand on and explain the requirements of Chapter 2.2.10. Applications to the OIE for recognition of freedom should follow the format and answer all the questions posed by the “Questionnaire on FMD” available from the OIE Central Bureau.

The impact and epidemiology of FMD differ widely in different regions of the world and therefore it is impossible to provide specific guidelines for all situations. It is axiomatic that the surveillance strategies employed for demonstrating freedom from FMD at an acceptable level of confidence will need to be adapted to the local situation. For example, the approach to proving freedom from FMD following an outbreak caused by a pig-adapted strain of FMD virus (FMDV) should differ significantly from an application designed to prove freedom from FMD for a country or zone where African buffaloes (Syncerus caffer) provide a potential reservoir of infection. It is incumbent upon the applicant country to submit a dossier to the OIE in support of its application that not only explains the epidemiology of FMD in the region concerned but also demonstrates how all the risk factors are managed. This should include provision of scientifically-based supporting data. There is therefore considerable latitude available to Member Countries to provide a well-reasoned argument to prove that the absence of FMDV infection (in non-vaccinated populations) or circulation (in vaccinated populations) is assured at an acceptable level of confidence.

Surveillance for FMD should be in the form of a continuing programme designed to establish that the whole territory or part of it is free from FMDV infection/ circulation.

For the purposes of this Appendix, virus circulation means transmission of FMDV as demonstrated by clinical signs, serological evidence or virus isolation.

Article 3.8.7.2.

**General conditions and methods**

1. A surveillance system in accordance with Appendix 3.8.1. should be under the responsibility of the Veterinary Authority. A procedure should be in place for the rapid collection and transport of samples from suspect cases of FMD to a laboratory for FMD diagnoses as described in the Terrestrial Manual.

2. The FMD surveillance programme should:
Annex VII (contd)

a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers who have day-to-day contact with livestock, as well as diagnosticians, should report promptly any suspicion of FMD. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Authority. All suspect cases of FMD should be investigated immediately. Where suspicion cannot be resolved by epidemiological and clinical investigation, samples should be taken and submitted to an approved laboratory. This requires that sampling kits and other equipment are available for those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in FMD diagnosis and control;

b) implement, when relevant, regular and frequent clinical inspection and serological testing of high-risk groups of animals, such as those adjacent to an FMD infected country or zone (for example, bordering a game park in which infected wildlife are present).

An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is FMDV. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. Applications for freedom from FMDV infection/circulation should, in consequence, provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement standstill orders, etc.).

Article 3.8.7.3.

Surveillance strategies

1. Introduction

The target population for surveillance aimed at identifying disease and infection should cover all the susceptible species within the country or zone to be recognised as free from FMDV infection/circulation.

The strategy employed may be based on randomised sampling requiring surveillance consistent with demonstrating the absence of FMDV infection/circulation at an acceptable level of statistical confidence. The frequency of sampling should be dependent on the epidemiological situation. Targeted surveillance (e.g. based on the increased likelihood of infection in particular localities or species) may be an appropriate strategy. The applicant country should justify the surveillance strategy chosen as adequate to detect the presence of FMDV infection/circulation in accordance with Appendix 3.8.1. and the epidemiological situation. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clear clinical signs (e.g. cattle and pigs). If a Member wishes to apply for recognition of a specific zone within the country as being free from FMDV infection/circulation, the design of the survey and the basis for the sampling process would need to be aimed at the population within the zone.

For random surveys, the design of the sampling strategy will need to incorporate an epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect infection/circulation if it were to occur at a predetermined minimum rate. The sample size and expected disease prevalence determine the level of confidence in the results of the survey. The applicant country must justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Appendix 3.8.1. Selection of the design prevalence in particular clearly needs to be based on the prevailing or historical epidemiological situation.
Irrespective of the survey design selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and production class of animals in the target population.

Irrespective of the testing system employed, surveillance design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following-up positives to ultimately determine with a high level of confidence, whether they are indicative of infection/circulation or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as herds which may be epidemiologically linked to it.

The principles involved in surveillance for disease/infection are technically well defined. The design of surveillance programmes to prove the absence of FMDV infection/circulation needs to be carefully followed to avoid producing results that are either insufficiently reliable to be accepted by the OIE or international trading partners, or excessively costly and logistically complicated. The design of any surveillance programme, therefore, requires inputs from professionals competent and experienced in this field.

2. **Clinical surveillance**

Clinical surveillance aims at detecting clinical signs of FMD by close physical examination of susceptible animals. Whereas significant emphasis is placed on the diagnostic value of mass serological screening, surveillance based on clinical inspection should not be underrated. It may be able to provide a high level of confidence of detection of disease if a sufficiently large number of clinically susceptible animals is examined.

Clinical surveillance and laboratory testing should always be applied in series to clarify the status of FMD suspects detected by either of these complementary diagnostic approaches. Laboratory testing may confirm clinical suspicion, while clinical surveillance may contribute to confirmation of positive serology. Any sampling unit within which suspicious animals are detected should be classified as infected until contrary evidence is produced.

A number of issues must be considered in clinical surveillance for FMD. The often underestimated labour intensity and the logistical difficulties involved in conducting clinical examinations should not be underestimated and should be taken into account.

Identification of clinical cases is fundamental to FMD surveillance. Establishment of the molecular, antigenic and other biological characteristics of the causative virus, as well as its source, is dependent upon disclosure of such animals. It is essential that FMDV isolates are sent regularly to the regional reference laboratory for genetic and antigenic characterization.

3. **Virological surveillance**

Virological surveillance using tests described in the Terrestrial Manual should be conducted:

- **a)** to monitor at risk populations;
- **b)** to confirm clinically suspect cases;
- **c)** to follow up positive serological results;
- **d)** to test “normal” daily mortality, to ensure early detection of infection in the face of vaccination or in establishments epidemiologically linked to an outbreak.
Annex VII (contd)

4. Serological surveillance

Serological surveillance aims at detecting antibodies against FMDV. Positive FMDV antibody test results can have four possible causes:

a) natural infection with FMDV;

b) vaccination against FMD;

c) maternal antibodies derived from an immune dam (maternal antibodies in cattle are usually found only up to 6 months of age but in some individuals and in some species, maternal antibodies can be detected for considerably longer periods);

d) heterophile (cross) reactions.

It is important that serological tests, where applicable, contain antigens appropriate for detecting antibodies against viral variants (types, subtypes, lineages, topotypes, etc.) that have recently occurred in the region concerned. Where the probable identity of FMDVs is unknown or where exotic viruses are suspected to be present, tests able to detect representatives of all serotypes should be employed (e.g. tests based on nonstructural viral proteins – see below).

It may be possible to use serum collected for other survey purposes for FMD surveillance. However, the principles of survey design described in this Appendix and the requirement for a statistically valid survey for the presence of FMDV should not be compromised.

The discovery of clustering of seropositive reactions should be foreseen. It may reflect any of a series of events, including but not limited to the demographics of the population sampled, vaccinal exposure or the presence of field strain infection. As clustering may signal field strain infection, the investigation of all instances must be incorporated in the survey design. If vaccination cannot be excluded as the cause of positive serological reactions, diagnostic methods should be employed that detect the presence of antibodies to nonstructural proteins (NSPs) of FMDVs as described in the Terrestrial Manual.

The results of random or targeted serological surveys are important in providing reliable evidence that FMDV infection is not present in a country or zone. It is therefore essential that the survey be thoroughly documented.

Article 3.8.7.4.

Countries applying for freedom from FMD for the whole country or a zone where vaccination is not practised

In addition to the general conditions described in Chapter 2.2.10., a Member applying for recognition of FMD freedom for the country or a zone where vaccination is not practised should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and will be planned and implemented according to general conditions and methods in this Appendix, to demonstrate absence of FMDV infection, during the preceding 12 months in susceptible populations. This requires the support of a national or other laboratory able to undertake identification of FMDV infection through virus/antigen/genome detection and antibody tests described in the Terrestrial Manual.
Annex VII (contd)

Article 3.8.7.5.

**Countries or zones applying for freedom from FMD where vaccination is practised**

In addition to the general conditions described in Chapter 2.2.10., a Member applying for recognition of country or zone freedom from FMD with vaccination should show evidence of an effective surveillance programme planned and implemented according to general conditions and methods in this Appendix. Absence of clinical disease in the country or zone for the past 2 years should be demonstrated. Furthermore, surveillance should demonstrate that FMDV has not been circulating in any susceptible population during the past 12 months. This will require serological surveillance incorporating tests able to detect antibodies to NSPs as described in the Terrestrial Manual. Vaccination to prevent the transmission of FMDV may be part of a disease control programme. The level of herd immunity required to prevent transmission will depend on the size, composition (e.g. species) and density of the susceptible population. It is therefore impossible to be prescriptive. However, the aim should, in general, be to vaccinate at least 80% of the susceptible population. The vaccine must comply with the Terrestrial Manual. Based on the epidemiology of FMD in the country or zone, it may be that a decision is reached to vaccinate only certain species or other subsets of the total susceptible population. In that case, the rationale should be contained within the dossier accompanying the application to the OIE for recognition of status.

Evidence to show the effectiveness of the vaccination programme should be provided.

Article 3.8.7.6.

**Countries or zones re-applying for freedom from FMD where vaccination is either practised or not practised, following an outbreak**

In addition to the general conditions described in Chapter 2.2.10., a country re-applying for country or zone freedom from FMD where vaccination is practised or not practised should show evidence of an active surveillance programme for FMD as well as absence of FMDV infection/circulation. This will require serological surveillance incorporating, in the case of a country or a zone practising vaccination, tests able to detect antibodies to NSPs as described in the Terrestrial Manual.

Four strategies are recognised by the OIE in a programme to eradicate FMDV infection following an outbreak:

1. slaughter of all clinically affected and in-contact susceptible animals;
2. slaughter of all clinically affected and in-contact susceptible animals and vaccination of at-risk animals, with subsequent slaughter of vaccinated animals;
3. slaughter of all clinically affected and in-contact susceptible animals and vaccination of at-risk animals, without subsequent slaughter of vaccinated animals;
4. vaccination used without slaughter of affected animals or subsequent slaughter of vaccinated animals.

The time periods before which an application can be made for re-instatement of freedom from FMD depends on which of these alternatives is followed. The time periods are prescribed in Article 2.2.10.8.

In all circumstances, a Member re-applying for country or zone freedom from FMD with vaccination or without vaccination should report the results of an active surveillance programme implemented according to general conditions and methods in this Appendix.
Annex VII (contd)

Article 3.8.7.7.

The use and interpretation of serological tests (see Figure 1)

The recommended serological tests for FMD surveillance are described in the Terrestrial Manual.

Animals infected with FMDV produce antibodies to both the structural proteins (SP) and the nonstructural proteins (NSP) of the virus. Tests for SP antibodies to include SP-ELISAs and the virus neutralisation test (VNT). The SP tests are serotype specific and for optimal sensitivity should utilise an antigen or virus closely related to the field strain against which antibodies are being sought. Tests for NSP antibodies include NSP I-ELISA 3ABC and the electro-immunotransfer blotting technique (EITB) as recommended in the Terrestrial Manual or equivalent validated tests. In contrast to SP tests, NSP tests can detect antibodies to all serotypes of FMD virus. Animals vaccinated and subsequently infected with FMD virus develop antibodies to NSPs, but in some, the titre may be lower than that found in infected animals that have not been vaccinated. Both the NSP I-ELISA 3ABC and EITB tests have been extensively used in cattle. Validation in other species is ongoing. Vaccines used should comply with the standards of the Terrestrial Manual insofar as purity is concerned to avoid interference with NSP antibody testing.

Serological testing is a suitable tool for FMD surveillance. The choice of a serosurveillance system will depend on, amongst other things, the vaccination status of the country. A country, which is free from FMD without vaccination, may choose serosurveillance of high-risk subpopulations (e.g. based on geographical risk for exposure to FMDV). SP tests may be used in such situations for screening sera for evidence of FMDV infection/circulation if a particular virus of serious threat has been identified and is well characterised. In other cases, NSP testing is recommended in order to cover a broader range of strains and even serotypes. In both cases, serological testing can provide additional support to clinical surveillance. Regardless of whether SP or NSP tests are used in countries that do not vaccinate, a diagnostic follow-up protocol should be in place to resolve any presumptive positive serological test results.

In areas where animals have been vaccinated, SP antibody tests may be used to monitor the serological response to the vaccination. However, NSP antibody tests should be used to monitor for FMDV infection/circulation. NSP-ELISAs may be used for screening sera for evidence of infection/circulation irrespective of the vaccination status of the animal. All herds with seropositive reactors should be investigated. Epidemiological and supplementary laboratory investigation results should document the status of FMDV infection/circulation for each positive herd. Tests used for confirmation should be of high diagnostic specificity to eliminate as many false positive screening test reactors as possible. The diagnostic sensitivity of the confirmatory test should approach that of the screening test. The EITB or another OIE-accepted test should be used for confirmation.

Information should be provided on the protocols, reagents, performance characteristics and validation of all tests used.

1. The follow-up procedure in case of positive test results if no vaccination is used in order to establish or re-establish FMD free status without vaccination

Any positive test result (regardless of whether SP or NSP tests were used) should be followed up immediately using appropriate clinical, epidemiological, serological and, where possible, virological investigations of the reactor animal at hand, of susceptible animals of the same epidemiological unit and of susceptible animals that have been in contact or otherwise epidemiologically associated with the reactor animal. If the follow-up investigations provide no evidence for FMDV infection, the reactor animal shall be classified as FMD negative. In all other cases, including the absence of such follow-up investigations, the reactor animal should be classified as FMD positive.
2. The follow-up procedure in case of positive test results if vaccination is used in order to establish or re-establish FMD free status with vaccination

In case of vaccinated populations, one has to exclude that positive test results are indicative of virus circulation. To this end, the following procedure should be followed in the investigation of positive serological test results derived from surveillance conducted on FMD vaccinated populations.

The investigation should examine all evidence that might confirm or refute the hypothesis that the positive results to the serological tests employed in the initial survey were not due to virus circulation. All the epidemiological information should be substantiated, and the results should be collated in the final report.

It is suggested that in the primary sampling units where at least one animal reacts positive to the NSP test, the following strategy(ies) should be applied:

a) Following clinical examination, a second serum sample should be taken from the animals tested in the initial survey after an adequate interval of time has lapsed, on the condition that they are individually identified, accessible and have not been vaccinated during this period. Antibody titres against NSP at the time of retest should be statistically either equal to or lower than those observed in the initial test if virus is not circulating.

The animals sampled should remain in the holding pending test results and should be clearly identifiable. If the three conditions for retesting mentioned above cannot be met, a new serological survey should be carried out in the holding after an adequate period of time, repeating the application of the primary survey design and ensuring that all animals tested are individually identified. These animals should remain in the holding and should not be vaccinated, so that they can be retested after an adequate period of time.

b) Following clinical examination, serum samples should be collected from representative numbers of cattle that were in physical contact with the primary sampling unit. The magnitude and prevalence of antibody reactivity observed should not differ in a statistically significant manner from that of the primary sample if virus is not circulating.

c) Following clinical examination, epidemiologically linked herds should be serologically tested and satisfactory results should be achieved if virus is not circulating.

d) Sentinel animals can also be used. These can be young, unvaccinated animals or animals in which maternally conferred immunity has lapsed and belonging to the same species resident within the positive initial sampling units. They should be serologically negative if virus is not circulating. If other susceptible, unvaccinated ruminants (sheep, goats) are present, they could act as sentinels to provide additional serological evidence.

Laboratory results should be examined in the context of the epidemiological situation. Corollary information needed to complement the serological survey and assess the possibility of viral circulation includes but is not limited to:

- characterization of the existing production systems;
- results of clinical surveillance of the suspects and their cohorts;
- quantification of vaccinations performed on the affected sites;
- sanitary protocol and history of the establishments with positive reactors;
- control of animal identification and movements;
- other parameters of regional significance in historic FMDV transmission.

The entire investigative process should be documented as standard operating procedure within the surveillance programme.
Fig. 1. Schematic representation of laboratory tests for determining evidence of FMDV infection through or following serological surveys

Key:
- **ELISA**: Enzyme-linked immunosorbent assay
- **VNT**: Virus neutralisation test
- **NSP**: Nonstructural protein(s) of foot and mouth disease virus (FMDV)
- **3ABC**: NSP antibody test
- **EITB**: Electro-immuno transfer blotting technique (Western blot for NSP antibodies of FMDV)
- **SP**: Structural protein test
- **S**: No evidence of FMDV

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APPENDIX 3.6.2.
FOOT AND MOUTH DISEASE VIRUS INACTIVATION PROCEDURES

Article 3.6.2.1.

Meat

For the inactivation of viruses present in meat, one of the following procedures should be used:

1. **Canning**

   Meat is subjected to heat treatment in a hermetically sealed container to reach an internal core temperature of at least 70°C for a minimum of 30 minutes or to any equivalent treatment which has been demonstrated to inactivate the FMD virus.

2. **Thorough cooking**

   Meat, previously deboned and defatted, shall be subjected to heating so that an internal temperature of 70°C or greater is maintained for a minimum of 30 minutes.

   After cooking, it shall be packed and handled in such a way that it cannot be exposed to a source of virus.

3. **Drying after salting**

   When rigor mortis is complete, the meat must be deboned, salted with cooking salt (NaCl) and completely dried. It must not deteriorate at ambient temperature.

   ‘Drying’ is defined in terms of the ratio between water and protein which must not be greater than 2.25:1.

Article 3.6.2.2.

Wool and hair

For the inactivation of viruses present in wool and hair for industrial use, one of the following procedures should be used:

1. industrial washing, which consists of the immersion of the wool in a series of baths of water, soap and sodium hydroxyde (soda) or potassium hydroxyde (potash);

2. chemical depilation by means of slaked lime or sodium sulphide;

3. fumigation in formaldehyde in a hermetically sealed chamber for at least 24 hours. The most practical method is to place potassium permanganate in containers (which must NOT be made of plastic or polyethylene) and add commercial formalin; the amounts of formalin and potassium permanganate are respectively 53 ml and 35 g per cubic metre of the chamber;
Annex VII (contd)

4. industrial scouring which consists of the immersion of wool in a water-soluble detergent held at 60-70°C;

5. storage of wool at 18°C for 4 weeks, or 4°C for 4 months, or 37°C for 8 days.

Article 3.6.2.3.

Bristles

For the inactivation of viruses present in bristles for industrial use, one of the following procedures should be used:

1. boiling for at least one hour;

2. immersion for at least 24 hours in a 1% solution of formaldehyde prepared from 30 ml commercial formalin per litre of water.

Article 3.6.2.4.

Raw hides and skins

For the inactivation of viruses present in raw hides and skins for industrial use, the following procedure should be used: salting for at least 28 days in sea salt containing 2% sodium carbonate.

Article 3.6.2.5.

Milk and cream for human consumption

For the inactivation of viruses present in milk and cream for human consumption, one of the following procedures should be used:

1. a sterilisation process applying a minimum temperature of 132°C for at least one second (ultra-high temperature [UHT]), or

2. if the milk has a pH less than 7.0, a sterilisation process applying a minimum temperature of 72°C for at least 15 seconds (high temperature - short time pasteurisation [HTST]), or

3. if the milk has a pH of 7.0 or over, the HTST process applied twice.

Article 3.6.2.6.

Milk for animal consumption

For the inactivation of viruses present in milk for animal consumption, one of the following procedures should be used:

1. the HTST process applied twice;

2. HTST combined with another physical treatment, e.g. maintaining a pH 6 for at least one hour or additional heating to at least 72°C combined with dessication;
3. UHT combined with another physical treatment referred to in point 2 above.

Article 3.6.2.7.

**Skins and trophies from wild animals susceptible to foot and mouth disease**

For the inactivation of viruses present in skins and trophies from wild animals susceptible to FMD, one of the following procedures should be used prior to complete taxidermal treatment:

1. boiling in water for an appropriate time so as to ensure that any matter other than bone, horns, hooves, claws, antlers or teeth is removed;

2. gamma irradiation at a dose of at least 20 kiloGray at room temperature (20°C or higher);

3. soaking, with agitation, in a 4% (w/v) solution of washing soda (sodium carbonate - Na$_2$CO$_3$) maintained at pH 11.5 or above for at least 48 hours;

4. soaking, with agitation, in a formic acid solution (100 kg salt [NaCl] and 12 kg formic acid per 1,000 litres water) maintained at below pH 3.0 for at least 48 hours; wetting and dressing agents may be added;

5. in the case of raw hides, salting for at least 28 days with sea salt containing 2% washing soda (sodium carbonate - Na$_2$CO$_3$).

Article 3.6.2.8.

**Casings of small ruminants and pigs**

For the inactivation of viruses present in casings of small ruminants and pigs, the following procedures should be used: salting for at least 30 days either with dry salt (NaCl) or with saturated brine (Aw < 0.80), and kept at room temperature during this entire period.
CHAPTER 2.2.12.

RINDERPEST

Article 2.2.12.1.

For the purposes of the Terrestrial Code, the incubation period for rinderpest (RP) shall be 21 days.

For the purpose of this chapter, a case includes an animal infected with rinderpest virus (RPV).

For the purpose of this chapter, susceptible animals apply to both domestic and wild artiodactyls.

For the purposes of international trade, this chapter deals not only with the occurrence of clinical signs caused by RPV, but also with the presence of infection with RPV in the absence of clinical signs.

Ban on vaccination against rinderpest means a ban on administering a RP vaccine to any susceptible animal and a heterologous vaccine against RP to any large ruminants or pigs.

1. Animal not vaccinated against RP means:
   a) for large ruminants and pigs: an animal that has received neither a RP vaccine nor a heterologous vaccine against RP;
   b) for small ruminants: an animal that has not received a RP vaccine.

2. The following defines the occurrence of RPV infection:
   a) RPV has been isolated and identified as such from an animal or a product derived from that animal; or
   b) viral antigen or viral ribonucleic acid (RNA) specific to RP has been identified in samples from one or more animals showing one or more clinical signs consistent with RP, or epidemiologically linked to an outbreak of RP, or giving cause for suspicion of association or contact with RP; or
   c) antibodies to RPV antigens which are not the consequence of vaccination, have been identified in one or more animals with either epidemiological links to a confirmed or suspected outbreak of RP in susceptible animals, or showing clinical signs consistent with recent infection with RP.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.2.12.2.

RP free country

To qualify for inclusion in the existing list of RP free countries, a country should:

1. have a record of regular and prompt animal disease reporting;
2. send a declaration to the OIE stating that:
   a) there has been no outbreak of RP during the past 24 months,
   b) no evidence of RPV infection has been found during the past 24 months,
Annex VIII (contd)

c) no vaccination against RP has been carried out during the past 24 months,

and supply documented evidence that surveillance for both RP and RPV infection in accordance with
Appendix 3.8.2. is in operation and that regulatory measures for the prevention and control of RP
have been implemented;

3. not have imported since the cessation of vaccination any animals vaccinated against RP.

The country will be included in the list only after the submitted evidence has been accepted by the OIE.
Retention on the list requires that the information in points 2 and 3 above be re-submitted annually and
changes in the epidemiological situation or other significant events should be reported promptly to the
OIE.

Article 2.2.12.3.

Recovery of free status

When a RP outbreak or RPV infection occurs in a RP free country, one of the following waiting periods is
required to regain the status of RP free country:

1. 3 months after the last case where a stamping-out policy and serological surveillance are applied in
accordance with Appendix 3.8.2.; or

2. 3 months after the slaughter of all vaccinated animals where a stamping-out policy, emergency vaccination
and serological surveillance are applied in accordance with Appendix 3.8.2.; or

3. 6 months after the last case or the last vaccination (according to the event that occurs the latest), where
a stamping-out policy, emergency vaccination not followed by the slaughter of all vaccinated animals, and
serological surveillance are applied in accordance with Appendix 3.8.2.

Where a stamping-out policy is not practised, the above waiting periods do not apply but Article 2.2.12.2.
applies.

Article 2.2.12.4.

Infected country

When the requirements for acceptance as a RP free country are not fulfilled, a country shall be considered
as RP infected.

Article 2.2.12.5.

When importing from RP free countries, Veterinary Authorities should require:

for RP susceptible animals

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of RP on the day of shipment;

2. remained in a RP free country since birth or for at least 30 days prior to shipment.
Article 2.2.12.6.

When importing from RP infected countries, Veterinary Authorities should require:

for RP susceptible animals

the presentation of an international veterinary certificate attesting that:

1. RP is the subject of a national surveillance programme according to Appendix 3.8.2.;
2. RP has not occurred within a 10-kilometre radius of the establishment of origin of the animals destined for export for at least 21 days prior to their shipment to the quarantine station referred to in point 3b) below;
3. the animals:
   a) showed no clinical sign of RP on the day of shipment;
   b) were kept in the establishment of origin since birth or for at least 21 days before introduction into the quarantine station referred to in point c) below;
   c) have not been vaccinated against RP, were isolated in a quarantine station for the 30 days prior to shipment, and were subjected to a diagnostic test for RP on two occasions with negative results, at an interval of not less than 21 days;
   d) were not exposed to any source of infection during their transportation from the quarantine station to the place of shipment;
4. RP has not occurred within a ten-kilometre radius of the quarantine station for 30 days prior to shipment.

Article 2.2.12.7.

When importing from RP free countries, Veterinary Authorities should require:

for semen of RP susceptible animals

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of RP on the day of collection of the semen;
   b) were kept in a RP free country for at least 3 months prior to collection;
2. the semen was collected, processed and stored in conformity with the provisions of either Appendix 3.2.1. or Appendix 3.2.2., as relevant.

Article 2.2.12.8.

When importing from RP infected countries, Veterinary Authorities should require:

for semen of RP susceptible animals
Annex VIII (contd)

the presentation of an international veterinary certificate attesting that:

1. RP is the subject of a national surveillance programme according to Appendix 3.8.2.;

2. the donor animals:
   a) showed no clinical sign of RP on the day of collection of the semen;
   b) were kept in an establishment where no RP susceptible animals had been added in the 21 days before collection, and that RP has not occurred within 10 kilometres of the establishment for the 21 days before and after collection;
   c) were vaccinated against RP at least 3 months prior to collection; or
   d) have not been vaccinated against RP, and were subjected to a diagnostic test on two occasions with negative results, at an interval of not less than 21 days within the 30 days prior to collection;

3. the semen was collected, processed and stored in conformity with the provisions of either Appendix 3.2.1. or Appendix 3.2.2., as relevant.

Article 2.2.12.9.

When importing from RP free countries, Veterinary Authorities should require:

for in vivo derived embryos of RP susceptible animals

the presentation of an international veterinary certificate attesting that:

1. the donor females were kept in an establishment located in a RP free country at the time of collection;

2. the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1. or Appendix 3.3.3., as relevant.

Article 2.2.12.10.

When importing from RP infected countries, Veterinary Authorities should require:

for in vivo derived embryos of RP susceptible animals

the presentation of an international veterinary certificate attesting that:

1. RP is the subject of a national surveillance programme according to Appendix 3.8.2.;

2. the donor females:
   a) and all other animals in the establishment showed no clinical sign of RP at the time of collection and for the following 21 days;
   b) were kept in an establishment where no RP susceptible animals had been added in the 21 days before collection of the embryos;
   c) were vaccinated against RP at least 3 months prior to collection; or
d) have not been vaccinated against RP, and were subjected to a diagnostic test for RP on two occasions with negative results, at an interval of not less than 21 days within the 30 days prior to collection;

3. the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1. or Appendix 3.3.3., as relevant.

Article 2.2.12.11.

When importing from RP free countries, Veterinary Authorities should require:

for fresh meat or meat products of susceptible animals

the presentation of an international veterinary certificate attesting that the entire consignment comes from animals which have been kept in the country since birth or for at least 3 months prior to slaughter.

Article 2.2.12.12.

When importing from RP infected countries, Veterinary Authorities should require:

for fresh meat (excluding offal) of susceptible animals

the presentation of an international veterinary certificate attesting that the entire consignment of meat:

1. comes from a country where RP is the subject of a national surveillance programme according to Appendix 3.8.2.;

2. comes from animals which:

   a) showed no clinical sign of RP within 24 hours before slaughter;

   b) have remained in the country for at least 3 months prior to slaughter;

   c) were kept in the establishment of origin since birth or for at least 30 days prior to shipment to the approved abattoir, and that RP has not occurred within a ten-kilometre radius of the establishment during that period;

   d) were vaccinated against RP at least 3 months prior to shipment to the approved abattoir;

   e) had been transported, in a vehicle which was cleansed and disinfected before the animals were loaded, directly from the establishment of origin to the approved abattoir without coming into contact with other animals which do not fulfil the required conditions for export;

   f) were slaughtered in an approved abattoir in which no RP has been detected during the period between the last disinfection carried out before slaughter and the date on which the shipment has been dispatched.

Article 2.2.12.13.

When importing from RP infected countries, Veterinary Authorities should require:

for meat products of susceptible animals

the presentation of an international veterinary certificate attesting that:
Annex VIII (contd)

1. only fresh meat complying with the provisions of Article 2.2.12.12. has been used in the preparation of the meat products; or

2. the meat products have been processed to ensure the destruction of the RPV in conformity with one of the procedures referred to in Article 3.6.2.1.;

3. the necessary precautions were taken after processing to avoid contact of the meat products with any possible source of RPV.

Article 2.2.12.14.

When importing from RP free countries, Veterinary Authorities should require:

for milk and milk products intended for human consumption and for products of animal origin (from RP susceptible animals) intended for use in animal feeding or for agricultural or industrial use

the presentation of an international veterinary certificate attesting that these products come from animals which have been kept in the country since birth or for at least 3 months.

Article 2.2.12.15.

When importing from RP infected countries, Veterinary Authorities should require:

for milk and cream

the presentation of an international veterinary certificate attesting that:

1. these products:
   a) originate from herds or flocks which were not subjected to any restrictions due to RP at the time of milk collection;
   b) have been processed to ensure the destruction of the RPV in conformity with one of the procedures referred to in Article 3.6.2.5. and in Article 3.6.2.6.;

2. the necessary precautions were taken after processing to avoid contact of the products with any potential source of RPV.

Article 2.2.12.16.

When importing from RP infected countries, Veterinary Authorities should require:

for milk products

the presentation of an international veterinary certificate attesting that:

1. these products are derived from milk complying with the above requirements;

2. the necessary precautions were taken after processing to avoid contact of the milk products with a potential source of RPV.

Article 2.2.12.17.

When importing from RP infected countries, Veterinary Authorities should require:
for blood and meat-meals (from susceptible animals)

the presentation of an international veterinary certificate attesting that the manufacturing method for these products included heating to a minimum internal temperature of 70°C for at least 30 minutes.

**Article 2.2.12.18.**

When importing from RP infected countries, Veterinary Authorities should require:

for wool, hair, bristles, raw hides and skins (from susceptible animals)

the presentation of an international veterinary certificate attesting that:

1. these products have been processed to ensure the destruction of the RPV in conformity with one of the procedures referred to in Article 3.6.2.2., Article 3.6.2.3. and Article 3.6.2.4.;

2. the necessary precautions were taken after processing to avoid contact of the products with any potential source of RPV.

Veterinary Authorities can authorise, without restriction, the import or transit through their territory of semi-processed hides and skins (limed hides, pickled pelts, and semi-processed leather - e.g. wet blue and crust leather), provided that these products have been submitted to the usual chemical and mechanical processes in use in the tanning industry.

**Article 2.2.12.19.**

When importing from RP infected countries, Veterinary Authorities should require:

for hooves, claws, bones and horns, hunting trophies and preparations destined for museums (from susceptible animals)

the presentation of an international veterinary certificate attesting that these products:

1. were completely dried and had no trace on them of skin, flesh or tendon; and/or

2. have been adequately disinfected.

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1 [Note: International veterinary certificates for animal products coming from RP infected countries, may not be required if the products are transported in an approved manner to premises controlled and approved by the Veterinary Authority of the importing country for processing to ensure the destruction of the RPV as described in Article 3.6.2.2., Article 3.6.2.3. and Article 3.6.2.4.]

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APPENDIX 3.8.2.

GUIDELINES FOR THE SURVEILLANCE OF RINDERPEST

Article 3.8.2.1.

Purposes of the document

In order to receive OIE recognition of rinderpest freedom, a country's national authority must present for consideration a dossier of information relating to its livestock production systems, rinderpest vaccination and eradication history and the functioning of its Veterinary Services. The dossier must contain convincing evidence derived from an animal disease surveillance system that sufficient evidence has accrued to demonstrate that the presence of rinderpest virus would have been disclosed were it to be present. Guidelines for the structure and the functioning of Veterinary Services and diagnostic support services are provided in Chapters 1.3.3. and 1.3.4. of the Terrestrial Code. A Member must also be in compliance with its OIE reporting obligations (Chapter 1.1.2. of the Terrestrial Code).

Article 3.8.2.2.

Definitions

1. Rinderpest

For the purpose of this Appendix, rinderpest is defined as an infection of large ruminants (cattle, buffaloes, yaks, etc.), small ruminants, pigs and various wildlife species within the order Artiodactyla, caused by rinderpest virus. In small ruminants and various species of wildlife, particularly antelopes, infection generally passes without the development of frank clinical signs. Characteristic clinical signs and pathological lesions are described in Chapter 2.1.4. of the Terrestrial Manual.

Outbreaks of rinderpest in cattle may be graded as per-acute, acute or sub-acute. Differing clinical presentations reflect variations in levels of innate host resistance (Bos indicus breeds being more resistant than Bos taurus), and variations in the virulence of the attacking strain. It is generally accepted that unvaccinated populations of cattle are likely to promote the emergence of virulent strains and associated epidemics while partially vaccinated populations favour the emergence of mild strains associated with endemic situations. In the case of per-acute cases the presenting sign may be sudden death. In the case of sub-acute (mild) cases, clinical signs are irregularly displayed and difficult to detect.

Freedom from rinderpest means freedom from rinderpest virus infection.

2. Rinderpest vaccines

For the purpose of this Appendix and the Terrestrial Code, OIE-recognised rinderpest vaccines currently in use, or likely to become so in the foreseeable future, are considered to be commercial modified live vaccines produced from attenuated rinderpest virus (referred to as 'rinderpest vaccine') produced in accordance with Chapter 2.1.4. of the Terrestrial Manual.

Article 3.8.2.3.

Rinderpest surveillance

General guidelines on animal disease surveillance are outlined in Appendix 3.8.1. of the Terrestrial Code.
Rinderpest must be a notifiable disease i.e. notification of outbreaks of rinderpest as soon as detected or suspected must be brought to the attention of the Veterinary Authority.

The precise surveillance information required for establishing freedom will differ from country to country depending on factors such as the former rinderpest status of the country, the regional rinderpest situation and accreditation status, the time elapsing since the last occurrence of rinderpest, livestock husbandry systems (e.g. extensive pastoralism, nomadism and transhumance versus sedentary agropastoralism) and trading patterns.

Evidence of efficiency of the surveillance system can be provided by the use of performance indicators.

Surveillance results presented will be expected to have accrued from a combination of surveillance activities including some or all of the following:

1. **A routine national animal disease reporting system supported by evidence of its efficiency and follow-up – an on-going, statutory, centrally organised system of reporting**

   Ideally disease reports should be expressed in a Geographical Information System environment and analysed for clustering of observations and followed up.

2. **Emergency disease reporting systems and investigation of epidemiologically significant events (stomatitis-enteritis syndrome)**

   Emergency reporting systems can be devised to short-circuit normal passive reporting systems to bring suspicious events to the fore and lead to rapid investigation and tracing. All such investigations should be well documented for presentation as an outcome of the surveillance system.

3. **Detection and thorough investigation of epidemiologically significant events (stomatitis-enteritis syndrome) which raise suspicion of rinderpest supported by evidence of efficiency of the system**

   Laboratory examination undertaken to confirm or rule out rinderpest is given extra credibility if it is accompanied by the results of differential diagnostic examinations.

4. **Searching for evidence of clinical rinderpest**

   Active search for disease might include participatory disease searching combined with village disease searching, tracing backwards and forwards, follow-up and investigation.

5. **Serosurveillance**

   a) **Randomised serosurveys**

   Statistically selected samples from relevant strata within the host populations are examined to detect serological evidence of possible virus circulation.

   A sampling unit for the purposes of disease investigation and surveillance is defined as a group of animals in sufficiently close contact that individuals within the group are at approximately equal risk of coming in contact with the virus if there should be an infectious animal within the group. In most circumstances, the sampling unit will be a herd which is managed as a unit by an individual or a community, but it may also be other epidemiologically appropriate groupings which are subject to regular mixing, such as all animals belonging to residents of a village. In the areas where nomadic or transhumant movements exist, the sampling unit can be the permanent bore holes, wells or water points. Sampling units should normally be defined so that their size is generally between 50 and 1,000 animals.
i) Criteria for stratification of host populations

Strata are homogeneously mixing sub-populations of livestock. Any disease surveillance activities must be conducted on populations stratified according to the management system, and by herd size where this is variable. Herds, or other sampling units, should be selected by proper random statistical selection procedures from each stratum.

ii) Field procedures and sample sizes

Annual sample sizes shall be sufficient to provide 95% probability of detecting evidence of rinderpest if present at a prevalence of 1% of herds or other sampling units and 5% within herds or other sampling units. This can typically be achieved by examining 300 herds per stratum per year, but procedures for sampling should be in accordance with the “Guide to Epidemiological Surveillance for Rinderpest”\(^1\), or another procedure that would achieve the same probability of detection.

Where the sampling frame of herds is known, herds shall be selected for examination by the use of random number tables. Otherwise, samples of herds can be selected by taking the nearest herd to a randomly selected map reference, provided that the herds are evenly distributed. Failing this, any herd(s) within a fixed radius of randomly selected map references should be sampled. It must be compulsory for any selected herd to be examined or tested as required.

In carrying out clinical surveillance for evidence of rinderpest, all animals in selected herds or sampling units will be examined by a veterinarian for signs of the disease, especially mouth lesions. Any positive result shall be evaluated using epidemiological and laboratory methods to confirm or refute the suspicion of rinderpest virus activity. All animals born after the cessation of vaccination and more than one year old will be eligible for serological testing.

Where operational considerations require it, the number of eligible animals tested within each sampled herd may be reduced. This will reduce the probability of within-herd detection and there must be at least a compensatory increase in the number of herds sampled, so that the required 95% probability of detecting 1% between-herd prevalence is maintained.

b) Risk-focussed serosurveillance

Risk-focussed serosurveillance differs from randomised serosurveillance in that it increases detection sensitivity by obtaining samples from areas/populations determined to be at higher risk of infection, so as to detect serological evidence of possible virus circulation. The operational modalities for risk-based focussing of surveillance require definition (randomisation within defined focus, high risk animals, etc.). The extent to which randomisation needs to be retained in the generation of risk-focussed serosurveillance data needs to be established.

Focussing can be achieved by reference to some or all of the following:

i) Historical disease patterns (prior probability mapping) - clinical, participatory and laboratory-based

ii) Critical population size, structure and density

iii) Livestock husbandry and farming systems
iv) Movement and contact patterns — markets and other trade-related movements

v) Transmission parameters (e.g. virulence of the strain, animal movements)

vi) Wildlife and other species demography.

Article 3.8.2.4.

Selection of cattle and buffaloes for serosurveillance

Ageing cattle and Asian buffaloes for the purpose of serosurveillance:

Mis-ageing of cattle selected for serosurveillance is the most common source of error. Colostral immunity can persist almost up to one year of age when measured by the H c-ELISA. Thus, it is essential to exclude from sampling buffaloes and cattle less than one year of age. In addition, it is frequently necessary to be able to exclude those which are older than a certain age, for example, to select only those born after cessation of vaccination.

Accounts of the ages for eruption of the incisor teeth vary markedly and are clearly dependent on species, breed, nutritional status and nature of the feed.

Pragmatically, and solely for the purposes of serosurveillance, it can be accepted that:

a) cattle having only one pair of erupted permanent central incisor teeth are aged between 21 and 36 months (Asian buffaloes 24-48 months);

b) cattle having only two pairs of erupted permanent central incisor teeth are aged between 30 and 48 months (Asian buffaloes 48-60 months).

Thus selecting a cohort of cattle possessing only one pair of permanent incisors will preclude any interference from maternal immunity derived from earlier vaccination or infection and ensure that vaccinated cattle are not included if vaccination ceased 3 years or more previously (for Asian buffaloes 4 years or more).

Although it is stressed here that animals with milk teeth only are not suitable for surveillance based on serology, they are of particular interest and importance in surveillance for clinical disease. After the loss of colostral immunity, by about one year of age, these are the animals which are most likely to suffer the more severe disease form and in which to look for lesions indicative of rinderpest.

Article 3.8.2.5.

Wildlife surveillance where a significant susceptible wildlife population exists

There are some key wildlife populations, especially African buffaloes, which act as sentinels for rinderpest infection. Where a significant population of a susceptible wildlife species exists, serosurveillance data are required to support absence of infection. These populations should be monitored purposively to support the dossiers to be submitted for freedom from rinderpest virus infection. Detection of virus circulation in wildlife can be undertaken indirectly by sampling contiguous livestock populations.

Obtaining meaningful data from wildlife surveillance can be enhanced by close coordination of activities in the regions and countries. Both purposive and opportunistic samplings are used to obtain material for analysis in national and reference laboratories. The latter are required because most countries are unable to perform the full testing protocol for detecting rinderpest antibodies in wildlife sera.
Purposive sampling is the preferred method to provide wildlife data to evaluate the status of rinderpest infection. In reality, the capacity to perform purposive work in the majority of countries remains minimal. Opportunistic sampling (hunting) is feasible and it provides useful background information.

Wildlife form transboundary populations; therefore, any data from the population could be used to represent the result for the ecosystem and be submitted by more than one country in a dossier (even if the sampling was not obtained in the country submitting). It is therefore recommended that the countries represented in a particular ecosystem should coordinate their sampling programmes.

The standards for serosurveillance are different from that set for cattle because the serological tests are not fully validated for wildlife species and financial and logistic constraints of sampling prevent collection of large numbers of samples.

From the collective experience of the laboratories and experts over the years, an appropriate test protocol is based on the high expected sero-prevalence in a previously infected buffalo herd (99% seroconversion of eligible animals within a herd), which is detected using a test, which is 100% sensitive. No single test can achieve this; however, combining H c-ELISA to VNT raises sensitivity close to 100%.

In the order of 1-2% of a herd of African buffaloes must be sampled to ensure that no positive case is missed. For example in a herd of 300 buffaloes, five animals should be sampled and the above multiple test protocol followed. Where the serological history of the herd is known from previous work (as might be the case for a sentinel herd), repeat sampling need only focus on the untested age groups, born since the last known infection. Appropriate sampling fraction for other wildlife species are less well defined, as social organization (herd structure, likely contact rates, etc.) vary. The sample needs to be taken according to the known epidemiology of the disease in a given species. Opportunistic samples, which are positive, should not be interpreted without a purposive survey to confirm the validity of these results. Opportunistic sampling cannot follow a defined protocol and therefore can only provide background information.

**Article 3.8.2.6.**

**Evaluation of applications for accreditation of freedom from rinderpest**

Evaluation of applications for the status of freedom from rinderpest will be the responsibility of the OIE Scientific Commission for Animal Diseases which can request the Director General if the OIE to appoint an **ad hoc** group in order to assist in reaching an informed decision to present to the OIE International Committee for approval.

The composition and method of selection of the **ad hoc** group shall be such as to ensure both a high level of expertise in evaluating the evidence and total independence of the group in reaching conclusions concerning the disease status of a particular country.

**Article 3.8.2.7.**

**Steps to be taken to declare a country to be free from rinderpest**

Recognition of the status 'free from rinderpest' is given to a Member. Where traditionally managed livestock move freely across international borders, groups of Members may usefully associate themselves into a group for the purposes of obtaining data to be used for mutually supportive applications for individual country accreditation.

For the purpose of this Appendix, the following assumptions are made:
Annex VIII (contd)

a) that within most previously infected countries, rinderpest vaccine will have been used to control the rate of infection;

b) that within an endemically infected population there will be a large number of immune hosts (both vaccines and recovered animals);

c) that the presence of a proportion of immune hosts within a vaccinated population could have led to a slowing of the rate of virus transmission and possibly the concomitant emergence of strains of reduced virulence, difficult to detect clinically;

d) that the virulence of the virus (and therefore the ease of clinical detection) may or may not increase as the herd immunity declines following withdrawal of vaccination; however, continuing transmission will generate serological evidence of their persistence.

Before accreditation can be considered, countries which have controlled the disease by the use of rinderpest vaccine must wait until an unvaccinated cohort is available to allow meaningful serological surveillance to be conducted.

The OIE has concluded that the majority of countries have stopped vaccinating for a sufficient length of time for it now to be feasible that a single submission of evidence gained over 2 years of appropriate surveillance shall be sufficient to gain rinderpest free accreditation.

A Member accredited as free from rinderpest must thereafter submit annual statements to the Director General of the OIE indicating that surveillance has failed to disclose the presence of rinderpest, and that all other criteria continue to be met.

A country previously infected with rinderpest which has not employed rinderpest vaccine for at least 25 years and has throughout that period detected no evidence of rinderpest virus disease or infection may be accredited as free from rinderpest by the OIE based on historical grounds, provided that the country:

- has had throughout at least the last 10 years and maintains permanently an adequate animal disease surveillance system along with the other requirements outlined in Article 3.8.1.6.;

- is in compliance with OIE reporting obligations (Chapter 1.1.2.).

The Veterinary Authorities of the Member must submit a dossier containing evidence supporting their claim to be free from rinderpest on a historical basis to the Director General of the OIE for evaluation by the OIE Scientific Commission for Animal Diseases and accreditation by the OIE International Committee. The dossier should contain at least the following information:

- a description of livestock populations, including wildlife;

- the history of rinderpest occurrence in the country and its control;

- an affirmation that rinderpest has not occurred for 25 years, that vaccine has not been used during that time, and that rinderpest is a notifiable disease;

- evidence that in the last 10 years the disease situation throughout the Member has been constantly monitored by a competent and effective veterinary infrastructure that has operated a national animal disease reporting system submitting regular (monthly) disease occurrence reports to the Veterinary Authority;

- the structure and functioning of the Veterinary Services;
- the Member operates a reliable system of risk analysis based importation of livestock and livestock products.

Evidence in support of these criteria must accompany the Member’s accreditation application dossier. In the event that satisfactory evidence is not forthcoming, the OIE may seek clarification or refer the dossier back to the originators, giving its reasons for so doing. Under such circumstances a fresh dossier would be entertained in due course.

OR

A Member having eradicated rinderpest within the last 25 years, wishing to be accredited free from rinderpest and having ended rinderpest vaccination must initiate a two-year surveillance programme to demonstrate freedom from rinderpest whilst banning further use of rinderpest vaccine. The step of accreditation as free from rinderpest is subject to meeting stringent criteria with international verification under the auspices of the OIE.

A country historically infected with rinderpest but which has convincing evidence that the disease has been excluded for at least two years and is not likely to return, may apply to OIE to be accredited as free from rinderpest. The conditions which apply include that an adequate animal disease surveillance system has been maintained throughout at least that period.

The Veterinary Authority of the Member must submit a dossier containing evidence supporting their claim to be free from rinderpest to the Director General of the OIE for evaluation by the OIE Scientific Commission for Animal Diseases and accreditation by the OIE International Committee showing that they comply with:

- the provisions outlined in Chapter 2.2.12. of the Terrestrial Code;
- OIE reporting obligations outlined in Chapter 1.1.2. of the Terrestrial Code.

Other conditions that apply are:

- The Member affirms that rinderpest has not occurred for at least 2 years, that vaccine has not been used during that time, and that rinderpest is a notifiable disease.
- The Veterinary Authority has issued orders curtailing the distribution and use of rinderpest vaccine in livestock.
- The Veterinary Authority has issued orders for the recall and destruction of rinderpest vaccine already issued.
- The Veterinary Authority has issued orders restricting the importation of rinderpest vaccine into, or the further manufacture of rinderpest vaccine within, the territory under his jurisdiction. An exception can be made for establishing a safeguarded rinderpest emergency vaccine bank under the control of the Chief Veterinary Officer who can demonstrate that no calls have been made on that vaccine bank.
- The Veterinary Authority has set in place a rinderpest contingency plan.
- Over the previous 2 years at least, the disease situation throughout the Member has been constantly monitored by a competent and effective infrastructure that has operated a national animal disease reporting system submitting regular (monthly) disease occurrence reports to the Veterinary Authority.
All outbreaks of disease with a clinical resemblance to rinderpest have been thoroughly investigated and routinely subjected to laboratory testing by an OIE recognised rinderpest-specific test within the national rinderpest laboratory or at a recognised reference laboratory.

The dossier shall contain:

- the results of a continuous surveillance programme, including appropriate serological surveys conducted during at least the last 24 months, providing convincing evidence for the absence of rinderpest virus circulation;

- a description of livestock populations including wildlife;

- the history of rinderpest occurrence in the country and its control;

- an affirmation that rinderpest has not occurred for at least 2 years, that vaccine has not been used during that time, and that rinderpest is a notifiable disease;

- evidence that in the last 2 years the disease situation throughout the Member has been constantly monitored by a competent and effective veterinary infrastructure that has operated a national animal disease reporting system submitting regular (monthly) disease occurrence reports to the Veterinary Authority;

- the structure and functioning of the Veterinary Services;

- the Member operates a reliable system of risk analysis based importation of livestock and livestock products.

In the event that satisfactory evidence in support of the application is not forthcoming, the OIE may seek clarification or refer the dossier back to the originators, giving its reasons for so doing. Under such circumstances a fresh dossier would be entertained in due course.

**Article 3.8.2.8.**

**Rinderpest outbreaks after the accreditation process and recovery of rinderpest free status**

Should there be an outbreak, or outbreaks, of rinderpest in a Member at any time after recognition of rinderpest freedom, the origin of the virus strain must be thoroughly investigated. In particular it is important to determine if this is due to the re-introduction of virus or re-emergence from an undetected focus of infection. The virus must be isolated and compared with historical strains from the same area as well as those representatives of other possible sources. The outbreak itself must be contained with the utmost rapidity using the resources and methods outlined in the Contingency Plan.

After elimination of the outbreak, a Member wishing to regain the status ‘free from rinderpest’ must undertake serosurveillance to determine the extent of virus spread.

If investigations show the outbreak virus originated from outside the country, provided the outbreak was localised, rapidly contained and speedily eliminated, and provided there was no serological evidence of virus spread outside the index infected area, accreditation of freedom could proceed rapidly. The country must satisfy the OIE Scientific Commission for Animal Diseases that the outbreaks were contained, eliminated and did not represent endemic infection.
An application to regain the status free from rinderpest shall not generally be accepted until both clinical and serological evidence shows that there has been no virus transmission for at least 3 or 6 months, depending on whether or not stamping-out or vaccination respectively has been applied.

CHAPTER 2.3.15.

CONTAGIOUS BOVINE PLEUROPNEUMONIA

Article 2.3.15.1.

For the purposes of the Terrestrial Code, the incubation period for contagious bovine pleuropneumonia (CBPP) shall be 6 months.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.3.15.2.

CBPP free country

To be declared free from either disease or infection by the OIE, a country should meet the requirements contained in Appendix 3.8.3.

The country will be included in the list of free country only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information above be re-submitted annually and changes in the epidemiological situation or other significant events should be reported promptly to the OIE.

Article 2.3.15.3.

CBPP free zone

To be declared free from either disease or infection by the OIE, a zone defined according to the provisions of Chapter 1.3.5. should meet the requirements contained in Appendix 3.8.3.

The zone will be included in the list of free zone only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information above be re-submitted annually and changes in the epidemiological situation or other significant events should be reported promptly to the OIE.

Article 2.3.15.4.

CBPP infected country or zone

When the requirements for acceptance as a CBPP free country or zone are not fulfilled, a country or zone shall be considered as infected.

Article 2.3.15.5.

Veterinary Authorities of CBPP free countries may prohibit importation or transit through their territory, from countries considered infected with CBPP, of domestic and wild bovidae.

Article 2.3.15.6.

When importing from CBPP free countries, Veterinary Authorities should require:

for domestic bovidae
Annex IX (contd)

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CBPP on the day of shipment;
2. were kept in a CBPP free country since birth or for at least the past 6 months.

Article 2.3.15.7.

When importing from CBPP free countries, Veterinary Authorities should require:

for wild bovidae

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CBPP on the day of shipment;
2. come from a CBPP free country;

if the country of origin has a common border with a country considered infected with CBPP:

3. were kept in a quarantine station for the 6 months prior to shipment.

Article 2.3.15.8.

When importing from CBPP infected countries, Veterinary Authorities should require:

for bovidae for breeding

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CBPP on the day of shipment;
2. were subjected to the complement fixation test for CBPP with negative results, on two occasions, with an interval of not less than 21 days and not more than 30 days between each test, the second test being performed within 14 days prior to shipment;
3. were isolated from other domestic bovidae from the day of the first complement fixation test until shipment;
4. were kept since birth, or for the past 6 months, in an establishment where no case of CBPP was officially reported during that period, and that the establishment was not situated in a CBPP infected zone;
5. have not been vaccinated against CBPP; or
6. were vaccinated using a vaccine complying with the standards described in the Terrestrial Manual not more than 4 months prior to shipment. In this case, the condition laid down in point 2 above is not required.

Article 2.3.15.9.

When importing from CBPP infected countries, Veterinary Authorities should require:

for bovidae for slaughter
the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CBPP on the day of shipment;

2. were kept since birth, or for the past 6 months, in an establishment where no case of CBPP was officially reported during that period, and that the establishment was not situated in a CBPP infected zone.

Article 2.3.15.10.

When importing from CBPP infected countries, Veterinary Authorities should require:

for wild bovidae

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CBPP on the day of shipment;

2. were kept, for the 180 days prior to shipment, in a quarantine station where no case of CBPP was officially reported during that period, and that the quarantine station was not situated in a CBPP infected zone;

3. have not been vaccinated against CBPP; or

4. were vaccinated using a vaccine complying with the standards described in the Terrestrial Manual not more than 4 months prior to shipment. In this case, the condition laid down in point 2 above is not required.

Article 2.3.15.11.

When importing from CBPP infected countries, Veterinary Authorities should require:

for fresh meat of bovidae

the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals:

1. which showed no lesion of CBPP;

2. which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections to rule out the presence of CBPP with favourable results.

Article 2.3.15.12.

When importing from CBPP free countries, Veterinary Authorities should require:

for in vivo derived or in vitro produced embryos/oocytes of bovidae

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of CBPP on the day of collection of the embryos/oocytes;
Annex IX (contd)

b) were kept in a CBPP free country since birth or for at least the past 6 months;

2. the oocytes were fertilised with semen meeting the conditions referred to in points a) and b) above and in Appendix 3.2.1.;

3. the embryos/oocytes were collected, processed and stored in conformity with the provisions of Appendix 3.3.1., Appendix 3.3.2. or Appendix 3.3.3., as relevant.

Article 2.3.15.13.

When importing from CBPP infected countries, Veterinary Authorities should require:

for in vivo derived or in vitro produced embryos/oocytes of bovidae

the presentation of an international veterinary certificate attesting that:

1. the donor animals:

   a) showed no clinical sign of CBPP on the day of collection of the embryos/oocytes;

   b) were subjected to the complement fixation test for CBPP with negative results, on two occasions, with an interval of not less than 21 days and not more than 30 days between each test, the second test being performed within 14 days prior to collection;

   c) were isolated from other domestic bovidae from the day of the first complement fixation test until collection;

   d) were kept since birth, or for the past 6 months, in an establishment where no case of CBPP was reported during that period, and that the establishment was not situated in a CBPP infected zone;

   e) have not been vaccinated against CBPP; or

   f) were vaccinated using a vaccine complying with the standards described in the Terrestrial Manual not more than 4 months prior to collection; in this case, the condition laid down in point b) above is not required;

2. the oocytes were fertilised with semen meeting the conditions referred to in points a) to f) above and in Appendix 3.2.1.;

3. the embryos/oocytes were collected, processed and stored in conformity with the provisions of Appendix 3.3.1., Appendix 3.3.2. or Appendix 3.3.3., as relevant.
 GUIDELINES ON SURVEILLANCE FOR CONTAGIOUS BOVINE PLEUROPNEUMONIA

1. Introduction

The Ad hoc Group on Contagious Bovine Pleuropneumonia (CBPP) Surveillance Systems held a meeting on 7-9 June 1993 with the purpose of formulating these standards, which describe surveillance systems suited to the declaration of countries and zones free of disease and free of infection. Background information is contained in the report of the meeting. In order to write these standards, the Group reviewed the following:

a) epidemiological and non-disease factors influencing the choice of CBPP surveillance systems;

b) sampling and surveillance strategies;

c) diagnostic methods applicable to CBPP surveillance systems;

d) the implications of CBPP vaccination for surveillance systems.

This last point was the subject of lengthy discussions during the meeting of the OIE Committee in May 1994. A revised text was submitted at the following meeting of the Committee (May 1995), which requested that a small group of experts formulate revised proposals. The present text is the product of their consensus.

2. Definition and purposes of surveillance

Disease surveillance is necessary to provide evidence that a country or zone is free from a disease or infection. Disease surveillance should be implemented by both:

a) a system of reporting any signs of disease activity which come to the notice of Veterinary Services or livestock owners; and

b) an active programme of examination of statistically selected samples from host populations in order to detect clinical signs or other indications of the occurrence of disease or transmission of infection.

In either case, suspicion of disease activity should be followed by quarantine, confirmatory diagnostic work and any necessary disease control measures. Surveillance thus implies that official action will follow from the discovery of evidence of disease or infection. It can be contrasted with monitoring, in which the gathering of data from the field takes place similarly, but no official action based on the findings is implied in the data-gathering activity.

Within the context of pleuropneumonia, specific measures need to be implemented, such as an exhaustive inspection of all lungs of bovines throughout the country or zone.
Annex IX (contd)

3. **Steps to be taken to declare a country free from contagious bovine pleuropneumonia**

The current goal in CBPP control is to achieve freedom from disease in particular countries and later of entire world regions, with the ultimate aim of achieving global eradication. It is therefore necessary to institute a system for verifying the steps towards these short and long-term aims, and to assist countries which wish to trade in livestock or livestock products, but face difficulties due to the presence or past occurrence of CBPP.

In conformity with the general principles for assessing disease status developed by the OIE, a four-stage process should be applied:

- intention to eradicate pleuropneumonia: the longest phase, depending on prevalence of the disease in the country or zone, geographical, socio-economic and administrative conditions, and the capacity of the animal health infrastructure;

- once a country is free from CBPP and that disease is unlikely to be re-introduced, the country can declare itself provisionally free from disease, provided it meets the criteria listed below;

- declaration of freedom from clinical CBPP, after international verification carried out under the auspices of the OIE;

- declaration of freedom from CBPP, where a country meets more stringent surveillance and control criteria.

The last three stages are strictly covered by the epidemiological surveillance methods of the OIE.

The sequence of operations differs both in terms of tactics and duration depending on whether or not the country wishing to eradicate CBPP practises vaccination.

'Disease' in the context of declaration of freedom means that the particular pathogenic agent is present and causes significant pathological effects on animals which become infected with the agent. Thus 'freedom from disease' means that there is no evidence in animals within the country or zone of any pathological effects occurring (including clinical signs) due to the presence of the agent, and from all the evidence pathogenic strains of the particular agent have been eliminated.

**COUNTRIES PRACTISING VACCINATION**

The process is summarised in the following chart:

The specific criteria proposed for each stage of this process are as follows:

a) **Provisional freedom from disease**

   For a country to declare the whole or a part of its territory provisionally free from disease, it must fulfil certain conditions, which are:

   i) no clinical or pathological evidence of CBPP should have been detected for at least 3 years;

   ii) there is an effective Veterinary Service which is able to monitor the animal health situation in the country;

   iii) there is effective meat inspection at approved abattoirs, and effective surveillance of populations in which significant numbers of slaughtered susceptible livestock are not subject to meat inspection;
iv) all evidence suggestive of CBPP is investigated by field and laboratory methods (including serological and microbiological assessment) to refute a possible diagnosis of CBPP;

v) there is an effective reporting system, both from the field to the central veterinary authority, and by that body to the OIE;

vi) there is an effective system to prevent the introduction of infection, including appropriate border control, quarantine, etc.;

vii) if vaccination has been used, all vaccination against CBPP has ceased by the date of declaration; the OIE and neighbouring countries having been notified in writing, giving the date from which vaccination was discontinued.

b) Freedom from clinical CBPP

A country which has declared itself or a zone to be provisionally free from disease may be declared by the OIE free from clinical CBPP, provided that the following criteria are met:

i) no clinical or pathological evidence of CBPP has been detected for at least 5 years;

ii) no CBPP vaccination has taken place for at least 2 years;
Annex IX (contd)

iii) the country operates surveillance and disease reporting systems for CBPP adequate to detect disease if it were present, and ensures that veterinary staff are adequately trained in the recognition of CBPP;

iv) all susceptible livestock at recognised abattoirs are subject to meat inspection procedures adequate to detect lung lesions, with diagnostic procedures to refute a possible diagnosis of CBPP;

v) there has been a programme of surveillance (using serological, pathological and microbiological techniques) for at least 2 years on any populations of susceptible domestic livestock where more than 10% of slaughtering is not subject to adequate meat inspection procedures;

vi) all evidence suggestive of CBPP is investigated by field and laboratory methods (including serological and microbiological assessment) to refute a possible diagnosis of CBPP;

vii) there are effective measures in force to prevent re-introduction of the disease.

On meeting these criteria, a country may apply to the OIE for all, or a zone, of its territory to be declared free from clinical CBPP.

An Expert Panel for the Verification of Disease Status of the OIE will evaluate the application and decide whether or not to approve it. In coming to its decision, the Expert Panel will consider evidence presented by the country and will gather information on the extent to which the criteria are met. This information-gathering will usually include sending members of the Panel to make a field visit to the country. The Expert Panel will report its findings to the OIE Scientific Commission for Animal Diseases. The Commission will report its conclusions annually to the International Committee for endorsement.

To maintain this status, a country must continue to meet these requirements until it is declared free from clinical CBPP, and must report to the OIE an annual summary of developments.

Should there be a localised temporary outbreak of disease due to re-introduction of CBPP to a country which has met, or is within 2 years of meeting, the requirements for a declaration of freedom from clinical CBPP, that country should implement a stamping-out policy, which may be supported by intensive perifocal vaccination, to eradicate the outbreak. In such circumstances if no vaccination was carried out, it will then require at least one year from the date of the last case before the country becomes eligible to apply for a declaration of freedom from clinical CBPP. If vaccination was used, this period is extended to 2 years from the date of the last case or the last vaccination (whichever occurs later). In making an application under these special circumstances, it must be shown that the outbreak did not represent endemic infection, and that the disease has been eradicated by the actions taken.

The declaration of zones to be free from clinical CBPP will not remove the requirement for the country subsequently to meet the criteria for declaration of freedom from clinical CBPP for the country as a whole; if it wishes to achieve that status, it will have to meet all of the requirements specified above before it can apply for a declaration of freedom from clinical CBPP for the entire country.

C) Freedom from CBPP

A country or a zone of its territory which has within the last 10 years either vaccinated against CBPP, or found clinical or pathological evidence of CBPP, may be declared by the OIE to be free from CBPP if the following criteria are met:
Annex IX (contd)

i) it has been declared free from clinical CBPP at least 2 years earlier, and continues to meet the requirements for this status;

ii) there has been effective abattoir surveillance for at least 4 years, covering all susceptible domestic livestock;

iii) use has been made of diagnostic procedures capable of differentiating Mycoplasma mycoides from other bovine Mycoplasma infections in the investigation of respiratory disease, and the findings are consistent with freedom from M. mycoides infection;

iv) there has been a programme of surveillance, including serological, pathological and microbiological components, for at least 3 years on any populations of susceptible domestic livestock where more than 10% of slaughter stock are not subject to adequate meat inspection procedures.

On satisfying these criteria, a country may apply to the OIE to be declared free from CBPP.

An Expert Panel for the Verification of Disease Status of the OIE will evaluate the application and decide whether or not to approve it. In coming to its decision, the Expert Panel will consider evidence presented by the country and will gather information on the extent to which the criteria are met. This information-gathering will usually include sending members of the Panel to make a field visit to the country.

The Expert Panel will report its findings to the OIE Scientific Commission for Animal Diseases. The Commission will report its conclusions annually to the International Committee for endorsement.

In the special case of a country or zone which has been considered to be continuously free from CBPP for at least 10 years, and meets all of the following requirements:

v) has not vaccinated against CBPP for at least 10 years;

vi) throughout that period found no clinical or pathological evidence of CBPP infection;

vii) had throughout that period, and undertakes to maintain permanently, an adequate disease surveillance and reporting system, covering all susceptible domestic livestock;

viii) in appropriate circumstances, made use of diagnostic procedures capable of differentiating Mycoplasma mycoides from other bovine Mycoplasma infections in the investigation of respiratory disease, with findings consistent with freedom from M. mycoides infection;

the country or zone may be declared by the OIE to be free from CBPP without the necessity to proceed through the normal intermediate steps. This declaration will be based on the conclusions of the Expert Panel for the Verification of Disease Status.

Declaration of freedom from CBPP can be made for the country as a whole, or for zones within a country.

Should there be a localised temporary outbreak of disease due to re-introduction of CBPP to a country which has met, or is within one year of meeting, the requirements for a declaration of freedom from CBPP, that country may take special measures (excluding the use of vaccination) to eradicate the outbreak. In such circumstances, it will then require at least 2 years from the date of the last case before the country becomes eligible to apply for a declaration of freedom from CBPP. In making an application under these special circumstances, the country must demonstrate that the outbreak did not represent endemic infection, and that the disease has been eradicated by the actions taken.
Annex IX (contd)

In order to maintain this status, the country must continue to operate an efficient disease surveillance and reporting system, which would detect CBPP if it occurred.

**COUNTRIES NOT PRACTISING VACCINATION**

These are generally countries with a solid animal health infrastructure (with a system for individually identifying animals) where CBPP has been accidentally introduced.

The specific criteria proposed for each stage of this process are as follows:

a)  **Provisional freedom from disease**

   A country may declare the whole or a zone of its territory provisionally free from disease one year after the last infected herds and in-contact herds have been slaughtered, on condition that:

   i)  there has been no vaccination in the country or zone for at least 2 years;

   ii) all treatment against CBPP is prohibited for sick animals or suspected cases;

   iii) a stamping-out policy is implemented after any CBPP outbreaks. Within the framework of the declaration, a minimum period of 12 months will be required after the last sick or in-contact herd has been slaughtered;

   iv) an epidemiological investigation, including serological tests, has been carried out to determine the prevalence of the disease in the country or infected zone. Special attention should be given to screening animals transported into or out of the infected herds during the 6 months preceding detection of the outbreak(s);

   v)  a system of livestock identification and movement control has been set up in the country or zone for the purposes of CBPP control and surveillance as follows:

      - all herds are officially registered and all animals of susceptible species aged over 12 months are individually identified;

      - before being moved, other than for immediate slaughter, all animals of susceptible species are to be clinically inspected and serologically tested for CBPP;

   vi) all animals of susceptible species in herds or establishments within a 3 km radius of an outbreak, and any animals with a possible epidemiological link, are individually identified, placed in quarantine for at least 6 months, and

      - all animals of susceptible species in the aforementioned herds or establishments are serological tested on two occasions at an interval of 2 to 8 weeks; microbiological investigations are to be carried out on any serologically positive animal;

      - during the quarantine period, animals in the aforementioned herds or establishments are not to be moved other than to an officially approved abattoir, where they are to be immediately slaughtered and subjected to sanitary inspection after slaughter;

      - microbiological tests should be carried out on animals presenting lesions suggestive of CBPP;
vii) surveillance is carried out in abattoirs in the contaminated country. Any lesion suggestive of CBPP should be examined microbiologically and, if the result is positive, the herd of origin must be found and subjected to serological testing;

viii) the diagnostic tests used in the country or zone comply with OIE standards and are conducted in a nationally approved laboratory.

b) **Freedom from contagious bovine pleuropneumonia**

A country or zone may be declared by the OIE to be free from CBPP 2 years after the last infected and in-contact herds have been slaughtered if the conditions listed in points a)i) to a)iviii) continue to be met.

4. **Epidemiological methods**

a) **Surveillance systems**

In demonstrating that a country or zone is free of disease, it is necessary to conduct a surveillance programme which would have a very high probability of detecting the disease if it were present. Surveillance for CBPP will include a combination of clinical, pathological, serological and microbiological methods, built around an epidemiological surveillance approach. The mix of procedures used will depend on the specific circumstances of the country or zone.
The most efficient means of detecting CBPP is through effective meat inspection procedures at abattoirs followed by laboratory examination of suspect lesions. Where a very high proportion of susceptible domestic livestock are slaughtered in controlled abattoirs, this will provide a very sensitive surveillance system covering the whole population. It is possible that structured investigation of a statistical sample of carcasses might be used to augment the routine meat inspection procedures.

Where large numbers of susceptible livestock are exported for slaughter, it may be necessary to obtain meat inspection data from the importing country.

Where a significant proportion of susceptible domestic livestock are not subject to meat inspection at the abattoir, then it will be necessary to use alternative surveillance methods based on the examination of samples of herds so as to achieve a standard probability of detection. Animals in sampled herds would be subjected to clinical examination for signs of CBPP, but not all infected animals exhibit clinical signs. Serological testing can be useful in identifying infected herds, but due to the limitations of the currently available serological tests, and the possibility that the disease may be present at very low prevalence, such surveillance systems are not very efficient in proving freedom from disease, and require large numbers of herds to be sampled.

b) Definition of sampling units

A sampling unit for the purposes of disease investigation and surveillance is defined as a group of animals in sufficiently close contact that individuals in the group would be at approximately equal risk of coming into contact with the disease agent if there were an infectious animal within the group. In most circumstances, the sampling unit would be a herd which is managed as a unit by an individual or a community, but there may be other epidemiologically appropriate groupings which are subject to regular mixing, such as all the animals belonging to residents of a village. Sampling units should normally be defined so that the majority of units contain between 50 and 1,000 animals.

c) Criteria for the stratification and sampling of host populations

'Serological surveillance would only be adopted for CBPP in circumstances where the preferred slaughterhouse surveillance system described in item 3(c) of this document could not be carried out on an adequate scale because too low a proportion of animals was slaughtered in a slaughterhouse. Thus the following system would be used as an exceptional case, rather than as the usual procedure'.

Any disease surveillance activities must be conducted on populations stratified according to disease risk, which depends principally upon the environment and management system. The cattle production systems of most countries would be categorised into between two and six strata.

Annual sample sizes must be sufficient to provide 95% probability of detecting evidence of CBPP if it were present at a prevalence of 1% of herds or other sampling units. Given perfect sensitivity of the within-herd testing procedure, this would require the examination of 300 herds from each stratum per year. However, the currently available serological tests have rather low sensitivity. The sensitivity of the test procedure at herd level is further reduced when only a sample of the herd is tested. It is possible to compensate for lower sensitivity by increasing the numbers of herds examined. The required sample size is determined by adjusting the prevalence to allow for the lack of sensitivity. For example, if there was 50% probability of detecting a sampled infected herd (sensitivity 0.5), then a true disease prevalence of 1% of herds would result in a detectable prevalence of 0.5%, and this detectable prevalence would be used to determine the required sample size.
Herds, or other sampling units, must be selected from each stratum by proper random methods, which are described in the Guide to Epidemiological Surveillance for Rinderpest published by the OIE. Any randomly selected herd must be examined in order to achieve the required probability of detection. However, this probability can often be increased by an important but unquantifiable margin by sampling additional herds based on subjective assessment of risk, or information gained during field work.

5. **Contagious bovine pleuropneumonia vaccines**

T1 strain (and its streptomycin-resistant variant) is the recommended vaccine, and the following facts are relevant to disease surveillance activities:

Current vaccines do not induce life-long immunity; the duration of protection after vaccination is about one year.

A significant proportion of vaccinated animals do not develop a serological response detectable by currently used techniques, although such animals may be protected against challenge. Where the serological response to vaccination is detectable by the complement fixation test, it usually persists for less than 3 months.

As their immunity wanes, vaccinated cattle are more likely to develop chronic lesions (sequestra) after infection.

6. **Diagnostic methods**

The diagnosis of CBPP depends on:

- clinical signs in the live animal;
- gross pathological findings;
- serological tests;
- culture and identification of the causative organism.

a) **Clinical diagnosis**

The clinical signs of CBPP may be slight or non-existent. Furthermore, the use of anti-microbial or anti-inflammatory drugs can mask the clinical expression of the disease. For these reasons, clinical signs are an unreliable indicator of the presence of the disease. However, if respiratory disease is observed in a livestock population, then the diagnosis of CBPP should be considered and confirmed or rejected on the basis of further pathological, microbiological or serological investigations.

b) **Gross pathology**

The lung lesions of CBPP are distinctive. Consequently, abattoir meat inspection is the most practical single method for maintaining CBPP surveillance. The pleura and lungs should be examined by palpation and section. A mixture of acute lesions and chronic lesions (sequestra) may be found in the same herd or even the same animal. In case of chronic infection, post-mortem diagnosis may be the only way of detecting asymptomatic animals, which may not react to serological tests.
Annex IX (contd)

c) **Serological diagnosis**

The serological test of choice is the complement fixation test (CFT). The specificity of this test can be as high as 99.5%, but the frequency of false positive reactions may temporarily be higher in certain herds. The sensitivity of the test is limited, and it may fail to identify four classes of animals:

i) animals in the very early stages of the disease;

ii) animals in the very late stages of the disease (the CFT appears to fail to detect 30% of animals containing sequestra);

iii) animals with massive lesions, where the antibodies produced are overwhelmed by the antigen;

iv) animals which have been treated in the early stages of the disease may fail to develop a detectable serological response.

Despite these limitations, the CFT is a useful herd test.

The CFT reaction after vaccination is inconstant and short-lived (generally less than 3 months).

An indirect enzyme linked immunosorbent assay (ELISA) is under field evaluation in several countries. It is at least as sensitive as the CFT, but as with other ELISA systems, increased sensitivity can only be achieved at the expense of specificity, and vice versa. It is a useful tool to measure the efficacy of vaccination programmes, as the detectable response is more reliable than the CFT, and may persist for as long as one year after vaccination.

Monoclonal and competitive ELISA systems are being developed and should offer higher specificity.

The passive haemagglutination test, while not used routinely, may have a place in serological diagnosis. It is more sensitive than the CFT in early and late stages of disease, but the specificity is lower. It has a potential role as a screening test.

The slide agglutination test is simple to perform and could be used as a pen-side test. It is more sensitive than the CFT in the early stages of the disease, but it lacks specificity.

d) **Culture and identification of the causative organism**

It is desirable that all diagnoses are confirmed by isolation of the causative organism. It may prove difficult to isolate *Mycoplasma* from chronic lesions and also after animals have been treated with anti-microbial drugs.

The causative organism is normally identified by growth inhibition tests and/or the immunofluorescence test. Closely related *Mycoplasma* may cause cross-reactions in these tests. Several new techniques which may overcome this problem are being developed, and these include immunobinding, immunoperoxidase and polymerase chain reaction (PCR) tests. These need further evaluation.
e) **Testing imported animals**

In formulating its recommendations for a system of declaration of freedom, the Group acknowledged that existing serological tests for CBPP are quite variable in sensitivity and specificity. Hence serological methods alone are unlikely to prevent the introduction of infection if live animals are imported from CBPP-infected countries. The chronic course of the disease may mean that diagnosis following introduction of CBPP may be delayed by a number of years. In the longer term there is a need for more sensitive and specific diagnostic tests. Pending the development of such tests, serological methods are necessary, but not sufficient to prevent introduction of the disease in live animals.
APPENDIX 3.8.10.

GUIDELINES FOR THE SURVEILLANCE OF BLUETONGUE

Article 3.8.10.1.

Introduction

This Appendix defines the principles and provides a guide on the surveillance for bluetongue (BT) in accordance with Appendix 3.8.1., applicable to countries seeking to demonstrate recognition for a declared BT status, with or without the use of vaccination. This may be for the entire country or zone. Guidance for countries seeking free status following an outbreak and for the maintenance of BT status is also provided. This Appendix complements Chapter 2.2.13.

BT is a vector-borne infection transmitted by different species of Culicoides insects in a range of ecosystems. An important component of BT epidemiology is vectorial capacity which provides a measure of disease risk that incorporates vector competence, abundance, biting rates, survival rates and extrinsic incubation period. However, methods and tools for measuring some of these vector factors remain to be developed, particularly in a field context. Therefore, surveillance for BT should focus on transmission in domestic ruminants.

Susceptible wild ruminant populations should be included in surveillance when these animals are intended for trade.

The impact and epidemiology of BT differ widely in different regions of the world and therefore it is impossible to provide specific guidelines for all situations. It is incumbent upon Member Countries to provide scientific data that explain the epidemiology of BT in the region concerned and adapt the surveillance strategies for defining their infection status (free, seasonally free or infected country or zone) to the local conditions. There is considerable latitude available to Member Countries to justify their infection status at an acceptable level of confidence.

Surveillance for BT should be in the form of a continuing programme.

Article 3.8.10.2.

Case definition

For the purposes of surveillance, a case refers to an animal infected with BT virus (BTV).

For the purposes of international trade, a distinction must be made between a case as defined below and an animal that is potentially infectious to vectors. The conditions for trade are defined in Chapter 2.2.13. of the Terrestrial Code.

The purpose of surveillance is the detection of virus circulation in a country or zone and not determination of the status of an individual animal or herds. Surveillance deals not only with the occurrence of clinical signs caused by BTV, but also with the evidence of infection with BTV in the absence of clinical signs.

The following defines the occurrence of BTV infection:

1. BTV has been isolated and identified as such from an animal or a product derived from that animal, or
Annex X (contd)

2. viral antigen or viral ribonucleic acid (RNA) specific to one or more of the serotypes of BTV has been identified in samples from one or more animals showing clinical signs consistent with BT, or epidemiologically linked to a confirmed or suspected case, or giving cause for suspicion of previous association or contact with BTV, or

3. antibodies to structural or nonstructural proteins of BTV that are not a consequence of vaccination have been identified in one or more animals that either show clinical signs consistent with BT, or epidemiologically linked to a confirmed or suspected case, or give cause for suspicion of previous association or contact with BTV

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 3.8.10.3.

General conditions and methods

1. A surveillance system in accordance with Appendix 3.8.1. should be under the responsibility of the Veterinary Authority. In particular:

   a) a formal and ongoing system for detecting and investigating outbreaks of disease should be in place;

   b) a procedure should be in place for the rapid collection and transport of samples from suspect cases of BT to a laboratory for BT diagnosis as described in the Terrestrial Manual;

   c) a system for recording, managing and analysing diagnostic and surveillance data should be in place.

2. The BT surveillance programme should:

   a) in a country/zone free or seasonally free, include an early warning system for reporting suspicious cases. Farmers and workers, who have day-to-day contact with domestic ruminants, as well as diagnosticians, should report promptly any suspicion of BT to the Veterinary Authority. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Authority. An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is BTV. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. All suspected cases of BT should be investigated immediately and samples should be taken and submitted to an approved laboratory. This requires that sampling kits and other equipment are available for those responsible for surveillance;

   b) conduct random or targeted serological and virological surveillance appropriate to the infection status of the country or zone.

Generally, the conditions to prevent exposure of susceptible animals to BTV infected vectors will be difficult to apply. However, under specific situations, establishments such as artificial insemination centres or quarantine stations exposure to vectors may be preventable. The testing requirements for animals kept in these facilities are described in Articles 2.2.13.11. and 2.2.13.15.
Article 3.8.10.4.

Surveillance strategies

The target population for surveillance aimed at identification of disease and/or infection should cover susceptible domestic ruminants within the country or zone. Active and passive surveillance for BTV infection should be ongoing. Surveillance should be composed of random or targeted approaches using virological, serological and clinical methods appropriate for the infection status of the country or zone.

The strategy employed may be based on surveillance using randomised sampling that would demonstrate the absence of BTV infection at an acceptable level of confidence. The frequency of sampling should be dependent on the epidemiological situation. Random surveillance is conducted using serological tests described in the Terrestrial Manual. Positive serological results may be followed up with virological methods as appropriate.

Targeted surveillance (e.g. based on the increased likelihood of infection in particular localities or species) may be an appropriate strategy. Virological and serological methods may be used concurrently to define the BTV status of targeted populations.

A country should justify the surveillance strategy chosen as being adequate to detect the presence of BTV infection in accordance with Appendix 3.8.1. and the prevailing epidemiological situation. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clinical signs (e.g. sheep). Similarly, virological and serological testing may be targeted to species that rarely show clinical signs (e.g. cattle).

In vaccinated populations, serological and virological surveillance is necessary to detect the BTV types circulating to ensure that all circulating types are included in the vaccination programme.

If a Member wishes to declare freedom from BTV infection in a specific zone, the design of the surveillance strategy would need to be aimed at the population within the zone.

For random surveys, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect evidence of infection if it were to occur at a predetermined minimum rate. The sample size and expected prevalence determine the level of confidence in the results of the survey. The applicant country must justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Appendix 3.8.1. Selection of the design prevalence in particular needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey approach selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and the different species in the target population.

Irrespective of the testing system employed, surveillance system design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of infection or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as those which may be epidemiologically linked to it.
Annex X (contd)

The principles involved in surveillance for disease infection are technically well defined. The design of surveillance programmes to prove the absence of BTV infection/circulation needs to be carefully followed to avoid producing results that are either insufficiently reliable to be accepted by international trading partners, or excessively costly and logistically complicated. The design of any surveillance programme, therefore, requires inputs from professionals competent and experienced in this field.

1. Clinical surveillance

Clinical surveillance aims at the detection of clinical signs of BT at the flock/herd level. Whereas significant emphasis is placed on the diagnostic value of mass serological screening, surveillance based on clinical inspection should not be underrated, particularly during a newly introduced infection. In sheep and occasionally goats, clinical signs may include oedema, hyperaemia of mucosal membranes, coronitis and cyanotic tongue.

BT suspects detected by clinical surveillance should always be confirmed by laboratory testing.

2. Serological surveillance

An active programme of surveillance of host populations to detect evidence of BTV transmission is essential to establish BTV status in a country or zone. Serological testing of ruminants is one of the most effective methods of detecting the presence of BTV. The species tested depends on the epidemiology of BTV infection, and the species available, in the local area. Cattle are usually the most sensitive indicator species. Management variables that may influence likelihood of infection, such as the use of insecticides and animal housing, should be considered.

Surveillance may include serological surveys, for example abattoir surveys, the use of cattle as sentinel animals (which must be individually identifiable), or a combination of methods.

The objective of serological surveillance is to detect evidence of BTV circulation. Samples should be examined for antibodies against BTV using tests prescribed in the Terrestrial Manual. Positive BTV antibody tests results can have four possible causes:

a) natural infection with BTV,

b) vaccination against BTV,

c) maternal antibodies,

d) positive results due to the lack of specificity of the test.

It may be possible to use sera collected for other survey purposes for BTV surveillance. However, the principles of survey design described in these guidelines and the requirements for a statistically valid survey for the presence of BTV infection should not be compromised.

The results of random or targeted serological surveys are important in providing reliable evidence that no BTV infection is present in a country or zone. It is, therefore, essential that the survey is thoroughly documented. It is critical to interpret the results in light of the movement history of the animals being sampled.

Serological surveillance in a free zone should target those areas that are at highest risk of BTV transmission, based on the results of previous surveillance and other information. This will usually be towards the boundaries of the free zone. In view of the epidemiology of BTV infection, either random or targeted sampling is suitable to select herds and/or animals for testing.
A surveillance zone within a free country or zone should separate it from a potentially infected country or zone. Serological surveillance in a free country or zone should be carried out over an appropriate distance from the border with a potentially infected country or zone, based upon geography, climate, history of infection and other relevant factors.

Serological surveillance in infected zones will identify changes in the boundary of the zone, and can also be used to identify the BTV types circulating. In view of the epidemiology of BTV infection, either random or targeted sampling is suitable.

3. **Virological surveillance**

Isolation and genetic analysis of BTV from a proportion of infected animals is beneficial in terms of providing information on serotype and genetic characteristics of the viruses concerned.

Virological surveillance using tests described in the Terrestrial Manual can be conducted:

a) to identify virus circulation in at risk populations,

b) to confirm clinically suspect cases,

c) to follow up positive serological results,

d) to better characterize the genotype of circulating virus in a country or zone.

4. **Sentinel animals**

Sentinel animals are a form of targeted surveillance with a prospective study design. They are the preferred strategy for BTV surveillance. They comprise groups of unexposed animals managed at fixed locations and sampled regularly to detect new BTV infections.

The primary purpose of a sentinel animal programme is to detect BTV infections occurring at a particular place, for instance sentinel groups may be located on the usual boundaries of infected zones to detect changes in distribution of BTV. In addition, sentinel animal programmes allow the timing and dynamics of infections to be observed.

A sentinel animal programme should use animals of known source and history of exposure, control management variables such as use of insecticides and animal housing (depending on the epidemiology of BTV in the area under consideration), and be flexible in its design in terms of sampling frequency and choice of tests.

Care is necessary in choosing the sites for the sentinel groups. The aim is to maximise the chance of detecting BTV activity at the geographical location for which the sentinel site acts as a sampling point. The effect of secondary factors that may influence events at each location, such as climate, may also be analysed. To avoid confounding factors, sentinel groups should comprise animals selected to be of similar age and susceptibility to BTV infection. Cattle are the most appropriate sentinels but other domestic ruminant species may be used. The only feature distinguishing groups of sentinels should be their geographical location.

Sera from sentinel animal programmes should be stored methodically in a serum bank to allow retrospective studies to be conducted in the event of new serotypes being isolated.
The frequency of sampling will depend on the reason for choosing the sampling site. In endemic areas, virus isolation will allow monitoring of the serotypes and genotypes of BTV circulating during each time period. The borders between infected and non infected areas can be defined by serological detection of infection. Monthly sampling intervals are frequently used. Sentinels in declared free zones add to confidence that BTV infections are not occurring unobserved. In such cases, sampling prior to and after the possible period of transmission is sufficient.

Definitive information on BTVs circulating in a country or zone is provided by isolation and identification of the viruses. If virus isolation is required, sentinels should be sampled at sufficiently frequent intervals to ensure that samples are collected during the period of viraemia.

5. Vector surveillance

BTV is transmitted between ruminant hosts by species of Culicoides which vary across the world. It is therefore important to be able to identify potential vector species accurately although many such species are closely related and difficult to differentiate with certainty.

The main purpose of vector surveillance is to define high, medium and low-risk areas and local details of seasonality by determining the various species present in an area, their respective seasonal occurrence, and abundance. Vector surveillance has particular relevance to potential areas of spread. Long term surveillance can also be used to assess vector suppression measures.

The most effective way of gathering this information should take account of the biology and behavioural characteristics of the local vector species of Culicoides and may include the use of Onderstepoort-type light traps or similar, operated from dusk to dawn in locations adjacent to domestic ruminants, or the use of drop traps over ruminant animals.

Vector surveillance should be based on scientific sampling techniques. The choice of the number and type of traps to be used in vector surveillance and the frequency of their use should take into account the size and ecological characteristics of the area to be surveyed.

The operation of vector surveillance sites at the same locations as sentinel animals is advisable.

The use of a vector surveillance system to detect the presence of circulating virus is not recommended as a routine procedure as the typically low vector infection rates mean that such detections can be rare. Other surveillance strategies (e.g. the use of sentinel animals of domestic ruminants) are preferred to detect virus circulation.

Article 3.8.10.5.

Documentation of BTV infection free status

1. Countries declaring freedom from BTV infection for the country or zone

In addition to the general conditions described in Chapter 2.2.13. of the Terrestrial Code, a Member declaring freedom from BTV infection for the entire country or a zone should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and should be planned and implemented according to general conditions and methods described in this Appendix, to demonstrate absence of BTV infection during the preceding 24 months in susceptible domestic ruminant populations. This requires the support of a laboratory able to undertake identification of BTV infection through virus detection and antibody tests described in the Terrestrial Manual. This surveillance should be targeted to non-vaccinated animals. Clinical surveillance may be effective in sheep while serological surveillance is more appropriate in cattle.
2. Additional requirements for countries or zones that practise vaccination

Vaccination to prevent the transmission of BTV may be part of a disease control programme. The level of flock or herd immunity required to prevent transmission will depend on the flock or herd size, composition (e.g. species) and density of the susceptible population. It is therefore impossible to be prescriptive. The vaccine must also comply with the provisions stipulated for BTV vaccines in the Terrestrial Manual. Based on the epidemiology of BTV infection in the country or zone, it may be that a decision is reached to vaccinate only certain species or other subpopulations.

In countries or zones that practise vaccination, there is a need to perform virological and serological tests to ensure the absence of virus circulation. These tests should be performed on non-vaccinated subpopulations or on sentinels. The tests have to be repeated at appropriate intervals according to the purpose of the surveillance programme. For example, longer intervals may be adequate to confirm endemicity, while shorter intervals may allow on-going demonstration of absence of transmission.

Article 3.8.10.6.

The use and interpretation of serological and virus detection tests

1. Serological testing

Ruminants infected with BTV produce antibodies to structural and non-structural viral proteins, as do animals vaccinated with current modified live virus vaccines. Antibodies to the BTV serogroup antigen are detected with high sensitivity and specificity by competitive ELISA (c-ELISA) and to a lesser extent by AGID as described in the Terrestrial Manual. Positive c-ELISA results can be confirmed by neutralization assay to identify the infecting serotype(s); however, BTV infected ruminants can produce neutralizing antibodies to serotypes of BTV other than those to which they were exposed (false positive results), especially if they have been infected with multiple serotypes.

2. Virus detection

The presence of BTV in ruminant blood and tissues can be detected by virus isolation or polymerase chain reaction (PCR) as described in the Terrestrial Manual.

Interpretation of positive and negative results (both true and false) differs markedly between these tests because they detect different aspects of BTV infection, specifically (1) infectious BTV (virus isolation) and (2) nucleic acid (PCR). The following are especially relevant to interpretation of PCR assays:

a) The nested PCR assay detects BTV nucleic acid in ruminants long after the clearance of infectious virus. Thus positive PCR results do not necessarily coincide with active infection of ruminants. Furthermore, the nested PCR assay is especially prone to template contamination, thus there is considerable risk of false positive results.

b) PCR procedures other than real time PCR allow sequence analysis of viral amplicons from ruminant tissues, insect vectors or virus isolates. These sequence data are useful for creating data bases to facilitate important epidemiological studies, including the possible distinction of field and vaccine virus strains of BTV, genotype characterization of field strains of BTV, and potential genetic divergence of BTV relevant to vaccine and diagnostic testing strategies.

It is essential that BTV isolates are sent regularly to the OIE Reference Laboratories for genetic and antigenic characterization.
Annex X (contd)

**Fig. 1. Application of laboratory tests in veterinary surveillance**

**Fig. 2. Application of laboratory tests in veterinary surveillance**

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CHAPTER 2.3.3.

BOVINE TUBERCULOSIS

Article 2.3.3.1.

The recommendations in this Chapter are intended to manage the human and animal health risks associated with Mycobacterium bovis (M. bovis) infection in domestic (permanently captive and owned free-range) bovines including cattle (Bos taurus, B. indicus and B. grunniens), water buffaloes (Bubalus bubalis) and wood bison (Bison bison and B. bison) and in farmed deer (red, wapiti, sika, samba, rusa, fallow, white-tailed, black-tailed and mule deer Cervus elaphus, C. canadensis, C. nippon, C. unicolor unicolor, C. timorensis, Dama dama dama, Odocoileus virginianus borealis, Odocoileus hemionus columbianus and Odocoileus hemionus hemionus).

When authorising import or transit of the following commodities, Veterinary Authorities should comply with the requirements prescribed in this Chapter relevant to the status of bovine tuberculosis in the exporting country, zone or compartment:

1. live animals;
2. semen, ova and in vivo derived embryos collected and handled in accordance with the recommendations of the International Embryo Transfer Society;
3. meat and meat products;
4. milk and milk products;
5. antler velvet.

Standards for diagnostic tests are described in the Terrestrial Manual.

Article 2.3.3.2.

Country, zone or compartment free from bovine tuberculosis

To qualify as free from bovine tuberculosis, a country, zone or compartment should satisfy the following requirements:

1. M. bovis infection in domestic (permanently captive and owned free-range) bovines including cattle (Bos taurus, B. indicus and B. grunniens), water buffalo (Bubalus bubalis) and wood bison (Bison bison and B. bison) and in farmed deer as specified in Article 2.3.3.1 is a notifiable disease in the country;
2. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of clinical tuberculosis;
3. surveillance programme, involving regular and periodic testing of all cattle, water buffalo, and wood bison and farmed deer herds and capable of detecting infection at an annual period prevalence of 0.2% of herds and 0.1% of animals with 95% confidence has failed to detect infection shown that at least 99.8% of the herds and 99.9% of the animals in the country, zone or compartment have been found free from bovine tuberculosis and the percentage of herds confirmed infected with M. bovis has not exceeded 0.1% per year for 3 consecutive years;
4. a surveillance programme should be in place to detect bovine tuberculosis in the country, zone or compartment, through monitoring at slaughter ante-mortem and post-mortem inspection based on the inspection as described in Article Appendix 2.3.3.8. 3.10.1.;

5. if the surveillance programme described in points 3 and 4 above has failed to detect M. bovis for 3 consecutive years, surveillance may be maintained through monitoring at slaughter alone;

56. cattle, water buffalo and wood bison and farmed deer introduced into a country, zone or compartment free from bovine tuberculosis should be accompanied by a certificate from an official veterinarian attesting that they come from a country, zone or compartment or herd free from bovine tuberculosis or comply with the relevant provisions in Article 2.3.3.4. or in Article 2.3.3.5.

Article 2.3.3.3.

Herd free from bovine tuberculosis

To qualify as free from bovine tuberculosis, a herd of cattle, water buffalo, and wood bison or farmed deer should satisfy the following requirements:

1. the herd is in a country, zone or compartment free from bovine tuberculosis and is certified free by the veterinary authority; or

2. cattle, water buffalo and, wood bison and farmed deer in the herd:
   a) showed no clinical sign of bovine tuberculosis;
   b) over 6 weeks of age, have shown a negative result to at least two tuberculin tests carried out at an interval of a minimum of 6 months, the first test being performed at 6 months following the slaughter of the last affected animal;
   c) showed a negative result to an annual tuberculin test to ensure the continuing absence of bovine tuberculosis; or
      i) showed a negative result to a tuberculin test every 2 years to ensure the continuing absence of bovine tuberculosis if the annual percentage of herds confirmed as infected with tuberculosis is not more than 1% of all herds in the country or zone during the last 2 years; or
      ii) showed a negative result to a tuberculin test every 3 years to ensure the continuing absence of bovine tuberculosis if the annual percentage of herds confirmed as infected with tuberculosis is not more than 0.2% of all herds in the country or zone during the last 4 years; or
      iii) showed a negative result to a tuberculin test every 4 years to ensure the continuing absence of bovine tuberculosis if the annual percentage of herds confirmed as infected with tuberculosis is not more than 0.1% of all herds in the country or zone during the last 6 years;

3. cattle, water buffalo, and wood bison and farmed deer introduced into the herd come from a herd free from bovine tuberculosis. This condition may be waived for animals which have been isolated and which, prior to entry into the herd, were subjected to at least two tuberculin tests carried out at a 6-month interval with negative results.

Article 2.3.3.4.

Veterinary authorities of importing countries should require:
 Annex XI (contd)

for cattle, water buffalo and, wood bison and farmed deer for breeding or rearing

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of bovine tuberculosis on the day of shipment;

2. originate from a herd free from bovine tuberculosis that is in a country, zone or compartment free from bovine tuberculosis; or

3. were subjected to the tuberculin test for bovine tuberculosis with negative results during the 30 days prior to shipment and come from a herd free from bovine tuberculosis; or

4. have been isolated and prior to entry into the herd were subjected to at least two tuberculin tests carried out at a six-month interval with negative results.

Article 2.3.3.5.

Veterinary Authorities of importing countries should require:

for cattle, water buffalo and, wood bison and farmed deer for slaughter

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of bovine tuberculosis on the day of shipment;

2. originated from a herd free from bovine tuberculosis or were subjected to a tuberculin test for bovine tuberculosis with negative results during the 30 days prior to shipment; or

3. were not being eliminated as part of an eradication programme against bovine tuberculosis.

Article 2.3.3.6.

Veterinary Authorities of importing countries should require:

for semen of cattle, water buffalo and, wood bison and farmed deer

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of bovine tuberculosis on the day of collection of the semen;
   b) were kept in an artificial insemination centre free from bovine tuberculosis in a country, zone or compartment free from bovine tuberculosis and which only accepts animals from free herds in a free country, zone or compartment; or
   c) showed negative results to tuberculin tests carried out annually and were kept in a herd free from bovine tuberculosis;

2. the semen was collected, processed and stored in conformity with the provisions of Appendix 3.2.1.

Article 2.3.3.7.

Veterinary Authorities of importing countries should require:
Annex XI (contd)

for embryos/ova of cattle, water buffalo and wood bison and farmed deer

the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) and all other susceptible animals in the herd of origin showed no clinical sign of bovine tuberculosis during the 24 hours prior to embryo collection;
   b) originated from a herd free from bovine tuberculosis in a country, zone or compartment free from bovine tuberculosis; or
   c) were kept in a herd free from bovine tuberculosis, and were subjected to a tuberculin test for bovine tuberculosis with negative results during an isolation period of 30 days in the establishment of origin prior to departure to the collection centre;

2. the embryos/ova were collected, processed and stored in conformity with the provisions of Appendix 3.3.1., Appendix 3.3.2. or Appendix 3.3.3., as relevant.

Article 2.3.3.8.

Veterinary Authorities of importing countries should require:

for fresh meat and meat products of cattle, water buffalo and wood bison and farmed deer

the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which have been subjected to ante-mortem and post-mortem inspections as described in Appendix 3.10.1.

Article 2.3.3.9.

Veterinary Authorities of importing countries should require:

for milk and milk products of cattle, water buffalo and wood bison

the presentation of an international veterinary certificate attesting that the consignment:

1. has been derived from animals in a herd free from bovine tuberculosis; or
2. was subjected to pasteurization; or
3. was subjected to a combination of control measures with equivalent performance as described in the Codex Alimentarius Code of Hygienic Practice for Milk and Milk Products.

Article 2.3.3.10.

Veterinary Authorities of importing countries should require:

for antler velvet of farmed deer

the presentation of an international veterinary certificate attesting that the consignment:

1. has been derived from animals in a herd free from bovine tuberculosis; or
2. has been cooked at 60º C for 3 hours, or an equivalent temperature/time treatment.

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CHAPTER 2.3.13.

BOVINE SPONGIFORM ENCEPHALOPATHY

Article 2.3.13.1.

The recommendations in this Chapter are intended to manage the human and animal health risks associated with the presence of the bovine spongiform encephalopathy (BSE) agent in cattle (Bos taurus and B. indicus) only.

1. When authorising import or transit of the following commodities and any products made from these commodities and containing no other tissues from cattle, Veterinary Authorities should not require any BSE related conditions, regardless of the BSE risk status of the cattle population of the exporting country, zone or compartment:

   a) milk and milk products;

   b) semen and in vivo derived cattle embryos collected and handled in accordance with the recommendations of the International Embryo Transfer Society;

   c) hides and skins;

   d) gelatine and collagen prepared exclusively from hides and skins;

   e) protein-free tallow (maximum level of insoluble impurities of 0.15% in weight) and derivatives made from this tallow;

   f) dicalcium phosphate (with no trace of protein or fat);

   g) deboned skeletal muscle meat (excluding mechanically separated meat) from cattle 30 months of age or less, which were not subjected to a stunning process prior to slaughter, with a device injecting compressed air or gas into the cranial cavity or to a pithing process, and which passed ante-mortem and post-mortem inspections and which has been prepared in a manner to avoid contamination with tissues listed in Article 2.3.13.14.;

   h) blood and blood by-products, from cattle which were not subjected to a stunning process, prior to slaughter, with a device injecting compressed air or gas into the cranial cavity, or to a pithing process.

2. When authorising import or transit of other commodities listed in this Chapter, Veterinary Authorities should require the conditions prescribed in this Chapter relevant to the BSE risk status of the cattle population of the exporting country, zone or compartment.

Standards for diagnostic tests are described in the Terrestrial Manual.
Article 2.3.13.2.

The BSE risk status of the cattle population of a country, zone or compartment should be determined on the basis of the following criteria:

1. the outcome of a risk assessment, based on Section 1.3., identifying all potential factors for BSE occurrence and their historic perspective. Countries should review the risk assessment annually to determine whether the situation has changed.

   a) Release assessment

   Release assessment consists of assessing, through consideration of the following, the likelihood that the BSE agent has either been introduced into the country, zone or compartment via commodities potentially contaminated with it, or is already present in the country, zone or compartment:

   i) the presence or absence of the BSE agent in the indigenous ruminant population of the country, zone or compartment and, if present, evidence regarding its prevalence;

   ii) production of meat-and-bone meal or greaves from the indigenous ruminant population;

   iii) imported meat-and-bone meal or greaves;

   iv) imported cattle, sheep and goats;

   v) imported animal feed and feed ingredients;

   vi) imported products of ruminant origin for human consumption, which may have contained tissues listed in Article 2.3.13.14. and may have been fed to cattle;

   vii) imported products of ruminant origin intended for in vivo use in cattle.

   The results of any epidemiological investigation into the disposition of the commodities identified above should be taken into account in carrying out the assessment.

   b) Exposure assessment

   If the release assessment identifies a risk factor, an exposure assessment should be conducted, consisting of assessing the likelihood of cattle being exposed to the BSE agent, through a consideration of the following:

   i) recycling and amplification of the BSE agent through consumption by cattle of meat-and-bone meal or greaves of ruminant origin, or other feed or feed ingredients contaminated with these;

   ii) the use of ruminant carcasses (including from fallen stock), by-products and slaughterhouse waste, the parameters of the rendering processes and the methods of animal feed manufacture;

   iii) the feeding or not of ruminants with meat-and-bone meal and greaves derived from ruminants, including measures to prevent cross-contamination of animal feed;

   iv) the level of surveillance for BSE conducted on the cattle population up to that time and the results of that surveillance;
2. on-going awareness programme for veterinarians, farmers, and workers involved in transportation, marketing and slaughter of cattle to encourage reporting of all cases showing clinical signs consistent with BSE in target sub-populations as defined in Appendix 3.8.4.;

3. the compulsory notification and investigation of all cattle showing clinical signs consistent with BSE;

4. the examination carried out in accordance with the Terrestrial Manual in an approved laboratory of brain or other tissues collected within the framework of the aforementioned surveillance and monitoring system.

When the risk assessment demonstrates negligible risk, the country should conduct Type B surveillance in accordance with Appendix 3.8.4.

When the risk assessment fails to demonstrate negligible risk, the country should conduct Type A surveillance in accordance with Appendix 3.8.4.

Article 2.3.13.3.

Negligible BSE risk

Commodities from the cattle population of a country, zone or compartment pose a negligible risk of transmitting the BSE agent if the following conditions are met:

1. a risk assessment, as described in point 1 of Article 2.3.13.2., has been conducted in order to identify the historical and existing risk factors, and the country has demonstrated that appropriate specific measures have been taken for the relevant period of time defined below to manage each identified risk;

2. the country has demonstrated that Type B surveillance in accordance with Appendix 3.8.4. is in place and the relevant points target, in accordance with Table 1, has been met;

3. EITHER:
   a) there has been no case of BSE or, if there has been a case, every case of BSE has been demonstrated to have been imported and has been completely destroyed, and
      i) the criteria in points 2 to 4 of Article 2.3.13.2. have been complied with for at least 7 years; and
      ii) it has been demonstrated through an appropriate level of control and audit that for at least 8 years neither meat-and-bone meal nor greaves derived from ruminants has been fed to ruminants;
   OR
   b) if there has been an indigenous case, every indigenous case was born more than 11 years ago; and
      i) the criteria in points 2 to 4 of Article 2.3.13.2. have been complied with for at least 7 years; and
      ii) it has been demonstrated through an appropriate level of control and audit that for at least 8 years neither meat-and-bone meal nor greaves derived from ruminants has been fed to ruminants; and
Annex XII (contd)

iii) all BSE cases, as well as:

- all cattle which, during their first year of life, were reared with the BSE cases during their first year of life, and which investigation showed consumed the same potentially contaminated feed during that period, or

- if the results of the investigation are inconclusive, all cattle born in the same herd as, and within 12 months of the birth of, the BSE cases,

if alive in the country, zone or compartment, are permanently identified, and their movements controlled, and, when slaughtered or at death, are completely destroyed.

The country or zone will be included in the list of negligible risk only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information above be re-submitted annually and changes in the epidemiological situation or other significant events should be reported promptly to the OIE.

Article 2.3.13.4.

Controlled BSE risk

Commodities from the cattle population of a country, zone or compartment pose a controlled risk of transmitting the BSE agent if the following conditions are met:

1. a risk assessment, as described in point 1 of Article 2.3.13.2., has been conducted in order to identify the historical and existing risk factors and the country has demonstrated that appropriate measures are being taken to manage all identified risks, but these measures have not been taken for the relevant period of time;

2. the country has demonstrated that Type A surveillance in accordance with Appendix 3.8.4. has been carried out and the relevant points target, in accordance with Table 1, has been met; Type B surveillance may replace Type A surveillance once the relevant points target is met;

3. EITHER:

   a) there has been no case of BSE or, if there has been a case, every case of BSE has been demonstrated to have been imported and has been completely destroyed, the criteria in points 2 to 4 of Article 2.3.13.2. are complied with and it can be demonstrated through an appropriate level of control and audit that neither meat-and-bone meal nor greaves derived from ruminants has been fed to ruminants, but at least one of the following two conditions applies:

      i) the criteria in points 2 to 4 of Article 2.3.13.2. have not been complied with for 7 years;

      ii) it cannot be demonstrated that controls over the feeding of meat-and-bone meal or greaves derived from ruminants to ruminants have been in place for 8 years;

   OR

   b) there has been an indigenous case of BSE, the criteria in points 2 to 4 of Article 2.3.13.2. are complied with, and it can be demonstrated through an appropriate level of control and audit that neither meat-and-bone meal nor greaves derived from ruminants has been fed to ruminants, but at least one of the following two conditions applies:
Annex XII (contd)

i) the criteria in points 2 to 4 of Article 2.3.13.2. have not been complied with for 7 years;
ii) it cannot be demonstrated that controls over the feeding of meat-and-bone meal and greaves derived from ruminants to ruminants have been in place for 8 years;

AND

iii) and all BSE cases, as well as:
- all cattle which, during their first year of life, were reared with the BSE cases during their first year of life, and which investigation showed consumed the same potentially contaminated feed during that period, or
- if the results of the investigation are inconclusive, all cattle born in the same herd as, and within 12 months of the birth of, the BSE cases, if alive in the country, zone or compartment, are permanently identified, and their movements controlled, and, when slaughtered or at death, are completely destroyed.

The country or zone will be included in the list of controlled risk only after the submitted evidence has been accepted by the OIE. Retention on the list requires that the information above be re-submitted annually and changes in the epidemiological situation or other significant events should be reported promptly to the OIE.

Article 2.3.13.5.

Undetermined BSE risk

The cattle population of a country, zone or compartment poses an undetermined BSE risk if it cannot be demonstrated that it meets the requirements of another category.

Article 2.3.13.6.

When importing from a country, zone or compartment posing a negligible BSE risk, Veterinary Authorities should require:

for all commodities from cattle not listed in point 1 of Article 2.3.13.1, the presentation of an international veterinary certificate attesting that the country, zone or compartment complies with the conditions in Article 2.3.13.3.

Article 2.3.13.7.

When importing from a country, zone or compartment posing a negligible BSE risk, but where there has been an indigenous case, Veterinary Authorities should require:

for cattle selected for export, the presentation of an international veterinary certificate attesting that the animals:
1. are identified by a permanent identification system in such a way as to demonstrate that they are not exposed cattle as described in point 3.b(iii) of Article 2.3.13.3;
2. were born after the date from which the ban on the feeding of ruminants with meat-and-bone meal and greaves derived from ruminants had been effectively enforced.
Annex XII (contd)

Article 2.3.13.8.

When importing from a country, zone or compartment posing a controlled BSE risk, Veterinary Authorities should require:

for cattle

the presentation of an international veterinary certificate attesting that:

1. the country, zone or compartment complies with the conditions referred to in Article 2.3.13.4.;

2. cattle selected for export are identified by a permanent identification system in such a way as to demonstrate that they are not exposed cattle as described in point 3.b)iii) of Article 2.3.13.4.;

3. cattle selected for export were born after the date from which the ban on the feeding of ruminants with meat-and-bone meal and greaves derived from ruminants was effectively enforced.

Article 2.3.13.9.

When importing from a country, zone or compartment with an undetermined BSE risk, Veterinary Authorities should require:

for cattle

the presentation of an international veterinary certificate attesting that:

1. the feeding of ruminants with meat-and-bone meal and greaves derived from ruminants has been banned and the ban has been effectively enforced;

2. all BSE cases, as well as:

   a) all cattle which, during their first year of life, were reared with the BSE cases during their first year of life, and, which investigation showed consumed the same potentially contaminated feed during that period, or

   b) if the results of the investigation are inconclusive, all cattle born in the same herd as, and within 12 months of the birth of, the BSE cases,

      if alive in the country, zone or compartment, are permanently identified, and their movements controlled, and, when slaughtered or at death, are completely destroyed;

3. cattle selected for export:

   a) are identified by a permanent identification system in such a way as to demonstrate that they are not exposed cattle as demonstrated in point 2 above;

   b) were born at least 2 years after the date from which the ban on the feeding of ruminants with meat-and-bone meal and greaves derived from ruminants was effectively enforced.
Annex XII (contd)

Article 2.3.13.10.
When importing from a country, zone or compartment posing a negligible BSE risk, Veterinary Authorities should require:

for fresh meat and meat products from cattle (other than those listed in point 1 of Article 2.3.13.1.)

the presentation of an international veterinary certificate attesting that:

1. the country, zone or compartment complies with the conditions in Article 2.3.13.3.;

2. the cattle from which the fresh meat and meat products were derived passed ante-mortem and post-mortem inspections;

3. in countries with negligible BSE risk where there have been indigenous cases, the cattle from which the fresh meat and meat products were derived were born after the date from which the ban on the feeding of ruminants with meat-and-bone meal and greaves derived from ruminants had been effectively enforced.

Article 2.3.13.11.
When importing from a country, zone or compartment with a controlled BSE risk, Veterinary Authorities should require:

for fresh meat and meat products from cattle (other than those listed in point 1 of Article 2.3.13.1.)

the presentation of an international veterinary certificate attesting that:

1. the country, zone or compartment complies with the conditions referred to in Article 2.3.13.4.;

2. the cattle from which the fresh meat and meat products were derived passed ante-mortem and post-mortem inspections;

3. cattle from which the fresh meat and meat products destined for export were derived were not subjected to a stunning process, prior to slaughter, with a device injecting compressed air or gas into the cranial cavity, or to a pithing process;

4. the fresh meat and meat products were produced and handled in a manner which ensures that such products do not contain and are not contaminated with:

   a) the tissues listed in points 1 and 2 of Article 2.3.13.14.;

   b) mechanically separated meat from the skull and vertebral column from cattle over 30 months of age.

Article 2.3.13.12.
When importing from a country, zone or compartment with an undetermined BSE risk, Veterinary Authorities should require:

for fresh meat and meat products from cattle (other than those listed in point 1 of Article 2.3.13.1.)

the presentation of an international veterinary certificate attesting that:
Annex XII (contd)

1. the cattle from which the fresh meat and meat products originate:
   a) have not been fed meat-and-bone meal or gravis derived from ruminants;
   b) passed ante-mortem and post-mortem inspections;
   c) were not subjected to a stunning process, prior to slaughter, with a device injecting compressed air or gas into the cranial cavity, or to a pithing process;

2. the fresh meat and meat products were produced and handled in a manner which ensures that such products do not contain and are not contaminated with:
   a) the tissues listed in points 1 and 3 of Article 2.3.13.14.;
   b) nervous and lymphatic tissues exposed during the deboning process;
   c) mechanically separated meat from the skull and vertebral column from cattle over 12 months of age.

Article 2.3.13.13.

1. Ruminant-derived meat-and-bone meal or gravis, or any commodities containing such products, which originate from a country, zone or compartment defined in Article 2.3.13.3., but where there has been an indigenous case of BSE, should not be traded if such products were derived from cattle born before the date from which the ban on the feeding of ruminants with meat-and-bone meal and gravis derived from ruminants had been effectively enforced.

2. Ruminant-derived meat-and-bone meal or gravis, or any commodities containing such products, which originate from a country, zone or compartment defined in Articles 2.3.13.4. and 2.3.13.5. should not be traded between countries.

Article 2.3.13.14.

1. From cattle of any age originating from a country, zone or compartment defined in Articles 2.3.13.4. and 2.3.13.5., the following commodities, and any commodity contaminated by them, should not be traded for the preparation of food, feed, fertilisers, cosmetics, pharmaceuticals including biologicals, or medical devices: tonsils and distal ileum. Protein products, food, feed, fertilisers, cosmetics, pharmaceuticals or medical devices prepared using these commodities (unless covered by other Articles in this Chapter) should also not be traded.

2. From cattle that were at the time of slaughter over 30 months of age originating from a country, zone or compartment defined in Article 2.3.13.4., the following commodities, and any commodity contaminated by them, should not be traded for the preparation of food, feed, fertilisers, cosmetics, pharmaceuticals including biologicals, or medical devices: brains, eyes, spinal cord, skull and vertebral column. Protein products, food, feed, fertilisers, cosmetics, pharmaceuticals or medical devices prepared using these commodities (unless covered by other Articles in this Chapter) should also not be traded.

3. From cattle that were at the time of slaughter over 12 months of age originating from a country, zone or compartment defined in Article 2.3.13.5., the following commodities, and any commodity contaminated by them, should not be traded for the preparation of food, feed, fertilisers, cosmetics, pharmaceuticals including biologicals, or medical devices: brains, eyes, spinal cord, skull and vertebral column. Protein products, food, feed, fertilisers, cosmetics, pharmaceuticals or medical devices prepared using these commodities (unless covered by other Articles in this Chapter) should also not be traded.
Annex XII (contd)

Article 2.3.13.15.

Veterinary Authorities of importing countries should require:

for gelatine and collagen prepared from bones and intended for food or feed, cosmetics, pharmaceuticals including biologicals, or medical devices

the presentation of an international veterinary certificate attesting that:

1. the commodities came from a country, zone or compartment posing a negligible BSE risk;

OR

2. they originate from a country, zone or compartment posing a controlled or undetermined BSE risk and are derived from cattle which have passed ante-mortem and post-mortem inspections; and that

a) skulls from cattle over 30 12 months of age at the time of slaughter have been excluded;

b) the bones have been subjected to a process which includes all of the following steps:

i) degreasing,

ii) acid demineralisation,

iii) acid or alkaline treatment,

iv) filtration,

v) sterilisation at =138°C for a minimum of 4 seconds,

or to an equivalent or better process in terms of infectivity reduction (such as high pressure heating);

OR

3. they originate from a country, zone or compartment posing an undetermined BSE risk and are derived from cattle which have passed ante-mortem and post-mortem inspections; and that

a) skulls and vertebrae (except tail vertebrae) from cattle over 12 months of age at the time of slaughter have been excluded;

b) the bones have been subjected to a process which includes all of the following steps:

i) degreasing,

ii) acid demineralisation,

iii) acid or alkaline treatment,

iv) filtration,

v) sterilisation at =138°C for a minimum of 4 seconds,

or to an equivalent or better process in terms of infectivity reduction (such as high pressure heating).
Annex XII (contd)

Article 2.3.13.16.

Veterinary Authorities of importing countries should require:

for tallow and dicalcium phosphate (other than as defined in Article 2.3.13.1.) intended for food, feed, fertilisers, cosmetics, pharmaceuticals including biologicals, or medical devices

the presentation of an international veterinary certificate attesting that:

1. the commodities tallow came from a country, zone or compartment posing a negligible BSE risk; or

2. they originate from a country, zone or compartment posing a controlled BSE risk, are derived from cattle which have passed ante-mortem and post-mortem inspections, and have not been prepared using the tissues listed in points 1 and 2 of Article 2.3.13.14.

Article 2.3.13.16. bis

Veterinary Authorities of importing countries should require:

for dicalcium phosphate (other than as defined in Article 2.3.13.1.) intended for food, feed, fertilisers, cosmetics, pharmaceuticals including biologicals, or medical devices

the presentation of an international veterinary certificate attesting that:

1. the dicalcium phosphate came from a country, zone or compartment posing a negligible BSE risk; or

2. it originates from a country, zone or compartment posing a controlled or undetermined BSE risk and is a by-product of bone gelatine produced according to Article 2.3.13.15.

Article 2.3.13.17.

Veterinary Authorities of importing countries should require:

for tallow derivatives (other than those made from protein-free tallow as defined in Article 2.3.13.1.) intended for food, feed, fertilisers, cosmetics, pharmaceuticals including biologicals, or medical devices

the presentation of an international veterinary certificate attesting that:

1. the commodities originate from a country, zone or compartment posing a negligible BSE risk; or

2. they are derived from tallow meeting the conditions referred to in Article 2.3.13.16.; or

3. they have been produced by hydrolysis, saponification or transesterification using high temperature and pressure.

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OIE Terrestrial Animal Health Standards Commission/September 2007
APPENDIX 3.8.4.

GUIDELINES ON SURVEILLANCE FOR BOVINE SPONGIFORM ENCEPHALOPATHY

Article 3.8.4.1.

Introduction

1. Depending on the risk category of a country, zone or compartment with regard to bovine spongiform encephalopathy (BSE), surveillance for BSE may have one or more goals:
   a) detecting BSE, to a pre-determined design prevalence, in a country, zone or compartment;
   b) monitoring the evolution of BSE in a country, zone or compartment;
   c) monitoring the effectiveness of a feed ban and/or other risk mitigation measures, in conjunction with auditing;
   d) supporting a claimed BSE status;
   e) gaining or regaining a higher BSE status.

2. When the BSE agent is present in a country or zone, the cattle population will comprise the following sectors, in order of decreasing size:
   a) cattle not exposed to the infective agent;
   b) cattle exposed but not infected;
   c) infected cattle, which may lie within one of three stages in the progress of BSE:
      i) the majority will die or be killed before reaching a stage at which BSE is detectable by current methods;
      ii) some will progress to a stage at which BSE is detectable by testing before clinical signs appear;
      iii) the smallest number will show clinical signs.

3. The BSE status of a country, zone or compartment cannot be determined only on the basis of a surveillance programme but should be determined in accordance with all the factors listed in Article 2.3.13.2. The surveillance programme should take into account the diagnostic limitations associated with the above sectors and the relative distributions of infected cattle among them.

4. With respect to the distribution and expression of the BSE agent within the sectors described above, the following four subpopulations of cattle have been identified for surveillance purposes:
   a) cattle over 30 months of age displaying behavioural or clinical signs consistent with BSE (clinical suspects);
   b) cattle over 30 months of age that are non-ambulatory, recumbent, unable to rise or to walk without assistance; cattle over 30 months of age sent for emergency slaughter or condemned at ante-mortem inspection (casualty or emergency slaughter or downer cattle);
c) cattle over 30 months of age which are found dead or killed on farm, during transport or at an abattoir (fallen stock);

d) cattle over 36 months of age at routine slaughter.

5. A gradient is used to describe the relative value of surveillance applied to each subpopulation. Surveillance should focus on the first subpopulation, but investigation of other subpopulations will help to provide an accurate assessment of the BSE situation in the country, %me or compartment. This approach is consistent with Appendix 3.8.1. on general guidelines for animal health surveillance.

6. When establishing a surveillance strategy, authorities need to take into account the inherent difficulties of obtaining samples on farm, and overcome them. These difficulties include higher cost, the necessity to educate and motivate owners, and counteracting potentially negative socio-economic implications.

Article 3.8.4.2.

Description of cattle subpopulations

1. Cattle over 30 months of age displaying behavioural or clinical signs consistent with BSE (clinical suspects)

Cattle affected by illnesses that are refractory to treatment, and displaying progressive behavioural changes such as excitability, persistent kicking when milked, changes in herd hierarchical status, hesitation at doors, gates and barriers, as well as those displaying progressive neurological signs without signs of infectious illness are candidates for examination. These behavioural changes, being very subtle, are best identified by those who handle animals on a daily basis. Since BSE causes no pathognomonic clinical signs, all countries with cattle populations will observe individual animals displaying clinical signs consistent with BSE. It should be recognised that cases may display only some of these signs, which may also vary in severity, and such animals should still be investigated as potential BSE affected animals. The rate at which such suspicious cases are likely to occur will differ among epidemiological situations and cannot therefore be predicted reliably.

This subpopulation is the one exhibiting the highest prevalence. The accurate recognition, reporting and classification of such animals will depend on the ongoing owner/veterinarian awareness programme. This and the quality of the investigation and laboratory examination systems (Article 2.3.13.2.), implemented by the Veterinary Services, are essential for the credibility of the surveillance system.

2. Cattle over 30 months of age that are non-ambulatory, recumbent, unable to rise or to walk without assistance; cattle over 30 months of age sent for emergency slaughter or condemned at ante-mortem inspection (casualty or emergency slaughter, or downer cattle)

These cattle may have exhibited some of the clinical signs listed above which were not recognised as being consistent with BSE. Experience in countries where BSE has been identified indicates that this subpopulation is the one demonstrating the second highest prevalence. For that reason, it is the second most appropriate population to target in order to detect BSE.

3. Cattle over 30 months of age which are found dead or killed on farm, during transport or at an abattoir (fallen stock)

These cattle may have exhibited some of the clinical signs listed above prior to death, but were not recognised as being consistent with BSE. Experience in countries where BSE has been identified indicates that this subpopulation is the one demonstrating the third highest prevalence.
4. Cattle over 36 months of age at routine slaughter

Experience in countries where BSE has been identified indicates that this subpopulation is the one demonstrating the lowest prevalence. For that reason, it is the least appropriate population to target in order to detect BSE. However, sampling in this subpopulation may be an aide in monitoring the progress of the epizootic and the efficacy of control measures applied, because it offers continuous access to a cattle population of known class, age structure and geographical origin. Testing of routine slaughter cattle 36 months of age or less is of relatively very little value (Table 2).

Article 3.8.4.3.

Implementation of surveillance

In order to implement efficiently a surveillance strategy for BSE, a country must use documented records or reliable estimates of the age distribution of the adult cattle population and the number of cattle tested for BSE stratified by age and by subpopulation within the country, zone or compartment.

The approach assigns 'point values' to each sample, based on the subpopulation from which it was collected and the likelihood of detecting infected cattle in that subpopulation. The number of points a sample is assigned is determined by the subpopulation from which the sample is collected and the age of the animal sampled. The total points accumulation is then periodically compared to the target number of points for a country, zone or compartment.

A surveillance strategy should be designed to ensure that samples are representative of the herd of the country, zone or compartment, and include consideration of demographic factors such as production type and geographic location, and the potential influence of culturally unique husbandry practices. The approach used and the assumptions made should be fully documented, and the documentation retained for 7 years.

The points targets and surveillance point values in this Appendix were obtained by applying the following factors to a statistical model:

a) the design prevalence for Type A or Type B surveillance;

b) a confidence level of 95%;

c) the pathogenesis, and pathological and clinical expression of BSE:

i) sensitivity of diagnostic methods used;

ii) relative frequency of expression by age;

iii) relative frequency of expression within each subpopulation;

iv) interval between pathological change and clinical expression;

d) demographics of the cattle population, including age distribution;

e) influence of BSE on culling or attrition of animals from the cattle population via the four subpopulations;

f) percentage of infected animals in the cattle population which are not detected.
Although the procedure accepts very basic information about a cattle population, and can be used with estimates and less precise data, careful collection and documentation of the data significantly enhance their value. Since samples from clinical suspect animals provide many times more information than samples from healthy or dead-of-unknown-cause animals, careful attention to the input data can substantially decrease the procedure's cost and the number of samples needed. The essential input data are:

\[ g \] cattle population numbers stratified by age;

\[ h \] the number of cattle tested for BSE stratified by age and by subpopulation.

This Appendix utilises Tables 1 and 2 to determine a desired surveillance points target and the point values of surveillance samples collected.

Within each of the subpopulations above in a country, zone or compartment, a country may wish to target cattle identifiable as imported from countries or zones not free from BSE and cattle which have consumed potentially contaminated feedstuffs from countries or zones not free from BSE.

All clinical suspects should be investigated, regardless of the number of points accumulated. In addition, animals from the other subpopulations should be tested.

1. **Type A surveillance**

The application of Type A surveillance will allow the detection of BSE around a design prevalence of at least one case per 100,000 in the adult cattle population in the country, zone or compartment of concern, at a confidence level of 95%.

2. **Type B surveillance**

The application of Type B surveillance will allow the detection of BSE around a design prevalence of at least one case per 50,000 in the adult cattle population in the country, zone or compartment of concern, at a confidence level of 95%.

Type B surveillance may be carried out by countries, zones or compartments of negligible BSE risk status (Article 2.3.13.3.) to confirm the conclusions of the risk assessment, for example by demonstrating the effectiveness of the measures mitigating any risk factors identified, through surveillance targeted to maximise the likelihood of identifying failures of such measures.

Type B surveillance may also be carried out by countries, zones or compartments of controlled BSE risk status (Article 2.3.13.4.), following the achievement of the relevant points target using Type A surveillance, to maintain confidence in the knowledge gained through Type A surveillance.

**Article 3.8.4.4.**

1. **Selecting the points target**

The surveillance points target should be selected from Table 1, which shows target points for adult cattle populations of different sizes. The size of the adult cattle population of a country, zone or compartment may be estimated or may be set at one million because, for statistical reasons, one million is the point beyond which sample size does not further increase with population size.
Table 1. Points targets for different adult cattle population sizes in a country, zone or compartment

<table>
<thead>
<tr>
<th>Adult cattle population size (24 months and older)</th>
<th>Type A surveillance</th>
<th>Type B surveillance</th>
</tr>
</thead>
<tbody>
<tr>
<td>&gt;1,000,000</td>
<td>300,000</td>
<td>150,000</td>
</tr>
<tr>
<td>800,000-1,000,000</td>
<td>240,000</td>
<td>120,000</td>
</tr>
<tr>
<td>600,000-800,000</td>
<td>180,000</td>
<td>90,000</td>
</tr>
<tr>
<td>400,000-600,000</td>
<td>120,000</td>
<td>60,000</td>
</tr>
<tr>
<td>200,000-400,000</td>
<td>60,000</td>
<td>30,000</td>
</tr>
<tr>
<td>100,000-200,000</td>
<td>30,000</td>
<td>15,000</td>
</tr>
<tr>
<td>50,000-100,000</td>
<td>15,000</td>
<td>7,500</td>
</tr>
</tbody>
</table>

2. Determining the point values of samples collected

Table 2 can be used to determine the point values of the surveillance samples collected. The approach assigns point values to each sample according to the likelihood of detecting infection based on the subpopulation from which the sample was collected and the age of the animal sampled. This approach takes into account the general principles of surveillance described in Appendix 3.8.1. and the epidemiology of BSE.

Because precise aging of the animals that are sampled may not be possible, Table 2 combines point values into five age categories. The point estimates for each category were determined as an average for the age range comprising the group. The age groups were selected on their relative likelihoods of expressing BSE according to scientific knowledge of the incubation of the disease and the world BSE experience. Samples may be collected from any combination of subpopulations and ages but should reflect the demographics of the cattle herd of the country, zone or compartment. In addition, countries should sample at least three of the four subpopulations.

If a country, zone or compartment determines, based on the demographics and epidemiological characteristics of its cattle population, that precise classification of the subpopulations ‘casualty or emergency slaughter, or downer cattle’ and ‘fallen stock’ is not possible, these subpopulations may be combined. In such a case, the surveillance point values accorded to the combined subpopulation would be that of ‘fallen stock’.

The total points for samples collected may be accumulated over a period of a maximum of 7 consecutive years to achieve the target number of points determined in Table 1.
Annex XII (contd)

Table 2. Surveillance point values for samples collected from animals in the given subpopulation and age category

<table>
<thead>
<tr>
<th>Surveillance subpopulation</th>
<th>Routine slaughter¹</th>
<th>Fallen stock²</th>
<th>Casually slaughter³</th>
<th>Clinical suspect⁴</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age = 1 year and = 2 years</td>
<td>0.01</td>
<td>0.2</td>
<td>0.4</td>
<td>N/A</td>
</tr>
<tr>
<td>Age = 2 years and = 4 years (young adult)</td>
<td>0.1</td>
<td>0.2</td>
<td>0.4</td>
<td>260</td>
</tr>
<tr>
<td>Age = 4 years and = 7 years (middle adult)</td>
<td>0.2</td>
<td></td>
<td>0.9</td>
<td>1.6</td>
</tr>
<tr>
<td>Age = 7 years and = 9 years (older adult)</td>
<td>0.4</td>
<td></td>
<td>0.7</td>
<td>220</td>
</tr>
<tr>
<td>Age = 9 years (aged)</td>
<td>0.0</td>
<td>0.1</td>
<td>0.2</td>
<td>45</td>
</tr>
</tbody>
</table>

Surveillance points remain valid for 7 years (the 95th percentile of the incubation period).

1. See point 4) of Article 3.8.4.2.
2. See point 3) of Article 3.8.4.2.
3. See point 2) of Article 3.8.4.2.
4. See point 1) of Article 3.8.4.2

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CHAPTER 2.5.5.

EQUINE INFLUENZA

Article 2.5.5.1.

For the purposes of the Terrestrial Code, equine influenza (EI) is defined as an infection of domestic horses, donkeys and mules.

For the purposes of international trade, this Chapter deals not only with the occurrence of clinical signs caused by equine influenza virus (EIV), but also with the presence of infection with EIV in the absence of clinical signs.

For the purposes of this chapter, isolation is defined as 'the separation of horses from horses of a different equine influenza health status, utilising appropriate biosecurity measures, with the purpose of preventing the transmission of infection'.

For the purposes of the Terrestrial Code, the infective period for equine influenza is 21 days.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.5.5.2.

The EI status of a country, a zone or a compartment can be determined on the basis of the following criteria:

1. the outcome of a risk assessment identifying all potential factors for EI occurrence and their historic perspective;

2. whether EI is notifiable in the whole country, an on-going EI awareness programme is in place, and all notified suspect occurrences of EI are subjected to field and, where applicable, laboratory investigations;

3. appropriate surveillance is in place to demonstrate the presence of infection in the absence of clinical signs in horses.

Article 2.5.5.3.

Equine influenza free country, zone or compartment

A country or zone or compartment may be considered free from EI provided the disease is notifiable in the whole country and it shows evidence of an effective surveillance programme, planned and implemented according to the general principles in Appendix 3.8.1. The surveillance may need to be adapted to parts of the country, zone or compartment depending on historical or geographical factors, industry structure, population data, movements of equids into the country, zone or compartment, wild equid populations or proximity to recent outbreaks.

A country, zone or compartment seeking freedom from EI, in which vaccination is practised, should also demonstrate that EIV has not been circulating in the domestic horse population during the past 12 months, through surveillance, in accordance with Appendix 3.8.1., at a level sufficient to provide at least a 95% level of confidence of detecting infection if it is present at a prevalence rate exceeding 1%. In a country in which vaccination is not practised, surveillance could be conducted using serological testing. In countries where vaccination is practised, the surveillance should include methods of virus detection.
If an outbreak of clinical equine influenza occurs in a previously free country, zone or compartment, free status can be regained 12 months after the last clinical case, providing that surveillance for evidence of infection has been carried out during that 12-month period at a level in accordance with Appendix 3.8.1, sufficient to provide at least a 95% level of confidence of detecting infection if it is present at a prevalence rate exceeding 1%.

Article 2.5.5.4.
(under study)

Regardless of the EI status of the exporting country, zone or compartment, the Veterinary Authority of a country, zone or compartment should authorise without restriction on account of EI the importation into their territory of the following commodities:

1. semen;
2. in vivo derived equine embryos collected, processed and stored in conformity with the provisions of Appendix 3.3.1.

Article 2.5.5.5

When importing horses for immediate slaughter, the Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the horses showed no clinical sign of EI on the day of shipment.

Article 2.5.5.6.

When importing horses for unrestricted movement, Veterinary Authorities should require:

the presentation of an international veterinary certificate attesting that the horses:

1. came from an EI free country, zone or compartment in which they had been resident for at least 21 days; in the case of a vaccinated horse, information on its vaccination status should be included in the veterinary certificate;

OR

2. came from a country, zone or compartment not known to be free from EI, were subjected to pre-export isolation for 21 days and showed no clinical sign of EI during isolation nor on the day of shipment; and
3. were vaccinated according to the manufacturer’s instructions between 21 and 90 days before shipment either with a primary course or a booster.

Article 2.5.5.7.

When importing horses which will be kept in isolation (see Article 2.5.5.1.), Veterinary Authorities should require:

the presentation of an international veterinary certificate attesting that the horses:

1. came from an EI free country, zone or compartment in which they had been resident for at least 21 days; in the case of a vaccinated horse, information on its vaccination status should be included in the veterinary certificate;
OR

2. showed no clinical sign of EI in any premises in which the horses had been resident for the 21 days prior to shipment nor on the day of shipment; and

3. were vaccinated according to the manufacturer’s instructions.

Article 2.5.5.8.

When importing fresh meat of horses, mules or donkeys, Veterinary Authorities should require the presentation of an international veterinary certificate attesting that the fresh meat came from horses, mules or donkeys which had been subjected to ante-mortem and post-mortem inspections as described in Appendix 3.10.1.
CHAPTER 2.5.7.

EQUINE RHINOPNEUMONITIS
(Equine herpes virus type 1 infection)

Article 2.5.7.1.

Standards for diagnostic tests are described in the Terrestrial Manual.

Article 2.5.7.2.

Veterinary Authorities of importing countries should require:

for equines

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of equine herpes virus type 1 infection, on the day of shipment and during the 21 days prior to shipment;

2. were kept for the 21 days prior to shipment in an establishment where no case of equine herpes virus type 1 infection was reported during that period.

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CHAPTER 2.5.10.

EQUINE VIRAL ARTERITIS

Article 2.5.10.1.

The infective period for equine viral arteritis (EVA) shall be 28 days for all categories of equine except sexually mature stallion where the infective period may be for the life of the animal. Because the infective period may be extended in the case of virus shedding in semen, the status of seropositive stallions should be checked to ensure that they do not shed virus in their semen.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.5.10.2.

Veterinary Authorities of importing countries should require:

for uncastrated male equines imported on a temporary basis for breeding or on a permanent basis:

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of EVA on the day of shipment and during the 28 days prior to shipment;

2. were isolated and subjected, to a test for EVA, as prescribed in the Terrestrial Manual, carried out either:
   a) on a single blood sample collected during the 28 days prior to shipment with negative result, or
   b) on blood samples taken on two occasions at least 14 days apart within 28 days prior to shipment, which demonstrated stable or declining antibody titres; or

3. were isolated and subjected between 6 and 9 months of age to a test for EVA, as prescribed in the Terrestrial Manual, carried out on two blood samples collected at least 14 days apart with stable or decreasing titre, immediately vaccinated for EVA and regularly revaccinated according to the manufacturer’s instructions; or

4. were isolated and subjected to a test for EVA, as prescribed in the Terrestrial Manual, on a blood sample with negative results, immediately vaccinated for EVA, kept for 21 days following vaccination separated from other equidae and regularly revaccinated according to the manufacturer’s instructions; or

5. have been subjected to a test for EVA, as prescribed in the Terrestrial Manual, carried out on a blood sample with positive results and then: either
   a) were subsequently test mated to two mares within 12 months prior to shipment which were subjected to two tests for EVA as prescribed in the Terrestrial Manual with negative results on blood samples collected at the time of test mating and again 28 days after the mating; or
   b) were subjected to a test for equine arteritis virus as prescribed in the Terrestrial Manual with negative results, carried out on semen collected during the 28 days prior to shipment.
Annex XIV (contd)

Article 2.5.10.3.

Veterinary Authorities of importing countries should require:

for uncastrated male equines imported on a temporary basis other than for breeding, and for equines other than uncastrated males:

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of EVA on the day of shipment and were kept in an establishment where no animals have shown any signs of EVA for the 28 days prior to shipment;

2. were isolated and subjected to a test for EVA, as prescribed in the Terrestrial Manual, carried out either:
   a) on a single blood sample collected during the 28 days prior to shipment with negative results, or
   b) on blood samples collected on two occasions at least 14 days apart within 28 days prior to shipment, which demonstrated stable or declining antibody titres;

OR

3. were isolated and subjected, between 6 and 9 months of age, to a diagnostic test for EVA, as prescribed in the Terrestrial Manual, on a blood sample with stable or decreasing titre, immediately vaccinated for EVA and regularly revaccinated.

Article 2.5.10.4.

Veterinary Authorities of importing countries should require:

for semen

the presentation of an international veterinary certificate attesting that the animal donors:

1. were kept for the 28 days prior to semen collection in an establishment where no equine has shown any clinical sign of EVA during that period;

2. showed no clinical sign of EVA on the day of semen collection;

3. were subjected between 6 and 9 months of age to a test for EVA as prescribed in the Terrestrial Manual on a blood sample with stable or decreasing titre, immediately vaccinated for EVA and regularly revaccinated according to the manufacturer's instructions; or

4. were subjected to a test for EVA as prescribed in the Terrestrial Manual on a blood sample with negative results, immediately vaccinated for EVA, kept for 21 days following vaccination separated from other equidae and regularly revaccinated according to the manufacturer's instructions; or

5. were subjected to a test for EVA as prescribed in the Terrestrial Manual on a blood sample with negative results within 14 days prior to semen collection, and had been separated from other equidae from the time of the taking of the blood sample to the time of semen collection; or

6. have been subjected to a test for EVA as prescribed in the Terrestrial Manual on a blood sample with positive results and then: either
a) were subsequently test mated to two mares within 12 months prior to semen collection, which were subjected to two tests for EVA as prescribed in the Terrestrial Manual with negative results on blood samples collected at the time of test mating and again 28 days after the test mating, or

b) were subjected to a test for equine arteritis virus as prescribed in the Terrestrial Manual with negative results, carried out on semen collected within one year prior to collection of the semen to be exported.
CHAPTER 2.5.14.

AFRICAN HORSE SICKNESS

Article 2.5.14.1.

For the purposes of the Terrestrial Code, the infective period for African horse sickness virus (AHSV) shall be 40 days for domestic horses. Although critical information is lacking for some species, this Chapter applies to all equidae.

All countries or zones neighbouring, or considered to be at risk from, a country or zone not having free status should determine their AHSV status from an ongoing surveillance programme. Throughout the Chapter surveillance is in all cases understood as being conducted as described in Appendix 3.8.X.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.5.14.2.

AHSV free country or zone

1. A country or zone may be considered free from AHSV when African horse sickness (AHS) is notifiable in the whole country, systematic vaccination is prohibited, importation of equidae, their semen, and oocytes or embryos, and pathological material and biological products from these species are carried out in accordance with this chapter, and either:

   a) historical freedom as described in Appendix 3.8.1. has demonstrated no evidence of AHSV in the country or zone; or

   b) the country or zone has not reported any case of AHS for at least 2 years and is not adjacent to a country or zone not having a free status; or

   c) a surveillance programme has demonstrated no evidence of AHSV in the country or zone for at least 12 months; or

   d) the country or zone has not reported any case of AHS and a surveillance programme has demonstrated no evidence of Culicoides likely to be competent AHSV vectors in the country or zone.

2. An AHSV free country or zone will not lose its free status through the importation of vaccinated or seropositive equidae, their semen, oocytes or embryos from infected countries or infected zones, provided these imports are carried out in accordance with this chapter.

Article 2.5.14.3.

AHSV seasonally free zone

1. An AHSV seasonally free zone is a part of an infected country or an infected zone for which for part of a year, ongoing surveillance and monitoring demonstrate no evidence of AHSV transmission and of the presence of adult Culicoides likely to be competent AHSV vectors.

2. For the application of Articles 2.5.14.6., 2.5.14.8. and 2.5.14.9., the seasonally free period is:
Annex XV (contd)

a) taken to commence the day following the last evidence of AHSV transmission and of the cessation of activity of adult Culicoides likely to be competent AHSV vectors as demonstrated by an ongoing surveillance programme, and

b) taken to conclude either:

i) at least 28 days before the earliest date that historical data show AHSV activity has recommenced; or

ii) immediately when current climatic data or data from a surveillance and monitoring programme indicate an earlier resurgence of activity of adult Culicoides likely to be competent AHSV vectors.

3. An AHSV seasonally free zone will not lose its free status through the importation of vaccinated or seropositive equidae, their semen, oocytes or embryos from infected countries or infected zones, provided these imports are carried out in accordance with this chapter.

Article 2.5.14.4.

**AHSV infected country or zone**

An AHSV infected country or infected zone is a clearly defined area where the conditions of Article 2.5.14.2. or Article 2.5.14.3. do not apply.

Article 2.5.14.5.

When importing from AHSV free countries that are neither neighbouring nor considered to be at risk from an AHSV infected country or infected zone, Veterinary Administrations should require:

for equidae

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of AHS on the day of shipment;

2. have not been vaccinated against AHS within the last 40 days;

3. were kept in an AHSV free country since birth or for at least 40 days prior to shipment;

4. either:

   a) did not transit through an infected country or infected zone; or

   b) were protected from attack by Culicoides likely to be competent AHSV vectors at all times when transiting through an infected country or infected zone.

Article 2.5.14.6.

When importing from AHSV free countries or free zones or from AHSV seasonally free zones (during the seasonally free period) that are neighbouring or are considered to be at risk from an AHSV infected country or infected zone, Veterinary Administrations should require:

for equidae
the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical signs of AHS on the day of shipment;
2. have not been vaccinated against AHS within the last 40 days;
3. were kept in an AHSV free country, free zone or seasonally free zone during the seasonally free period since birth or for at least 40 days prior to shipment; or
4. in a country or zone considered to be at risk, were held in quarantine for at least 40 days prior to shipment and protected at all times from attack by Culicoides likely to be competent AHSV vectors; and
   a) a serological test according to the Terrestrial Manual to detect antibodies to the AHSV group, was carried out with a negative result on a blood sample collected at least 28 days after introduction into the quarantine station; or
   b) serological tests according to the Terrestrial Manual to detect serotype specific antibodies against the AHSV serotypes known to occur within the region were carried out with no significant increase in antibody titre on blood samples collected on two occasions, with an interval of not less than 21 days, the first sample being collected at least 7 days after introduction into the quarantine station; or
   c) agent identification tests according to the Terrestrial Manual were carried out with negative results on blood samples collected on two occasions with an interval of not less than 14 days between collection, the first sample being collected at least 7 days after introduction into the quarantine station;
5. were protected from attack by Culicoides likely to be competent AHSV vectors at all times during transportation (including to and at the place of shipment).

Article 2.5.14.7.

When importing from an AHSV infected country or an AHSV infected zone, Veterinary Authorities should require:

for equidae

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of AHS on the day of shipment;
2. have not been vaccinated against AHS within the last 40 days;
3. were held continuously during the quarantine period of at least 40 days, in a vector-proof quarantine station and protected at all times from attack by Culicoides likely to be competent AHSV vectors; and
   a) a serological test according to the Terrestrial Manual to detect antibodies to the AHSV group, was carried out with a negative result on a blood sample collected at least 28 days after introduction into the quarantine station; or
Annex XV (contd)

b) serological tests according to the Terrestrial Manual to detect serotype-specific antibodies against the AHSV serotypes known to occur within the region were carried out with no significant increase in antibody titre on blood samples collected on two occasions, with an interval of not less than 21 days, the first sample being collected at least 7 days after introduction into the quarantine station; or

c) agent identification tests according to the Terrestrial Manual were carried out with negative results on blood samples collected on two occasions with an interval of not less than 14 days between collection, the first sample being collected at least 7 days after introduction into the quarantine station;

4. were protected from attack by Culicoides likely to be competent AHSV vectors at all times during transportation (including during transportation to and at the place of shipment).

Article 2.5.14.8.

Veterinary Administrations Authorities of importing countries should require:

for equid semen

the presentation of an international veterinary certificate attesting that the donor animals:

1. showed no clinical sign of AHS on the day of collection of the semen and for the following 40 days;
2. had not been vaccinated against AHS within 40 days prior to the day of collection;
3. were either:
   a) kept in an AHSV free country or free zone or from an AHSV seasonally free zone (during the seasonally free period) for at least 40 days before commencement of, and during collection of the semen, or
   b) kept in an AHSV free vector-proof artificial insemination centre throughout the collection period, and subjected to either:
      i) a serological test according to the Terrestrial Manual to detect antibody to the AHSV group, carried out with a negative result on a blood sample collected at least 28 days and not more than 90 days after the last collection of semen; or
      ii) agent identification tests according to the Terrestrial Manual carried out with negative results on blood samples collected at commencement and conclusion of, and at least every 7 days, during semen collection for this consignment.

Article 2.5.14.9.

Veterinary Administrations Authorities of importing countries should require:

for in vivo derived equid embryos/oocytes

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) showed no clinical sign of AHS on the day of collection of the semen embryos/oocytes and for the following 40 days;
b) had not been vaccinated against AHS within 40 days prior to the day of collection;

c) were either:

   i) kept in an AHSV free country or free zone or from an AHSV seasonally free zone (during the seasonally free period) for at least 40 days before commencement of, and during collection of the embryos/oocytes, or

   ii) kept in an AHSV free vector-proof collection centre throughout the collection period, and subjected to either:

       - a serological test according to the Terrestrial Manual to detect antibody to the AHSV group carried out with a negative result on a blood sample collected at least 28 days and not more than 90 days after the last collection of semen embryos/oocytes; or

       - agent identification tests according to the Terrestrial Manual carried out with negative results on blood samples collected at commencement and conclusion of, and at least every 7 days during semen embryos/oocytes collection for this consignment;

2. the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1.;

3. semen used to fertilize the oocytes, complies at least with the requirements in Article 2.5.14.8.

Article 2.5.14.10.

**Protecting animals from Culicoides attack**

When transporting equines through AHSV infected countries or AHSV infected zones, Veterinary Administrations Authorities should require strategies to protect animals from attack by Culicoides likely to be competent AHSV vectors during transport, taking into account the local ecology of the vector.

Potential risk management strategies include a combination of:

1. treating animals with chemical repellents prior to and during transportation, in insecticide treated and sanitized vehicles treated with appropriate residual contact insecticide;

2. loading, transporting and unloading animals at times of low vector activity (i.e. bright sunshine and low temperature);

3. ensuring vehicles do not stop en route during dawn or dusk, or overnight, unless the animals are held behind insect proof netting;

4. darkening the interior of the vehicle, for example by covering the roof and/or sides of vehicles with shade cloth;

5. monitoring for vectors at common stopping and offloading points to gain information on seasonal variations;

6. using historical, ongoing and/or AHS modelling information to identify low risk ports and transport routes.
APPENDIX 3.8.X.

GUIDELINES ON SURVEILLANCE FOR AFRICAN HORSE SICKNESS

Article 3.8.X.1.

Introduction

This Appendix defines the principles and provides a guide on the surveillance for African horse sickness (AHS), complementary to Appendix 3.8.1., applicable to countries seeking to demonstrate recognition for a declared African horse sickness virus (AHSV) status. This may be for the entire country or zone. Guidance for countries seeking free status following an outbreak and for the maintenance of AHS status is also provided.

AHS is a vector-borne infection transmitted by a limited number of species of Culicoides insects. Unlike the related bluetongue virus, AHSV is so far geographically restricted to sub-Saharan Africa with periodic excursions into North Africa, southwest Europe, the Middle East and adjacent regions of Asia. An important component of AHSV epidemiology is vectorial capacity which provides a measure of disease risk that incorporates vector competence, abundance, seasonal incidence, biting rates, survival rates and the extrinsic incubation period. However, methods and tools for measuring some of these vector factors remain to be developed, particularly in a field context.

In addition to the general conditions described in Chapter 2.5.14. of the Terrestrial Code, a Member demonstrating freedom from AHSV infection for the entire country, or a zone should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and should be planned and implemented according to general conditions and methods described in this Appendix. This requires the support of a laboratory able to undertake identification of AHSV infection through the virus detection and antibody tests described in the Terrestrial Manual.

Susceptible wild equid populations should be included in the surveillance programme when these animals are intended for trade.

Case definition

For the purposes of surveillance, a case refers to an equid infected with AHSV.

The purpose of surveillance is to determine if a country or zone is free from AHSV or if a zone is seasonally free from AHSV. Surveillance deals not only with the occurrence of clinical signs caused by AHSV, but also with evidence of infection with AHSV in the absence of clinical signs.

The following defines the occurrence of AHSV infection:

1. AHSV has been isolated and identified as such from an equid or a product derived from that equid, or
2. viral antigen or viral RNA specific to one or more of the serotypes of AHSV has been identified in samples from one or more equids showing clinical signs consistent with AHS, or epidemiologically linked to a confirmed or suspected case, or giving cause for suspicion of previous association or contact with AHSV, or
3. serological evidence of active infection with AHSV by detection of seroconversion with production of antibodies to structural or nonstructural proteins of AHSV that are not a consequence of vaccination have been identified in one or more equids that either show clinical signs consistent with AHS, or epidemiologically linked to a confirmed or suspected case, or give cause for suspicion of previous association or contact with AHSV.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

**Article 3.8.X.2.**

**General conditions and methods**

1. A surveillance system should be under the responsibility of the Veterinary Administration. In particular the following should be in place:
   a) a formal and ongoing system for detecting and investigating outbreaks of disease;
   b) a procedure for the rapid collection and transport of samples from suspect cases of AHS to a laboratory for AHS diagnosis as described in the Terrestrial Manual;
   c) a system for recording, managing and analysing diagnostic, epidemiologic and surveillance data.

2. The AHS surveillance programme should:
   a) in a country/zone, free or seasonally free, include an early warning system for reporting suspicious cases. Persons who have regular contact with equids, as well as diagnosticians, should report promptly any suspicion of AHS to the Veterinary Authority. An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is AHS. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. All suspected cases of AHS should be investigated immediately and samples should be taken and submitted to an approved laboratory. This requires that sampling kits and other equipment are available for those responsible for surveillance;
   b) conduct random or targeted serological and virological surveillance appropriate to the infection status of the country or zone in accordance with Appendix 3.8.1.

**Article 3.8.X.3.**

**Surveillance strategies**

The target population for surveillance aimed at identification of disease and/or infection should cover susceptible domestic equids within the country or zone. Active and passive surveillance for AHSV infection should be ongoing. Surveillance should be composed of random or targeted approaches using virological, serological and clinical methods appropriate for the infection status of the country or zone.

A country should justify the surveillance strategy chosen as appropriate to detect the presence of AHSV infection in accordance with Appendix 3.8.1. and the prevailing epidemiological situation. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clinical signs (e.g. horses). Similarly, virological and serological testing may be targeted to species that rarely show clinical signs (e.g. donkeys).

In vaccinated populations serological and virological surveillance is necessary to detect the AHSV types circulating to ensure that all circulating types are included in the vaccination programme.
If a Member wishes to declare freedom from AHSV infection in a specific zone, the design of the surveillance strategy would need to be aimed at the population within the zone.

For random surveys, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect infection if it were to occur at a predetermined minimum rate. The sample size, expected prevalence and diagnostic sensitivity of the tests determine the level of confidence in the results of the survey. The applicant country must justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Appendix 3.8.1. Selection of the design prevalence, in particular, needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey approach selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and the different species in the target population.

Irrespective of the testing system employed, surveillance system design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of infection or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as those which may be epidemiologically linked to it.

The principles for surveillance for disease/infection are technically well defined. Surveillance programmes to prove the absence of AHSV infection/circulation, need to be carefully designed to avoid producing results that are either insufficiently reliable to be accepted by international trading partners, or excessively costly and logistically complicated. The design of any surveillance programme, therefore, requires inputs from professionals competent and experienced in this field.

1. **Clinical surveillance**

   Clinical surveillance aims at the detection of clinical signs of AHS in equids particularly during a newly introduced infection. In horses, clinical signs may include pyrexia, oedema, hyperaemia of mucosal membranes and dyspnoea.

   AHS suspects detected by clinical surveillance should always be confirmed by laboratory testing.

2. **Serological surveillance**

   Serological surveillance of equid populations is useful to confirm absence of AHSV transmission in a country or zone. The species tested should reflect the local epidemiology of AHSV infection, and the equine species available. Management variables that may reduce the likelihood of infection, such as the use of insecticides and animal housing, should be taken into account when selecting equids to be included in the surveillance system.

   Samples should be examined for antibodies against AHSV using tests prescribed in the Terrestrial Manual. Positive AHSV antibody test results can have four possible causes:

   a) natural infection with AHSV;

   b) vaccination against AHSV;
c) maternal antibodies;
d) positive results due to the lack of specificity of the test.

It may be possible to use sera collected for other purposes for AHSV surveillance. However, the principles of survey design described in these guidelines and the requirements for a statistically valid survey for the presence of AHSV infection should not be compromised.

The results of random or targeted serological surveys are important in providing reliable evidence that no AHSV infection is present in a country or zone. It is, therefore, essential that the survey is thoroughly documented. It is critical to interpret the results in light of the movement history of the animals being sampled.

Serological surveillance in a free zone should target those areas that are at highest risk of AHSV transmission, based on the results of previous surveillance and other information. This will usually be towards the boundaries of the free zone. In view of the epidemiology of AHSV, either random or targeted sampling is suitable to select herds and/or animals for testing.

Serological surveillance in a free country or zone should be carried out over an appropriate distance from the border with an infected country or infected zone, based upon geography, climate, history of infection and other relevant factors. The surveillance should be carried out over a distance of at least 100 kilometres from the border with that country or zone, but a lesser distance could be acceptable if there are relevant ecological or geographical features likely to interrupt the transmission of AHSV. An AHSV free country or zone may be protected from an adjacent infected country or infected zone by a buffer zone.

Serological surveillance in infected zones will identify changes in the boundary of the zone, and can also be used to identify the AHSV types circulating. In view of the epidemiology of AHSV infection, either random or targeted sampling is suitable.

3. Virological surveillance

Isolation and genetic analysis of AHSV from a proportion of infected animals is beneficial in terms of providing information on serotype and genetic characteristics of the viruses concerned.

Virological surveillance using tests described in the Terrestrial Manual can be conducted:
a) to identify virus circulation in at risk populations;
b) to confirm clinically suspect cases;
c) to follow up positive serological results;
d) to better characterize the genotype of circulating virus in a country or zone.

4. Sentinel animals

Sentinel animals are a form of targeted surveillance with a prospective study design. They comprise groups of unexposed equids managed at fixed locations and sampled regularly to detect new AHSV infections.

The primary purpose of a sentinel equid programme is to detect AHSV infections occurring at a particular place, for instance sentinel groups may be located on the boundaries of infected zones to detect changes in distribution of AHSV. In addition, sentinel equid programmes allow the timing and dynamics of infections to be observed.
A sentinel equid programme should use animals of known source and history of exposure, control management variables such as use of insecticides and animal housing (depending on the epidemiology of AHSV in the area under consideration), and be flexible in its design in terms of sampling frequency and choice of tests.

Care is necessary in choosing the sites for the sentinel groups. The aim is to maximise the chance of detecting AHSV activity at the geographical location for which the sentinel site acts as a sampling point. The effect of secondary factors that may influence events at each location, such as climate, may also be analysed. To avoid confounding factors sentinel groups should comprise animals selected to be of similar age and susceptibility to AHSV infection. The only feature distinguishing groups of sentinels should be their geographical location. Sera from sentinel animal programmes should be stored methodically in a serum bank to allow retrospective studies to be conducted in the event of new serotypes being isolated.

The frequency of sampling should reflect the equid species used and the reason for choosing the sampling site. In endemic areas virus isolation will allow monitoring of the serotypes and genotypes of AHSV circulating during each time period. The borders between infected and non infected areas can be defined by serological detection of infection. Monthly sampling intervals are frequently used. Sentinels in declared free zones add to confidence that AHSV infections are not occurring unobserved. Here sampling prior to and after the possible period of transmission is sufficient.

Definitive information on AHSV circulating in a country or zone is provided by isolation and identification of the viruses. If virus isolation is required sentinels should be sampled at sufficiently frequent intervals to ensure that some samples are collected during the period of viraemia.

5. Vector surveillance

AHSV is transmitted between equine hosts by species of Culicoides which vary across the world. It is therefore important to be able to identify potential vector species accurately although many such species are closely related and difficult to differentiate with certainty.

The main purpose of vector surveillance is to define high, medium and low-risk areas and local details of seasonality by determining the various species present in an area, their respective seasonal occurrence, and abundance. Vector surveillance has particular relevance to potential areas of spread. Long term surveillance can also be used to assess vector abatement measures.

The most effective way of gathering this information should take account of the biology and behavioural characteristics of the local vector species of Culicoides and may include the use of Onderstepoort-type light traps or similar, operated from dusk to dawn in locations adjacent to equids.

Vector surveillance should be based on scientific sampling techniques. The choice of the number and types of traps to be used in vector surveillance and the frequency of their use should take into account the size and ecological characteristics of the area to be surveyed.

The operation of vector surveillance sites at the same locations as sentinel animals is advisable.

The use of a vector surveillance system to detect the presence of circulating virus is not recommended as a routine procedure as the typically low vector infection rates mean that such detections can be rare. Other surveillance strategies are preferred to detect virus circulation.
CHAPTER 2.6.6.

AFRICAN SWINE FEVER

Article 2.6.6.1.

The pig is the only natural host for African swine fever (ASF) virus. The definition of pig includes all varieties of Sus scrofa, both domestic and wild, warthogs Phacochoerus spp., bushpigs Potamochoerus spp. and giant forest hog Hylochoerus meinertzhageni. For the purposes of this chapter, a distinction is made between domestic pigs (permanently captive and farmed free-range pigs) and wild pigs (including feral pigs and wild boar) as well as between Sus scrofa and African pig species.

All varieties of Sus scrofa are susceptible to the pathogenic effects of ASF virus, while the African wild pigs are not and act as reservoirs of the infection. Ticks of the genus Ornithodoros are natural hosts of the virus and act as biological vectors of the infection.

For the purpose of the Terrestrial Code, the incubation period in Sus scrofa is 15 days.

Standards for diagnostic tests are described in the Terrestrial Manual.

Article 2.6.6.2.

The ASF status of a country, zone or compartment can only be determined after considering the following criteria in domestic and wild pigs, as applicable:

1. ASF should be notifiable in the whole country, and all clinical signs suggestive of ASF should be subjected to appropriate field and/or laboratory investigations;

2. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of ASF;

3. the Veterinary Administration Authority should have current knowledge of, and authority over, all domestic pigs in the country, zone or compartment;

4. the Veterinary Administration Authority should have current knowledge about the population and habitat of wild pigs in the country or zone.

Article 2.6.6.3.

ASF free country, zone or compartment

1. ASF free status

   a) Historically free status

      A country or zone may be considered free from ASF without formally applying a specific surveillance programme if the provisions of Article 3.8.1.6. are complied with.

   b) Free status as a result of an eradication programme

      A country or zone which does not meet the conditions of point a) above or a compartment may be considered free from ASF when:
Annex XVI (contd)

i) there has been no outbreak of ASF during the past 3 years; this period can be reduced to 12 months when there is no evidence of tick involvement in the epidemiology of the infection;

ii) no evidence of ASFV infection has been found during the past 12 months;

iii) surveillance in accordance with Appendix 3.8.8. has been in place in domestic pigs for the past 12 months;

iii) no evidence of ASFV infection has been found during the past 12 months

AND

in the case of a country or zone, surveillance in accordance with Appendix 3.8.8. has been in place to determine the ASF status of the wild pig population and:

civ) there has been no clinical evidence, nor virological evidence of ASF in wild pigs during the past 12 months;

dv) no seropositive wild pigs have been detected in the age class 6-12 months during the past 12 months;

ev) imported wild pigs comply with the relevant requirements in Article 2.6.6.7.

Article 2.6.6.4.

Recovery of free status

Should an ASF outbreak occur in a free country, zone or compartment, the free status of the country, zone or compartment may be restored where surveillance in accordance with Appendix 3.8.8. has been carried out with negative results, either:

1. 3 months after the last case where a stamping-out policy is practised and there is no evidence of tick involvement in the epidemiology of the infection;

OR

2. in the case where ticks are suspected to be involved in the epidemiology of the infection, 3 months after the last case where a stamping-out policy, followed by acaricide treatment and the use of sentinel pigs, is practised;

OR

3. where a stamping-out policy is not practised, the provisions of point b) of Article 2.6.6.3. should be followed;

AND

4. in the case of a country or zones, based on surveillance in accordance with Appendix 3.8.8., ASF infection is not known to occur in any wild pig population in the country or zone.

Article 2.6.6.5.

When importing from ASF free countries, zones or compartments, Veterinary Authorities should require:
Annex XVI (contd)

for domestic pigs

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of ASF on the day of shipment;
2. were kept in an ASF free country, zone or compartment since birth or for at least the past 40 days.

Article 2.6.6.6.

When importing from ASF infected countries or infected zones with ASF infection in domestic pigs, Veterinary Administrations should require:

for domestic pigs

the presentation of an international veterinary certificate attesting that the animals:

1. were kept since birth or for the past 40 days in a ASF free compartment showed no clinical sign of ASF on the day of shipment;
2. showed no clinical sign of ASF on the day of shipment were kept since birth or for the past 40 days in an ASF free compartment.

Article 2.6.6.7.

When importing fromASF free countries or zones, Veterinary Administrations should require:

for wild pigs

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of ASF on the day of shipment;
2. have been captured in an ASF free country or zone.

Article 2.6.6.8.

When importing from ASF free countries, zones or compartments, Veterinary Administrations should require:

for semen of domestic pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in an ASF free country, zone or compartment since birth or for at least 40 days prior to collection in accordance with 2.6.6.6.;
   b) showed no clinical sign of ASF on the day of collection of the semen;
2. the semen was collected, processed and stored in conformity with the provisions of Appendix 3.2.2.
Annex XVI (contd)

Article 2.6.6.9.

When importing from ASF infected countries or infected zones considered infected with ASF in domestic pigs, Veterinary Administrations should require:

for semen of domestic pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in an ASF free compartment and since birth or for at least 40 days prior to collection;
   b) showed no clinical sign of ASF on the day of collection of the semen and for the following 40 days;

2. the semen was collected in accordance with 2.6.6.8, processed and stored in conformity with the provisions of Appendix 3.2.2.

Article 2.6.6.10.

When importing from ASF free countries, zones or compartments, Veterinary Administrations should require:

for in vivo derived embryos of pigs

the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) were kept in an ASF free country, zone or compartment in domestic pigs since birth or for at least 40 days in accordance with 2.6.6.6 prior to collection;
   b) showed no clinical sign of ASF on the day of collection of the embryos;

2. the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1.

Article 2.6.6.11.

When importing from ASF infected countries or infected zones considered infected with ASF in domestic pigs, Veterinary Administrations should require:

for in vivo derived embryos of pigs

the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) were kept in an ASF free compartment and since birth or for at least 40 days prior to collection;
   b) showed no clinical sign of ASF on the day of collection of the embryos and for the following 40 days;
2. the embryos were collected in accordance with 2.6.6.10, processed and stored in conformity with the provisions of Appendix 3.3.1.

Article 2.6.6.12.

When importing from ASF free countries, zone or compartments, Veterinary Administrations should require:

for fresh meat of domestic pigs

the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in an ASF free country, zone or compartment since birth or for at least the past 40 days, or which have been imported in accordance with Article 2.6.6.5. or Article 2.6.6.6.

2. have been slaughtered in an approved abattoir, have been subjected to ante-mortem and post-mortem inspections in accordance with Appendix 3.10.1, and have been found free of any sign suggestive of ASF.

Article 2.6.6.13.

When importing from ASF free countries or zones, Veterinary Administrations should require:

for fresh meat of wild pigs

the presentation of an international veterinary certificate attesting that:

1. the entire consignment of meat comes from animals which:
   1. a) have been killed in an ASF free country or zone;
   2. b) have been subjected to a post-mortem inspection in accordance with Appendix 3.10.1, in an approved examination centre, and have been found free of any sign suggestive of ASF.

and, if the zone where the animal has been killed is adjacent to a zone with infection in wild pigs:

2. a sample has been collected from every animal killed and has been subjected to a virological test and a serological test for ASF, with negative results.

Article 2.6.6.14.

Veterinary Administrations of importing countries should require:

for meat products of pigs (either domestic or wild), or for products of animal origin (from fresh meat of pigs) intended for use in animal feeding, for agricultural or industrial use, or for pharmaceutical or surgical use, or for trophies derived from wild pigs

the presentation of an international veterinary certificate attesting that the products:

1. have been prepared:
   a) exclusively from fresh meat meeting the conditions laid down in Articles 2.6.6.12. or 2.6.6.13., as relevant;
 Annex XVI (contd)

b) in a processing establishment:
   i) approved by the Veterinary Administration Authority for export purposes;
   ii) processing only meat meeting the conditions laid down in Articles 2.6.6.12. or 2.6.6.13., as relevant;

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2. have been processed in an establishment approved by the Veterinary Administration Authority for export purposes so as to ensure the destruction of the ASF virus and that the necessary precautions were taken after processing to avoid contact of the product with any source of ASF virus.

Article 2.6.6.15.

Veterinary Authorities of importing countries should require:

for products of animal origin (from pigs, but not derived from fresh meat) intended for use in animal feeding and for agricultural or industrial use

the presentation of an international veterinary certificate attesting that the products:

1. have been prepared:
   a) exclusively from products meeting the conditions laid down for fresh meat in Articles 2.6.6.12. or 2.6.6.13., as relevant;
   b) in a processing establishment:
      i) approved by the Veterinary Administration Authority for export purposes;
      ii) processing only products meeting the conditions laid down in point a) above;

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2. have been processed in an establishment approved by the Veterinary Administration Authority for export purposes so as to ensure the destruction of the ASF virus and that the necessary precautions were taken after processing to avoid contact of the product with any source of ASF virus.

Article 2.6.6.15.bis

Veterinary Authorities of importing countries should require:

for bristles (from pigs)

the presentation of an international veterinary certificate attesting that the products:

1. come from an ASF free country, zone or compartment; or

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the ASF virus and that the necessary precautions were taken after processing to avoid contact of the product with any source of ASF virus.
Article 2.6.6.16.

Veterinary Administrations Authorities of importing countries should require:

for litter and manure (from pigs)

the presentation of an international veterinary certificate attesting that the products:

1. come from an ASF free country, zone or compartment; or

2. have been processed in an establishment approved by the Veterinary Administration Authority for export purposes so as to ensure the destruction of the ASF virus and that the necessary precautions were taken after processing to avoid contact of the product with any source of ASF virus.

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CHAPTER 2.6.7.

CLASSICAL SWINE FEVER

Article 2.6.7.1.

The pig is the only natural host for classical swine fever (CSF) virus. The definition of pig includes all varieties of Sus scrofa, both domestic breeds and wild boar. For the purposes of this chapter, a distinction is made between domestic pigs (permanently captive and owned farmed, free-range pigs) and wild pigs (including feral pigs).

Pigs exposed to CSF virus prenatally may be persistently infected throughout life and may have an incubation period of several months before showing signs of disease. Pigs exposed postnatally have an incubation period of 7-10 days, and are usually infective between post-infection days 5 and 14, but up to 3 months in cases of chronic infections.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

Article 2.6.7.2.

The CSF status of a country, zone or compartment can only be determined after considering the following criteria in domestic and wild pigs, as applicable:

1. a risk assessment has been conducted, identifying all potential factors for CSF occurrence and their historic perspective;

2. CSF should be notifiable in the whole country, and all clinical signs suggestive of CSF should be subjected to appropriate field and/or laboratory investigations;

3. an on-going awareness programme should be in place to encourage reporting of all cases suggestive of CSF;

4. the Veterinary Administration Authority should have current knowledge of, and authority over, all domestic pigs in the country, zone or compartment;

5. the Veterinary Administration Authority should have current knowledge about the population and habitat of wild pigs in the country or zone.

Article 2.6.7.3.

CSF free country, zone or compartment

1. CSF free status in the absence of an outbreak

   a) Historically free status

   A country, or zone or compartment may be considered free from CSF after conducting a risk assessment as referred to in Article 2.6.7.2, but without formally applying a specific surveillance programme, if the provisions of Article 3.8.1.6. are complied with.
Annex XVII (contd)

b) Free status as a result of a specific surveillance programme

A country, zone or compartment which does not meet the conditions of point 1 above may be considered free from CSF when a risk assessment as referred to in Article 2.6.7.2. has been conducted, surveillance in accordance with Appendix 3.8.8. has been in place for at least 12 months, and when no outbreak has been observed for at least 12 months.

2. CSF free status following an outbreak b) Free status as a result of an eradication programme

A country, or zone or compartment which does not meet the conditions of point a) or b) above or a compartment may be considered free from CSF when; if surveillance in accordance with Appendix 3.8.8. has been in place and after a risk assessment as referred to in Article 2.6.7.2. has been conducted, and

a) where a stamping-out policy without vaccination is practised and no outbreak has been observed in domestic pigs for at least 6 months;

OR

b) where a stamping-out policy with vaccination is practised, and either:

i) vaccinated pigs are slaughtered, and no outbreak has been observed in domestic pigs for at least 6 months after the last vaccinated pig was slaughtered; or

ii) where there are validated means of distinguishing between vaccinated and infected pigs, no outbreak has been observed in domestic pigs for at least 6 months;

OR

c) where a vaccination strategy is practised without a stamping-out policy;

i) vaccination has been banned in all domestic pigs in the country, zone or compartment for at least 12 months, unless there are validated means of distinguishing between vaccinated and infected pigs;

ii) if vaccination has been practised within the past 5 years, surveillance in accordance with Appendix 3.8.8. has been in place for at least 6 months to demonstrate the absence of infection within the population of domestic pigs 6 months to one year old; and

iii) no outbreak has been observed in domestic pigs for at least 12 months;

AND

in all cases, based on surveillance in accordance with Appendix 3.8.8., CSF infection is not known to occur in any wild pig population in the country or zone

i) there has been no outbreak of CSF during the past 12 months;

ii) no evidence of CSFV infection has been found during the past 12 months;

iii) no vaccination against CSF has been carried out during the past 12 months;

iv) surveillance in accordance with Appendix 3.8.8. has been in place in domestic pigs for the past 12 months;
AND

in the case of a country or zone, surveillance in accordance with Appendix 3.8.8. has been in place to determine the CSF status of the wild pig population, and:

v) there has been no clinical evidence or virological evidence of CSF in wild pigs during the past 12 months;

vi) no seropositive wild pigs have been detected in the age class 6-12 months during the past 12 months;

vii) there has been no vaccination in wild pigs for the past 12 months;

viii) imported wild pigs comply with the relevant requirements in Article 2.6.7.7.

Article 2.6.7.4.

Country free of CSF in domestic pigs but with a wild pig population

Requirements in points 2a to 2c of Article 2.6.7.3., as relevant, are complied with. As CSF infection may be present in the wild pig population, the following additional conditions are complied with:

1. a programme for the management of CSF in wild pigs is in place, taking into account the measures in place to manage the disease in the wild pig population, the presence of natural boundaries, the ecology of the wild pig population, and an assessment of the risk of disease spread;

2. zoning or compartmentalisation is applied the domestic pig population must be separated from the infected wild pig population through biosecurity measures to prevent transmission of CSF from wild pigs to domestic pigs.

Article 2.6.7.5.

Recovery of free status

Should a CSF outbreak occur in a previously free country, zone or compartment, the free status of the country, zone or compartment may be restored not less than 30 days after completion of a stamping-out policy where surveillance in accordance with Appendix 3.8.8. has been carried out with negative results, either:

If emergency vaccination has been practised within the CSF domestic pig control area, recovery of the free status cannot occur before all the vaccinated pigs have been slaughtered, unless there are validated means of distinguishing between vaccinated and infected pigs.

1. 3 months after the last case where a stamping-out policy without vaccination is practised;

OR

2. where a stamping-out policy with emergency vaccination is practised:

   a) 3 months after the last case and the slaughter of all vaccinated animals, or

   b) 3 months after the last case without the slaughter of vaccinated animals where there are validated means, validated to OIE standards (Chapter I.1.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs.
Annex XVII (contd)

OR

3. where a stamping-out policy is not practised, the provisions of point b) of Article 2.6.7.3 should be followed;

AND

in the case of a country or zone, based on surveillance in accordance with Appendix 3.8.8., CSFV infection is not known to occur in any wild pig population in the country or zone.

Article 2.6.7.6.

Country or zone free of CSF in wild pigs

A country or zone may be considered free from CSF in wild pigs when:

1. the domestic pig population in the country or zone is free from CSF infection;

2. surveillance in accordance with Appendix 3.8.8. has been in place to determine the CSF status of the wild pig population in the country, and in the country or zone

   a) there has been no clinical evidence, nor virological evidence of CSF in wild pigs during the past 12 months;

   b) no seropositive wild pigs have been detected in the age class 6-12 months during the past 12 months;

3. there has been no vaccination in wild pigs for the past 12 months;

4. the feeding of swill to wild pigs is forbidden, unless the swill has been treated to destroy any CSF virus that may be present, in conformity with one of the procedures referred to in Article 3.6.4.1.;

5. imported wild pigs comply with the relevant requirements set forth in the present chapter.

Article 2.6.7.7.

When importing from CSF free countries, zones or compartments, Veterinary Administrations should require:

for domestic pigs

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CSF on the day of shipment;

2. were kept in a country, zone or compartment free of CSF since birth or for at least the past 3 months;

3. have not been vaccinated against CSF, nor are they the progeny of vaccinated sows, unless there are validated means, validated to OIE standards (Chapter I.1.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs.
Article 2.6.7.8.

When importing from countries free of CSF in domestic pigs but wild pig population, Veterinary Administrations should require:

for domestic pigs

the presentation of an international veterinary certificate attesting that the animals:

1. were kept in a country or zone free of CSF in domestic pigs since birth or for at least the past 3 months;
2. have not been vaccinated against CSF, nor are they the progeny of vaccinated sows, unless there are validated means of distinguishing between vaccinated and infected pigs;
3. come from a CSF free zone or compartment;
4. showed no clinical sign of CSF on the day of shipment.

Article 2.6.7.9.

When importing from CSF infected countries or infected zones with CSF infection in domestic pigs, Veterinary Administrations should require:

for domestic pigs

the presentation of an international veterinary certificate attesting that the animals:

1. have not been vaccinated against CSF nor are they the progeny of vaccinated sows, unless there are validated means of distinguishing between vaccinated and infected pigs showed no clinical sign of CSF on the day of shipment;
2. were kept since birth or for the past 3 months in a CSF free compartment;
3. showed no clinical sign of CSF on the day of shipment have not been vaccinated against CSF nor are they the progeny of vaccinated sows, unless there are validated means, validated to OIE standards (Chapter I.1.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs.

Article 2.6.7.10.

When importing from CSF free countries or zones, Veterinary Administrations should require:

for wild pigs

the presentation of an international veterinary certificate attesting that the animals:

1. showed no clinical sign of CSF on the day of shipment;
2. have been captured in a country or zone free from CSF;
3. have not been vaccinated against CSF, unless there are validated means, validated to OIE standards (Chapter I.1.3. of the Terrestrial Manual), of distinguishing between vaccinated and infected pigs and, if the zone where the animal has been captured is adjacent to a zone with infection in wild pigs:
Annex XVII (contd)

4. were kept in a quarantine station for 40 days prior to shipment, and were subjected to a virological test and a serological test performed at least 21 days after entry into the quarantine station, with negative results.

Article 2.6.7.11a.

When importing from CSF free countries, zones or compartments, Veterinary Administrations should require:

for semen of domestic pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in a country, zone or compartment free of CSF since birth or for at least 3 months prior to collection;
   b) showed no clinical sign of CSF on the day of collection of the semen;

2. the semen was collected, processed and stored in conformity with the provisions of Appendix 3.2.2.

Article 2.6.7.12.

When importing from countries free of CSF in domestic pigs but with infection in the wild pig population, Veterinary Administrations should require:

for semen of domestic pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in a country, zone or compartment free of CSF in domestic pigs since birth or for at least 3 months prior to collection;
   b) showed no clinical sign of CSF on the day of collection of the semen and for the following 40 days;

2. the semen was collected, processed and stored in conformity with the provisions of Appendix 3.2.2.

Article 2.6.7.13a.

When importing from CSF infected countries or infected zones considered infected with CSF in domestic pigs, Veterinary Administrations should require:

for semen of domestic pigs

the presentation of an international veterinary certificate attesting that:

1. the donor animals:
   a) were kept in a compartment free of CSF in domestic pigs since birth or for at least 3 months prior to collection;
b) showed no clinical sign of CSF on the day of collection of the semen and for the following 40 days;

c) have not been vaccinated against CSF and were subjected to a serological test performed at least 21 days after collection, with negative results;

or

d) have been vaccinated against CSF and were subjected to a serological test performed at least 21 days after collection and it has been conclusively demonstrated by means, validated to OIE standards (Chapter I.1.3. of the Terrestrial Manual), that any antibody is due to the vaccine.

2. the semen was collected, processed and stored in conformity with the provisions of Appendix 3.2.2.

Article 2.6.7.1410.

When importing from CSF free countries, zones or compartments, Veterinary Administrations should require:

for in vivo derived embryos of pigs

the presentation of an international veterinary certificate attesting that:

1. the donor females showed no clinical sign of CSF on the day of collection of the embryos;

2. the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1.

Article 2.6.7.15.

When importing from countries free of CSF in domestic pigs but with infection in the wild pig population, Veterinary Administrations should require:

for in vivo derived embryos of pigs

the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) were kept in a country, zone or compartment free of CSF in domestic pigs since birth or for at least 3 months prior to collection;
   b) showed no clinical sign of CSF on the day of collection of the embryos;

2. the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1.

Article 2.6.7.1611.

When importing from CSF infected countries or infected zones considered infected with CSF in domestic pigs, Veterinary Administrations should require:

for in vivo derived embryos of pigs

Annex XVII (contd)
the presentation of an international veterinary certificate attesting that:

1. the donor females:
   a) were kept in a CSF free compartment in domestic pigs since birth or for at least 3 months prior to collection;
   b) showed no clinical sign of CSF on the day of collection of the embryos and for the following 40 days;
   c) have not been vaccinated against CSF and were subjected, with negative results, to a serological test performed at least 21 days after collection;
   or
   d) have been vaccinated against CSF and were subjected to a serological test performed at least 21 days after collection and it has been conclusively demonstrated by means, validated to OIE standards (Chapter I.1.3. of the Terrestrial Manual), that any antibody is due to the vaccine;

2. the embryos were collected, processed and stored in conformity with the provisions of Appendix 3.3.1.

Article 2.6.7.17(12).

When importing from CSF free countries, zones or compartments, Veterinary Administrations should require:

for fresh meat of domestic pigs

the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. have been kept in a country, zone or compartment free of CSF since birth or for at least the past 3 months, or which have been imported in accordance with Article 2.6.7.5. or Article 2.6.7.6.;

2. have been slaughtered in an approved abattoir, have been subjected to ante-mortem and post-mortem inspections in accordance with Appendix 3.10.1. and have been found free of any sign suggestive of CSF.

Article 2.6.7.18.

When importing from countries or zones free of CSF in domestic pigs but wild pig population, Veterinary Administrations should require:

for fresh meat of domestic pigs

the presentation of an international veterinary certificate attesting that the entire consignment of meat comes from animals which:

1. were kept in a country, zone or compartment free of CSF in domestic pigs since birth or for at least the past 3 months;
2. have been slaughtered in an approved abattoir, have been subjected to ante mortem and post mortem inspections as described in the Codex Alimentarius Code of Hygienic Practice for Meat and have been found free of any sign suggestive of CSF.

Article 2.6.7.1913.

When importing from CSF free countries or zones, Veterinary Authorities should require:

for fresh meat of wild pigs

the presentation of an international veterinary certificate attesting that:

1. the entire consignment of meat comes from animals which:
   a) have been killed in a CSF free country or zone;
   b) have been subjected to a post-mortem inspection as described in the Codex Alimentarius Code of Hygienic Practice for Meat in accordance with Appendix 3.10.1, in an approved examination centre, and have been found free of any sign suggestive of CSF;

and, if the zone where the animal has been killed is adjacent to a zone with infection in wild pigs:

2. a sample has been collected from every animal shot, and has been subjected to a virological test and a serological test for CSF, with negative results.

Article 2.6.7.2014

Veterinary Authorities of importing countries should require:

for meat products of pigs (either domestic or wild), or for products of animal origin (from fresh meat of pigs) intended for use in animal feeding, for agricultural or industrial use, or for pharmaceutical or surgical use, or for trophies derived from wild pigs

the presentation of an international veterinary certificate attesting that the products:

1. have been prepared:
   a) exclusively from fresh meat meeting the conditions laid down in Articles 2.6.7.1712, 2.6.7.18, or 2.6.7.1913, as relevant;
   b) in a processing establishment:
      i) approved by the Veterinary Authority for export purposes;
      ii) processing only meat meeting the conditions laid down in Articles 2.6.7.1712, 2.6.7.18, or 2.6.7.1913, as relevant;

OR

2. have been processed in an establishment approved by the Veterinary Authority for export purposes so as to ensure the destruction of the CSF virus in conformity with one of the procedures referred to in Article 3.6.4.2. and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.
Veterinary Administrations of importing countries should require:

for products of animal origin (from pigs, but not derived from fresh meat) intended for use in animal feeding and for agricultural or industrial use

the presentation of an international veterinary certificate attesting that the products:

1. have been prepared:
   a) exclusively from products meeting the conditions laid down for fresh meat in Articles 2.6.7.12, 2.6.7.18, or 2.6.7.19, as relevant;
   b) in a processing establishment:
      i) approved by the Veterinary Administration Authority for export purposes;
      ii) processing only products meeting the conditions laid down in point a) above;
   OR

2. have been processed in an establishment approved by the Veterinary Administration Authority for export purposes so as to ensure the destruction of the CSF virus in conformity with one of the procedures referred to in Article 3.6.4.2. and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.

Veterinary Administrations of importing countries should require:

for bristles (from pigs)

the presentation of an international veterinary certificate attesting that the products:

1. come from a CSF free country, zone or compartment; or

2. have been processed in an establishment approved by the Veterinary Administration Authority for export purposes so as to ensure the destruction of the CSF virus and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.

Veterinary Administrations of importing countries should require:

for litter and manure (from pigs)

the presentation of an international veterinary certificate attesting that the products:

1. come from a country, zone or compartment free of CSF; or
2. have been processed in an establishment approved by the Veterinary Administration Authority for export purposes so as to ensure the destruction of the CSF virus and that the necessary precautions were taken after processing to avoid contact of the product with any source of CSF virus.
APPENDIX 3.8.8.

GUIDELINES FOR THE ON-SURVEILLANCE OF CLASSICAL SWINE FEVER

Introduction

This Appendix defines the principles and provides a guide on the surveillance for classical swine fever (CSF), complementary to in accordance with Appendix 3.8.1., applicable to countries seeking to demonstrate recognition of freedom from CSF status. This may be for the entire country or a zone within the country. Guidance for countries seeking reestablishment of freedom from CSF for the whole country or a zone following an outbreak, as well as guidelines and for demonstrating the maintenance of CSF free status are also provided. This Appendix complements Chapter 2.6.7.

The impact and epidemiology of CSF differ widely in different regions of the world, and it is, therefore, impossible to provide specific guidelines for all situations. It is axiomatic that the surveillance strategies employed for demonstrating freedom from CSF at an acceptable level of confidence will need to be adapted to the local situation. For example, the approach must be tailored in order to prove freedom from CSF for a country or zone where wild pigs provide a potential reservoir of infection, or where CSF is present in adjacent countries. The method must examine the epidemiology of CSF in the region concerned and adapt to the specific risk factors encountered. This should include provision of scientifically based supporting data. There is, therefore, latitude available to Members to provide a well-reasoned argument to prove that absence of classical swine fever virus (CSFV) infection is assured at an acceptable level of confidence.

Surveillance for CSF should be in the form of a continuing programme designed to either establish that a population is free from CSFV infection (either the whole country, or a zone within the country is free from CSFV infection or a compartment) or to detect the introduction of CSFV into a population already recognized as free. Consideration should be given to the specific characteristics of CSF epidemiology which include: the role of swill feeding and the impact of different production systems on disease spread, the role of semen in transmission of the virus, the lack of pathognomonic gross lesions and clinical signs, the frequency of clinically inapparent infections, the occurrence of persistent and chronic infections, and the genotypic, antigenic, and virulence variability exhibited by different strains of CSFV. Serological cross-reactivity with other pestiviruses has to be taken into consideration when interpreting data from serological surveys. A common route by which ruminant pestiviruses can infect pigs is the use of vaccines contaminated with bovine viral diarrhoea virus (BVDV).

For the purposes of this Appendix, virus infection means presence of CSFV as demonstrated directly by virus isolation, the detection of virus antigen or virus nucleic acid, or indirectly by seroconversion which is not the result of vaccination.

General conditions and methods

1. A surveillance system in accordance with Appendix 3.8.1. should be under the responsibility of the Veterinary Administration Authority. A procedure should be in place for the rapid collection and transport of samples to an accredited laboratory as described in the Terrestrial Manual.
Annex XVII (contd)

2. The CSF surveillance programme should:

a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers, who have day-to-day contact with livestock, as well as diagnosticians, should report promptly any suspicion of CSF to the Veterinary Authority. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Administration. Since many strains of CSFV do not induce pathognomonic gross lesions or clinical signs, cases in which CSF cannot be ruled out should be immediately investigated employing clinical, pathological, and laboratory diagnosis. This requires that sampling kits and other equipment are available to those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in CSF diagnosis, epidemiological evaluation, and control;

b) implement, when relevant, regular and frequent clinical inspections and serological testing of high-risk groups of animals (for example, where swill feeding is practised), or those adjacent to a CSF infected country or zone (for example, bordering areas where infected wild pigs are present).

An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is CSFV. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot, therefore, be reliably predicted. Recognitions for freedom from CSFV infection should, as a consequence, provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement standstill orders, etc.).

Article 3.8.8.3.

Surveillance strategies

1. Introduction

There are two basic strategies that can be employed for CSF surveillance depending on the purpose of the country for seeking recognition of freedom from CSF. In countries historically free of CSF, surveillance programmes should be designed to detect the introduction of CSFV into domestic or wild swine. The optimal strategy to meet this objective is most often targeted surveillance.

The target population for covered by surveillance aimed at identifying detecting disease and infection should include domestic and wild pig populations within the country or zone to be recognised as free from CSFV infection. Such surveillance may involve opportunistic testing of samples submitted for other purposes, but a more efficient and effective strategy is one which includes targeted surveillance.

Depending on the local epidemiological situation, targeted surveillance could be considered as more effective than a randomized surveillance strategy. Surveillance is targeted to the pig population which presents the highest risk of infection (for example, swill fed farms, pigs reared outdoors or farms in proximity to infected wild pigs). Each country will need to identify its individual risk factors. These may include: temporal and spatial distribution of past outbreaks, pig movements and demographics, etc.
For reasons of cost, the longevity of antibody levels, as well as the existence of clinically inapparent infections and difficulties associated with differential diagnosis of other diseases, serology is often the most effective and efficient surveillance methodology. In some circumstances, which will be discussed later, clinical and virological surveillance may also have value.

The country should justify the surveillance strategy chosen as adequate to detect the presence of CSFV infection in accordance with Appendix 3.8.1. and the epidemiological situation. Cumulative survey results in combination with the results of passive surveillance, over time, will increase the level of confidence in the surveillance strategy. If a Member wishes to apply for recognition by other Members of a specific zone within the country as being free from CSFV infection, the design of the surveillance strategy and the basis for any sampling process would need to be aimed at the population within the zone.

For random surveys, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect infection if it were to occur at a predetermined minimum rate. The sample size and expected disease prevalence determine the level of confidence in the results of the survey. The country must justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Appendix 3.8.1. Selection of the design prevalence in particular clearly needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey design selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and production class of animals in the target population.

Irrespective of the testing system employed, the surveillance system design should anticipate the occurrence of false positive reactions. This is especially true of the serological diagnosis of CSF because of the recognized cross-reactivity with ruminant pestiviruses. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether or not they are indicative of CSFV infection. This should involve confirmatory and differential tests for pestiviruses, as well as further investigations concerning the original sampling unit as well as animals which may be epidemiologically linked.

2. Clinical and virological surveillance

Beyond their role in targeted surveillance, clinical and virological surveillance for CSF has two aims: a) to shorten the period between introduction of CSF virus into a disease free country or zone and its detection, and b) to confirm that no unnoticed outbreaks have occurred.

In the past, clinical identification of cases was the cornerstone of early detection of CSF. However, emergence of low virulence strains of CSF, as well as new diseases - in particular post-weaning multisystemic wasting syndrome and porcine dermatitis and nephropathy syndrome have made such reliance less effective, and in countries where such diseases are common, can add significant risk of masking the presence of CSF.
One element of clinical surveillance involves the detection of clinical signs of CSF by close physical examination of susceptible animals. The spectrum of disease signs and gross pathology seen in CSF infections, along with the plethora of other agents that can mimic CSF, renders the value of clinical examination alone somewhat inefficient as a surveillance tool. These factors, along with the compounding effects of concurrent infections and disease caused by ruminant pestiviruses, dictate the need for laboratory testing in order to clarify the status of CSF suspects detected by clinical monitoring.

Nevertheless, clinical presentation should not be ignored as a tool for early detection; in particular, any cases where clinical signs or lesions consistent with CSF are accompanied by high morbidity and/or mortality should be investigated without delay. In CSFV infections involving low virulence strains, high mortality may only be seen in young animals. Otherwise close physical examination of susceptible animals is useful as a selection criteria for CSF surveillance, particularly in diagnostic laboratories or slaughter establishments or when applied to high risk populations such as swill feeding operations.

In the past, clinical identification of cases was the cornerstone of early detection of CSF. However, emergence of low virulence strains of CSF, as well as new diseases—in particular post-weaning multisystemic wasting syndrome and porcine dermatitis and nephropathy syndrome have made such reliance less effective, and, in countries where such diseases are common, can add significant risk of masking the presence of CSF. In zones or countries where such diseases exist, careful clinical and virological surveillance of such cases should be applied.

Clinical signs and pathology of CSF infection will also vary considerably, depending on the strain of virus as well as host factors, such as age, nutrition and health status. These factors, along with the compounding effects of concurrent infections and disease caused by ruminant pestiviruses, dictate the need for laboratory testing in order to clarify the status of CSF suspects detected by clinical monitoring. The difficulties in detecting chronic disease manifested by non-specific clinical signs and delayed seroconversion and seronegativity, in persistently infected piglets, both of which may be clinically normal, makes virological investigation essential. As part of a herd investigation, such animals are likely to be in a minority and would not confound a diagnosis based on serology. Individually or as part of recently mixed batches, such animals may, however, escape detection by this method. A holistic approach to investigation, taking note of herd history, pig, personnel and vehicle movements and disease status in neighbouring zones or countries, can also assist in targeting surveillance in order to increase efficiency and enhance the likelihood of early detection.

The labour-intensive nature of clinical, pathological and virological investigations, along with the smaller ‘window of opportunity’ inherent in virus, rather than antibody detection, has, in the past, resulted in greater emphasis being placed on mass serological screening as the best method for surveillance. However, surveillance based on clinical and pathological inspection and virological testing should not be underrated. If targeted at high risk groups in particular, it provides an opportunity for early detection that can considerably reduce the subsequent spread of disease. Herds predominated by adult animals, such as nucleus herds and artificial insemination studs, are particularly useful groups to monitor, since infection by low virulence viruses in such groups may be clinically inapparent, yet the degree of spread may be high.

Clinical and virological monitoring may also provide a high level of confidence of rapid detection of disease if a sufficiently large number of clinically susceptible animals is examined. In particular, molecular detection methods are increasingly able to offer the possibility of such large-scale screening for the presence of virus, at reasonable cost.
Wild pigs and, in particular, those with a wholly free-living existence, rarely present the opportunity for clinical observation, but should form part of any surveillance scheme and should, ideally, be monitored for virus as well as antibody.

Vaccine design and diagnostic methodologies, and in particular methods of virus detection, are increasingly reliant on up-to-date knowledge of the molecular, antigenic and other biological characteristics of viruses currently circulating and causing disease. Furthermore, epidemiological understanding of the pathways of spread of CSFV can be greatly enhanced by molecular analyses of viruses in endemic areas and those involved in outbreaks in disease-free areas. It is therefore essential that CSFV isolates are sent regularly to the regional OIE Reference Laboratory for genetic and antigenic characterisation.

3. Serological surveillance

Serological surveillance aims at detecting antibodies against CSFV. Positive CSFV antibody test results can have five possible causes:

a) natural infection with CSFV;

b) legal or illegal vaccination against CSF;

c) maternal antibodies derived from an immune sow (maternal antibodies) are usually found only up to 4.5 months of age, but, in some individuals, maternal antibodies can be detected for considerably longer periods;

d) cross-reactions with other pestiviruses;

e) non-specific reactors.

The infection of pigs with other pestiviruses may complicate a surveillance strategy based on serology. Antibodies to bovine viral diarrhoea virus (BVDV) and Border disease virus (BDV) can give positive results in serological tests for CSF, due to common antigens. Such samples will require differential tests to confirm their identity. Although persistently infected immunotolerant pigs are themselves seronegative, they continuously shed virus, so the prevalence of antibodies at the herd level will be high. Chronically infected pigs may have undetectable or fluctuating antibody levels.

It may be possible to use sera collected for other survey purposes for CSF surveillance. However, the principles of survey design described in this Appendix and the requirement for statistical validity should not be compromised.

The discovery of clustering of seropositive reactions should be foreseen. It may reflect any of a series of events, including but not limited to the demographics of the population sampled, vaccinal exposure or the presence of infection by field strains or other pestiviruses. Because clustering may signal field strain infection, the investigation of all instances must be incorporated in the survey design. Clustering of positive animals is always epidemiologically significant and therefore should be investigated.

In countries or zones that are moving towards freedom, serosurveillance can provide valuable information on the disease status and efficacy of any control programme. Targeted serosurveillance of young stock will indicate whether newly circulating virus is present, although the presence of maternal antibody will also need to be considered. If conventional attenuated vaccine is currently being used or has been used in the recent past, serology aimed at detecting the presence of field virus will likewise need to be targeted at unvaccinated animals and after the disappearance of maternal antibody. General usage in such situations may also be used to assess levels of vaccine coverage.
Annex XVII (contd)

Vaccines also exist which, when used in conjunction with dedicated serological tests, may allow discrimination between vaccinal antibody and that induced by field infection. Such tools, described in the Terrestrial Manual, will need to be fully validated. They do not confer the same degree of protection as that provided by conventional vaccines, particularly with respect to preventing transplacental infections. Furthermore, serosurveillance using such differentiation requires cautious interpretation on a herd basis.

The results of random or targeted serological surveys are important in providing reliable evidence that no CSFV infection is present in a country or zone. It is therefore essential that the survey be thoroughly documented.

Article 3.8.8.4.

Country or zone historically free of CSF in domestic and wild pigs

1. Historically free status

The free status should be reviewed whenever evidence emerges to indicate that changes which may alter the underlying assumption of continuing historical freedom, has occurred. Such changes include but are not limited to:

a) an emergence or an increase in the prevalence of CSF in countries or zones from which live pigs or products are imported;

b) an increase in the volume of imports or a change in their country or zone of origin;

c) an increase in the prevalence of CSF in the domestic or wild pigs of adjacent countries or zones;

d) an increased entry from, or exposure to, infected wild pig populations of adjacent countries or zones.

2. Free status as a result of an eradication programme

In addition to the general conditions described in Chapter 2.6.7., a Member Country seeking recognition of CSF freedom for the country or a zone, whether or not vaccination had been practised, should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and will be planned and implemented according to the general conditions and methods described in this Appendix, to demonstrate the absence of CSFV infection in domestic and wild pig populations. This requires the support of a national or other laboratory able to undertake identification of CSFV infection through virus detection and serological tests described in the Terrestrial Manual.

Article 3.8.8.5

Countries, zones or compartments applying for freedom from CSF where vaccination is practised

1. Country or zone free of CSF

In addition to the general conditions described in Chapter 2.6.7., a Member seeking recognition of CSF freedom for the country or a zone, whether or not vaccination had been practised, should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances in and around the country or zone and will be planned and implemented according to the general conditions and methods described in this Appendix, to demonstrate the absence of CSFV infection in domestic and wild pig populations. This requires the support of a national or other laboratory able to undertake identification of CSFV infection through virus detection and serological tests described in the Terrestrial Manual.
2. **Compartment free of CSF**

The objective of surveillance in this instance is to demonstrate that the two subpopulations are effectively separated by measures that ensure the biosecurity of domestic pigs is to demonstrate the absence of CSFV infection in the compartment. The provisions of Chapter 1.3.5. should be followed. The effective separation of the two subpopulations should be demonstrated. To this end, a biosecurity programme which includes but is not limited to the following provisions should be implemented:

- a) a programme for the management of CSF in wild pigs;
- b) delineation of CSF wild pig control areas around every CSF case reported in wild pigs;
- c) assessment of the presence and mitigative role of natural boundaries;
- d) documentation of the ecology of the wild pig population;
- e) proper containment of domestic pigs;
- f) control of movement of vehicles with cleaning and disinfection as appropriate;
- g) control of personnel entering into the establishments and awareness of risk of fomite spread;
- h) prohibition of introduction to the establishments of hunted wild caught animals and their products;
- i) registry record of animal movements into and out of establishments;
- j) information and training programmes for farmers, hunters, processors, veterinarians, etc.

3. The biosecurity programme implemented would also requires internal and external monitoring by the Veterinary Authority. These elements should include but are not limited to:

- a) periodic clinical and serological monitoring of herds in the country or zone, and adjacent wild pig populations following these guidelines;
- b) herd registration;
- c) official accreditation of biosecurity programme plan;
- d) periodic monitoring and review.

4. Monitoring the CSF status of wild and domestic pig populations outside the compartment will be of value in assessing the degree of risk they pose to the CSF free domestic population compartment. The design of a monitoring system for wild pigs is dependent on several factors such as the size and distribution of the population, the organisation of the Veterinary Services and resources available. The occurrence of CSF in wild and domestic pigs may vary considerably among countries. Surveillance design should be epidemiologically based, and the Member must justify its choice of design prevalence and level of confidence based on Appendix 3.8.1.
Annex XVII (contd)

5. The geographic distribution and approximate size of wild pig populations need to be assessed as a prerequisite for designing a monitoring system. Sources of information may include wildlife conservation organisations, hunter associations and other available sources. The objective of a surveillance programme when the disease is already known to exist should be to determine the geographic distribution and the extent of the infection.

Article 3.8.8.6.

Recovery of free status

1. Countries or zones seeking re-establishment of freedom from CSF following an outbreak

In addition to the general conditions described in Chapter 2.6.7., a country seeking reestablishment of country or zone freedom from CSF should show evidence of an active surveillance programme for CSF as well as to demonstrate absence of CSFV infection.

Populations under this surveillance programme should include, but not be limited to:

a) establishments in the area proximity of the outbreak;

b) establishments epidemiologically linked to the outbreak;

c) animals used to re-populate affected establishments and any establishments where contiguous culling is carried out;

d) wild pig populations in the area of the outbreak.

In all circumstances, a Member seeking reestablishment of country or zone freedom from CSF with vaccination or without vaccination should report the results of an active and a passive surveillance programme in which the pig population undergoes regular clinical, pathological, virological, and/or serological examination, planned and implemented according to the general conditions and methods described in these guidelines. The surveillance should be based on a statistically representative sample of the populations at risk.

2. Country or zone free of Surveillance for CSF in wild pigs

While the same principles apply, surveillance in wild pigs presents challenges beyond those encountered in domestic populations in each of the following areas:

a) determination of the distribution, size and movement patterns associated with the wild pig population;

b) assessment of the possible presence of CSF within the population;

c) determination of the practicability of establishing a zone.

c) determination of the practicability of establishing a zone.

The design of a monitoring system for wild pigs is dependent on several factors such as the organisation of the Veterinary Services and resources available. The geographic distribution and approximate size of wild pig populations need to be assessed as a prerequisite for designing a monitoring system. Sources of information may include wildlife conservation organisations, hunter associations and other available sources. The objective of a surveillance programme is to determine the geographic distribution and estimation of target population.
The design of a monitoring system for wild pigs is dependent on several factors such as the organisation of the Veterinary Services and resources available. The geographic distribution and approximate size of wild pig populations need to be assessed as a prerequisite for designing a monitoring system. Sources of information may include wildlife conservation organisations, hunter associations and other available sources. The objective of a surveillance programme is to determine the geographic distribution and estimation of a target population.

Estimates of wild pig populations can be made using advanced methods (radio tracking, linear transect method, capture/recapture) or traditional methods based on the number of animals that can be hunted to allow for natural restocking (hunting bags).

For implementation of the monitoring programme, it will be necessary to define the limits of the territory over which wild pigs range in order to delineate the epidemiological units within the monitoring programme. It is often difficult to define epidemiological units for wild animals. The most practical approach is based on natural and artificial barriers.

The monitoring programme should also include animals found dead, road kills, animals showing abnormal behaviour or exhibiting gross lesions during dressing.

There may be situations where a more targeted surveillance programme can provide additional assurance. The criteria to define high risk areas for targeted surveillance can include:

a) areas with past history of CSF;
b) sub-regions with high wild pig density;
c) border regions with CSF affected countries or zones;
d) areas of contact interface between wild and domestic pig sub-populations;
e) picnic and camping areas;
f) around farms with free-ranging pigs;
g) garbage dumps;
h) special other risk areas determined by local the Veterinary Authorities.

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CHAPTER 2.7.12.

AVIAN INFLUENZA

Article 2.7.12.1.

1. For the purposes of international trade, avian influenza in its notifiable form (NAI) is defined as an infection of poultry caused by any influenza A virus of the H5 or H7 subtypes or by any AI virus with an intravenous pathogenicity index (IVPI) greater than 1.2 (or as an alternative at least 75% mortality) as described below. NAI viruses can be divided into highly pathogenic notifiable avian influenza (HPNAI) and low pathogenicity notifiable avian influenza (LPNAI):

a) HPNAI viruses have an IVPI in 6-week-old chickens greater than 1.2 or, as an alternative, cause at least 75% mortality in 4 to 8-week-old chickens infected intravenously. H5 and H7 viruses which do not have an IVPI of greater than 1.2 or cause less than 75% mortality in an intravenous lethality test should be sequenced to determine whether multiple basic amino acids are present at the cleavage site of the haemagglutinin molecule (HA0); if the amino acid motif is similar to that observed for other HPNAI isolates, the isolate being tested should be considered as HPNAI;

b) LPNAI are all influenza A viruses of H5 and H7 subtype that are not HPNAI viruses.

2. Poultry is defined as ‘all domesticated birds, including backyard poultry, used for the production of meat or eggs for consumption, for the production of other commercial products, for restocking supplies of game, or for breeding these categories of birds, as well as fighting cocks used for any purpose’.

Birds that are kept in captivity for any reason other than those reasons referred to in the preceding paragraph, including those that are kept for shows, races, exhibitions, competitions, breeding or selling these categories of birds as well as pet birds, are not considered to be poultry.

3. For the purposes of international trade, this chapter deals not only with the occurrence of clinical signs caused by NAI virus, but also with the presence of infection with NAI virus in the absence of clinical signs.

4. For the purposes of international trade, a country should not impose immediate trade bans in response to a notification of infection with HPAI and LPAI virus in birds other than poultry according to Article 2.1.1.3. of the Terrestrial Code.
Annex XVIII (contd)

5. Antibodies to H5 or H7 subtype of NAI virus, which have been detected in poultry and are not a consequence of vaccination, have to be further investigated. In the case of isolated serological positive results, NAI infection may be ruled out on the basis of a thorough epidemiological investigation that does not demonstrate further evidence of NAI infection.

6. The following defines the occurrence of infection with NAI virus:

a) HPNAI virus has been isolated and identified as such or viral RNA specific for HPNAI has been detected in poultry or a product derived from poultry; or

b) LPNAI virus has been isolated and identified as such or viral RNA specific for LPNAI has been detected in poultry or a product derived from poultry.

For the purposes of the Terrestrial Code, ‘NAI free establishment’ means an establishment in which the poultry have shown no evidence of NAI infection, based on surveillance in accordance with Appendix 3.8.9.

For the purposes of the Terrestrial Code, the incubation period for NAI shall be 21 days.

Standards for diagnostic tests, including pathogenicity testing, are described in the Terrestrial Manual. Any vaccine used should comply with the standards described in the Terrestrial Manual.

Article 2.7.12.2.

The NAI status of a country, a zone or a compartment can be determined on the basis of the following criteria:

1. the outcome of a risk assessment identifying all potential factors for NAI occurrence and their historic perspective;

21. NAI is notifiable in the whole country, an on-going NAI awareness programme is in place, and all notified suspect occurrences of NAI are subjected to field and, where applicable, laboratory investigations;

32. appropriate surveillance is in place to demonstrate the presence of infection in the absence of clinical signs in poultry, and the risk posed by birds other than poultry; this may be achieved through an NAI surveillance programme in accordance with Appendix 3.8.9.

3. consideration of all epidemiological factors for NAI occurrence and their historical perspective.

Article 2.7.12.3.

NAI free country, zone or compartment

A country, zone or compartment may be considered free from NAI when it has been shown that neither HPNAI nor LPNAI infection has been present in the country, zone or compartment for the past 12 months, based on surveillance in accordance with Appendix 3.8.9. The surveillance may need to be adapted to parts of the country or existing zones or compartments depending on historical or geographical factors, industry structure, population data, or proximity to recent outbreaks.
If infection has occurred in a previously free country, zone or compartment, NAI free status can be regained:

1. In the case of HPNAI infections, 3 months after a stamping-out policy (including disinfection of all affected establishments) is applied, providing that surveillance in accordance with Appendix 3.8.9. has been carried out during that three-month period.

2. In the case of LPNAI infections, poultry may be kept for slaughter for human consumption subject to conditions specified in Article 2.7.12.18. or 2.7.12.19. or a stamping-out policy may be applied; in either case, 3 months after the disinfection of all affected establishments, providing that surveillance in accordance with Appendix 3.8.9. has been carried out during that three-month period.

Article 2.7.12.4.

**HPNAI free country, zone or compartment**

A country, zone or compartment may be considered free from HPNAI when it has been shown that HPNAI infection has not been present in the country, zone or compartment for the past 12 months, although its LPNAI status may be unknown, or when, based on surveillance in accordance with Appendix 3.8.9., it does not meet the criteria for freedom from NAI but any NAI virus detected has not been identified as HPNAI virus. The surveillance may need to be adapted to parts of the country or existing zones or compartments depending on historical or geographical factors, industry structure, population data, or proximity to recent outbreaks.

If infection has occurred in a previously free country, zone or compartment, HPNAI free status can be regained 3 months after a stamping-out policy (including disinfection of all affected establishments) is applied, providing that surveillance in accordance with Appendix 3.8.9. has been carried out during that three-month period.

Article 2.7.12.5.

When importing from an NAI free country, zone or compartment, Veterinary Authorities should require:

for live poultry (other than day-old poultry)

the presentation of an international veterinary certificate attesting that:

1. the poultry showed no clinical sign of NAI on the day of shipment;

2. the poultry were kept in an NAI free country, zone or compartment since they were hatched or for at least the past 21 days;
Annex XVIII (contd)

3. the required surveillance, in accordance with Appendix 3.8.9., has been carried out on the establishment within at least the past 21 days;

4. if vaccinated, the poultry have been vaccinated in accordance with Appendix 3.8.9., and the relevant information is attached.

**Article 2.7.12.6.**

Regardless of the NAI status of the country, zone or compartment of origin, Veterinary Authorities should require:

for live birds other than poultry

the presentation of an international veterinary certificate attesting that:

1. the birds showed no clinical sign of infection with a virus which would be considered NAI in poultry on the day of shipment;

2. the birds were kept in isolation approved by the Veterinary Services since they were hatched or for at least the 21 days prior to shipment and showed no clinical sign of infection with a virus which would be considered NAI in poultry during the isolation period;

3. the birds were subjected to a diagnostic test 7 to 14 days prior to shipment to demonstrate freedom from infection with a virus which would be considered NAI in poultry;

4. the birds are transported in new containers.

If the birds have been vaccinated, the relevant information should be attached to the certificate.

**Article 2.7.12.7.**

When importing from an NAI free country, zone or compartment, Veterinary Authorities should require:

for day-old live poultry

the presentation of an international veterinary certificate attesting that:

1. the poultry were kept in an NAI free country, zone or compartment since they were hatched;

2. the poultry were derived from parent flocks which had been kept in an NAI free country, zone or compartment for at least 21 days prior to and at the time of the collection of the eggs;

3. if the poultry or the parent flocks were vaccinated, vaccination was carried out in accordance with Appendix 3.8.9., and the relevant information is attached.

**Article 2.7.12.8.**

When importing from an HPNAI free country, zone or compartment, Veterinary Authorities should require:
for day-old live poultry

the presentation of an international veterinary certificate attesting that:

1. the poultry were kept in an HPNAI free country, zone or compartment since they were hatched;
2. the poultry were derived from parent flocks which had been kept in an NAI free establishment for at least 21 days prior to and at the time of the collection of the eggs;
3. the poultry are transported in new containers;
4. if the poultry or the parent flocks were vaccinated, vaccination was carried out in accordance with Appendix 3.8.9., and the relevant information is attached.

Article 2.7.12.9.

When importing from an NAI free country, zone or compartment, Veterinary Authorities should require:

for hatching eggs

the presentation of an international veterinary certificate attesting that:

1. the eggs came from an NAI free country, zone or compartment;
2. the eggs were derived from parent flocks which had been kept in an NAI free country, zone or compartment for at least 21 days prior to and at the time of the collection of the eggs;
3. if the parent flocks were vaccinated, vaccination was carried out in accordance with Appendix 3.8.9., and the relevant information is attached.

Article 2.7.12.10.

When importing from an HPNAI free country, zone or compartment, Veterinary Authorities should require:

for hatching eggs

the presentation of an international veterinary certificate attesting that:

1. the eggs came from an HPNAI free country, zone or compartment;
2. the eggs were derived from parent flocks which had been kept in an NAI free establishment for at least 21 days prior to and at the time of the collection of the eggs;
3. the eggs have had their surfaces sanitised (in accordance with Article 3.4.1.7.) and are transported in new packing material;
4. if the parent flocks were vaccinated, vaccination was carried out in accordance with Appendix 3.8.9., and the relevant information is attached.
Annex XVIII (contd)

Article 2.7.12.11.

When importing from an NAI free country, zone or compartment, Veterinary Authorities should require:

for eggs for human consumption

the presentation of an international veterinary certificate attesting that the eggs come from an NAI free country, zone or compartment.

Article 2.7.12.12.

When importing from an HPNAI free country, zone or compartment, Veterinary Authorities should require:

for eggs for human consumption

the presentation of an international veterinary certificate attesting that the eggs:

1. come from an HPNAI free country, zone or compartment;

2. have had their surfaces sanitised (in accordance with Article 3.4.1.7.) and are transported in new packing material.

Article 2.7.12.13.

When importing from an NAI free country, zone or compartment, Veterinary Authorities should require:

for egg products

the presentation of an international veterinary certificate attesting that the egg products come from, and were processed in, an NAI free country, zone or compartment.

Article 2.7.12.14.

Regardless of the NAI status of the country, zone or compartment of origin. When importing from a country, zone or compartment not considered free from NAI, Veterinary Authorities should require:

for egg products

the presentation of an international veterinary certificate attesting that:

1. the egg products are derived from eggs which meet the requirements of Articles 2.7.12.9., 2.7.12.10., 2.7.12.11. or 2.7.12.12.; or

2. the egg products were processed to ensure the destruction of NAI virus in accordance with Appendix 3.6.5.;

3. the necessary precautions were taken after processing to avoid contact of the commodity with any source of NAI virus.
Annex XVIII (contd)

Article 2.7.12.15.
When importing from an NAI free country, zone or compartment, Veterinary Authorities should require:

for poultry semen

the presentation of an international veterinary certificate attesting that the donor poultry:

1. showed no clinical sign of NAI on the day of semen collection;
2. were kept in an NAI free country, zone or compartment for at least the 21 days prior to and at the time of semen collection.

Article 2.7.12.16.
When importing from an HPNAI free country, zone or compartment, Veterinary Authorities should require:

for poultry semen

the presentation of an international veterinary certificate attesting that the donor poultry:

1. showed no clinical sign of HPNAI on the day of semen collection;
2. were kept in an HPNAI free country, zone or compartment for at least the 21 days prior to and at the time of semen collection.

Article 2.7.12.17.
Regardless of the NAI status of the country, zone or compartment of origin, Veterinary Authorities should require:

for semen of birds other than poultry

the presentation of an international veterinary certificate attesting that the donor birds:

1. were kept in isolation approved by the Veterinary Services for at least the 21 days prior to semen collection;
2. showed no clinical sign of infection with a virus which would be considered NAI in poultry during the isolation period;
3. were tested between 7 and 14 days prior to semen collection and shown to be free of NAI infection.

Article 2.7.12.18.
When importing from an NAI free country, zone or compartment, Veterinary Authorities should require:
Annex XVIII (contd)

for fresh meat of poultry

the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from birds:

1. which have been kept in an NAI free country, zone or compartment since they were hatched or for at least the past 21 days;

2. which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections to rule out the presence of NAI with favourable results.

Article 2.7.12.19.

When importing from an HPNAI free country, zone or compartment, Veterinary Authorities should require:

for fresh meat of poultry

the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from birds:

1. which have been kept in an HPNAI free country, zone or compartment since they were hatched or for at least the past 21 days;

2. which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections to rule out the presence of NAI with favourable results.

Article 2.7.12.20.

Regardless of the NAI status of the country, zone or compartment of origin, Veterinary Authorities should require:

for meat products of poultry

the presentation of an international veterinary certificate attesting that:

1. the commodity is derived from fresh meat which meets the requirements of Articles 2.7.12.18. or 2.7.12.19.; or

2. the commodity has been processed to ensure the destruction of avian influenza virus in accordance with Appendix 3.6.5.;

3. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

Article 2.7.12.21.

Regardless of the NAI status of the country, zone or compartment of origin, Veterinary Authorities should require:

for products of poultry origin intended for use in animal feeding, or for agricultural or industrial use
the presentation of an international veterinary certificate attesting that:

1. these commodities come from poultry which have been kept in an NAI free country, zone or compartment since they were hatched or for at least the past 21 days; or

2. these commodities have been processed to ensure the destruction of avian influenza virus (under study);

3. the necessary precautions were taken to avoid contact of the commodity with any source of NAI virus.

**Article 2.7.12.22.**

Regardless of the NAI status of the country, zone or compartment of origin, Veterinary Authorities should require:

for feathers and down (from poultry)

the presentation of an international veterinary certificate attesting that:

1. these commodities come from poultry which have been kept in an NAI free country, zone or compartment since they were hatched or for at least the past 21 days; or

2. these commodities have been processed to ensure the destruction of avian influenza virus (under study);

3. the necessary precautions were taken to avoid contact of the commodity with any source of avian influenza virus.

**Article 2.7.12.23.**

Regardless of the NAI status of the country, zone or compartment, Veterinary Authorities should require for the importation of:

meat or other products from birds other than poultry

the presentation of an international veterinary certificate attesting that:

1. the commodity has been processed to ensure the destruction of avian influenza virus (under study);

2. the necessary precautions were taken after processing to avoid contact of the commodity with any source of NAI virus.
APPENDIX 3.6.5.

GUIDELINES FOR ON THE INACTIVATION OF THE AVIAN INFLUENZA VIRUS

Article 3.6.5.1.

Eggs and egg products

The following times for industry standard temperatures are suitable for the inactivation of highly pathogenic notifiable avian influenza (HPNAI) virus present in eggs and egg products:

<table>
<thead>
<tr>
<th>Temperature (°C)</th>
<th>Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Whole egg</td>
<td>60</td>
</tr>
<tr>
<td>Whole egg blends</td>
<td>60</td>
</tr>
<tr>
<td>Whole egg blends</td>
<td>61.1</td>
</tr>
<tr>
<td>Liquid egg white</td>
<td>55.6</td>
</tr>
<tr>
<td>Liquid egg white</td>
<td>56.7</td>
</tr>
<tr>
<td>10% salted yolk</td>
<td>62.2</td>
</tr>
<tr>
<td>Dried egg white</td>
<td>67</td>
</tr>
<tr>
<td>Dried egg white</td>
<td>54.4</td>
</tr>
</tbody>
</table>

The listed temperatures are indicative of a range that achieves a 7-log kill. Where scientifically documented, variances from these times and temperatures may also be suitable when they achieve the inactivation of the virus.

Article 3.6.5.2.

Meat

A procedure which produces a core temperature of 70°C for 3.5 seconds is suitable for the inactivation of HPNAI virus present in meat.

<table>
<thead>
<tr>
<th>Temperature (°C)</th>
<th>Time</th>
</tr>
</thead>
<tbody>
<tr>
<td>Poultry meat</td>
<td>60.0</td>
</tr>
<tr>
<td></td>
<td>65.0</td>
</tr>
<tr>
<td></td>
<td>70.0</td>
</tr>
<tr>
<td></td>
<td>73.9</td>
</tr>
</tbody>
</table>

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OIE Terrestrial Animal Health Standards Commission/September 2007
APPENDIX 3.8.9.

GUIDELINES FOR THE SURVEILLANCE OF AVIAN INFLUENZA

Introduction

This Appendix defines the principles and provides a guide on the surveillance for notifiable avian influenza (NAI) complementary to, in accordance with Appendix 3.8.1., applicable to countries seeking to demonstrate recognition for a declared NAI status, with or without the use of vaccination. This may be for the entire country, zone or compartment. Guidance for countries seeking free status following an outbreak and for the maintenance of NAI status is also provided. This Appendix complements Chapter 2.7.12.

The presence of avian influenza viruses in wild birds creates a particular problem. In essence, no country can declare itself free from avian influenza (AI) in wild birds. However, the definition of NAI in Chapter 2.7.12. refers to the infection in poultry only, and this Appendix was developed under this definition.

The impact and epidemiology of NAI differ widely in different regions of the world and therefore it is impossible to provide specific guidelines for all situations. It is axiomatic that the surveillance strategies employed for demonstrating freedom from NAI at an acceptable level of confidence will need to be adapted to the local situation. Variables such as the frequency of contacts of poultry with wild birds, different biosecurity levels and production systems and the commingling of different susceptible species including domestic waterfowl require specific surveillance strategies to address each specific situation. It is incumbent upon the country to provide scientific data that explains the epidemiology of NAI in the region concerned and also demonstrates how all the risk factors are managed. There is therefore considerable latitude available to Members to provide a well reasoned argument to prove that absence of NAI virus (NAIV) infection is assured at an acceptable level of confidence.

Surveillance for NAI should be in the form of a continuing programme designed to establish that the country, zone or compartment, for which application is made, is free from NAIV infection.

Article 3.8.9.2.

General conditions and methods

1. A surveillance system in accordance with Appendix 3.8.1. should be under the responsibility of the Veterinary Authority. In particular:
   a) a formal and ongoing system for detecting and investigating outbreaks of disease or NAI infection should be in place;
   b) a procedure should be in place for the rapid collection and transport of samples from suspect cases of NAI to a laboratory for NAI diagnosis as described in the Territorial Manual;
   c) a system for recording, managing and analysing diagnostic and surveillance data should be in place.
2. The NAI surveillance programme should:

   a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers, who have day-to-day contact with poultry, as well as diagnosticians, should report promptly any suspicion of NAI to the Veterinary Services. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Authority. All suspected cases of NAI should be investigated immediately. As suspicion cannot be resolved by epidemiological and clinical investigation alone, samples should be taken and submitted to an approved laboratory. This requires that sampling kits and other equipment are available for those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in NAI diagnosis and control. In cases where potential public health implications are suspected, notification to the appropriate public health authorities is essential;

   b) implement, when relevant, regular and frequent clinical inspection, serological and virological testing of high-risk groups of animals, such as those adjacent to an NAI infected country, zone or compartment, places where birds and poultry of different origins are mixed, such as live bird markets, poultry in close proximity to waterfowl or other sources of NAIV.

An effective surveillance system will periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is NAI. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. Applications for freedom from NAI infection should, in consequence, provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement standstill orders, etc.).

Article 3.8.9.3.

Surveillance strategies

1. Introduction

   The target population for surveillance aimed at identification of disease and infection should cover all the susceptible poultry species within the country, zone or compartment. Active and passive surveillance for NAI should be ongoing. The frequency of active surveillance should be at least every 6 months. Surveillance should be composed of random and targeted approaches using virological, serological and clinical methods.

   The strategy employed may be based on randomised sampling requiring surveillance consistent with demonstrating the absence of NAIV infection at an acceptable level of confidence. The frequency of sampling should be dependent on the epidemiological situation. Random surveillance is conducted using serological tests described in the Terrestrial Manual. Positive serological results should be followed up with virological methods.

   Targeted surveillance (e.g. based on the increased likelihood of infection in particular localities or species) may be an appropriate strategy. Virological and serological methods should be used concurrently to define the NAI status of high risk populations.
A country should justify the surveillance strategy chosen as adequate to detect the presence of NAIV infection in accordance with Appendix 3.8.1. and the prevailing epidemiological situation, including cases of HPNAI detected in any birds. It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clear clinical signs (e.g. chickens). Similarly, virological and serological testing could be targeted to species that may not show clinical signs (e.g. ducks).

If a Member wishes to declare freedom from NAIV infection in a specific zone or compartment, the design of the survey and the basis for the sampling process would need to be aimed at the population within the zone or compartment.

For random surveys, the design of the sampling strategy will need to incorporate epidemiologically appropriate design prevalence. The sample size selected for testing will need to be large enough to detect infection if it were to occur at a predetermined minimum rate. The sample size and expected disease prevalence determine the level of confidence in the results of the survey. The applicant country must justify the choice of design prevalence and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Appendix 3.8.1. Selection of the design prevalence in particular clearly needs to be based on the prevailing or historical epidemiological situation.

Irrespective of the survey approach selected, the sensitivity and specificity of the diagnostic tests employed are key factors in the design, sample size determination and interpretation of the results obtained. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and the different species in the target population.

Irrespective of the testing system employed, surveillance system design should anticipate the occurrence of false positive reactions. If the characteristics of the testing system are known, the rate at which these false positives are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of infection or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as flocks which may be epidemiologically linked to it.

The principles involved in surveillance for disease/infection are technically well defined. The design of surveillance programmes to prove the absence of NAIV infection/circulation needs to be carefully followed to avoid producing results that are either insufficiently reliable, or excessively costly and logistically complicated. The design of any surveillance programme, therefore, requires inputs from professionals competent and experienced in this field.

2. Clinical surveillance

Clinical surveillance aims at the detection of clinical signs of NAI at the flock level. Whereas significant emphasis is placed on the diagnostic value of mass serological screening, surveillance based on clinical inspection should not be underrated. Monitoring of production parameters, such as increased mortality, reduced feed and water consumption, presence of clinical signs of a respiratory disease or a drop in egg production, is important for the early detection of NAIV infection. In some cases, the only indication of LPNAIV infection may be a drop in feed consumption or egg production.

Clinical surveillance and laboratory testing should always be applied in series to clarify the status of NAI suspects detected by either of these complementary diagnostic approaches. Laboratory testing may confirm clinical suspicion, while clinical surveillance may contribute to confirmation of positive serology. Any sampling unit within which suspicious animals are detected should be classified as infected until evidence to the contrary is produced.
Identification of suspect flocks is vital to the identification of sources of NAIV and to enable the molecular, antigenic and other biological characteristics of the virus to be determined. It is essential that NAIV isolates are sent regularly to the regional Reference Laboratory for genetic and antigenic characterization.

3. **Virological surveillance**

Virological surveillance using tests described in the Terrestrial Manual should be conducted:

a) to monitor at risk populations;

b) to confirm clinically suspect cases;

c) to follow up positive serological results;

d) to test 'normal' daily mortality, to ensure early detection of infection in the face of vaccination or in establishments epidemiologically linked to an outbreak.

4. **Serological surveillance**

Serological surveillance aims at the detection of antibodies against NAIV. Positive NAIV antibody test results can have four possible causes:

a) natural infection with NAIV;

b) vaccination against NAIV;

c) maternal antibodies derived from a vaccinated or infected parent flock are usually found in the yolk and can persist in progeny for up to 4 weeks;

d) false positive results due to the lack of specificity of the test.

It may be possible to use serum collected for other survey purposes for NAI surveillance. However, the principles of survey design described in these guidelines and the requirement for a statistically valid survey for the presence of NAIV should not be compromised.

The discovery of clusters of seropositive flocks may reflect any of a series of events, including but not limited to the demographics of the population sampled, vaccinal exposure or infection. As clustering may signal infection, the investigation of all instances must be incorporated in the survey design. Clustering of positive flocks is always epidemiologically significant and therefore should be investigated.

If vaccination cannot be excluded as the cause of positive serological reactions, diagnostic methods to differentiate antibodies due to infection or vaccination should be employed.

The results of random or targeted serological surveys are important in providing reliable evidence that no NAIV infection is present in a country, zone or compartment. It is therefore essential that the survey be thoroughly documented.

5. **Virological and serological surveillance in vaccinated populations**

The surveillance strategy is dependent on the type of vaccine used. The protection against AI is haemagglutinin subtype specific. Therefore, two broad vaccination strategies exist: 1) inactivated whole AI viruses, and 2) haemagglutinin expression-based vaccines.

In the case of vaccinated populations, the surveillance strategy should be based on virological and/ or serological methods and clinical surveillance. It may be appropriate to use sentinel birds for this purpose. These birds should be unvaccinated, AI virus antibody free birds and clearly and permanently identified. The interpretation of serological results in the presence of vaccination is described in Article 3.8.9.7.
Article 3.8.9.4.

Documentation of NAI or HPNAI free status

1. **Countries declaring freedom from NAI or HPNAI for the country, zone or compartment**

   In addition to the general conditions described in the Terrestrial Code, a Member declaring freedom from NAI or highly pathogenic notifiable avian influenza (HPNAI) for the entire country, or a zone or a compartment should provide evidence for the existence of an effective surveillance programme. The strategy and design of the surveillance programme will depend on the prevailing epidemiological circumstances and should be planned and implemented according to general conditions and methods described in this Appendix, to demonstrate absence of NAIV or HPNAIV infection, during the preceding 12 months in susceptible poultry populations (vaccinated and non-vaccinated). This requires the support of a laboratory able to undertake identification of NAIV or HPNAIV infection through virus detection and antibody tests described in the Terrestrial Manual. This surveillance may be targeted to poultry population at specific risks linked to the types of production, possible direct or indirect contact with wild birds, multi-age flocks, local trade patterns including live bird markets, use of possibly contaminated surface water, and the presence of more than one species on the holding and poor biosecurity measures in place.

2. **Additional requirements for countries, zones or compartments that practise vaccination**

   Vaccination to prevent the transmission of HPNAI virus may be part of a disease control programme. The level of flock immunity required to prevent transmission will depend on the flock size, composition (e.g. species) and density of the susceptible poultry population. It is therefore impossible to be prescriptive. The vaccine must also comply with the provisions stipulated for NAI vaccines in the Terrestrial Manual. Based on the epidemiology of NAI in the country, zone or compartment, it may be that a decision is reached to vaccinate only certain species or other poultry subpopulations.

   In all vaccinated flocks there is a need to perform virological and serological tests to ensure the absence of virus circulation. The use of sentinel poultry may provide further confidence of the absence of virus circulation. The tests have to be repeated at least every 6 months or at shorter intervals according to the risk in the country, zone or compartment.

   Evidence to show the effectiveness of the vaccination programme should also be provided.

Article 3.8.9.5.

Countries, zones or compartments declaring that they have regained freedom from NAI or HPNAI following an outbreak

In addition to the general conditions described in Chapter 2.7.12., a country declaring that it has regained country, zone or compartment freedom from NAI or HPNAI virus infection should show evidence of an active surveillance programme depending on the epidemiological circumstances of the outbreak to demonstrate the absence of the infection. This will require surveillance incorporating virus detection and antibody tests described in the Terrestrial Manual. The use of sentinel birds may facilitate the interpretation of surveillance results.
Annex XVIII (contd)

A Member declaring freedom of country, zone or compartment after an outbreak of NAI or HPNAI (with or without vaccination) should report the results of an active surveillance programme in which the NAI or HPNAI susceptible poultry population undergoes regular clinical examination and active surveillance planned and implemented according to the general conditions and methods described in these guidelines. The surveillance should at least give the confidence that can be given by a randomized representative sample of the populations at risk.

Article 3.8.9.6.

NAI free establishments within HPNAI free compartments

The declaration of NAI free establishments requires the demonstration of absence of NAIV infection. Birds in these establishments should be randomly tested using virus detection or isolation tests, and serological methods, following the general conditions of these guidelines. The frequency of testing should be based on the risk of infection and at a maximum interval of 21 days.

Article 3.8.9.7.

The use and interpretation of serological and virus detection tests

Poultry infected with NAI virus produce antibodies to haemagglutinin (HA), neuraminidase (NA), non-structural proteins (NSPs), nucleoprotein/matrix (NP/M) and the polymerase complex proteins. Detection of antibodies against the polymerase complex proteins will not be covered in this Appendix. Tests for NP/M antibodies include direct and blocking ELISA, and agar gel immunodiffusion (AGID) tests. Tests for antibodies against NA include the neuraminidase inhibition (NI), indirect fluorescent antibody and direct ELISA tests. For the HA, antibodies are detected in haemagglutination inhibition (HI) and neutralization (SN) tests. The HI test is reliable in avian species but not in mammals. The SN test can be used to detect subtype specific antibodies to the haemagglutinin and is the preferred test for mammals and some avian species. The AGID test is reliable for detection of NP/M antibodies in chickens and turkeys, but not in other avian species. As an alternative, blocking ELISA tests have been developed to detect NP/M antibodies in all avian species.

The HI and NI tests can be used to subtype AI viruses into 16 haemagglutinin and 9 neuraminidase subtypes. Such information is helpful for epidemiological investigations and in categorization of AI viruses.

Poultry can be vaccinated with a variety of AI vaccines including inactivated whole AI virus vaccines, and haemagglutinin expression-based vaccines. Antibodies to the haemagglutinin confer subtype specific protection. Various strategies can be used to differentiate vaccinated from infected birds including serosurveillance in unvaccinated sentinel birds or specific serological tests in the vaccinated birds.

AI virus infection of unvaccinated birds including sentinels is detected by antibodies to the NP/M, subtype specific HA or NA proteins, or NSP. Poultry vaccinated with inactivated whole AI vaccines containing an influenza virus of the same H sub-type but with a different neuraminidase may be tested for field exposure by applying serological tests directed to the detection of antibodies to the NA of the field virus. For example, birds vaccinated with H7N3 in the face of a H7N1 epidemic may be differentiated...
from infected birds (DIVA) by detection of subtype specific NA antibodies of the N1 protein of the field virus. Alternatively, in the absence of DIVA, inactivated vaccines may induce low titres of antibodies to NSP and the titre in infected birds would be markedly higher. Encouraging results have been obtained experimentally with this system, but it has not yet been validated in the field. In poultry vaccinated with haemagglutinin expression-based vaccines, antibodies are detected to the specific HA, but not any of the other AI viral proteins. Infection is evident by antibodies to the NP/M or NSP, or the specific NA protein of the field virus. Vaccines used should comply with the standards of the Terrestrial Manual.

All flocks with seropositive results should be investigated. Epidemiological and supplementary laboratory investigation results should document the status of NAI infection/circulation for each positive flock. A confirmatory test should have a higher specificity than the screening test and sensitivity at least equivalent than that of the screening test.

Information should be provided on the performance characteristics and validation of tests used.

1. The follow-up procedure in case of positive test results if vaccination is used

   In case of vaccinated populations, one has to exclude the likelihood that positive test results are indicative of virus circulation. To this end, the following procedure should be followed in the investigation of positive serological test results derived from surveillance conducted on NAI-vaccinated poultry. The investigation should examine all evidence that might confirm or refute the hypothesis that the positive results to the serological tests employed in the initial survey were not due to virus circulation. All the epidemiological information should be substantiated, and the results should be collated in the final report.

   Knowledge of the type of vaccine used is crucial in developing a serological based strategy to differentiate infected from vaccinated animals.

   a) Inactivated whole AI virus vaccines can use either homologous or heterologous neuraminidase subtypes between the vaccine and field strains. If poultry in the population have antibodies to NP/M and were vaccinated with inactivated whole AI virus vaccine, the following strategies should be applied:

      i) sentinel birds should remain NP/M antibody negative. If positive for NP/M antibodies, indicating AI virus infection, specific HI tests should be performed to identify H5 or H7 AI virus infection;

      ii) if vaccinated with inactivated whole AI virus vaccine containing homologous NA to field virus, the presence of antibodies to NSP could be indicative of infection. Sampling should be initiated to exclude the presence of NAIV by either virus isolation or detection of virus specific genomic material or proteins;

      iii) if vaccinated with inactivated whole AI virus vaccine containing heterologous NA to field virus, presence of antibodies to the field virus NA or NSP would be indicative of infection. Sampling should be initiated to exclude the presence of NAIV by either virus isolation or detection of virus specific genomic material or proteins.

   b) Haemagglutinin expression-based vaccines contain the HA protein or gene homologous to the HA of the field virus. Sentinel birds as described above can be used to detect AI infection. In vaccinated or sentinel birds, the presence of antibodies against NP/M, NSP or field virus NA is indicative of infection. Sampling should be initiated to exclude the presence of NAIV by either virus isolation or detection of virus specific genomic material or proteins.
Annex XVIII (contd)

2. **The follow-up procedure in case of positive test results indicative of infection for determination of infection due to HPNAI or LPNAI virus**

The detection of antibodies indicative of a NAI virus infection as indicated in point a)(i) above will result in the initiation of epidemiological and virological investigations to determine if the infections are due to HPNAI or LPNAI viruses.

Virological testing should be initiated in all antibody-positive and at risk populations. The samples should be evaluated for the presence of AI virus, by virus isolation and identification, and/or detection of influenza A specific proteins or nucleic acids (Figure 2). Virus isolation is the gold standard for detecting infection by AI virus and the method is described in the Terrestrial Manual. All AI virus isolates should be tested to determine HA and NA subtypes, and in vivo tested in chickens and/or sequencing of HA proteolytic cleavage site of H5 and H7 subtypes for determination of classification as HPNAI, LPNAI or LPAI (not notifiable) viruses. As an alternative, nucleic acid detection tests have been developed and validated; these tests have the sensitivity of virus isolation, but with the advantage of providing results within a few hours. Samples with detection of H5 and H7 HA subtypes by nucleic acid detection methods should either be submitted for virus isolation, identification, and in vivo testing in chickens, or sequencing of nucleic acids for determination of proteolytic cleavage site as HPNAI or LPNAI viruses. The antigen detection systems, because of low sensitivity, are best suited for screening clinical field cases for infection by Type A influenza virus looking for NP/M proteins. NP/M positive samples should be submitted for virus isolation, identification and pathogenicity determination.

Laboratory results should be examined in the context of the epidemiological situation. Corollary information needed to complement the serological survey and assess the possibility of viral circulation includes but is not limited to:

a) characterization of the existing production systems;

b) results of clinical surveillance of the suspects and their cohorts;

c) quantification of vaccinations performed on the affected sites;

d) sanitary protocol and history of the affected establishments;

e) control of animal identification and movements;

f) other parameters of regional significance in historic NAIV transmission.

The entire investigative process should be documented as standard operating procedure within the epidemiological surveillance programme.
Fig. 1. Schematic representation of laboratory tests for determining evidence of NAI infection through or following serological surveys.
Annex XVIII (contd)

Fig. 2. **Schematic representation of laboratory tests for determining evidence of NAI infection using virological methods**

The above diagram indicates the tests which are recommended for use in the investigation of poultry flocks.

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CHAPTER 2.7.13.

NEWCASTLE DISEASE

Article 2.7.13.1.

1. For the purposes of the international trade, an outbreak of Newcastle disease (ND) for the purpose of the Terrestrial Code is defined in the Terrestrial Manual is defined as an infection of birds poultry caused by a virus of avian paramyxovirus serotype 1 (APMV-1), termed virulent Newcastle disease virus (vNDV), that meets one of the following criteria for virulence:

   a) the virus has an intracerebral pathogenicity index (ICPI) in day-old chicks (Gallus gallus) of 0.7 or greater; or

   b) multiple basic amino acids have been demonstrated in the virus (either directly or by deduction) at the C-terminus of the F2 protein and phenylalanine at residue 117, which is the N-terminus of the F1 protein. The term ‘multiple basic amino acids’ refers to at least three arginine or lysine residues between residues 113 and 116. Failure to demonstrate the characteristic pattern of amino acid residues as described above would require characterisation of the isolated virus by an ICPI test.

In this definition, amino acid residues are numbered from the N-terminus of the amino acid sequence deduced from the nucleotide sequence of the F0 gene, 113–116 corresponds to residues -4 to -1 from the cleavage site.

Viruses classified as APMV-1 are synonymous with Newcastle disease virus (NDV). Those viruses that meet the criteria of virulence to be the cause of ND are termed virulent Newcastle disease virus (vNDV). All other APMV-1s that do not meet the criteria for vNDV are termed low virulent NDV (loNDV).

2. Poultry is defined as ‘all domesticated birds including backyard poultry, used for the production of meat or eggs for consumption, for the production of other commercial products, for restocking supplies of game, or for breeding these categories of birds, as well as fighting cocks used for any purpose’. All backyard and game fowl regardless of use will be defined as poultry.

Birds that are kept in captivity for any reason other than those defined as poultry reasons referred to in the preceding paragraph, including those that are kept for shows, races, exhibitions, competitions, or sale breeding or selling these categories of birds as well as pet birds, are not considered to be poultry.

3. This chapter only deals with vNDV infection of birds poultry as defined in point 1 above, in the presence or absence of clinical signs. For the purposes of international trade, a country should interpret as not impose immediate trade bans in response to reports occurrence of infection with vNDV in birds other than poultry according to the Terrestrial Code and should not impose immediate trade bans, although such infections should be notified.

4. The occurrence of infection with vNDV is defined as the isolation and identification of:

   a) vNDV has been isolated and identified as such or the detection of viral RNA specific for vNDV has been detected.
Annex XIX (contd)

b) For the purposes of the Terrestrial Code, the incubation period for ND shall be 21 days.

c) Standards for diagnostic tests, including pathogenicity testing, are described in the Terrestrial Manual. When the use of ND vaccines is appropriate, those vaccines should comply with the standards described in the Terrestrial Manual.

Article 2.7.13.2.

The ND status of a country, a zone or a compartment can only be determined and certified on the basis of the following criteria:

1. ND is notifiable in the whole country, an on-going ND awareness programme is in place, and all notified suspect occurrences of ND are subjected to field and, where applicable, laboratory investigations;

2. appropriate surveillance is in place to demonstrate the presence of vNDV infection in the absence of clinical signs in poultry, this may be achieved through an ND surveillance programme in accordance with Appendix 3.8.x.;

3. consideration of all epidemiological factors for ND occurrence and their historical perspective.

Article 2.7.13.3.

ND free country, zone or compartment

A country, zone or compartment may be considered free from ND when it has been shown that vNDV infection has not been present in the country, zone or compartment for the past 12 months, based on surveillance in accordance with Appendix x.x.x. The surveillance may need to be adapted to parts of the country or existing zones or compartments depending on historical or geographical factors, industry structure, population data, or proximity to recent outbreaks.

If infection has occurred in a previously free country, zone or compartment, ND free status can be regained three months after a stamping-out policy (including disinfection of all affected establishments) is applied, providing that surveillance in accordance with Appendix x.x.x. has been carried out during that three-month period.

Article 2.7.13.4.

When importing from an ND free country, zone or compartment as defined in Article 2.7.13.3., Veterinary Administrations should require:

for live poultry (other than day-old poultry)

the presentation of an international veterinary certificate attesting that:

1. the poultry showed no clinical sign suggestive of ND on the day of shipment;

2. the poultry were kept in an ND free country, zone or compartment since they were hatched or for at least the past 21 days;

3. the poultry have not been vaccinated against ND; if the birds were vaccinated against ND, the nature of the vaccine used and the date of vaccination shall be attached to the certificate;

4. the birds are transported in new or appropriately sanitized containers.
Article 2.7.13.5.

Regardless of the ND status of the country, zone or compartment of origin, Veterinary Administrations Authorities should require:

for live birds other than poultry

the presentation of an international veterinary certificate attesting that:

1. the birds showed no clinical sign suggestive of ND on the day of shipment;

2. the birds were kept in isolation approved by the Veterinary Services since they were hatched or for at least the 21 days prior to shipment and showed no clinical sign of infection with a virus which would be considered ND in poultry during the isolation period;

3. the birds were subjected to a diagnostic test 7 to 14 days prior to shipment to demonstrate freedom from infection with vNDV;

4. the birds are transported in new or appropriately sanitized containers the birds have not been vaccinated against ND; if the birds were vaccinated against ND, the nature of the vaccine used and the date of vaccination shall also be attached to the certificate;

5. the birds have not been vaccinated against ND or if the birds were vaccinated against ND the nature of the vaccine used and the date of vaccination shall also be attached to the certificate the birds are transported in new or appropriately sanitized containers.

Article 2.7.13.6.

When importing from an ND free country, zone or compartment as defined in Article 2.7.13.3., Veterinary Administrations Authorities should require:

for day-old live poultry

the presentation of an international veterinary certificate attesting that:

1. the poultry were hatched and kept in an ND free country, zone or compartment;

2. the poultry were derived from parent flocks which had been kept in an ND free country, zone or compartment for at least 21 days prior to and at the time of the collection of the eggs;

3. the poultry have not been vaccinated against ND; if poultry or parent flocks were vaccinated against ND, the nature of the vaccine used and the date of vaccination shall also be attached to the certificate;

4. the birds poultry are transported in new or appropriately sanitized containers.

Article 2.7.13.7.

Regardless of the ND status of the country, zone or compartment, Veterinary Administrations Authorities should require:

for day-old live birds other than poultry

the presentation of an international veterinary certificate attesting that:
Annex XIX (contd)

1. the birds showed no clinical sign suggestive of ND on the day of shipment;
2. the birds were hatched and kept in isolation approved by the Veterinary Services;
3. the parent flock birds were subjected to a diagnostic test at the time of the collection of the eggs to demonstrate freedom from infection with vNDV;
4. the birds are transported in new or appropriately sanitized containers;
5. the birds have not been vaccinated against ND or if the birds or parent flocks were vaccinated against ND the nature of the vaccine used and the date of vaccination shall also be attached to the certificate.

When importing from an ND free country, zone or compartment as defined in Article 2.7.13.3., Veterinary Administrations should require:

for hatching eggs from poultry

the presentation of an international veterinary certificate attesting that:

1. the eggs came from an ND free country, zone or compartment;
2. the eggs were derived from parent flocks which had been kept in an ND free country, zone or compartment for at least 21 days prior to and at the time of the collection of the eggs;
3. the parent flocks have not been vaccinated against ND; or if parent flocks were vaccinated against ND the nature of the vaccine used and the date of vaccination shall also be attached to the certificate;
4. the eggs are transported in new or appropriately sanitized containers.

Regardless of the ND status of the country, zone or compartment origin, Veterinary Administrations should require:

for hatching eggs from birds other than poultry

the presentation of an international veterinary certificate attesting that:

1. the parent flock birds were subjected to a diagnostic test 7 days prior to and at the time of the collection of the eggs to demonstrate freedom from infection with vNDV;
2. the birds eggs are transported in new or appropriately sanitized containers;
3. the parent flocks have not been vaccinated against ND; if parent flocks were vaccinated against ND, the nature of the vaccine used and the date of vaccination shall also be attached to the certificate.

When importing from an ND free country, zone or compartment as defined in Article 2.7.13.3., Veterinary Administrations should require:

for poultry eggs for human consumption
Annex XIX (contd)

the presentation of an international veterinary certificate attesting that:

1. the eggs were produced and packed in an ND free country, zone or compartment;
2. the eggs are transported in new or appropriately sanitized packing material.

Article 2.7.13.11

When importing from an ND free country, zone or compartment as defined in Article 2.7.13.3., Veterinary Administrations Authorities should require:

for poultry egg products

the presentation of an international veterinary certificate attesting that:

1. the egg products come from, and were processed in, an ND free country, zone or compartment;
2. the egg products are transported in new or appropriately sanitized containers.

Article 2.7.13.12

Regardless of the ND status of the country, zone or compartment of origin

When importing from a country, zone or compartment not considered free from ND, Veterinary Administrations Authorities should require:

for poultry egg products

the presentation of an international veterinary certificate attesting that:

1. the commodity is processed to ensure the destruction of vNDV (under study);
2. the necessary precautions were taken after processing to avoid contact of the commodity with any source of vNDV;
3. the egg products are transported in new or appropriately sanitized containers.

Article 2.7.13.13

When importing from an ND free country, zone or compartment as defined in Article 2.7.13.3., Veterinary Administrations Authorities should require:

for poultry semen

the presentation of an international veterinary certificate attesting that the donor poultry:

1. showed no clinical sign suggestive of ND on the day of semen collection;
2. were kept in an ND free country, zone or compartment for at least the 21 days prior to and at the time of semen collection.

Article 2.7.13.14

Regardless of the ND status of the country, zone or compartment of origin, Veterinary Administrations Authorities should require:
Annex XIX (contd)

for semen of birds other than poultry

the presentation of an international veterinary certificate attesting that the donor birds:

1. were kept in isolation approved by the Veterinary Services for at least the 21 days prior to and on the day of semen collection;

2. showed no clinical sign suggestive of infection with vNDV during the isolation period and on the day of semen collection;

3. were subjected to a diagnostic test 7 to 14 days prior to semen collection to demonstrate freedom from infection with vNDV.

Article 2.7.13.15.14.

When importing from an ND free country, zone or compartment as defined in Article 2.7.13.3., Veterinary Administrations should require:

for fresh meat of poultry

the presentation of an international veterinary certificate attesting that the entire consignment of fresh meat comes from birds:

1. which have been kept and slaughtered in an ND free country, zone or compartment since they were hatched or for at least the past 21 days;

2. which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections in accordance with Appendix 3.10.1. and have been found free of any sign suggestive of ND.

Article 2.7.13.16.

When importing from an ND free country, zone or compartment, Veterinary Authorities should require:

for meat products of poultry

the presentation of an international veterinary certificate attesting that:

1. the commodity is derived from fresh meat which meets the requirements of Article 2.7.13.15. and has been processed in an ND free country, zone or compartment;

2. the necessary precautions were taken to avoid contact of the commodity with any source of vNDV.

Article 2.7.13.16.

When importing from a country, zone or compartment not considered free from ND. Regardless of the ND status of the country, zone or compartment of origin, Veterinary Administrations should require:

for meat products of poultry

the presentation of an international veterinary certificate attesting that:

1. the entire consignment of meat comes from animals which have been slaughtered in an approved abattoir and have been subjected to ante-mortem and post-mortem inspections and have been found free of any signs suggestive of ND;
Annex XIX (contd)

12. the commodity is derived from fresh meat which meet the requirements of Article 2.7.13.15 (fresh meat) and has been processed in an ND free country, zone or compartment, or the commodity has been processed to ensure the destruction of vNDV (under study);

23. the necessary precautions were taken to avoid contact of the commodity with any source of vNDV.

Article 2.7.13.17.
When importing from a ND free country, zone or compartment, Veterinary Authorities should require:
for products of poultry origin intended for use in animal feeding, or for agricultural or industrial use
the presentation of an international veterinary certificate attesting that:
1. the commodities come from poultry which have been kept and processed in an ND free country, zone or compartment since they were hatched or for at least the past 21 days;
2. the necessary precautions were taken to avoid contact of the commodity with any source of vNDV.

Article 2.7.13.17.bis
When importing from a country, zone or compartment not considered free from ND, Regardless of the ND status of the country, zone or compartment of origin, Veterinary Administrations should require:
for products of poultry origin intended for use in animal feeding, or for agricultural or industrial use
the presentation of an international veterinary certificate attesting that:
1. these commodities come from poultry which have been kept and processed in an ND free country, zone or compartment since they were hatched or for at least the past 21 days, or these commodities have been processed to ensure the destruction of vNDV (under study);
2. the necessary precautions were taken to avoid contact of the commodity with any source of vNDV.

Article 2.7.13.18.
Regardless of the ND status of the country, zone or compartment of origin, Veterinary Administrations should require:
for feathers and down
the presentation of an international veterinary certificate attesting that:
1. these commodities come from poultry which have been kept and processed in an ND free country, zone or compartment since they were hatched or for at least the past 21 days, or these commodities have been processed to ensure the destruction of vNDV (under study);
2. the necessary precautions were taken to avoid contact of the commodity with any source of vNDV.

Article 2.7.13.19.
Regardless of the ND status of the country, zone or compartment, Veterinary Administrations should require for the importation of:
meat or other products from birds other than poultry

the presentation of an international veterinary certificate attesting that:

1. the commodity has been processed to ensure the destruction of vNDV (under study);

2. the necessary precautions were taken after processing to avoid contact of the commodity with any source of vNDV.
Annex XIX (contd)

APPENDIX 3.8.X.

GUIDELINES ON SURVEILLANCE FOR NEWCASTLE DISEASE

Article 3.8.X.1.

Introduction

This Appendix defines the principles and provides a guide on the surveillance for Newcastle Disease (ND) complementary to in accordance with Appendix 3.8.1., applicable to countries seeking to demonstrate recognition for a declared ND status, with or without the use of vaccination. This may be for the entire country, zone or compartment. Guidance for countries seeking free status following an outbreak and for the maintenance of ND status is also provided. This Appendix complements Chapter 2.7.13.

Surveillance for ND is complicated by the known prevalence of avian paramyxovirus serotype 1 (APMV-1) infections in many bird species, both domestic and wild, and the widespread utilization of ND vaccines in domestic poultry. Consequently it is required that APMV-1 isolates synonymous with Newcastle disease virus (NDV) be characterized to differentiate those infections of virulent NDV (vNDV) that are notifiable as defined in Chapter 2.7.13, from those of low virulence (loNDV) which are not.

Newcastle Disease (ND) is described defined in Chapter 2.7.13 as an infection of birds with APMV-1, however this appendix is only concerned with vNDV infections of poultry.

The impact and epidemiology of ND differ widely in different regions of the world and therefore it is not possible to provide specific guidelines for all situations. Therefore surveillance strategies employed for demonstrating freedom from ND at an acceptable level of confidence will need to be adapted to the local situation. Variables such as the frequency of contacts of poultry with wild birds, different biosecurity levels, production systems and the commingling of different susceptible species require specific surveillance strategies to address each specific situation. It is incumbent upon the country to provide scientific data that explains the epidemiology of ND in the region concerned and also demonstrates how all the risk factors are managed. There is therefore considerable latitude available to Members to provide a well-reasoned argument to prove freedom from vNDV infection.

Surveillance for ND should be in the form of a continuing programme designed to establish that the country, zone or compartment, for which application is made, is free from vNDV infection.

Article 3.8.X.2.

General conditions and methods

1. A surveillance system in accordance with Appendix 3.8.1. should be under the responsibility of the Veterinary Administration Authority. In particular there should be in place:

a) a formal and ongoing system for detecting and investigating outbreaks of disease or vNDV infection;

b) a procedure for the rapid collection and transport of samples from suspect cases of ND to an approved laboratory for ND diagnosis as described in the Terrestrial Manual;

c) a system for recording, managing and analysing diagnostic and surveillance data.
Annex XIX (contd)

2. The ND surveillance programme should:

a) include an early warning system throughout the production, marketing and processing chain for reporting suspicious cases. Farmers and workers, who have day-to-day contact with poultry, as well as diagnosticians, should report promptly any suspicion of ND to the Veterinary Authority. They should be supported directly or indirectly (e.g. through private veterinarians or veterinary para-professionals) by government information programmes and the Veterinary Administration. All suspected cases of ND should be investigated immediately. As suspicion cannot be resolved by epidemiological and clinical investigation alone, samples should be taken and submitted to an approved laboratory. This requires that sampling kits and other equipment are available to those responsible for surveillance. Personnel responsible for surveillance should be able to call for assistance from a team with expertise in ND diagnosis and control;

b) implement, when relevant, regular and frequent clinical, virological and serological surveillance of high risk groups of poultry within the target population, (e.g. those adjacent to an ND infected population, country, zone, compartment, places where birds and poultry of different origins are mixed, or other sources of vNDV).

An effective surveillance system may periodically identify suspicious cases that require follow-up and investigation to confirm or exclude that the cause of the condition is due to vNDV infection. The rate at which such suspicious cases are likely to occur will differ between epidemiological situations and cannot therefore be predicted reliably. Applications for freedom from vNDV infection should provide details of the occurrence of suspicious cases and how they were investigated and dealt with. This should include the results of laboratory testing and the control measures to which the animals concerned were subjected during the investigation (quarantine, movement stand-still orders, etc.).

Article 3.8.X.3.

Surveillance strategies

1. Introduction

The principles involved in surveillance for disease / infection are technically well defined. Any surveillance programme requires inputs from professionals competent and experienced in this field and should be thoroughly documented. The design of surveillance programmes to prove the absence of vNDV infection / circulation needs to be carefully followed to avoid producing results that are either unreliable, or excessively costly and logistically complicated.

If a country wishes to declare freedom from vNDV infection in a country, zone or compartment, the sub-population used for surveillance of the disease / infection should be representative of all poultry within the country, zone or compartment. Multiple surveillance methods should be used concurrently to accurately define the true ND status of poultry populations. Active and passive surveillance for ND should be ongoing with the frequency of active surveillance being at least every 6 months. Surveillance should be composed of random and/or targeted approaches, dependent on the local epidemiological situation and using clinical, virological and serological methods as described in the Terrestrial Manual (Chapter x.x.x.x). If alternative tests are used they must have been validated as fit-for-purpose in accordance with OIE standards. A country should justify the surveillance strategy chosen as adequate to detect the presence of vNDV infection in accordance with Appendix 3.8.1. and the prevailing epidemiological situation.
For random surveillance surveys, the design of the sampling strategy will need to incorporate an epidemiologically appropriate design to demonstrate the prevalence of vNDV infection. The sample size selected for testing should be large enough to detect infection if it were to occur at a predetermined minimum rate. The sample size and expected disease prevalence determine the level of confidence in the results of the survey. The survey design and frequency of sampling should be dependent on the historical and current local epidemiological situation. The applicant country must justify the choice of survey design and confidence level based on the objectives of surveillance and the epidemiological situation, in accordance with Appendix 3.8.1.

Targeted surveillance (e.g. based on the increased likelihood of infection in a population) may be an appropriate strategy.

It may, for example, be appropriate to target clinical surveillance at particular species likely to exhibit clear clinical signs (e.g. unvaccinated chickens). Similarly, virological and serological testing could target species that may not show clinical signs (Article 2.7.13.2.) of ND and are not routinely vaccinated (e.g. ducks). Surveillance may also target poultry populations at specific risk, for example direct or indirect contact with wild birds, multi-age flocks, local trade patterns including live poultry markets, the presence of more than one species on the holding and poor biosecurity measures in place. In situations where wild birds have been shown to play a role in the local epidemiology of ND, surveillance of wild birds may be of value in alerting Veterinary Services to the possible exposure of free ranging poultry.

The sensitivity and specificity of the diagnostic tests are key factors in the choice of survey design, which should anticipate the occurrence of false positive and false negative reactions. Ideally, the sensitivity and specificity of the tests used should be validated for the vaccination/infection history and for the different species in the target population. If the characteristics of the testing system are known, the rate at which these false reactions are likely to occur can be calculated in advance. There needs to be an effective procedure for following up positives to ultimately determine with a high level of confidence, whether they are indicative of infection or not. This should involve both supplementary tests and follow-up investigation to collect diagnostic material from the original sampling unit as well as flocks which may be epidemiologically linked to it.

The results of active and passive surveillance are important in providing reliable evidence that no vNDV infection is present in a country, zone or compartment.

2. **Clinical surveillance**

Clinical surveillance aims to detect clinical signs suggestive of ND at the flock level and should not be underestimated as an early indication of infection. Monitoring of production parameters (e.g. a drop in feed or water consumption or egg production) is important for the early detection of vNDV infection in some populations, as there may be no, or mild clinical signs, particularly if they are vaccinated. Any sampling unit within which suspicious animals are detected should be considered as infected until evidence to the contrary is produced. Identification of infected flocks is vital to the identification of sources of vNDV.

A presumptive diagnosis of clinical ND in suspect infected populations should always be confirmed by virological testing in an approved laboratory. This will enable the molecular, antigenic and other biological characteristics of the virus to be determined.

It is desirable that NDV isolates are sent promptly to an OIE Reference Laboratory for archiving and further characterization if required.

3. **Virological surveillance**

Virological surveillance should be conducted using tests described in the Terrestrial Manual to:
Annex XIX (contd)

a) monitor at risk populations;
b) confirm suspect clinical cases;
c) follow up positive serological results in unvaccinated populations or sentinel birds;
d) test ‘normal’ daily mortalities (if warranted by an increased risk e.g. infection in the face of vaccination or in establishments epidemiologically linked to an outbreak).

4. Serological surveillance

Where systematic vaccination is carried out, serological surveillance is of limited value. Serological surveillance cannot be used to discriminate between vNDV and other NDV strains aims at the detection of antibodies against NDV but is not diagnostic of the presence of vNDV. Test procedures and interpretations of results are as described in Chapter x.x.x of the Terrestrial Manual. Positive NDV antibody test results can have four five possible causes:
a) natural infection with NDV;
b) vaccination against ND (whether intentional or not);
c) exposure to vaccine virus;
de) maternal antibodies derived from a vaccinated or infected parent flock are usually found in the yolk and can persist in progeny for up to 4 weeks;
d) non-specific test reactions.

It may be possible to use serum collected for other survey purposes for ND surveillance. However, the principles of survey design described in these guidelines and the requirement for a statistically valid survey for the presence of NDV should not be compromised.

Discovery of seropositive, unvaccinated flocks must be investigated further by conducting a thorough epidemiological investigation. Since seropositive results are not necessarily indicative of active infection, virological surveillance methods should be used to confirm the presence of vNDV in such populations. Until validated strategies and tools to differentiate vaccinated animals from those infected with field ND viruses are available, serological tools should not be used to identify NDV infection in vaccinated populations.

5. Use of sentinel poultry

There are various applications of the use of sentinel poultry as a surveillance tool in susceptible populations to detect virus circulation by the presence of clinical disease or seroconversion. They may be used to monitor vaccinated populations or species which are less susceptible to the development of clinical disease for the circulation of virus. Sentinel poultry should ideally be immunologically naïve and may be used in vaccinated flocks subject to a risk assessment. In case of the use of sentinel poultry, the structure and organisation of the poultry sector, the type of vaccine used and local epidemiological factors will determine the type of production systems where sentinels should be placed, the frequency of placement and monitoring of the sentinels.

Sentinel poultry must be in close contact with, but should be identified to be clearly differentiated from, the target population. Sentinel poultry must be observed regularly for evidence of clinical disease and any disease incidents investigated by prompt virological laboratory testing. The species to be used as sentinels should be proven to be highly susceptible to infection and ideally develop clear signs of clinical disease. Where the sentinel poultry do not necessarily develop overt clinical disease a programme of regular active testing by virological and serological tests should be used (the development of clinical disease may be dependent on the sentinel species used or use of live vaccine in the target population that may infect the sentinel poultry). The testing regime and the interpretation of the results will depend on the type of vaccine used in the target population.
Article 3.8.X.4.

Documentation of ND free status

The requirements for a country, zone or compartment to declare freedom from ND are given in Article x.x.13.3.

A country declaring freedom of a country, zone or compartment (with or without vaccination) should report the results of a surveillance programme in which the ND susceptible poultry population undergoes regular surveillance planned and implemented according to the general conditions and methods described in these guidelines.

A country, zone or compartment may be considered free from ND when it has been shown that vNDV infection has not been present in the country, zone or compartment for the past 12 months, based on surveillance in accordance with Appendix x.x.x. The surveillance may need to be adapted to parts of the country or existing zones or compartments depending on historical or geographical factors, industry structure, population data, or proximity to recent outbreaks.

If infection has occurred in a previously free country, zone or compartment, ND free status can be regained three months after a stamping-out policy (including disinfection of all affected establishments) is applied, providing that surveillance in accordance with Appendix x.x.x. has been carried out during that three-month period.

1. Countries declaring freedom from ND for the country, zone or compartment

   In addition to the general conditions described in the Terrestrial Code, a Member declaring freedom from ND for the entire country, or a zone or a compartment should provide evidence for the existence of an effective surveillance programme. The surveillance programme should be planned and implemented according to general conditions and methods described in this Appendix to demonstrate absence of vNDV infection in poultry during the preceding 12 months. This requires the support of an approved laboratory capable of identification of vNDV infection through virus detection and antibody tests described in the Terrestrial Manual.

2. Additional requirements for countries, zones or compartments that practice vaccination

   Vaccination against ND may be used for risk management (to reduce the risk of introduction and subsequent transmission) or as part of a disease prevention and control programme. The level of flock immunity required to prevent transmission will depend on the flock size, composition (e.g., species) and density of the susceptible poultry population. It is therefore impossible to be prescriptive. The vaccine used must also comply with the provisions stipulated for ND vaccines in of the Terrestrial Manual.

   In all vaccinated populations there is a need to perform surveillance (Article x.x.x.x.) to ensure the absence of vNDV circulation. The use of sentinel poultry may provide further confidence of the absence of virus circulation. The surveillance must be repeated at least every 6 months or at shorter intervals according to the risk in the country, zone or compartment. Evidence to show the effectiveness of the vaccination programme should also be provided.

Article 3.8.X.5.

Countries, zones or compartments regaining freedom from ND following an outbreak

In addition to the general conditions described in Chapter 2.7.13., a country regaining country, zone or compartment freedom from vNDV infection should show evidence of an active surveillance programme depending on the epidemiological circumstances of the outbreak to demonstrate the absence of the infection. This will require surveillance incorporating virus detection and antibody tests described in the Terrestrial Manual. The use of sentinel poultry may facilitate the interpretation of surveillance results.
Annex XIX (contd)

A country declaring freedom of a country, zone or compartment after an outbreak of ND (with or without vaccination) should report the results of an active surveillance programme in which the ND susceptible poultry population undergoes regular clinical examination and active surveillance planned and implemented according to the general conditions and methods described in these guidelines. The surveillance should give at least the same confidence that can be achieved by testing a randomized representative sample of the populations at risk.
CHAPTER 2.2.XX.

WEST NILE FEVER

Article 2.2.XX.1.

When authorising import or transit of the following commodities and any products made from these, Veterinary Authorities should not require any West Nile virus (WNV) related conditions, regardless of the WNF risk status of the animal population of the exporting country or zone:

a) hatching eggs;

b) eggs for human consumption;

c) egg products;

d) poultry semen;

e) fresh meat and meat products of poultry;

f) products of poultry origin intended for use in animal feeding, or for agricultural or industrial use;

g) feathers and down from poultry;

h) semen of horses;

i) fresh meat and meat products of horses.

Article 2.2.XX.2.

West Nile fever (WNF) is a zoonotic disease caused by certain strains of the mosquito-borne West Nile virus (WNV).

For the purpose of this Chapter, the susceptible species are equidae, geese, ducks (under study) and chicken and turkey chicks less than 12 days old and birds other than poultry.

Although most avian species are susceptible to infection, the outcome of the infection is highly variable according to the species. Chickens and turkeys, are usually resistant to disease and do not develop viremia sufficient to infect mosquitoes, with the exception of chicks less than 12 days old.

Birds are responsible for virus dispersal, including reintroduction of WNV from endemic areas into regions that may subsequently experience sporadic outbreaks.

WNV is maintained in a mosquito–bird–mosquito transmission cycle, whereas humans and equidae are considered dead-end hosts. Most human infections occur by natural transmission from mosquitoes.

Many animal species are known to be susceptible to WNV infection and outbreaks of a fatal neurological disease have been reported in humans, equidae, geese and wild birds.

In relation to domestic animal trade, geese and ducks might represent pose a risk for the spread of the WNV the WNF as some species have been documented to develop a viremia sufficient to infect mosquitoes.
Annex XX (contd)

WNV has been reported to date in a wide geographical range that includes portions of Europe, Asia, Africa, Australia and the Americas. Although competent vectors and susceptible bird species are nearly ubiquitous, WNV circulation in sylvatic cycles may spill over occasionally in domestic population.

Surveillance for WNF will be carried out according to Appendix 3.8.X.

The following criteria defines the occurrence of WNF case:

1. WNV has been isolated and identified as such from an animal, including human; or
2. viral antigen or viral ribonucleic acid (RNA) specific to WNV has been identified in samples from one or more animals including human that showing clinical signs consistent with WNF, or that is epidemiologically linked to a confirmed or suspected outbreak of WNF; or
3. antibodies to WNV that are not a consequence of vaccination, have been identified in an animal, that including human showing clinical signs consistent with WNF, or that is epidemiologically linked to a confirmed or suspected outbreak of WNF.

For the purposes of the Terrestrial Code, the incubation period for WNF shall be 3-15 days.

Standards for diagnostic tests and vaccines are described in the Terrestrial Manual.

**Article 2.2.XX.2.**

**WNF infected country, or zone or compartment**

A WNF infected country, or zone or compartment is a country, zone or compartment clearly defined where one in which a case of WNF has been reported during the past 2 years.

**Article 2.2.XX.3.**

**WNF free country, or zone or compartment**

1. A country, or zone or compartment may be considered free from WNF when WNF is notifiable in the whole country and either:
   a) no clinical occurrence of indigenous WNF cases have been recorded for the past 2 years; or
   b) a surveillance programme in accordance with Appendix 3.8.X. has demonstrated no evidence of WNV in the country or zone or compartment during the past 2 years; or
   c) a surveillance programme has demonstrated no evidence of Culex mosquitoes likely to be competent WNV vectors in the country, or zone or compartment.

2. A WNF free country, or zone or compartment will not lose its free status through the importation from WNF infected countries or infected zones or compartment of:
   a) seropositive animals;
   b) semen, embryo or ova;
   c) animals vaccinated in accordance with the Terrestrial Manual at least 30 days prior to dispatch, and that the animals are identified in the accompanying certification as having been vaccinated; or
Annex XX (contd)

d) animals not vaccinated if a surveillance programme in accordance with Appendix 3.8.X. has been in place in the source population for a period of 30 days immediately prior to dispatch, and no evidence of WNV transmission has been detected.

Article 2.2.XX.4.

WNF seasonally free country or zone

1. A WNF seasonally free country or zone is a country or a zone for one in which for part of a year, surveillance demonstrates no evidence either of WNV transmission or presence of adult Culex mosquitoes likely to be competent WNV vectors.

2. For the application of Article 2.2.XX.6., the seasonally free period is taken to commence 21 days following the last evidence of WNV transmission (as demonstrated by the surveillance programme), or the cessation of activity of adult Culex mosquitoes likely to be competent WNV vectors.

3. For the application of Article 2.2.XX.6., the seasonally free period is taken to conclude either:

   a) at least 21 days before the earliest date that historical data show WNV transmission cycle has recommenced; or

   b) immediately if current climatic data or data from a surveillance programme indicate an earlier resurgence of activity of adult Culex mosquitoes likely to be competent WNV vectors.

4. A WNF seasonally free country or zone will not lose its free status through the importation of animals, semen, embryo or ova from infected countries or zones of:

   a) seropositive animals;

   b) semen, embryo or ova;

   c) animals vaccinated in accordance with the Terrestrial Manual at least 30 days prior to dispatch, and are identified in the accompanying certification as having been vaccinated; or

   d) animals not vaccinated if a surveillance programme in accordance with Appendix 3.8.X. has been in place in the source population for a period of 30 days immediately prior to dispatch, and no evidence of WNV transmission has been detected.

Article 2.2.XX.4bis.

WNF infected country or zone

A WNF infected country or zone is one in which a case of WNF has been reported during the past 2 years.

Article 2.2.XX.5.

When importing from WNF free countries, or zones, or compartment Veterinary Administrations should require:

for susceptible species

the presentation of an international veterinary certificate attesting that:
Annex XX (contd)

1. the animals were kept in a WNF free country, or zone, or compartment since birth or for at least 30 days prior to shipment; or

2. the animals were kept in a WNF free country, or zone, or compartment for at least 7 days, were subjected, with negative results, to an agent identification test according to the Terrestrial Manual, with negative results, carried out on a sample collected at least 3 days after the commencement of the residence period and remained in the WNF free country, or zone, or compartment until shipment; or

3. the animals:
   a) were vaccinated in accordance with the Terrestrial Manual 30 days before introduction into the free country, or zone, or compartment; and
   b) were identified as having been vaccinated; and
   c) were kept in a WNF free country or zone for at least 7 days; and
   d) remained in the WNF free country or zone until shipment;

AND

4. if the animals were exported from a WNF free zone, either:
   a) did not transit through an infected zone during transportation to the place of shipment; or
   b) were protected from attack from WNV mosquito vectors at all times when transiting through an infected zone; or
   c) had been vaccinated in accordance with point 3 above.

Article 2.2.XX.6.

When importing from WNF seasonally free countries or zones, Veterinary Administrations Authorities should require:

for susceptible species

the presentation of an international veterinary certificate attesting that the animals:

1. were kept during the seasonally free period in a WNF seasonally free country or zone for at least 30 days prior to shipment; or

2. were kept during the WNF seasonally free period in a WNF seasonally free country or zone for at least 7 days prior to shipment, and were subjected during the residence period in the country or zone to an agent identification test according to the Terrestrial Manual, with negative results, carried out on a sample collected at least 3 days after the commencement of the residence period and remained in the WNF seasonally free country, or zone until shipment; or

3. were kept during the seasonally free period in a WNF seasonally free country or zone, and were vaccinated in accordance with the Terrestrial Manual 30 days before introduction into the free country or zone against WNF, were identified as having been vaccinated and remained in the WNF seasonally free country or zone until shipment;
AND

4. if the animals were exported from a WNF free country or zone, either:
   a) did not transit through an infected country or infected zone during transportation to the place of shipment; or
   b) were protected from attack from WNV mosquito vectors at all times when transiting through an infected country or infected zone; or
   c) were vaccinated in accordance with point 3 above.

Article 2.2.XX.7.

When importing from WNF infected countries or infected zones, Veterinary Administrations Authorities should require:

for susceptible species

the presentation of an international veterinary certificate attesting that the animals:

1. were protected from attack from WNV mosquito vectors for at least 30 days prior to shipment; or
2. were subjected to a serological test according to the Terrestrial Manual to detect WNV neutralizing antibodies with positive results; or
3. were protected from attack from WNV mosquito vectors for at least 15 days prior to shipment, and were subjected during that period to an agent identification test according to the Terrestrial Manual, with negative results, carried out on a sample collected at least 3 days after being introduced in the mosquito free zone; or
4. were vaccinated in accordance with the Terrestrial Manual at least 30 days before shipment, against WNV, and were identified in the accompanying certification as having been vaccinated; or
5. are not vaccinated and a surveillance programme in accordance with Appendix 3.8.X. has been in place in the source population for a period of 30 days immediately prior to shipment, and no evidence of WNV transmission has been detected;

AND

6. were protected from attack from WNV mosquito vectors during transportation to the place of shipment; or

7. were vaccinated 30 days before shipment or had antibodies against WNV.

Article 2.2.XX.8.

When importing wild birds, Veterinary Administrations Authorities should require the presentation of an international veterinary certificate attesting that:

1. the birds showed no clinical sign of WNF on the day of shipment; and
2. the birds were kept in a quarantine station in a mosquito-free environment for 30 days prior to shipment and they were subjected, with negative results, to an agent identification test according to the Terrestrial Manual carried out on samples collected at least 3 days after the commencement of the residence period.
Protecting animals from WNV mosquito vectors

When transporting animals through WNF infected countries or infected zones, Veterinary Administrations should require strategies to protect animals from attack from WNV mosquito vectors during transport, taking into account the local ecology of the vectors.

Potential risk management strategies include:

1. treating animals with chemical repellents prior to and during transportation;

2. ensuring vehicles do not stop en route unless the animals are held behind insect proof netting;

3. surveillance for vectors at common stopping and offloading points to gain information on seasonal variations;

4. integrated pest management practices at holding, common stopping and offloading points;

5. using historical, ongoing and/or WNF modelling information to identify low risk ports and transport routes.
DRAFT GUIDELINES ON THE DESIGN AND IMPLEMENTATION OF IDENTIFICATION SYSTEMS TO ACHIEVE ANIMAL TRACEABILITY

Article 1

Introduction and objectives

These guidelines are based on the general principles presented in Article 3.5.1.1. The Guidelines outline for Member Countries the basic elements that need to be taken into account in the design and implementation of an animal identification system to achieve animal traceability. Whatever animal identification system the country adopts, it should comply with relevant OIE standards. Each country should design a program in accordance with the scope and relevant performance criteria to ensure that the desired animal traceability outcomes can be achieved.

Article 2

Definitions

These following definitions apply for the purpose of this Appendix.

**Desired outcomes**: describe the overall goals of a programme and are usually expressed in qualitative terms, e.g. ‘to help ensure that animals and/or animal products are safe and suitable for use’. Safety and suitability for use could be defined in terms such as animal health, food safety, trade and husbandry aspects.

**Performance criteria**: are specifications for performance of a programme and are usually expressed in quantitative terms, such as ‘all animals can be traced to the establishment of birth within 48 hours of an enquiry’.

**Reporting**: means advising the Veterinary Administration Authority in accordance with the procedures listed in the programme.

**Scope**: specifies the targeted species, population and/or production/trade sector within a defined area (country, zone) or compartment that is the subject of the identification and traceability programme.

**Transhumance**: periodic/seasonal movements of animals between different pastures or premises within or between countries.

Article 3

Key elements of the animal identification system

1. **Desired outcomes**

   Desired outcomes should be defined through consultation between the Veterinary Administration Authority and other parties, which should include (depending on scope) animal producers and food processors, private sector veterinarians, scientific research organisations and other government agencies. Desired outcomes may be defined in terms of any or all of the following:

   a) animal health (e.g. disease surveillance and notification; detection and control of disease; vaccination programmes);
   b) public health (e.g. surveillance and control of zoonotic diseases and food safety);
   c) management of emergencies e.g. natural catastrophies or man-made events;
   d) trade (support for inspection and certification activities of Veterinary Services);
   e) animal husbandry aspects (e.g. animal performance, genetic data)
Annex XXI (contd)

2. **Scope**

Scope should also be defined through consultation between the Veterinary Administration and other parties, as discussed above. The scope of animal identification systems is often based on the definition of a species and sector, to take account of particular characteristics of the farming systems e.g. pigs in pork export production; cattle within a defined FMD free zone. Different systems will be appropriate according to the production systems used in countries and the nature of their industries and trade.

3. **Performance criteria**

Performance criteria are also designed in consultation with other parties, as discussed above. The performance criteria depend on the desired outcomes and scope of the program. They are usually described in quantitative terms. For example, some countries consider it necessary to trace susceptible animals within 24-48 hours when dealing with highly contagious diseases such as FMD and avian influenza. For food safety, animal tracing to support investigation of incidents may also be urgent. For chronic animal diseases, such as bovine paratuberculosis it may be considered appropriate that animals can be traced within 30 days.

4. **Preliminary studies**

In designing animal identification systems it is useful to conduct preliminary studies, which should take into account:

a) Animal populations, species, distribution, herd management
b) Farming and industry structures, production and location
c) Animal health
d) Public health
e) Trade issues

f) **Animal husbandry**
g) Zoning and compartmentalisation
h) Animal movement patterns (including transhumance)
i) Information management and communication
j) Availability of resources (human and financial)
k) Social and cultural aspects
l) Stakeholder knowledge of the issues and expectations
m) Gaps between current enabling legislation and what is needed long term
n) International experience
o) National experience
Available technology options

Existing identification system(s)

Benefits from the identification scheme and to whom they accrue.

Pilot projects may form part of the preliminary study to test the animal identification system and animal traceability and to gather information for the design and the implementation of the programme.

Economic analysis may consider costs, benefits, funding mechanisms and sustainability.

5. Design of the programme

a) General provisions

The programme should be designed in consultation with the stakeholders to facilitate the implementation of the animal identification system and animal traceability. It should take into account the scope, performance criteria and desired outcomes as well as the results of any preliminary study.

All the specified documentation should be standardised as to format, content and context.

To protect and enhance the integrity of the system, procedures should be incorporated into the design of the programme to prevent, detect and correct errors e.g. use of algorithms to prevent duplication of identification numbers in an electronic database.

b) Means of animal identification

The choice of a physical animal identifier should take into account elements such as the durability, human resources, species and age of the animals to be identified, required period of identification, animal welfare, cultural aspects, technology compatibility and relevant standards, farming practices, animal population, climatic conditions, resistance to tampering, trade considerations, cost, and retention and readability of the identification method.

The Veterinary Administration Authority is responsible for approving the materials and equipment chosen, to ensure that these means of animal identification comply with technical and field performance specifications, and for the supervision of their distribution. The Veterinary Administration Authority is also responsible for ensuring that identifiers are unique and are used in accordance with the requirements of the animal identification system.

The Veterinary Administration Authority should establish procedures for animal identification and animal traceability including:

i) The time period within which an animal born on an establishment should be identified.

ii) Animals imported into an establishment.

iii) When an animal loses its identification or the identifier becomes unusable.

iv) Arrangements and rules for the destruction and/ or reuse of identifiers.
Annex XXI (contd)

Where group identification without a physical identifier is adequate, documentation should be created specifying at least the number of animals in the group, the species, the date of identification, the person legally responsible for the animals and/or establishment. This documentation constitutes a unique group identifier.

Where all animals in the group are physically identified with a group identifier, documentation should also specify the unique group identifier.

c) Registration

Procedures need to be incorporated into the design of the programme in order to ensure that relevant events and information are registered in a timely and accurate manner.

Depending on the scope, performance criteria and desired outcomes, records as described below should specify, at least, the species, the unique animal or group identifier, the date of the event, the identifier of the establishment where the event took place, and the code for the event itself.

i) Establishments/ owners

Establishments where animals are kept should be identified and registered, including at least their physical location (such as geographical coordinates or street address), the type of establishment and the species kept. The register should include the name of the person legally responsible for the animals at the establishment.

The types of establishments that may need to be registered include holdings (farms), assembly centres (e.g. agriculture shows and fairs, sporting events, transit centres, breeding centres), markets, abattoirs, rendering plants, dead stock collection points, transhumance areas, centres for necropsy and diagnosis, research centres, zoos, border posts, quarantine stations.

In cases where the registration of establishments is not applicable e.g. some transhumance systems, the animal owner, the owner's place of residence and the species kept should be recorded.

ii) Animals

Animal identification and species should be registered for each establishment/owner. Other relevant information about the animals at each establishment/owner may also be recorded e.g. date of birth, production category, sex, breed, animal identification of the parents.

iii) Movements

The registration of animal movements is necessary to achieve animal traceability. When an animal is introduced into or leaves an establishment, these events constitute a movement.

Some countries classify birth, slaughter and death of the animal as movements.

The information registered should include the date of the movement, the establishment from which the animal or group of animals was dispatched, the number of animals moved, the destination establishment, and any in transit establishment.
When establishments are not registered as part of the animal identification system, ownership and location changes constitute a movement record. Movement recording may also include means of transport and the vehicle identifier.

Procedures should be in place to maintain animal traceability during transport and when animals arrive and leave an establishment.

iv) Events other than movements

The following events may also be registered:

- birth, slaughter and death of the animal (when not classified as a movement)
- attachment of the unique identifier to an animal
- change of ownership regardless of change of establishment
- observation of an animal on an establishment (testing, health investigation, health certification, etc.)
- animal imported: a record of the animal identification from the exporting country should be kept and linked with the animal identification assigned in the importing country.
- animal exported: a record of the animal identification from the exporting country should be provided to the Veterinary Administration Authority in the importing country.
- animal identifier lost or replaced
- animal missing (lost, stolen, etc)
- animal identifier retired (at slaughter, following loss of the identifier or death of the animal on a farm, at diagnostic laboratories, etc)

D) Documentation

Documentation requirements should be clearly defined and standardised, according to the scope, performance criteria and desired outcomes and supported by the legal framework.

E) Reporting

Depending on the scope, performance criteria and desired outcomes, relevant information (such as animal identification, movement, events, changes in numbers of livestock, establishments) should be reported to the Veterinary Administration Authority by the person responsible for the animals.

F) Information system

An information system should be designed according to the scope, performance criteria and desired outcomes. This may be paper based or electronic. The system should provide for the collection, compilation, storage and retrieval of information on matters relevant to registration. The following considerations are important:

- Have the potential for linkage to traceability in the other parts of the food chain.
Minimise duplication.

Relevant components, including databases, should be compatible.

Confidentiality of data.

Appropriate safeguards to avoid loss of data, including backup system.

The Veterinary Administration should have access to this information system as appropriate to meet the scope, performance criteria and desired outcomes.

g) Laboratories

The results of diagnostic tests should record the animal identifier or the group identifier and the establishment where the sample was collected.

h) Abattoirs, rendering plants, dead stock collection points, markets, assembly centres

Abattoirs, rendering plants, dead stock collection points, markets and assembly centres should document arrangements for the maintenance of animal identification and animal traceability in compliance with the legal framework.

These establishments are critical points for control of animal health and food safety.

Animal identification should be recorded on documents accompanying samples collected for analysis.

The components of the animal identification system operating within abattoirs should complement and be compatible with arrangements for tracking animal products throughout the food chain. At an abattoir, animal identification should be maintained during the processing of the animal’s carcass until the carcass is deemed fit for human consumption.

The animal identification and the establishment from which the animal was dispatched should be registered by the abattoir, rendering plant and dead stock collection points.

Abattoirs, rendering plants and dead stock collection points should ensure that identifiers are collected and disposed of according to the procedures established and regulated within the legal framework. These procedures should minimize the risk of unauthorized reuse and, if appropriate, should establish arrangements and rules for the reuse of identifiers.

Reporting of movement by abattoirs, rendering plants and dead stock collection points should occur according to the scope, performance criteria and desired outcomes and the legal framework.

i) Penalties

Different levels and types of penalties should be defined in the programme and supported by the legal framework.

j) Commercial arrangements

An animal identification system requires producers, processors and others (depending on the design of the system) to purchase equipment. There are many possible commercial arrangements that will have a variety of implications for the uptake of the animal identification system.
k) Transition planning

Any transition from an existing animal identification system needs to be designed to ensure it is easy for users of the existing system to make the change and to ensure that data integrity is maintained during the transition and integrated into the new animal identification system.

l) Use of incentives

Depending on the drivers for participation in the animal identification scheme, incentives may be useful to encourage early adoption of the system or to fill capability, capacity or technology gaps.

6. Legal framework

The Veterinary Administration Authority, with other relevant governmental agencies and in consultation with stakeholders, should establish a legal framework for the implementation and enforcement of animal identification system and animal traceability in the country. The structure of this framework will vary from country to country.

Animal identification, animal traceability and animal movement should be under the responsibility of the Veterinary Administration Authority.

This legal framework should address:

i) desired outcomes and scope

ii) obligations of the Veterinary Administration Authority and other parties

iii) organisational arrangements, including the choice of technologies and methods used for the animal identification system and animal traceability

iv) management of animal movement

v) confidentiality of data

vi) data access / accessibility

vii) checking, verification, inspection and penalties

viii) where relevant, funding mechanisms

ix) where relevant, arrangements to support a pilot project.

7. Implementation

a) Action plan

For implementing the animal identification system, an action plan should be prepared specifying the timetable and including the milestones and performance indicators, the human and financial resources, and checking, enforcement and verification arrangements.

The following activities should be addressed in the action plan:
i) Communication

The scope, performance criteria, desired outcomes, responsibilities, movement and registration requirements and sanctions need to be communicated to all parties. Communication strategies need to be targeted to the audience, taking into account elements such as the level of literacy (including technology literacy) and spoken languages.

ii) Training programmes

It is desirable to implement training programmes to assist the Veterinary Services and other parties.

iii) Technical support

Technical support should be provided to address practical problems.

b) Checking and Verification

Checking activities should start at the beginning of the implementation to detect, prevent and correct errors and to provide feedback on programme design.

Verification should begin after a preliminary period as determined by the Veterinary Administration Authority in order to determine compliance with the legal framework and operational requirements.

c) Auditing

Auditing should be carried out under the authority of the Veterinary Administration Authority to detect any problems with the animal identification system and animal traceability and to identify possible improvements.

d) Review

The programme should be subject to periodic review, taking into account the results of checking, verification and auditing activities.
GUIDELINES FOR THE CONTROL OF HAZARDS OF ANIMAL HEALTH AND PUBLIC HEALTH IMPORTANCE IN ANIMAL FEED

Article 1

Introduction

Animal feed is a critical component of the food-chain that has a direct impact on animal health and welfare and also on food safety and public health.

Historically, the OIE primarily addressed animal feed as an important pathway for the entry and spread of contagious epidemic diseases, such as foot and mouth disease, swine vesicular disease and avian influenza. In recent years, the role of feed as a vector for disease agents, including zoonotic organisms, was a focus of standards development in regards to bovine spongiform encephalopathy. Animal feed and feed ingredients are widely traded internationally and trade disruptions have the potential to impact economies in both developed and developing countries. Since 2002 the OIE has expanded its zoonotic disease mandate to encompass animal production food safety, working in collaboration with the Codex Alimentarius Commission (CAC) and other international organisations. In 2006 the International Committee resolved that the OIE should develop guidance on foodborne zoonoses and animal feeding, complementing relevant CAC texts.

Article 2

Objective and scope

The objective of this OIE guideline is to provide guidance on animal feeding in relation to animal health and to complement the guidance provided by the Codex Code of Practice on Good Animal Feeding (CAC/RCP 54-2004) which deals primarily with food safety.

This guideline aims at ensuring the control of animal and public health hazards through adherence to recommended practices during the production (procurement, handling, storage, processing and distribution) and use of both commercial and on-farm produced animal feed and feed ingredients for food producing animals.

Scope

This guideline applies to the production and use of all products destined for animal feed and feed ingredients at all levels whether produced commercially or on farm. It also includes grazing or free-range feeding, forage crop production and water for drinking. Swill feeding is a particular aspect of on-farm practice that is specifically addressed because of its recognised role in disease transmission.

These guidelines deals with food or feed for terrestrial food-producing animals, other than aquatic animals (i.e. livestock and poultry).

Article 3

Definitions

Hazard

means a biological, chemical or physical agent in, or a condition of, feed or a feed ingredient on animal or animal product with the potential to cause an adverse effect on animal or public health.

Feed

means any material (single or multiple), whether processed, semi-processed or raw, which is intended to be fed directly to food-producing animals.
Feed additives means any intentionally added ingredient not normally consumed as feed by itself, whether or not it has nutritional value, which affects the characteristics of feed, or health of the animal and the characteristics of products. Microorganisms, enzymes, acidity regulators, trace elements, vitamins and other products fall within the scope of this definition depending on the purpose of use and method of administration. This excludes veterinary drugs.

Medicated feed means any feed which contains a veterinary drug administered to food producing animals, for therapeutic or prophylactic purposes or for modification of physiological functions.

Feed ingredient means a component part or constituent of any combination or mixture making up a feed, whether or not it has a nutritional value in the animal’s diet, including feed additives. Ingredients are of plant, or animal or aquatic origin, or other organic or inorganic substances.

Undesirable substance means a contaminant or other substance material which is present in and/ or on feed and feed ingredients and which constitute a risk whose presence is potentially harmful to animal or public health and/ or is restricted under current regulations.

Commercial feed means all materials that are sold and distributed as feed, or to be mixed with feed, for animals except: unmixed seed, whole, processed, or unprocessed; straw, stover, silage, cobs, husks, and hulls; or individual chemical compounds not mixed with other ingredients.

Cross contamination means contamination the presence of a material or product with another material or product containing a component that is a feed or feed additive and whose presence in that feed or feed additive is potentially harmful for animal or public health or is restricted under the regulatory framework current regulations.

General principles

1. Roles and responsibilities

The Competent Authority has the legal power to set and enforce regulatory animal feeding requirements, and has final responsibility for verifying that these requirements are met. The Competent Authority may establish regulatory requirements for relevant parties to provide it with information and assistance. Refer to Chapters 1.3.3. and 1.3.4. of the OIE Terrestrial Code.

Those involved in the production and use of animal feed and feed ingredients have the responsibility to ensure that these products meet regulatory requirements. All personnel involved in the manufacture, storage and handling of feed and feed ingredients should be adequately trained and aware of their role and responsibility in preventing the spread of animal health and public health hazards. Appropriate contingency plans should be developed. Equipment should be maintained in good working order and in a sanitary condition.

It is a particular responsibility of Veterinary Services to set and enforce the regulatory requirements pertaining to the use of veterinary drugs, animal disease control and the food safety aspects that relate to the management of live animals on farm.
Those providing specialist services to producers and to the feed industry (e.g. private veterinarians and laboratories) may be required to meet specific regulatory requirements pertaining to the services they provide (e.g. disease reporting, quality standards, transparency).

2. Regulatory safety standards

All feed and feed ingredients should meet regulatory safety standards. In defining limits and tolerances for hazards, scientific evidence, including the sensitivity of analytical methods and on the characterisation of risks, should be taken into account.

3. Risk analysis (risk assessment, risk management and risk communication)

Internationally accepted principles and practices on risk analysis (Section 1.3. of the OIE Terrestrial Code; and relevant Codex texts) should be used in developing and applying the regulatory framework.

Application of a generic framework should provide a systematic and consistent process for managing all biosecurity risks, while recognising the different risk assessment methodologies used in animal and public health.

4. Good practices

Where national guidelines exist, good agricultural practices and good manufacturing practices (including good hygienic practices) should be followed. Countries without such guidelines are encouraged to develop them.

Where appropriate, Hazard Analysis and Critical Control Point (HACCP) principles should be followed to control hazards that may occur in the manufacture of feed and feed additives.

5. Geographic and environmental considerations

Land and facilities used for production of animal feed and feed ingredients and water sources should not be located in close proximity to sources of hazards for animal health or food safety. Animal health considerations include factors such as disease status, location of quarantined premises and existence of zones/compartments of specified health status. Food safety considerations include factors such as industrial operations that generate pollutants and waste treatment plants.

6. Zoning and compartmentalisation

Feed is an important component of biosecurity and needs to be considered when defining a compartment or zone in accordance with Chapter 1.3.5. of the OIE Terrestrial Code.

7. Sampling and analysis

Sampling and analytical protocols should be based on scientifically recognized principles and procedures.

8. Labelling

Labelling on how the feed or feed ingredients should be handled, stored and used should be clear and informative as to how the feed and feed ingredients should be handled, stored and used unambiguously, legible and conspicuously placed on the package if sold in bagged form and on the waybill and other sales documents if sold in bulk, un-bagged form, and should comply with regulatory requirements.

See Codex Code of practice on good animal feeding (CAC/ RCP 54-2004).
9. **Design and management of inspection programmes**

   In meeting animal and public health objectives prescribed in national legislation or required by importing countries, Competent Authorities contribute through the direct performance of some tasks or through the auditing of animal and public health activities conducted by other agencies or the private sector.

   Feed and feed ingredients business operators and other relevant parts of industry should practice self-regulation to secure compliance with required standards for procurement, handling, storage, processing, distribution and use. Operators have the primary responsibility for implementing systems for process control. Where such systems are applied, the Competent Authority should verify that they achieve all regulatory requirements.

10. **Assurance and certification**

    Competent Authorities are responsible for providing assurances domestically and to trading partners that regulatory requirements, safety standards, have been met. For international trade in animal product based feeds, Veterinary Services are required to provide international veterinary certificates.

11. **Hazards associated with animal feed**

    a) **Biological hazards**

       Biological hazards that may occur in feed and feed ingredients include agents such as bacteria, viruses, prions, fungi and parasites.

    b) **Chemical hazards**

       Chemical hazards that may occur in feed and feed ingredients include naturally occurring chemicals (such as mycotoxins and gossypol), industrial and environmental contaminants (such as dioxins and PCBs), residues of veterinary drugs and pesticides and also radionuclides.

    c) **Physical hazards**

       Physical hazards that may occur in feed and feed ingredients include foreign objects (such as pieces of glass, metal, plastic or wood).

12. **Cross contamination**

    It is important to avoid cross-contamination during the manufacture, storage, distribution (including transport) and use of feed and feed ingredients and relevant provisions should be included in the regulatory framework. Scientific evidence, including the sensitivity of analytical methods and on the characterisation of risks, should be drawn upon in developing this framework.

    Procedures, such as flushing, sequencing and physical clean-out, should be used to avoid cross-contamination between batches of feed or feed ingredients.

13. **Antimicrobial resistance**

    Concerning the use of antimicrobials in animal feed refer to Section 3.9. of the OIE Terrestrial Code.

14. **Management of information**

    The Competent Authority should establish clear requirements for the provision of information by the private sector as this relates to regulatory requirements.

    Records should be maintained in a readily accessible form regarding the production, distribution and use of feed and feed ingredients. These records are required to facilitate the prompt trace-back of feed and feed ingredients to the immediate previous source, and trace-forward to the next subsequent recipients, to address identified animal health or public health concerns.
Animal identification and animal traceability are tools for addressing animal health (including zoonoses), and food safety risks arising from animal feed (see Section 3.5. of the OIE Terrestrial Code; Section 4.3. of CAC/RCP 54-2004).
APPENDIX 3.7.2.

GUIDELINES FOR THE TRANSPORT OF ANIMALS BY SEA

Preamble: These guidelines apply to the following live domesticated animals: cattle, buffalo, deer, camelids, sheep, goats, pigs and equines. They may also be applicable to other domesticated animals.

Article 3.7.2.1.

The amount of time animals spend on a journey should be kept to the minimum.

Article 3.7.2.2

1. Animal behaviour

Animal handlers should be experienced and competent in handling and moving farm livestock and understand the behaviour patterns of animals and the underlying principles necessary to carry out their tasks.

The behaviour of individual animals or groups of animals will vary depending on their breed, sex, temperament and age and the way in which they have been reared and handled. Despite these differences, the following behaviour patterns, which are always present to some degree in domestic animals, should be taken into consideration in handling and moving the animals.

Most domestic livestock are kept in herds and follow a leader by instinct.

Animals which are likely to be hostile to each other in a group situation should not be mixed.

The desire of some animals to control their personal space should be taken into account in designing loading and unloading facilities, transport vessels and containers.

Domestic animals will try to escape if any person approaches closer than a certain distance. This critical distance, which defines the flight zone, varies among species and individuals of the same species, and depends upon previous contact with humans. Animals reared in close proximity to humans (i.e. tame) have a smaller flight zone, whereas those kept in free range or extensive systems may have flight zones which may vary from one metre to many metres. Animal handlers should avoid sudden penetration of the flight zone which may cause a panic reaction which could lead to aggression or attempted escape.
Annex XXIII (contd)

An example of a flight zone (cattle)

Animal handler movement pattern to move cattle forward

Animal handlers should use the point of balance at the animal’s shoulder to move animals, adopting a position behind the point of balance to move an animal forward and in front of the point of balance to move it backward.

Domestic animals have a wide-angle vision but only have a limited forward binocular vision and poor perception of depth. This means that they can detect objects and movements beside and behind them, but can only judge distances directly ahead.

Domestic animals can hear over a greater range of frequencies than humans and are more sensitive to higher frequencies. They tend to be alarmed by constant loud noises and by sudden noises, which may cause them to panic. Sensitivity to such noises should also be taken into account when handling animals.
2. **Distractions and their removal**

Design of new loading and unloading facilities or modification of existing facilities should aim to minimise the potential for distractions that may cause approaching animals to stop, baulk or turn back. Below are examples of common distractions and methods for eliminating them:

- **a)** reflections on shiny metal or wet floors - move a lamp or change lighting;
- **b)** dark entrances - illuminate with indirect lighting which does not shine directly into the eyes of approaching animals;
- **c)** animals seeing moving people or equipment up ahead - install solid sides on chutes and races or install shields;
- **d)** dead ends - avoid if possible by curving the passage, or make an illusionary passage;
- **e)** chains or other loose objects hanging in chutes or on fences - remove them;
- **f)** uneven floors or a sudden drop in floor levels - avoid uneven floor surfaces or install a solid false floor to provide an illusion of a solid and continuous walking surface;
- **g)** sounds of air hissing from pneumatic equipment - install silencers or use hydraulic equipment or vent high pressure to the external environment using flexible hosing;
- **h)** clanging and banging of metal objects - install rubber stops on gates and other devices to reduce metal to metal contact;
- **i)** air currents from fans or air curtains blowing into the face of animals - redirect or reposition equipment.

**Article 3.7.2.3.**

**Responsibilities**

Once the decision to transport the animals by sea has been made, the welfare of the animals during their journey is the paramount consideration and is the joint responsibility of all people involved. The individual responsibilities of persons involved will be described in more detail in this Article. These guidelines may also be applied to the transport of animals by water within a country.

The management of animals at post-discharge facilities is outside the scope of this Appendix.

1. **General considerations**

- **a)** Exporters, importers, owners of animals, business or buying/selling agents, shipping companies, masters of vessels and managers of facilities are jointly responsible for the general health of the animals and their fitness for the journey, and for their overall welfare during the journey, regardless of whether duties are subcontracted to other parties during transport.

- **b)** Exporters, shipping companies, business or buying/selling agents, and masters of vessels are jointly responsible for planning the journey to ensure the care of the animals, including:

  - **i)** choosing appropriate vessels and ensuring that animal handlers are available to care for the animals;
ii) developing and keeping up to date contingency plans to address emergencies (including adverse weather conditions) and minimise stress during transport;

iii) correct loading of the ship, provision of appropriate food, water, ventilation and protection from adverse weather, regular inspections during the journey and for appropriate responses to problems arising;

iv) disposal of carcasses according to international law.

c) To carry out the above mentioned responsibilities, the parties involved should be competent regarding transport regulations, equipment usage, and the humane handling and care of animals.

2. Specific considerations

a) The responsibilities of the exporters include:

   i) the organisation, carrying out and completion of the journey, regardless of whether duties are subcontracted to other parties during transport;

   ii) ensuring that equipment and medication are provided as appropriate for the species and the journey;

   iii) securing the presence of the appropriate number of animal handlers competent for the species being transported;

   iv) ensuring compliance of the animals with any required veterinary certification, and their fitness to travel;

   v) in case of animals for export, ensuring compliance with any requirements of the importing and exporting countries.

b) The responsibilities of the importers include:

   (under study)

c) The responsibilities of the owners of the animals include the selection of animals that are fit to travel based on veterinary recommendations.

d) The responsibilities of the business or buying/selling agent include:

   i) selection of animals that are fit to travel based on veterinary recommendations;

   ii) availability of suitable facilities for the assembly, loading, transport, unloading and holding of animals at the start and at the end of the journey, and for emergencies.

e) The responsibilities of shipping companies include:

   (under study)

f) The responsibilities of masters of vessels include the provision of suitable premises for animals on the vessel.

g) The responsibilities of managers of facilities during loading include:

   i) providing suitable premises for loading the animals;
Annex XXIII (contd)

ii) providing an appropriate number of animal handlers to load the animals with minimum stress and the avoidance of injury;

iii) minimising the opportunities for disease transmission while the animals are in the facilities;

iv) providing appropriate facilities for emergencies;

v) providing facilities, veterinarians or animal handlers capable of killing animals humanely when required.

h) The responsibilities of managers of facilities during unloading include:

i) providing suitable facilities for unloading the animals onto transport vehicles for immediate movement or securely holding the animals in lairage, with shelter, water and feed, when required, for transit;

ii) providing animal handlers to unload the animals with minimum stress and injury;

iii) minimising the opportunities for disease transmission while the animals are in the facilities;

iv) providing appropriate facilities for emergencies;

v) providing facilities, and veterinarians or animal handlers capable of killing animals humanely when required.

i) The responsibilities of the animal handlers include humane handling and care of the animals, especially during loading and unloading.

j) The responsibilities of the Competent Authority of the exporting country include:

i) establishing minimum standards for animal welfare, including requirements for inspection of animals before and during their travel, and for certification and record keeping;

ii) approving facilities, containers, vehicles/ vessels for the holding and transport of animals;

iii) setting competence standards for animal handlers and managers of facilities;

v) implementation of the standards, including through accreditation of / interaction with other organisations and Competent Authorities;

vi) monitor and evaluate health and welfare performance, including the use of any veterinary medications.

k) The responsibilities of the Competent Authority of the importing country include:

i) establishing minimum standards for animal welfare, including requirements for inspection of animals after their travel, and for certification and record keeping;

ii) approve facilities, containers, vehicles/ vessels for the holding and transport of animals;

iii) setting competence standards for animal handlers and managers of facilities;

iv) implementation of the standards, including through accreditation of / interaction with other organisations and Competent Authorities;
v) ensuring that the exporting country is aware of the required standards for the vessel transporting the animals;

vi) monitor and evaluate health and welfare performance, including the use of any veterinary medications.

vii) give animal consignments priority to allow import procedures to be completed without unnecessary delay.

m) The responsibilities of veterinarians or in the absence of a veterinarian, the animal handlers travelling on the vessel with the animals include:

i) humane handling and treatment of animals during the journey, including in emergencies, such as humane killing of the animals;

ii) possess ability to report and act independently;

iii) meet daily with the master of the vessel to obtain up-to-date information on animal health and welfare status.

n) The receiving Competent Authority should report back to the sending Competent Authority on significant animal welfare problems which occurred during the journey.

Article 3.7.2.4.

Competence

1. All people responsible for animals during journeys, should be competent to carry out the relevant responsibilities listed in Article 3.7.2.3. Competence in areas other than animal welfare would need to be addressed separately. Competence may be gained through formal training and/or practical experience.

2. The assessment of competence of animal handlers should at a minimum address knowledge, and ability to apply that knowledge, in the following areas:

a) planning a journey, including appropriate space allowance, feed, water and ventilation requirements;

b) responsibilities for the welfare of animals during the journey, including loading and unloading;

c) sources of advice and assistance;

d) animal behaviour, general signs of disease, and indicators of poor animal welfare such as stress, pain and fatigue, and their alleviation;

e) assessment of fitness to travel; if fitness to travel is in doubt, the animal should be examined by a veterinarian;

f) relevant authorities and applicable transport regulations, and associated documentation requirements;

g) general disease prevention procedures, including cleaning and disinfection;

h) appropriate methods of animal handling during transport and associated activities such as assembling, loading, and unloading;
i) methods of inspecting animals, managing situations frequently encountered during transport such as adverse weather conditions, and dealing with emergencies, including euthanasia;

j) species-specific aspects and age-specific aspects of animal handling and care, including feeding, watering and inspection; and

k) maintaining a journey log and other records.

5. Assessment of competence for exporters should at a minimum address knowledge, and ability to apply that knowledge, in the following areas:

a) planning a journey, including appropriate space allowances, and feed, water and ventilation requirements;

b) relevant authorities and applicable transport regulations, and associated documentation requirements;

c) appropriate methods of animal handling during transport and associated activities such as cleaning and disinfection, assembling, loading, and unloading;

d) species-specific aspects of animal handling and care, including appropriate equipment and medication;

e) sources of advice and assistance;

f) appropriate record keeping; and

5) managing situations frequently encountered during transport, such as adverse weather conditions, and dealing with emergencies.

Article 3.7.2.5.

Planning the journey

1. General considerations

a) Adequate planning is a key factor affecting the welfare of animals during a journey.

b) Before the journey starts, plans should be made in relation to:

i) preparation of animals for the journey;

ii) type of transport vessel required;

iii) route, taking into account distance, expected weather and sea conditions;

iv) nature and duration of journey;

v) daily care and management of the animals, including the appropriate number of animal handlers, to help ensure the health and welfare of all the animals;

vi) avoiding the mixing of animals from different sources in a single pen group;
vii) provision of appropriate equipment and medication for the numbers and species carried; and

viii) emergency response procedures.

2. Preparation of animals for the journey
   
a) When animals are to be provided with a novel diet or unfamiliar methods of supplying of feed or water, they should be preconditioned.

b) There should be planning for water and feed availability during the journey. Feed should be of appropriate quality and composition for the species, age, condition of the animals, etc.

c) Extreme weather conditions are hazards for animals undergoing transport and require appropriate vessel design to minimise risks. Special precautions should be taken for animals that have not been acclimatised or which are unsuited to either hot or cold conditions. In some extreme conditions of heat or cold, animals should not be transported at all.

d) Animals more accustomed to contact with humans and with being handled are likely to be less fearful of being loaded and transported. Animals should be handled and loaded in a manner that reduces their fearfulness and improves their approachability.

e) Behaviour-modifying (such as tranquillisers) or other medication should not be used routinely during transport. Such medicines should only be administered when a problem exists in an individual animal, and should be administered by a veterinarian or other person who has been instructed in their use by a veterinarian. Treated animals should be placed in a dedicated area.

3. Control of disease
   
As animal transport is often a significant factor in the spread of infectious diseases, journey planning should take into account the following:

a) When possible and agreed by the Veterinary Authority of the importing country, animals should be vaccinated against diseases to which they are likely to be exposed at their destination.

b) Medications used prophylactically or therapeutically should only be administered by a veterinarian or other person who has been instructed in their use by a veterinarian.

c) Mixing of animals from different sources in a single consignment should be minimized.

4. Vessel and container design and maintenance
   
a) Vessels used for the sea transport of animals should be designed, constructed and fitted as appropriate to the species, size and weight of the animals to be transported. Special attention should be paid to the avoidance of injury to animals through the use of secure smooth fittings free from sharp protrusions and the provision of non-slip flooring. The avoidance of injury to animal handlers while carrying out their responsibilities should be emphasised.

b) Vessels should be properly illuminated to allow animals to be observed and inspected.

c) Vessels should be designed to permit thorough cleaning and disinfection, and the management of faeces and urine.

d) Vessels and their fittings should be maintained in good mechanical and structural condition.
e) Vessels should have adequate ventilation to meet variations in climate and the thermo-regulatory needs of the animal species being transported. The ventilation system should be effective when the vessel is stationary. An emergency power supply should be available to maintain ventilation in the case of primary machinery breakdown.

f) The feeding and watering system should be designed to permit adequate access to feed and water appropriate to the species, size and weight of the animals, and to minimise soiling of pens.

g) Vessels should be designed so that the faeces or urine from animals on upper levels do not soil animals on lower levels, or their feed or water.

h) Loading and stowage of feed and bedding should be carried out in such a way to ensure protection from fire hazards, the elements and sea water.

i) Where appropriate, suitable bedding, such as straw or sawdust, should be added to vessel floors to assist absorption of urine and faeces, provide better footing for animals and protect animals (especially young animals) from hard or rough flooring surfaces and adverse weather conditions.

j) The above principles apply also to containers used for the transport of animals.

5. Special provisions for transport in road vehicles on roll-on/roll-off vessels or for containers

   a) Road vehicles and containers should be equipped with a sufficient number of adequately designed, positioned and maintained securing points enabling them to be securely fastened to the vessel.

   b) Road vehicles and containers should be secured to the ship before the start of the sea journey to prevent them being displaced by the motion of the vessel.

   c) Vessels should have adequate ventilation to meet variations in climate and the thermo-regulatory needs of the animal species being transported, especially where the animals are transported in a secondary vehicle/container on enclosed decks.

   d) Due to the risk of limited airflow on certain decks of a vessel, a road vehicle or container may require a forced ventilation system of greater capacity than that provided by natural ventilation.

6. Nature and duration of the journey

   The maximum duration of a journey should be determined taking into account factors that determine the overall welfare of animals, such as:

   a) the ability of the animals to cope with the stress of transport (such as very young, old, lactating or pregnant animals);

   b) the previous transport experience of the animals;

   c) the likely onset of fatigue;

   d) the need for special attention;

   e) the need for feed and water;

   f) the increased susceptibility to injury and disease;

   g) space allowance and vessel design;
Annex XXIII (contd)

h) weather conditions;

i) vessel type used, method of propulsion and risks associated with particular sea conditions.

7. Space allowance

a) The number of animals which should be transported on a vessel and their allocation to different pens on the vessel should be determined before loading.

b) The amount of space required, including headroom, depends on the species of animal and should allow the necessary thermoregulation. Each animal should be able to assume its natural position for transport (including during loading and unloading) without coming into contact with the roof or upper deck of the vessel. When animals lie down, there should be enough space for every animal to adopt a normal lying posture.

c) Calculations for the space allowance for each animal should be carried out in reference to a relevant national or international document. The size of pens will affect the number of animals in each.

d) The same principles apply when animals are transported in containers.

8. Ability to observe animals during the journey

Animals should be positioned to enable each animal to be observed regularly and clearly by animal handler or other responsible person, during the journey to ensure their safety and good welfare.

9. Emergency response procedures

There should be an emergency management plan that identifies the important adverse events that may be encountered during the journey, the procedures for managing each event and the action to be taken in an emergency. For each important event, the plan should document the actions to be undertaken and the responsibilities of all parties involved, including communications and record keeping.

Article 3.7.2.6.

Documentation

1. Animals should not be loaded until the documentation required to that point is complete.

2. The documentation accompanying the consignment should include:

   a) journey travel plan and an emergency management plan;

   b) time, date and place of loading;

   c) the journey log – a daily record of inspection and important events which includes records of morbidity and mortality and actions taken, climatic conditions, food and water consumed, medication provided, mechanical defects;

   d) expected time, date and place of arrival and unloading;

   e) veterinary certification, when required;
f) animal identification to allow animal traceability of animals to the premises of departure, and, where possible, to the premises of origin;

g) details of any animals considered at particular risk of suffering poor welfare during transport (point 3e) of Article 3.7.2.7.;

h) number of animal handlers on board, and their competencies; and

i) stocking density estimate for each load in the consignment.

3. When veterinary certification is required to accompany consignments of animals, it should address:

a) when required, details of disinfection carried out;

b) fitness of the animals to travel;

c) animal identification (description, number, etc.); and

d) health status including any tests, treatments and vaccinations carried out.

Article 3.7.2.7.

Pre-journey period

1. General considerations

a) Before each journey, vessels should be thoroughly cleaned and, if necessary, treated for animal and public health purposes, using chemicals approved by the Competent Authority. When cleaning is necessary during a journey, this should be carried out with the minimum of stress and risk to the animals.

b) In some circumstances, animals may require pre-journey assembly. In these circumstances, the following points should be considered:

i) Pre-journey rest is necessary if the welfare of animals has become poor during the collection period because of the physical environment or the social behaviour of the animals.

ii) For animals such as pigs which are susceptible to motion sickness, and in order to reduce urine and faeces production during the journey, a species-specific short period of feed deprivation prior to loading is desirable.

iii) When animals are to be provided with a novel diet or unfamiliar methods of supplying feed or water, they should be preconditioned.

c) Where an animal handler believes that there is a significant risk of disease among the animals to be loaded or significant doubt as to their fitness to travel, the animals should be examined by a veterinarian.

d) Pre-journey assembly / holding areas should be designed to:

i) securely contain the animals;

ii) maintain an environment safe from hazards, including predators and disease;
Annex XXIII (contd)

iii) protect animals from exposure to adverse weather conditions;

iv) allow for maintenance of social groups; and

v) allow for rest, watering and feeding.

2. Selection of compatible groups

Compatible groups should be selected before transport to avoid adverse animal welfare consequences. The following guidelines should be applied when assembling groups of animals:

a) animals of different species should not be mixed unless they are judged to be compatible;

b) animals of the same species can be mixed unless there is a significant likelihood of aggression; aggressive individuals should be segregated (recommendations for specific species are described in detail in Article 3.7.2.12.). For some species, animals from different groups should not be mixed because poor welfare occurs unless they have established a social structure;

c) young or small animals may need to be separated from older or larger animals, with the exception of nursing mothers with young at foot;

d) animals with horns or antlers should not be mixed with animals lacking horns or antlers, unless judged to be compatible; and

e) animals reared together should be maintained as a group; animals with a strong social bond, such as a dam and offspring, should be transported together.

3. Fitness to travel

a) Animals should be inspected by a veterinarian or an animal handler to assess fitness to travel. If its fitness to travel is in doubt, it is the responsibility of a veterinarian to determine its ability to travel. Animals found unfit to travel should not be loaded onto a vessel.

b) Humane and effective arrangements should be made by the owner or agent for the handling and care of any animal rejected as unfit to travel.

c) Animals that are unfit to travel include, but may not be limited to:

i) those that are sick, injured, weak, disabled or fatigued;

ii) those that are unable to stand unaided or bear weight on each leg;

iii) those that are blind in both eyes;

iv) those that cannot be moved without causing them additional suffering;

v) newborn with an unhealed navel;

vi) females travelling without young which have given birth within the previous 48 hours;

vii) pregnant animals which would be in the final 10% of their gestation period at the planned time of unloading;

viii) animals with unhealed wounds from recent surgical procedures such as dehorning.
Annex XXIII (contd)

d) Risks during transport can be reduced by selecting animals best suited to the conditions of travel and those that are acclimatised to expected weather conditions.

e) Animals at particular risk of suffering poor welfare during transport and which require special conditions (such as in the design of facilities and vehicles, and the length of the journey) and additional attention during transport, may include:

i) very large or obese individuals;

ii) very young or old animals;

iii) excitable or aggressive animals;

iv) animals subject to motion sickness;

v) animals which have had little contact with humans;

vi) females in the last third of pregnancy or in heavy lactation.

f) Hair or wool length should be considered in relation to the weather conditions expected during transport.

Article 3.7.2.8.

Loading

1. Competent supervision

a) Loading should be carefully planned as it has the potential to be the cause of poor welfare in transported animals.

b) Loading should be supervised by the Competent Authority and conducted by animal handler(s). Animal handlers should ensure that animals are loaded quietly and without unnecessary noise, harassment or force, and that untrained assistants or spectators do not impede the process.

2. Facilities

a) The facilities for loading, including the collecting area at the wharf, races and loading ramps should be designed and constructed to take into account of the needs and abilities of the animals with regard to dimensions, slopes, surfaces, absence of sharp projections, flooring, sides, etc.

b) Ventilation during loading and the journey should provide for fresh air, and the removal of excessive heat, humidity and noxious fumes (such as ammonia and carbon monoxide). Under warm and hot conditions, ventilation should allow for the adequate convective cooling of each animal. In some instances, adequate ventilation can be achieved by increasing the space allowance for animals.

c) Loading facilities should be properly illuminated to allow the animals to be easily inspected by animal handlers, and to allow the ease of movement of animals at all times. Facilities should provide uniform light levels directly over approaches to sorting pens, chutes, loading ramps, with brighter light levels inside vehicles/containers, in order to minimise baulking. Dim light levels may be advantageous for the catching of some animals. Artificial lighting may be required.
Annex XXIII (contd)

3. Goads and other aids

When moving animals, their species specific behaviour should be used (see Article 3.7.2.12.). If goads and other aids are necessary, the following principles should apply:

a) Animals that have little or no room to move should not be subjected to physical force or goads and other aids which compel movement. Electric goads and prods should only be used in extreme cases and not on a routine basis to move animals. The use and the power output should be restricted to that necessary to assist movement of an animal and only when an animal has a clear path ahead to move. Goads and other aids should not be used repeatedly if the animal fails to respond or move. In such cases it should be investigated whether some physical or other impediment is preventing the animal from moving.

b) The use of such devices should be limited to battery-powered goads on the hindquarters of pigs and large ruminants, and never on sensitive areas such as the eyes, mouth, ears, anogenital region or belly. Such instruments should not be used on horses, sheep and goats of any age, or on calves or piglets.

c) Useful and permitted goads include panels, flags, plastic paddles, flappers (a length of cane with a short strap of leather or canvas attached), plastic bags and rattles; they should be used in a manner sufficient to encourage and direct movement of the animals without causing undue stress.

d) Painful procedures (including whipping, tail twisting, use of nose twitches, pressure on eyes, ears or external genitalia), or the use of goads or other aids which cause pain and suffering (including large sticks, sticks with sharp ends, lengths of metal piping, fencing wire or heavy leather belts), should not be used to move animals.

e) Excessive shouting at animals or making loud noises (e.g. through the cracking of whips) to encourage them to move should not occur as such actions may make the animals agitated, leading to crowding or falling.

f) The use of well trained dogs to help with the loading of some species may be acceptable.

g) Animals should be grasped or lifted in a manner which avoids pain or suffering and physical damage (e.g. bruising, fractures, dislocations). In the case of quadrupeds, manual lifting by a person should only be used in young animals or small species, and in a manner appropriate to the species; grasping or lifting animals only by their wool, hair, feathers, feet, neck, ears, tails, head, horns, limbs causing pain or suffering should not be permitted, except in an emergency where animal welfare or human safety may otherwise be compromised.

h) Conscious animals should not be thrown, dragged or dropped.

i) Performance standards should be established in which numerical scoring is used to evaluate the use of such instruments, and to measure the percentage of animals moved with an electric instrument and the percentage of animals slipping or falling as a result of their usage.

Article 3.7.2.9.

Travel

1. General considerations

a) Aimal handler(s) should check the consignment immediately before departure to ensure that the animals have been loaded according to the load plan. Each consignment should be checked following any incident or situation likely to affect their welfare and in any case within 12 hours of departure.
b) If necessary and where possible adjustments should be made to the stocking density as appropriate during the journey.

c) Each pen of animals should be observed on a daily basis for normal behaviour, health and welfare, and the correct operation of ventilation, watering and feeding systems. There should also be a night patrol. Any necessary corrective action should be undertaken promptly.

d) Adequate access to suitable feed and water should be ensured for all animals in each pen.

e) Where cleaning or disinfection is necessary during travel, it should be carried out with the minimum of stress to the animals.

2. Sick or injured animals

a) Sick or injured animals should be segregated.

b) Sick or injured animals should be appropriately treated or humanely killed, in accordance with a predetermined emergency response plan (Article 3.7.2.5.). Veterinary advice should be sought if necessary. All drugs and products should be used according to recommendations from a veterinarian and in accordance with the manufacturer’s instructions.

c) A record of treatments carried out and their outcomes should be kept.

d) When humane killing is necessary, the animal handler must ensure that it is carried out humanely. Recommendations for specific species are described in Appendix 3.7.6. on killing of animals for disease control purposes. Veterinary advice regarding the appropriateness of a particular method of euthanasia should be sought as necessary.

Article 3.7.2.10.

Unloading and post-journey handling

1. General considerations

a) The required facilities and the principles of animal handling detailed in Article 3.7.2.8. apply equally to unloading, but consideration should be given to the likelihood that the animals will be fatigued.

b) Unloading should be carefully planned as it has the potential to be the cause of poor welfare in transported animals.

c) A livestock vessel should have priority attention when arriving in port and have priority access to a berth with suitable unloading facilities. As soon as possible after the vessel’s arrival at the port and acceptance of the consignment by the Competent Authority, animals should be unloaded into appropriate facilities.

d) The accompanying veterinary certificate and other documents should meet the requirements of the importing country. Veterinary inspections should be completed as quickly as possible.

e) Unloading should be supervised by the Competent Authority and conducted by animal handler(s). The animal handlers should ensure that animals are unloaded as soon as possible after arrival but sufficient time should be allowed for unloading to proceed quietly and without unnecessary noise, harassment or force, and that untrained assistants or spectators do not impede the process.
Annex XXIII (contd)

2. Facilities

a) The facilities for unloading including the collecting area at the wharf, races and unloading ramps should be designed and constructed to take into account of the needs and abilities of the animals with regard to dimensions, slopes, surfaces, absence of sharp projections, flooring, sides, etc.

b) All unloading facilities should have sufficient lighting to allow the animals to be easily inspected by the animal handlers and to allow the ease of movement of animals at all times.

c) There should be facilities to provide animals with appropriate care and comfort, adequate space, access to quality feed and clean drinking water, and shelter from extreme weather conditions.

3. Sick or injured animals

a) An animal that has become sick, injured or disabled during a journey should be appropriately treated or humanely killed (see Appendix 3.7.6.). When necessary, veterinary advice should be sought in the care and treatment of these animals.

b) In some cases, where animals are non-ambulatory due to fatigue, injury or sickness, it may be in the best welfare interests of the animal to be treated or humanely killed aboard the vessel.

c) If unloading is in the best welfare interests of animals that are fatigued, injured or sick, there should be appropriate facilities and equipment for the humane unloading of such animals. These animals should be unloaded in a manner that causes the least amount of suffering. After unloading, separate pens and other appropriate facilities and treatments should be provided for sick or injured animals.

4. Cleaning and disinfection

a) Vessels and containers used to carry the animals should be cleaned before re-use through the physical removal of manure and bedding, by scraping, washing and flushing vessels and containers with water until visibly clean. This should be followed by disinfection when there are concerns about disease transmission.

b) Manure, litter and bedding should be disposed of in such a way as to prevent the transmission of disease and in compliance with all relevant health and environmental legislation.

Actions in the event of a refusal to allow the importation of a shipment

1. The welfare of the animals should be the first consideration in the event of a refusal to import.

2. When animals have been refused import, the Competent Authority of the importing country should make available suitable isolation facilities to allow the unloading of animals from a vessel and their secure holding, without posing a risk to the health of the national herd, pending resolution of the situation. In this situation, the priorities should be:

a) The Competent Authority of the importing country should provide urgently in writing the reasons for the refusal.

b) In the event of a refusal for animal health reasons, the Competent Authority of the importing country should provide urgent access to an OIE-appointed veterinarian(s) to assess the health status of the animals with regard to the concerns of the importing country, and the necessary facilities and approvals to expedite the required diagnostic testing.
c) The Competent Authority of the importing country should provide access to allow continued assessment of the ongoing health and welfare situation.

d) If the matter cannot be promptly resolved, the Competent Authority of the exporting and importing countries should call on the OIE to mediate.

3. In the event that the animals are required to remain on the vessel, the priorities should be:

a) The Competent Authority of the importing country should allow provisioning of the vessel with water and feed as necessary.

b) The Competent Authority of the importing country should provide urgently in writing the reasons for the refusal.

c) In the event of a refusal for animal health reasons, the Competent Authority of the importing country should provide urgent access to an OIE-appointed veterinarian(s) to assess the health status of the animals with regard to the concerns of the importing country, and the necessary facilities and approvals to expedite the required diagnostic testing.

d) The Competent Authority of the importing country should provide access to allow continued assessment of the ongoing health and other aspects of the welfare of the animals, and the necessary actions to deal with any issues which arise.

e) If the matter cannot be urgently resolved, the Competent Authorities of the exporting and importing countries should call on the OIE to mediate.

4. The OIE should utilise its dispute settlement mechanism to identify a mutually agreed solution which will address the animal health and welfare issues in a timely manner.

Species specific issues

**Camelids** of the new world in this context comprise llamas, alpacas, guanaco and vicuna. They have good eyesight and, like sheep, can negotiate steep slopes, though ramps should be as shallow as possible. They load most easily in a bunch as a single animal will strive to rejoin the others. Whilst they are usually docile, they have an unnerving habit of spitting in self-defence. During transport, they usually lie down. They frequently extend their front legs forward when lying, so gaps below partitions should be high enough so that their legs are not trapped when the animals rise.

**Cattle** are sociable animals and may become agitated if they are singled out. Social order is usually established at about two years of age. When groups are mixed, social order has to be re-established and aggression may occur until a new order is established. Crowding of cattle may also increase aggression as the animals try to maintain personal space. Social behaviour varies with age, breed and sex; Bos indicus and B. indicus-cross animals are usually more temperamental than European breeds. Young bulls, when moved in groups, show a degree of playfulness (pushing and shoving) but become more aggressive and territorial with age. Adult bulls have a minimum personal space of six square metres. Cows with young calves can be very protective, and handling calves in the presence of their mothers can be dangerous. Cattle tend to avoid “dead end” in passages.

**Goats** should be handled calmly and are more easily led or driven than if they are excited. When goats are moved, their gregarious tendencies should be exploited. Activities which frighten, injure or cause agitation to animals should be avoided. Bullying is particularly serious in goats. Housing strange goats together could result in fatalities, either through physical violence, or subordinate goats being refused access to food and water.
**Horses** in this context include all solipeds, donkeys, mules, hinnies and zebra. They have good eyesight and a very wide angle of vision. They may have a history of loading resulting in good or bad experiences. Good training should result in easier loading, but some horses can prove difficult, especially if they are inexperienced or have associated loading with poor transport conditions. In these circumstances, two experienced animal handlers can load an animal by linking arms or using a strop below its rump. Blindfolding may even be considered. Ramps should be as shallow as possible. Steps are not usually a problem when horses mount a ramp, but they tend to jump a step when descending, so steps should be as low as possible. Horses benefit from being individually stalled, but may be transported in compatible groups. When horses are to travel in groups, their shoes should be removed.

**Pigs** have poor eyesight, and may move reluctantly in unfamiliar. They benefit from well lit loading bays. Since they negotiate ramps with difficulty, these should be as level as possible and provided with secure footholds. Ideally, a hydraulic lift should be used for greater heights. Pigs also negotiate steps with difficulty. A good ‘rule-of-thumb’ is that no step should be higher than the pig’s front knee. Serious aggression may result if unfamiliar animals are mixed. Pigs are highly susceptible to heat stress.

**Sheep** are sociable animals with good eyesight and tend to “flock together”, especially when they are agitated. They should be handled calmly and their tendency to follow each other should be exploited when they are being moved. Sheep may become agitated if they are singled out for attention and will strive to rejoin the group. Activities which frighten, injure or cause agitation to sheep should be avoided. They can negotiate steep ramps.
APPENDIX 3.7.3.

GUIDELINES FOR THE TRANSPORT OF ANIMALS BY LAND

**Preamble**: These guidelines apply to the following live domesticated animals: cattle, buffalo, camels, sheep, goats, pigs, poultry and equines. They will also be largely applicable to some other animals (e.g., deer, other camelids and ratites). Wild, feral and partly domesticated animals may need different conditions.

Article 3.7.3.1.

The amount of time animals spend on a journey should be kept to the minimum.

Article 3.7.3.2.

1. **Animal behaviour**

   Animal handlers should be experienced and competent in handling and moving farm livestock and understand the behaviour patterns of animals and the underlying principles necessary to carry out their tasks.

   The behaviour of individual animals or groups of animals will vary, depending on their breed, sex, temperament and age and the way in which they have been reared and handled. Despite these differences, the following behaviour patterns which are always present to some degree in domestic animals, should be taken into consideration in handling and moving the animals.

   Most domestic livestock are kept in herds and follow a leader by instinct.

   Animals which are likely to harm each other in a group situation should not be mixed.

   The desire of some animals to control their personal space should be taken into account in designing loading and unloading facilities, transport vehicles and containers.

   Domestic animals will try to escape if any person approaches closer than a certain distance. This critical distance, which defines the flight zone, varies among species and individuals of the same species, and depends upon previous contact with humans. Animals reared in close proximity to humans (i.e. tame) have a smaller flight zone, whereas those kept in free range or extensive systems may have flight zones which may vary from one metre to many metres. Animal handlers should avoid sudden penetration of the flight zone which may cause a panic reaction which could lead to aggression or attempted escape.
Animal handlers should use the point of balance at the animal’s shoulder to move animals, adopting a position behind the point of balance to move an animal forward and in front of the point of balance to move it backward.

Domestic animals have wide-angle vision but only have limited forward binocular vision and poor perception of depth. This means that they can detect objects and movements beside and behind them, but can only judge distances directly ahead.

Although all domestic animals have a highly sensitive sense of smell, they may react differently to the smells encountered during travel. Smells which cause fear or other negative responses should be taken into consideration when managing animals.

Domestic animals can hear over a greater range of frequencies than humans and are more sensitive to higher frequencies. They tend to be alarmed by constant loud noise and by sudden noises, which may cause them to panic. Sensitivity to such noises should also be taken into account when handling animals.
2. **Distractions and their removal**

Distractions that may cause approaching animals to stop, baulk or turn back should be designed out from new loading and unloading facilities or removed from existing ones. Below are examples of common distractions and methods for eliminating them:

- **a)** reflections on shiny metal or wet floors - move a lamp or change lighting;
- **b)** dark entrances - illuminate with indirect lighting which does not shine directly into the eyes of approaching animals;
- **c)** animals seeing moving people or equipment up ahead - install solid sides on chutes and races or install shields;
- **d)** dead ends - avoid if possible by curving the passage, or make an illusory passage;
- **e)** chains or other loose objects hanging in chutes or on fences - remove them;
- **f)** uneven floors or a sudden drop in floor levels - avoid uneven floor surfaces or install a solid false floor to provide an illusion of a solid and continuous walking surface;
- **g)** sounds of air hissing from pneumatic equipment - install silencers or use hydraulic equipment or vent high pressure to the external environment using flexible hosing;
- **h)** clanging and banging of metal objects - install rubber stops on gates and other devices to reduce metal to metal contact;
- **i)** air currents from fans or air curtains blowing into the face of animals - redirect or reposition equipment.

**Article 3.7.3.3.**

**Responsibilities**

Once the decision to transport the animals has been made, the welfare of the animals during their journey is the paramount consideration and is the joint responsibility of all people involved. The individual responsibilities of persons involved will be described in more detail in this Article.

The roles of each of those responsible are defined below:

1. **The owners and managers of the animals are responsible for:**

   - **a)** the general health, overall welfare and fitness of the animals for the journey;
   - **b)** ensuring compliance with any required veterinary or other certification;
   - **c)** the presence of an animal handler competent for the species being transported during the journey with the authority to take prompt action; in case of transport by individual trucks, the truck driver may be the sole animal handler during the journey;
   - **d)** the presence of an adequate number of animal handlers during loading and unloading;
   - **e)** ensuring that equipment and veterinary assistance are provided as appropriate for the species and the journey.
2. Business agents or buying/selling agents are responsible for:

a) selection of animals that are fit to travel;

b) availability of suitable facilities at the start and at the end of the journey for the assembly, loading, transport, unloading and holding of animals, including for any stops at resting points during the journey and for emergencies.

3. Animal handlers are responsible for the humane handling and care of the animals, especially during loading and unloading, and for maintaining a journey log. To carry out their responsibilities, they should have the authority to take prompt action. In the absence of a separate animal handler, the driver is the animal handler.

4. Transport companies, vehicle owners and drivers are responsible for planning the journey to ensure the care of the animals; in particular they are responsible for:

a) choosing appropriate vehicles for the species transported and the journey;

b) ensuring that properly trained staff are available for loading/unloading of animals;

c) ensuring adequate competency of the driver in matters of animal welfare for the species being transported in case a separate animal handler is not assigned to the truck;

d) developing and keeping up-to-date contingency plans to address emergencies (including adverse weather conditions) and minimise stress during transport;

e) producing a journey plan which includes a loading plan, journey duration, itinerary and location of resting places;

f) loading only those animals which are fit to travel, for their correct loading into the vehicle and their inspection during the journey, and for appropriate responses to problems arising. If its fitness to travel is in doubt, the animal should be examined by a veterinarian in accordance with point 3a) of Article 3.7.3.7.;

g) welfare of the animals during the actual transport.

5. Managers of facilities at the start and at the end of the journey and at resting points are responsible for:

a) providing suitable premises for loading, unloading and securely holding the animals, with water and feed when required, until further transport, sale or other use (including rearing or slaughter);

b) providing an adequate number of animal handlers to load, unload, drive and hold animals in a manner that causes minimum stress and injury; in the absence of a separate animal handler, the driver is the animal handler.

c) minimising the opportunities for disease transmission;

d) providing appropriate facilities, with water and feed when required;

e) providing appropriate facilities for emergencies;

f) providing facilities for washing and disinfecting vehicles after unloading;

g) providing facilities and competent staff to allow the humane killing of animals when required.
h) ensuring proper rest times and minimal delay during stops.

6. The responsibilities of Competent Authorities include:

a) establishing minimum standards for animal welfare, including requirements for inspection of animals before, during and after their travel, defining ‘fitness to travel’ and appropriate certification and record keeping;

b) setting standards for facilities, containers and vehicles for the transport of animals;

c) setting standards for the competence of animal handlers, drivers and managers of facilities in relevant issues in animal welfare;

d) ensuring appropriate awareness and training of animal handlers, drivers and managers of facilities in relevant issues in animal welfare;

e) implementation of the standards, including through accreditation of / interaction with other organisations;

f) monitoring and evaluating the effectiveness of standards of health and other aspects of welfare;

g) monitoring and evaluating the use of veterinary medications;

h) giving animal consignments priority at frontiers in order to allow them to pass without unnecessary delay.

7. All individuals, including veterinarians, involved in transporting animals and the associated handling procedures should receive appropriate training and be competent to meet their responsibilities.

8. The receiving Competent Authority should report back to the sending Competent Authority on significant animal welfare problems which occurred during the journey.

Article 3.7.3.4.

Competence

1. All people responsible for animals during journeys, should be competent according to their responsibilities listed in Article 3.7.3.3. Competence may be gained through formal training and/or practical experience.

2. The assessment of the competence of animal handlers should at a minimum address knowledge, and ability to apply that knowledge, in the following areas:

a) planning a journey, including appropriate space allowance, and feed, water and ventilation requirements;

b) responsibilities for animals during the journey, including loading and unloading;

c) sources of advice and assistance;

d) animal behaviour, general signs of disease, and indicators of poor animal welfare such as stress, pain and fatigue, and their alleviation;
Annex XXIII (contd)

e) assessment of fitness to travel; if fitness to travel is in doubt, the animal should be examined by a veterinarian;

f) relevant authorities and applicable transport regulations, and associated documentation requirements;

g) general disease prevention procedures, including cleaning and disinfection;

h) appropriate methods of animal handling during transport and associated activities such as assembling, loading, and unloading;

i) methods of inspecting animals, managing situations frequently encountered during transport such as adverse weather conditions, and dealing with emergencies, including humane killing;

j) species-specific aspects and age-specific aspects of animal handling and care, including feeding, watering and inspection; and

k) maintaining a journey log and other records.

Article 3.7.3.5.

Planning the journey

1. General considerations

   a) Adequate planning is a key factor affecting the welfare of animals during a journey.

   b) Before the journey starts, plans should be made in relation to:

      i) preparation of animals for the journey;

      ii) choice of road, or rail, roll-on roll-off vessels or containers;

      iii) nature and duration of the journey;

      iv) vehicle/container design and maintenance, including roll-on roll-off vessels;

      v) required documentation;

      vi) space allowance;

      vii) rest, water and feed;

      viii) observation of animals en route;

      ix) control of disease;

      x) emergency response procedures;

      xi) forecast weather conditions (e.g. conditions being too hot or too cold to travel during certain periods of the day);

      xii) transfer time when changing mode of transport, and
xiii) waiting time at frontiers and inspection points.

c) Regulations concerning drivers (for example, maximum driving periods) should take into account animal welfare whenever is possible.

2. Preparation of animals for the journey

a) When animals are to be provided with a novel diet or method of water provision during transport, an adequate period of adaptation should be planned. For all animals it is extra important that the rest stops during long journeys are long enough to fulfill the needs of the animals of feed and water. Species-specific short period of feed deprivation prior to loading may be desirable.

b) Animals more accustomed to contact with humans and with being handled are likely to be less fearful of being loaded and transported. Animal handlers should handle and load animals in a manner that reduces their fearfulness and improves their approachability.

c) Behaviour-modifying compounds (such as tranquillisers) or other medication should not be used routinely during transport. Such compounds should only be administered when a problem exists in an individual animal, and should be administered by a veterinarian or other person who has been instructed in their use by a veterinarian.

3. Nature and duration of the journey

The maximum duration of a journey should be determined taking into account factors, such as:

a) the ability of the animals to cope with the stress of transport (such as very young, old, lactating or pregnant animals);

b) the previous transport experience of the animals;

c) the likely onset of fatigue;

d) the need for special attention;

e) the need for feed and water;

f) the increased susceptibility to injury and disease;

g) space allowance, vehicle design, road conditions and driving quality;

h) weather conditions;

i) vehicle type used, terrain to be traversed, road surfaces and quality, skill and experience of the driver.

4. Vehicle and container design and maintenance

a) Vehicles and containers used for the transport of animals should be designed, constructed and fitted as appropriate for the species, size and weight of the animals to be transported. Special attention should be paid to avoid injury to animals through the use of secure smooth fittings free from sharp protrusions. The avoidance of injury to drivers, and animal handlers while carrying out their responsibilities should be emphasised.
b) Vehicles and containers should be designed with the structures necessary to provide protection from adverse weather conditions and to minimise the opportunity for animals to escape.

c) In order to minimise the likelihood of the spread of infectious disease during transport, vehicles and containers should be designed to permit thorough cleaning and disinfection, and the containment of faeces and urine during a journey.

d) Vehicles and containers should be maintained in good mechanical and structural condition.

e) Vehicles and containers should have adequate ventilation to meet variations in climate and the thermo-regulatory needs of the animal species being transported; the ventilation system (natural or mechanical) should be effective when the vehicle is stationary, and the airflow should be adjustable.

f) Vehicles should be designed so that the faeces or urine from animals on upper levels do not soil animals on lower levels, nor their feed and water.

g) When vehicles are carried on board ferries, facilities for adequately securing them should be available.

h) If feeding or watering while the vehicle is moving is required, adequate facilities on the vehicle should be available.

i) When appropriate, suitable bedding should be added to vehicle floors to assist absorption of urine and faeces, to minimise slipping by animals, and protect animals (especially young animals) from hard flooring surfaces and adverse weather conditions.

5. Special provisions for transport in vehicles (road and rail) on roll-on/roll-off vessels or for containers

a) Vehicles and containers should be equipped with a sufficient number of adequately designed, positioned and maintained securing points enabling them to be securely fastened to the vessel.

b) Vehicles and containers should be secured to the vessel before the start of the sea journey to prevent them being displaced by the motion of the vessel.

c) Roll-on/roll-off vessels should have adequate ventilation to meet variations in climate and the thermo-regulatory needs of the animal species being transported, especially where the animals are transported in a secondary vehicle/container on enclosed decks.

6. Space allowance

a) The number of animals which should be transported on a vehicle or in a container and their allocation to compartments should be determined before loading.

b) The space required on a vehicle or in a container depends upon whether or not the animals need to lie down (for example, pigs, camels and poultry), or to stand (horses). Animals which will need to lie down often stand when first loaded or when the vehicle is driven with too much lateral movement or sudden braking.

c) When animals lie down, they should all be able to adopt a normal lying posture which allows necessary thermoregulation.

d) When animals are standing, they should have sufficient space to adopt a balanced position as appropriate to the climate and species transported.
e) The amount of headroom necessary depends on the species of animal. Each animal should be able to assume its natural position for transport (including during loading and unloading) without coming into contact with the roof or upper deck of the vehicle, and there should be sufficient headroom to allow adequate airflow over the animals.

f) Calculations for the space allowance for each animal should be carried out using the figures given in a relevant national or international document. The number and size of pens on the vehicle should be varied to where possible accommodate already established groups of animals while avoiding group sizes which are too large.

g) Other factors which may influence space allowance include:
   i) vehicle/container design;
   ii) length of journey;
   iii) need to provide feed and water on the vehicle;
   iv) quality of roads;
   v) expected weather conditions;
   vi) category and sex of the animals.

7. Rest, water and feed

   a) Suitable water and feed should be available as appropriate and needed for the species, age, and condition of the animals, as well as the duration of the journey, climatic conditions, etc.

   b) Animals should be allowed to rest at resting points at appropriate intervals during the journey. The type of transport, the age and species of the animals being transported, and climatic conditions should determine the frequency of rest stops and whether the animals should be unloaded. Water and feed should be available during rest stops.

8. Ability to observe animals during the journey

   a) Animals should be positioned to enable each animal to be observed regularly during the journey to ensure their safety and good welfare.

   b) If the animals are in crates or on multi-tiered vehicles which do not allow free access for observation, for example where the roof of the tier is too low, animals cannot be inspected adequately, and serious injury or disease could go undetected. In these circumstances, a shorter journey duration should be allowed, and the maximum duration will vary according to the rate at which problems arise in the species and under the conditions of transport.

9. Control of disease

   As animal transport is often a significant factor in the spread of infectious diseases, journey planning should take the following into account:

   a) mixing of animals from different sources in a single consignment should be minimised;

   b) contact at resting points between animals from different sources should be avoided;
Annex XXIII (contd)

c) when possible, animals should be vaccinated against diseases to which they are likely to be exposed at their destination;

d) medications used prophylactically or therapeutically should be approved by the Veterinary Authority of the importing country and should only be administered by a veterinarian or other person who has been instructed in their use by a veterinarian.

10. Emergency response procedures

There should be an emergency management plan that identifies the important adverse events that may be encountered during the journey, the procedures for managing each event and the action to be taken in an emergency. For each important event, the plan should document the actions to be undertaken and the responsibilities of all parties involved, including communications and record keeping.

11. Other considerations

a) Extreme weather conditions are hazardous for animals undergoing transport and require appropriate vehicle design to minimise risks. Special precautions should be taken for animals that have not been acclimatised or which are unsuited to either hot or cold conditions. In some extreme conditions of heat or cold, animals should not be transported at all.

b) In some circumstances, transportation during the night may reduce thermal stress or the adverse effects of other external stimuli.

Article 3.7.3.6.

Documentation

1. Animals should not be loaded until the documentation required to that point is complete.

2. The documentation accompanying the consignment should include:

   a) journey travel plan and an emergency management plan;

   b) date, time, and place of loading and unloading;

   c) veterinary certification, when required;

   d) animal welfare competencies of the driver; (under study)

   e) animal identification to allow animal traceability to the premises of departure and, where possible, to the premises of origin;

   f) details of any animals considered at particular risk of suffering poor welfare during transport (point 3e) of Article 3.7.3.7.);

   g) documentation of the period of rest, and access to feed and water, prior to the journey;

   h) stocking density estimate for each load in the consignment;

   i) the journey log - daily record of inspection and important events, including records of morbidity and mortality and actions taken, climatic conditions, rest stops, travel time and distance, feed and water offered and estimates of consumption, medication provided, and mechanical defects.
3. When veterinary certification is required to accompany consignments of animals, it should address:
   a) fitness of animals to travel;
   b) animal identification (description, number, etc.);
   c) health status including any tests, treatments and vaccinations carried out;
   d) when required, details of disinfection carried out.

At the time of certification, the veterinarian should notify animal handler or the driver of any factors affecting the fitness of animals to travel for a particular journey.

Article 3.7.3.7.

Pre-journey period

1. General considerations
   a) Pre-journey rest is necessary if the welfare of animals has become poor during the collection period because of the physical environment or the social behaviour of the animals. The need for rest should be judged by a veterinarian or other competent person.
   b) Pre-journey assembly/holding areas should be designed to:
      i) securely hold the animals;
      ii) maintain a safe environment from hazards, including predators and disease;
      iii) protect animals from exposure to severe weather conditions;
      iv) allow for maintenance of social groups;
      v) allow for rest, and appropriate water and feed;
   c) Consideration should be given to the previous transport experience, training and conditioning of the animals, if known, as these may reduce fear and stress in animals.
   d) Feed and water should be provided pre-journey if the journey duration is greater than the normal inter-feeding and drinking interval for the animal. Recommendations for specific species are described in detail in Article 3.7.3.12.
   e) When animals are to be provided with a novel diet or method of feed or water provision during the journey, an adequate period of adaptation should be allowed.
   f) Before each journey, vehicles and containers should be thoroughly cleaned and, if necessary, treated for animal health and public health purposes, using methods approved by the Competent Authority. When cleaning is necessary during a journey, this should be carried out with the minimum of stress and risk to the animals.
   g) Where an animal handler believes that there is a significant risk of disease among the animals to be loaded or significant doubt as to their fitness to travel, the animals should be examined by a veterinarian.
Annex XXIII (contd)

2. **Selection of compatible groups**

Compatible groups should be selected before transport to avoid adverse animal welfare consequences. The following guidelines should be applied when assembling groups of animals:

a) Animals reared together should be maintained as a group; animals with a strong social bond, such as a dam and offspring, should be transported together.

b) Animals of the same species can be mixed unless there is a significant likelihood of aggression; aggressive individuals should be segregated (recommendations for specific species are described in detail in Article 3.7.3.12.). For some species, animals from different groups should not be mixed because poor welfare occurs unless they have established a social structure.

c) Young or small animals should be separated from older or larger animals, with the exception of nursing mothers with young at foot.

d) Animals with horns or antlers should not be mixed with animals lacking horns or antlers unless judged to be compatible.

e) Animals of different species should not be mixed unless they are judged to be compatible.

3. **Fitness to travel**

a) Each animal should be inspected by a veterinarian or an animal handler to assess fitness to travel. If its fitness to travel is in doubt, the animal should be examined by a veterinarian. Animals found unfit to travel should not be loaded onto a vehicle, except for transport to receive veterinary treatment.

b) Humane and effective arrangements should be made by the owner and the agent for the handling and care of any animal rejected as unfit to travel.

c) Animals that are unfit to travel include, but may not be limited to:

i) those that are sick, injured, weak, disabled or fatigued;

ii) those that are unable to stand unaided and bear weight on each leg;

iii) those that are blind in both eyes;

iv) those that cannot be moved without causing them additional suffering;

v) newborn with an unhealed navel;

vi) pregnant animals which would be in the final 10% of their gestation period at the planned time of unloading;

vii) females travelling without young which have given birth within the previous 48 hours;

viii) those whose body condition would result in poor welfare because of the expected climatic conditions.

d) Risks during transport can be reduced by selecting animals best suited to the conditions of travel and those that are acclimatised to expected weather conditions.
e) Animals at particular risk of suffering poor welfare during transport and which require special conditions (such as in the design of facilities and vehicles, and the length of the journey) and additional attention during transport, may include:

i) large or obese individuals;
ii) very young or old animals;
iii) excitable or aggressive animals;
iv) animals which have had little contact with humans;
v) animal subject to motion sickness;
vii) females in late pregnancy or heavy lactation, dam and offspring;
vii) animals with a history of exposure to stressors or pathogenic agents prior to transport;
viii) animals with unhealed wounds from recent surgical procedures such as dehorning.

4. Specific species requirements

Transport procedures should be able to take account of variations in the behaviour of the species. Flight zones, social interactions and other behaviour vary significantly among species and even within species. Facilities and handling procedures that are successful with one species are often ineffective or dangerous with another.

Recommendations for specific species are described in detail in Article 3.7.3.12.

Article 3.7.3.8.

Loading

1. Competent supervision

a) Loading should be carefully planned as it has the potential to be the cause of poor welfare in transported animals.

b) Loading should be supervised and/or conducted by animal handlers. The animals are to be loaded quietly and without unnecessary noise, harassment or force. Untrained assistants or spectators should not impede the process.

c) When containers are loaded onto a vehicle, this should be carried out in such a way to avoid poor animal welfare.

2. Facilities

a) The facilities for loading including the collecting area, races and loading ramps should be designed and constructed to take into account the needs and abilities of the animals with regard to dimensions, slopes, surfaces, absence of sharp projections, flooring, etc.

b) Loading facilities should be properly illuminated to allow the animals to be observed by animal handler(s), and to allow the ease of movement of the animals at all times. Facilities should provide uniform light levels directly over approaches to sorting pens, chutes, loading ramps, with brighter light levels inside vehicles/containers, in order to minimise baulking. Dim light levels may be advantageous for the catching of poultry and some other animals. Artificial lighting may be required.
Annex XXIII (contd)

c) Ventilation during loading and the journey should provide for fresh air, the removal of excessive heat, humidity and noxious fumes (such as ammonia and carbon monoxide), and the prevention of accumulations of ammonia and carbon dioxide. Under warm and hot conditions, ventilation should allow for the adequate convective cooling of each animal. In some instances, adequate ventilation can be achieved by increasing the space allowance for animals.

3. Goads and other aids

When moving animals, their species specific behaviour should be used (see Article 3.7.3.12.). If goads and other aids are necessary, the following principles should apply:

a) Animals that have little or no room to move should not be subjected to physical force or goads and other aids which compel movement. Electric goads and prods should only be used in extreme cases and not on a routine basis to move animals. The use and the power output should be restricted to that necessary to assist movement of an animal and only when an animal has a clear path ahead to move. Goads and other aids should not be used repeatedly if the animal fails to respond or move. In such cases it should be investigated whether some physical or other impediment is preventing the animal from moving.

b) The use of such devices should be limited to battery-powered goads on the hindquarters of pigs and large ruminants, and never on sensitive areas such as the eyes, mouth, ears, anogenital region or belly. Such instruments should not be used on horses, sheep and goats of any age, or on calves or piglets.

c) Useful and permitted goads include panels, flags, plastic paddles, flappers (a length of cane with a short strap of leather or canvas attached), plastic bags and rattles; they should be used in a manner sufficient to encourage and direct movement of the animals without causing undue stress.

d) Painful procedures (including whipping, tail twisting, use of nose twitches, pressure on eyes, ears or external genitalia), or the use of goads or other aids which cause pain and suffering (including large sticks, sticks with sharp ends, lengths of metal piping, fencing wire or heavy leather belts), should not be used to move animals.

e) Excessive shouting at animals or making loud noises (e.g., through the cracking of whips) to encourage them to move should not occur, as such actions may make the animals agitated, leading to crowding or falling.

f) The use of well trained dogs to help with the loading of some species may be acceptable.

g) Animals should be grasped or lifted in a manner which avoids pain or suffering and physical damage (e.g. bruising, fractures, dislocations). In the case of quadrupeds, manual lifting by a person should only be used in young animals or small species, and in a manner appropriate to the species; grasping or lifting animals only by their wool, hair, feathers, feet, neck, ears, tails, head, horns, limbs causing pain or suffering should not be permitted, except in an emergency where animal welfare or human safety may otherwise be compromised.

h) Conscious animals should not be thrown, dragged or dropped.

i) Performance standards should be established in which numerical scoring is used to evaluate the use of such instruments, and to measure the percentage of animals moved with an electric instrument and the percentage of animals slipping or falling as a result of their usage.
Article 3.7.3.9.

Travel

1. General considerations

   a) Drivers and animal handlers should check the load immediately before departure to ensure that
   the animals have been properly loaded. Each load should be checked again early in the trip and
   adjustments made as appropriate. Periodic checks should be made throughout the trip,
   especially at rest or refuelling stops or during meal breaks when the vehicle is stationary.

   b) Drivers should utilise smooth, defensive driving techniques, without sudden turns or stops, to
   minimise uncontrolled movements of the animals.

2. Methods of restraining or containing animals

   a) Methods of restraining animals should be appropriate to the species and age of animals involved
   and the training of the individual animal.

   b) Recommendations for specific species are described in detail in Article 3.7.3.12.

3. Regulating the environment within vehicles or containers

   a) Animals should be protected against harm from hot or cold conditions during travel. Effective
   ventilation procedures for maintaining the environment within vehicles or containers will vary
   according to whether conditions are cold, hot and dry or hot and humid, but in all conditions a
   build-up of noxious gases should be prevented.

   b) The environment within vehicles or containers in hot and warm weather can be regulated by the
   flow of air produced by the movement of the vehicle. In warm and hot weather, the duration of
   journey stops should be minimised and vehicles should be parked under shade, with adequate and
   appropriate ventilation.

   c) To minimise slipping and soiling, and maintain a healthy environment, urine and faeces should
   be removed from floors when necessary and disposed of in such a way as to prevent the
   transmission of disease and in compliance with all relevant health and environmental legislation.

4. Sick, injured or dead animals

   a) A driver or animal handler finding sick, injured or dead animals should act according to a
   predetermined emergency response plan.

   b) Sick or injured animals should be segregated.

   c) Ferries (roll-on roll-off) should have procedures to treat sick or injured animals during the
   journey.

   d) In order to reduce the likelihood that animal transport will increase the spread of infectious
   disease, contact between transported animals, or the waste products of the transported animals,
   and other farm animals should be minimised.

   e) During the journey, when disposal of a dead animal becomes necessary, this should be carried out
   in such a way as to prevent the transmission of disease and in compliance with all relevant health
   and environmental legislation.
Annex XXIII (contd)

f) When killing is necessary, it should be carried out as quickly as possible and assistance should be sought from a veterinarian or other person(s) competent in humane killing procedures. Recommendations for specific species are described in Appendix 3.7.6. on killing of animals for disease control purposes.

5. Water and feed requirements

a) If journey duration is such that feeding or watering is required or if the species requires feed or water throughout, access to suitable feed and water for all the animals (appropriate for their species and age) carried in the vehicle should be provided. There should be adequate space for all animals to move to the feed and water sources and due account taken of likely competition for feed.

b) Recommendations for specific species are described in detail in Article 3.7.12.

6. Rest periods and conditions including hygiene

a) Animals that are being transported should be rested at appropriate intervals during the journey and offered feed and water, either on the vehicle or, if necessary, unloaded into suitable facilities.

b) Suitable facilities should be used en route, when resting requires the unloading of the animals. These facilities should meet the needs of the particular animal species and should allow access of all animals to feed and water.

7. In-transit observations

a) Animals being transported by road should be observed soon after a journey is commenced and whenever the driver has a rest stop. After meal breaks and refuelling stops, the animals should be observed immediately prior to departure.

b) Animals being transported by rail should be observed at each scheduled stop. The responsible rail transporter should monitor the progress of trains carrying animals and take all appropriate action to minimise delays.

c) During stops, it should be ensured that the animals continue to be properly confined, have appropriate feed and water, and their physical condition is satisfactory.

Article 3.7.3.10.

Unloading and post-journey handling

1. General considerations

a) The required facilities and the principles of animal handling detailed in Article 3.7.3.8. apply equally to unloading, but consideration should be given to the likelihood that the animals will be fatigued.

b) Unloading should be supervised and/or conducted by an animal handler with knowledge and experience of the behavioural and physical characteristics of the species being unloaded. Animals should be unloaded from the vehicle into appropriate facilities as soon as possible after arrival at the destination but sufficient time should be allowed for unloading to proceed quietly and without unnecessary noise, harassment or force.
c) Facilities should provide all animals with appropriate care and comfort, adequate space and ventilation, access to feed (if appropriate) and water, and shelter from extreme weather conditions.

d) For details regarding the unloading of animals at a slaughterhouse, see Appendix 3.7.5. on slaughter of animals for human consumption.

2. **Sick and/or injured animals**

   a) An animal that has become sick, injured or disabled during a journey should be appropriately treated or humanely killed (see Appendix 3.7.6. on killing of animals for disease control purposes). If necessary, veterinary advice should be sought in the care and treatment of these animals. In some cases, where animals are non-ambulatory due to fatigue, injury or sickness, it may be in the best welfare interests of the animal to be treated or killed aboard the vehicle. Assistance should be sought from a veterinarian or other person(s) competent in humane killing procedures.

   b) At the destination, the animal handler or the driver during transit should ensure that responsibility for the welfare of sick, injured or disabled animals is transferred to a veterinarian or other suitable person.

   c) If treatment or humane killing is not possible aboard the vehicle, there should be appropriate facilities and equipment for the humane unloading of animals that are non-ambulatory due to fatigue, injury or sickness. These animals should be unloaded in a manner that causes the least amount of suffering. After unloading, separate pens and other appropriate facilities should be available for sick or injured animals.

   d) Feed, if appropriate, and water should be available for each sick or injured animal.

3. **Addressing disease risks**

   The following should be taken into account in addressing the greater risk of disease due to animal transport and the possible need for segregation of transported animals at the destination:

   a) increased contact among animals, including those from different sources and with different disease histories;

   b) increased shedding of pathogens and increased susceptibility to infection related to stress and impaired defences against disease, including immunosuppression;

   c) exposure of animals to pathogens which may contaminate vehicles, resting points, markets, etc.

4. **Cleaning and disinfection**

   a) Vehicles, crates, containers, etc. used to carry the animals should be cleaned before re-use through the physical removal of manure and bedding by scraping, washing and flushing with water and detergent. This should be followed by disinfection when there are concerns about disease transmission.

   b) Manure, litter, bedding and the bodies of any animals which die during the journey should be disposed of in such a way as to prevent the transmission of disease and in compliance with all relevant health and environmental legislation.
Annex XXIII (contd)

c) Establishments like livestock markets, slaughterhouses, resting sites, railway stations, etc. where animals are unloaded should be provided with appropriate areas for the cleaning and disinfection of vehicles.

Article 3.7.3.11.

Actions in the event of a refusal to allow the completion of the journey

1. The welfare of the animals should be the first consideration in the event of a refusal to allow the completion of the journey.

2. When the animals have been refused import, the Competent Authority of the importing country should make available suitable isolation facilities to allow the unloading of animals from a vehicle and their secure holding, without posing a risk to the health of national herd or flock, pending resolution of the situation. In this situation, the priorities should be:
   a) The Competent Authority of the importing country should provide urgently in writing the reasons for the refusal.
   b) In the event of a refusal for animal health reasons, the Competent Authority of the importing country should provide urgent access to a veterinarian, where possible an OIE veterinarian(s) appointed by the Director General, to assess the health status of the animals with regard to the concerns of the importing country, and the necessary facilities and approvals to expedite the required diagnostic testing.
   c) The Competent Authority of the importing country should provide access to allow continued assessment of the health and other aspects of the welfare of the animals.
   d) If the matter cannot be promptly resolved, the Competent Authorities of the exporting and importing countries should call on the OIE to mediate.

3. In the event that a Competent Authority requires the animals to remain on the vehicle, the priorities should be:
   a) to allow re-provisioning of the vehicle with water and feed as necessary;
   b) to provide urgently in writing the reasons for the refusal;
   c) to provide urgent access to an independent veterinarian(s) to assess the health status of the animals, and the necessary facilities and approvals to expedite the required diagnostic testing in the event of a refusal for animal health reasons;
   d) to provide access to allow continued assessment of the health and other aspects of the welfare of the animals, and the necessary actions to deal with any animal issues which arise.

4. The OIE should utilise its dispute settlement mechanism to identify a mutually agreed solution which will address animal health and any other welfare issues in a timely manner.

Article 3.7.3.12.

Species specific issues

Camelids of the new world in this context comprise llamas, alpacas, guanaco and vicuna. They have good eyesight and, like sheep, can negotiate steep slopes, though ramps should be as shallow as possible. They load most easily in a bunch as a single animal will strive to rejoin the others. Whilst they are usually docile, they have an unnerving habit of spitting in self-defence. During transport, they usually lie down. They frequently extend their front legs forward when lying, so gaps below partitions should be high enough so that their legs are not trapped when the animals rise.
Cattle are sociable animals and may become agitated if they are singled out. Social order is usually established at about two years of age. When groups are mixed, social order has to be re-established and aggression may occur until a new order is established. Crowding of cattle may also increase aggression as the animals try to maintain personal space. Social behaviour varies with age, breed and sex; Bos indicus and B. indicus-cross animals are usually more temperamental than European breeds. Young bulls, when moved in groups, show a degree of playfulness (pushing and shoving) but become more aggressive and territorial with age. Adult bulls have a minimum personal space of six square metres. Cows with young calves can be very protective, and handling calves in the presence of their mothers can be dangerous. Cattle tend to avoid “dead end” in passages.

Goats should be handled calmly and are more easily led or driven than if they are excited. When goats are moved, their gregarious tendencies should be exploited. Activities which frighten, injure or cause agitation to animals should be avoided. Bullying is particularly serious in goats and can reflect demands for personal space. Housing strange goats together could result in fatalities, either through physical violence, or subordinate goats being refused access to food and water.

Horses in this context include, donkeys, mules and hinnies. They have good eyesight and a very wide angle of vision. They may have a history of loading resulting in good or bad experiences. Good training should result in easier loading, but some horses can prove difficult, especially if they are inexperienced or have associated loading with poor transport conditions. In these circumstances, two experienced animal handlers can load an animal by linking arms or using a strop below its rump. Blindfolding may even be considered. Ramps should be as shallow as possible. Steps are not usually a problem when horses mount a ramp, but they tend to jump a step when descending, so steps should be as low as possible. Horses benefit from being individually stalled, but may be transported in compatible groups. When horses are to travel in groups, their shoes should be removed. Horses are prone to respiratory disease if they are restricted by period by tethers that prevent the lowering and lifting of their heads.

Pigs have poor eyesight, and may move reluctantly in strange surroundings. They benefit from well lit loading bays. Since they negotiate ramps with difficulty, these should be as level as possible and provided with secure footholds. Ideally, a hydraulic lift should be used for greater heights. Pigs also negotiate steps with difficulty. A good ‘rule-of-thumb’ is that no step should be higher than the pig’s front knee. Serious aggression may result if unfamiliar animals are mixed. Pigs are highly susceptible to heat stress.

Sheep are sociable animals with good eyesight, a relatively subtle and undemonstrative behaviour and a tendency to “flock together”, especially when they are agitated. They should be handled calmly and their tendency to follow each other should be exploited when they are being moved. Crowding of sheep may lead to damaging aggressive and submissive behaviours as animals try to maintain personal space. Sheep may become agitated if they are singled out for attention, or kept alone, and will strive to rejoin the group. Activities which frighten, injure or cause agitation to sheep should be avoided. They can negotiate steep ramps.
APPENDIX 3.7.5.

GUIDELINES FOR THE SLAUGHTER OF ANIMALS

Article 3.7.5.1.

General principles

1. **Object**

   These guidelines address the need to ensure the welfare of food animals during pre-slaughter and slaughter processes, until they are dead.

   These guidelines apply to the slaughter in slaughterhouses of the following domestic animals: cattle, buffalo, bison, sheep, goats, camelids, deer, horses, pigs, ratites, rabbits and poultry. Other animals, wherever they have been reared, and all animals slaughtered outside slaughterhouses should be managed to ensure that their transport, lairage, restraint and slaughter is carried out without causing undue stress to the animals; the principles underpinning these guidelines apply also to these animals.

2. **Personnel**

   Persons engaged in the unloading, moving, lairage, care, restraint, stunning, slaughter and bleeding of animals play an important role in the welfare of those animals. For this reason, there should be a sufficient number of personnel, who should be patient, considerate, competent and familiar with the guidelines outlined in the present Appendix and their application within the national context.

   Competence may be gained through formal training and/or practical experience. This competence should be demonstrated through a current certificate from the Competent Authority or from an independent body accredited by the Competent Authority.

   The management of the slaughterhouse and the Veterinary Services should ensure that slaughterhouse staff are competent and carry out their tasks in accordance with the principles of animal welfare.

3. **Animal behaviour**

   Animal handlers should be experienced and competent in handling and moving farm livestock and understand the behaviour patterns of animals and the underlying principles necessary to carry out their tasks.

   The behaviour of individual animals or groups of animals will vary, depending on their breed, sex, temperament and age and the way in which they have been reared and handled. Despite these differences, the following behaviour patterns which are always present to some degree in domestic animals, should be taken into consideration in handling and moving the animals.

   Most domestic livestock are kept in herds and follow a leader by instinct.

   Animals which are likely to harm each other in a group situation should not be mixed at slaughterhouses.

   The desire of some animals to control their personal space should be taken into account in designing facilities.
Domestic animals will try to escape if any person approaches closer than a certain distance. This critical distance, which defines the flight zone, varies among species and individuals of the same species, and depends upon previous contact with humans. Animals reared in close proximity to humans (i.e. tame) have a smaller flight zone, whereas those kept in free range or extensive systems may have flight zones which may vary from one metre to many metres. Animal handlers should avoid sudden penetration of the flight zone which may cause a panic reaction which could lead to aggression or attempted escape.

**An example of a flight zone (cattle)**

Animal handlers should use the point of balance at the animal's shoulder to move animals, adopting a position behind the point of balance to move an animal forward and in front of the point of balance to move it backward.
Domestic animals have wide-angle vision but only have limited forward binocular vision and poor perception of depth. This means that they can detect objects and movements beside and behind them, but can only judge distances directly ahead.

Although all domestic animals have a highly sensitive sense of smell, they react in different ways to the smells of slaughterhouses. Smells which cause fear or other negative responses should be taken into consideration when managing animals.

Domestic animals can hear over a greater range of frequencies than humans and are more sensitive to higher frequencies. They tend to be alarmed by constant loud noise and by sudden noises, which may cause them to panic. Sensitivity to such noises should also be taken into account when handling animals.

4. Distractions and their removal

Distractions that may cause approaching animals to stop, balk or turn back should be designed out from new facilities or removed from existing ones. Below are examples of common distractions and methods for eliminating them:

a) reflections on shiny metal or wet floors - move a lamp or change lighting;

b) dark entrances to chutes, races, stun boxes or conveyor restrainers - illuminate with indirect lighting which does not shine directly into the eyes of approaching animals;

c) animals seeing moving people or equipment up ahead - install solid sides on chutes and races or install shields;

d) dead ends - avoid if possible by curving the passage, or make an illusory passage;

e) chains or other loose objects hanging in chutes or on fences - remove them;

f) uneven floors or a sudden drop in floor levels at the entrance to conveyor restrainers - avoid uneven floor surfaces or install a solid false floor under the restrainer to provide an illusion of a solid and continuous walking surface;

g) sounds of air hissing from pneumatic equipment - install silencers or use hydraulic equipment or vent high pressure to the external environment using flexible hosing;

h) clanging and banging of metal objects - install rubber stops on gates and other devices to reduce metal to metal contact;

i) air currents from fans or air curtains blowing into the face of animals - redirect or reposition equipment.

Article 3.7.5.2.

Moving and handling animals

1. General considerations

Animals should be transported to slaughter in a way that minimises adverse animal health and welfare outcomes, and the transport should be conducted in accordance with the OIE guidelines for the transportation of animals (Appendices 3.7.2 and 3.7.3).

The following principles should apply to unloading animals, moving them into lairage pens, out of the lairage pens and up to the slaughter point:
Annex XXIII (contd)

a) The conditions of the animals should be assessed upon their arrival for any animal welfare and health problems.

b) Injured or sick animals, requiring immediate slaughter, should be killed humanely and without delay, at the site where they are found in accordance with the OIE guidelines for the killing of animals for disease control purposes (Appendix 3.7.6.).

c) Animals should not be forced to move at a speed greater than their normal walking pace, in order to minimise injury through falling or slipping. Performance standards should be established where numerical scoring of the prevalence of animals slipping or falling is used to evaluate whether animal moving practices and/or facilities should be improved. In properly designed and constructed facilities with competent animal handlers, it should be possible to move 99% of animals without their falling.

d) Animals for slaughter should not be forced to walk over the top of other animals.

e) Animals should be handled in such a way as to avoid harm, distress or injury. Under no circumstances should animal handlers resort to violent acts to move animals, such as crushing or breaking tails of animals, grasping their eyes or pulling them by the ears. Animal handlers should never apply an injurious object or irritant substance to animals and especially not to sensitive areas such as eyes, mouth, ears, anogenital region or belly. The throwing or dropping of animals, or their lifting or dragging by body parts such as their tail, head, horns, ears, limbs, wool, hair or feathers, should not be permitted. The manual lifting of small animals is permissible.

f) When using goads and other aids, the following principles should apply:

i) Animals that have little or no room to move should not be subjected to physical force or goads and other aids which compel movement. Electric goads and prods should only be used in extreme cases and not on a routine basis to move animals. The use and the power output should be restricted to that necessary to assist movement of an animal and only when an animal has a clear path ahead to move. Goads and other aids should not be used repeatedly if the animal fails to respond or move. In such cases it should be investigated whether some physical or other impediment is preventing the animal from moving.

ii) The use of such devices should be limited to battery-powered goads on the hindquarters of pigs and large ruminants, and never on sensitive areas such as the eyes, mouth, ears, anogenital region or belly. Such instruments should not be used on horses, sheep and goats of any age, or on calves or piglets.

iii) Useful and permitted goads include panels, flags, plastic paddles, flappers (a length of cane with a short strap of leather or canvas attached), plastic bags and rattles; they should be used in a manner sufficient to encourage and direct movement of the animals without causing undue stress.

iv) Painful procedures (including whipping, tail twisting, use of nose twitches, pressure on eyes, ears or external genitalia), or the use of goads or other aids which cause pain and suffering (including large sticks, sticks with sharp ends, lengths of metal piping, fencing wire or heavy leather belts), should not be used to move animals.

v) Excessive shouting at animals or making loud noises (e.g. through the cracking of whips) to encourage them to move should not occur, as such actions may make the animals agitated, leading to crowding or falling.
vi) Animals should be grasped or lifted in a manner which avoids pain or suffering and physical damage (e.g. bruising, fractures, dislocations). In the case of quadrupeds, manual lifting by a person should only be used in young animals or small species, and in a manner appropriate to the species; grasping or lifting animals only by their wool, hair, feathers, feet, neck, ears, tails, head, horns, limbs causing pain or suffering should not be permitted, except in an emergency where animal welfare or human safety may otherwise be compromised.

vii) Conscious animals should not be thrown, dragged or dropped.

viii) Performance standards should be established to evaluate the use of such instruments. Numerical scoring may be used and to measure the percentage of animals moved with an electric instrument and the percentage of animals slipping or falling at a point in the slaughterhouse. Any risk of compromising animal welfare, for example slippery floor, should be investigated immediately and the defect rectified to eliminate the problem.

2. Provisions relevant to animals delivered in containers

a) Containers in which animals are transported should be handled with care, and should not be thrown, dropped or knocked over. Where possible, they should be horizontal while being loaded and unloaded mechanically, and stacked to ensure ventilation. In any case they should be moved and stored in an upright position as indicated by specific marks.

b) Animals delivered in containers with perforated or flexible bottoms should be unloaded with particular care in order to avoid injury. Where appropriate, animals should be unloaded from the containers individually.

c) Animals which have been transported in containers should be slaughtered as soon as possible; mammals and ratites which are not taken directly upon arrival to the place of slaughter should have drinking water available to them from appropriate facilities at all times. Delivery of poultry for slaughter should be scheduled such that they are not deprived of water at the premises for longer than 12 hours. Animals which have not been slaughtered within 12 hours of their arrival should be fed, and should subsequently be given moderate amounts of food at appropriate intervals.

3. Provisions relevant to restraining and containing animals

a) Provisions relevant to restraining animals for stunning or slaughter without stunning, to help maintain animal welfare, include:
   i) provision of a non-slippery floor;
   ii) avoidance of excessive pressure applied by restraining equipment that causes struggling or vocalisation in animals;
   iii) equipment engineered to reduce noise of air hissing and clanging metal;
   iv) absence of sharp edges in restraining equipment that would harm animals;
   v) avoidance of jerking or sudden movement of restraining device.

b) Methods of restraint causing avoidable suffering should not be used in conscious animals. Such methods include the following:
   i) suspending or hoisting animals (other than poultry) by the feet or legs;
   ii) indiscriminate and inappropriate use of stunning equipment;
   iii) mechanical clamping of the legs or feet of the animals (other than shackles used in poultry and ostriches) as the sole method of restraint;
iv) breaking legs, cutting leg tendons or blinding animals in order to immobilise them;
v) severing the spinal cord, for example using a puntilla or dagger, to immobilise animals using electric currents to immobilise animals, except for proper stunning.

Article 3.7.5.3.

Lairage design and construction

1. General considerations

The lairage should be designed and constructed to hold an appropriate number of animals in relation to the throughput rate of the slaughterhouse without compromising the welfare of the animals.

In order to permit operations to be conducted as smoothly and efficiently as possible without injury or undue stress to the animals, the lairage should be designed and constructed so as to allow the animals to move freely in the required direction, using their behavioural characteristics and without undue penetration of their flight zone.

The following guidelines may help to achieve this.

2. Design of lairages

a) The lairage should be designed to allow a one-way flow of animals from unloading to the point of slaughter, with a minimum number of abrupt corners to negotiate.

b) In red meat slaughterhouses, pens, passageways and races should be arranged in such a way as to permit inspection of animals at any time, and to permit the removal of sick or injured animals when considered to be appropriate, for which separate appropriate accommodation should be provided.

c) Each animal should have room to stand up and lie down and, when confined in a pen, to turn around, except where the animal is reasonably restrained for safety reasons (e.g. fractious bulls). Fractious animals should be slaughtered as soon as possible after arrival at the slaughterhouse to avoid welfare problems. The lairage should have sufficient accommodation for the number of animals intended to be held. Drinking water should always be available to the animals, and the method of delivery should be appropriate to the type of animal held. Troughs should be designed and installed in such a way as to minimise the risk of fouling by faeces, without introducing risk of bruising and injury in animals, and should not hinder the movement of animals.

d) Holding pens should be designed to allow as many animals as possible to stand or lie down against a wall. Where feed troughs are provided, they should be sufficient in number and feeding space to allow adequate access of all animals to feed. The feed trough should not hinder the movement of animals.

e) Where tethers, ties or individual stalls are used, these should be designed so as not to cause injury or distress to the animals and should also allow the animals to stand, lie down and access any food or water that may need to be provided.

f) Passageways and races should be either straight or consistently curved, as appropriate to the animal species. Passageways and races should have solid sides, but when there is a double race, the shared partition should allow adjacent animals to see each other. For pigs and sheep, passageways should be wide enough to enable two or more animals to walk side by side for as long as possible. At the point where passageways are reduced in width, this should be done by a means which prevents excessive bunching of the animals.
g) Animal handlers should be positioned alongside races and passageways on the inside radius of any curve, to take advantage of the natural tendency of animals to circle an intruder. Where one-way gates are used, they should be of a design which avoids bruising. Races should be horizontal but where there is a slope, they should be constructed to allow the free movement of animals without injury.

h) There should be a waiting pen, with a level floor and solid sides, between the holding pens and the race leading to the point of stunning or slaughter, to ensure a steady supply of animals for stunning or slaughter and to avoid having animal handlers trying to rush animals from the holding pens. The waiting pen should preferably be circular, but in any case, so designed that animals cannot be trapped or trampled.

i) Ramps or lifts should be used for loading and unloading of animals where there is a difference in height or a gap between the floor of the vehicle and the unloading area. Unloading ramps should be designed and constructed so as to permit animals to be unloaded from vehicles on the level or at the minimum gradient achievable. Lateral side protection should be available to prevent animals escaping or falling. They should be well drained, with secure footholds and adjustable to facilitate easy movement of animals without causing distress or injury.

3. Construction of lairages

a) Lairages should be constructed and maintained so as to provide protection from unfavourable climatic conditions, using strong and resistant materials such as concrete and metal which has been treated to prevent corrosion. Surfaces should be easy to clean. There should be no sharp edges or protuberances which may injure the animals.

b) Floors should be well drained and not slippery; they should not cause injury to the feet of the animals. Where necessary, floors should be insulated or provided with appropriate bedding. Drainage grids should be placed at the sides of pens and passageways and not where animals would have to cross them. Discontinuities or changes in floor patterns or texture which could cause baulking in the movement of animals should be avoided.

c) Lairages should be provided with adequate lighting, but care should be taken to avoid harsh lights and shadows, which frighten the animals or affect their movement. The fact that animals will move more readily from a darker area into a well-lit area might be exploited by providing for lighting that can be regulated accordingly.

d) Lairages should be adequately ventilated to ensure that waste gases (e.g. ammonia) do not build up and that draughts at animal height are minimised. Ventilation should be able to cope with the range of expected climatic conditions and the number of animals the lairage will be expected to hold.

e) Care should be taken to protect the animals from excessively or potentially disturbing noises, for example by avoiding the use of noisy hydraulic or pneumatic equipment, and muffling noisy metal equipment by the use of suitable padding, or by minimising the transmission of such a noise to the areas where animals are held and slaughtered.

f) Where animals are kept in outdoor lairages without natural shelter or shade, they should be protected from the effects of adverse weather conditions.

Article 3.7.5.4.

Care of animals in lairages

Animals in lairages should be cared for in accordance with the following guidelines:
Annex XXIII (contd)

1. As far as possible, established groups of animals should be kept together. Each animal should have enough space to stand up, lie down and turn around. Animals hostile to each other should be separated.

2. Where tethers, ties or individual stalls are used, they should allow animals to stand up and lie down without causing injury or distress.

3. Where bedding is provided, it should be maintained in a condition that minimises risks to the health and safety of the animals, and sufficient bedding should be used so that animals do not become soiled with manure.

4. Animals should be kept securely in the lairage, and care should be taken to prevent them from escaping and from predators.

5. Suitable drinking water should be available to the animals on their arrival and at all times to animals in lairages unless they are to be slaughtered without delay.

6. If animals are not to be slaughtered as soon as possible, suitable feed should be available to the animals on arrival and at intervals appropriate to the species. Unweaned animals should be slaughtered as soon as possible.

7. In order to prevent heat stress, animals subjected to high temperatures, particularly pigs and poultry, should be cooled by the use of water sprays, fans or other suitable means. However, the potential for water sprays to reduce the ability of animals to thermoregulate (especially poultry) should be considered in any decision to use water sprays. The risk of animals being exposed to very cold temperatures or sudden extreme temperature changes should also be considered.

8. The lairage area should be well lit in order to enable the animals to see clearly without being dazzled. During the night, the lights should be dimmed. Lighting should also be adequate to permit inspection of all animals. Subdued lighting, and for example blue light, may be useful in poultry lairages in helping to calm birds.

9. The condition and state of health of the animals in a lairage should be inspected at least every morning and evening by a veterinarian or, under the veterinarian’s responsibility, by another competent person, such as an animal handler. Animals which are sick, weak, injured or showing visible signs of distress should be separated, and veterinary advice should be sought immediately regarding treatment or euthanasia, or the animals should be humanely killed immediately if necessary.

10. Lactating dairy animals should be slaughtered as soon as possible. Dairy animals with obvious udder distension should be milked to minimise udder discomfort.

11. Animals which have given birth during the journey or in the lairage should be slaughtered as soon as possible or provided with conditions which are appropriate for suckling, for their welfare and the welfare of the newborn. Under normal circumstances, animals which are expected to give birth during a journey should not be transported.

12. Animals with horns, antlers or tusks capable of injuring other animals, if aggressive, should be penned separately.

Recommendations for specific species are described in detail in Articles 3.7.5.5. to 3.7.5.9.
Article 3.7.5.5.

Management of foetuses during slaughter of pregnant animals

Under normal circumstances, pregnant animals that would be in the final 10% of their gestation period at the planned time of unloading at the slaughterhouse should be neither transported nor slaughtered. If such an event occurs, an animal handler should ensure that females are handled separately and the specific procedures described below are applied. In all cases, the welfare of foetuses and dams during slaughter should be safeguarded.

1. Foetuses should not be removed from the uterus sooner than five minutes after the maternal neck or chest cut, to ensure absence of consciousness. A foetal heartbeat will usually still be present and foetal movements may occur at this stage, but these are only a cause for concern if the exposed foetus successfully breathes air.

2. If a live mature foetus is removed from the uterus, it should be prevented from inflating its lungs and breathing air (e.g. by clamping the trachea).

3. When uterine, placental or foetal tissues, including foetal blood, are not to be collected as part of the post-slaughter processing of pregnant animals, all foetuses should be left inside the unopened uterus until they are dead. When uterine, placental or foetal tissues are to be collected, where practical, foetuses should not be removed from the uterus until at least 15-20 minutes after the maternal neck or chest cut.

4. If there is any doubt about consciousness, the foetus should be killed with a captive bolt of appropriate size or a blow to the head with a suitable blunt instrument.

The above guidelines do not refer to foetal rescue. Foetal rescue, the practice of attempting to revive foetuses found alive at evisceration of the dam, should not be attempted during normal commercial slaughter as it may lead to serious welfare complications in the newborn animal. These include impaired brain function resulting from oxygen shortage before rescue is completed, compromised breathing and body heat production because of foetal immaturity, and an increased incidence of infections due to a lack of colostrums.
### Summary analysis of handling and restraining methods and the associated animal welfare issues

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### Article 3.7.5.6.

**Summary analysis of handling and restraining methods and the associated animal welfare issues**

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<td>Mechanical stunning methods Slaughter without stunning</td>
<td>Stress of resisting restraint; prolonged restraint, animal temperament; bruising. Keep restraint as short as possible</td>
<td>Competent animal handlers</td>
<td>Cattle, camelids</td>
</tr>
<tr>
<td></td>
<td>Tying of 3 or 4 legs</td>
<td>Mechanical stunning methods Slaughter without stunning</td>
<td>Stress of resisting restraint; prolonged restraint, animal temperament; bruising. Keep restraint as short as possible</td>
<td>Competent animal handlers</td>
<td>Sheep, goats, small camelids, pigs</td>
</tr>
</tbody>
</table>
Annex XXIII (contd)

Article 3.7.5.7.

Stunning methods

1. General considerations

The competence of the operators, and the appropriateness, and effectiveness of the method used for stunning and the maintenance of the equipment are the responsibility of the management of the slaughterhouse, and should be checked regularly by a Competent Authority.

Persons carrying out stunning should be properly trained and competent, and should ensure that:

a) the animal is adequately restrained;

b) animals in restraint are stunned as soon as possible;

c) the equipment used for stunning is maintained and operated properly in accordance with the manufacturer's recommendations, in particular with regard to the species and size of the animal;

d) the instrument is applied correctly;

e) stunned animals are bled out (slaughtered) as soon as possible;

f) animals should not be stunned when slaughter is likely to be delayed; and

g) backup stunning devices are available for immediate use if the primary method of stunning fails.

In addition, such persons should be able to recognise when an animal is not correctly stunned and should take appropriate action.

2. Mechanical stunning

A mechanical device should be applied usually to the front of the head and perpendicular to the bone surface. The following diagrams illustrate the proper application of the device for certain species.

Cattle

Figure source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).

The optimum position for cattle is at the intersection of two imaginary lines drawn from the rear of the eyes to the opposite horn buds.
**Pigs**

![Diagram of a pig head with an arrow indicating the shot placement.]

Figure source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).

The optimum position for pigs is on the midline just above eye level, with the shot directed down the line of the spinal cord.

**Sheep**

![Diagram of a sheep head with an arrow indicating the shot placement.]

Figure source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).

The optimum position for hornless sheep and goats is on the midline.
**Annex XXIII (contd)**

**Goats**

Figure Source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).

The optimum position for heavily horned sheep and horned goats is behind the poll, aiming towards the angle of the jaw.

**Horses**

Figure source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire AL4 8AN, United Kingdom (www.hsa.org.uk).

The optimum position for horses is at right angles to the frontal surface, well above the point where imaginary lines from eyes to ears cross.

Signs of correct stunning using a mechanical instrument are as follows:

a) the animal collapses immediately and does not attempt to stand up;
b) the body and muscles of the animal become tonic (rigid) immediately after the shot;
c) normal rhythmic breathing stops; and
d) the eyelid is open with the eyeball facing straight ahead and is not rotated.
3. **Electrical stunning**

a) **General considerations**

An electrical device should be applied to the animal in accordance with the following guidelines.

Electrodes should be designed, constructed, maintained and cleaned regularly to ensure that the flow of current is optimal and in accordance with manufacturing specifications. They should be placed so that they span the brain. The application of electrical currents which bypass the brain is unacceptable unless the animal has been stunned. The use of a single current leg-to-leg is unacceptable as a stunning method.

If, in addition, it is intended to cause cardiac arrest, the electrodes should either span the brain and immediately thereafter the heart, on the condition that it has been ascertained that the animal is adequately stunned, or span brain and heart simultaneously.

Electrical stunning equipment should not be applied on animals as a means of guidance, movement, restraint or immobilisation, and shall not deliver any shock to the animal before the actual stunning or killing.

Electrical stunning apparatus should be tested prior to application on animals using appropriate resistors or dummy loads to ensure the power output is adequate to stun animals.

The electrical stunning apparatus should incorporate a device that monitors and displays voltage (true RMS) and the applied current (true RMS) and that such devices are regularly calibrated at least annually.

Appropriate measures, such as removing excess wool or wetting the skin only at the point of contact, can be taken to minimise impedance of the skin and facilitate effective stunning.

The stunning apparatus required for electrical stunning should be provided with adequate power to achieve continuously the minimum current level recommended for stunning as indicated in the table below:

<table>
<thead>
<tr>
<th>Species</th>
<th>Minimum current levels for head-only stunning</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cattle</td>
<td>1.5 amps</td>
</tr>
<tr>
<td>Calves (bovines of less than 6 months of age)</td>
<td>1.0 amps</td>
</tr>
<tr>
<td>Pigs</td>
<td>1.25 amps</td>
</tr>
<tr>
<td>Sheep and goats</td>
<td>1.0 amps</td>
</tr>
<tr>
<td>Lambs</td>
<td>0.7 amps</td>
</tr>
<tr>
<td>Ostriches</td>
<td>0.4 amps</td>
</tr>
</tbody>
</table>

In all cases, the correct current level shall be attained within one second of the initiation of stun and maintained at least for three seconds and in accordance with the manufacturer's instructions.

b) **Electrical stunning of birds using a waterbath**

There should be no sharp bends or steep gradients in the shackle line and the shackle line should be as short as possible consistent with achieving acceptable line speeds, and ensuring that birds have settled by the time they reach the water bath. A breast comforter can be used effectively to reduce wing flapping and calm birds. The angle at which the shackle line approaches the entrance to the water bath, and the design of the entrance to the water bath, and the draining of excess 'live' water from the bath are all important considerations in ensuring birds are calm as they enter the bath, do not flap their wings, and do not receive pre-stun electric shocks.
In the case of birds suspended on a moving line, measures should be taken to ensure that the birds are not wing flapping at the entrance of the stunner. The birds should be secure in their shackle, but there should not be undue pressure on their shanks.

Waterbaths for poultry should be adequate in size and depth for the type of bird being slaughtered, and their height should be adjustable to allow for the head of each bird to be immersed. The electrode immersed in the bath should extend the full length of the waterbath. Birds should be immersed in the bath up to the base of their wings.

The waterbath should be designed and maintained in such a way that when the shackles pass over the water, they are in continuous contact with the earthed rubbing bar.

The control box for the waterbath stunner should incorporate an ammeter which displays the total current flowing through the birds.

The shackles-to-leg contact should be wetted preferably before the birds are inserted in the shackles. In order to improve electrical conductivity of the water it is recommended that salt be added in the waterbath as necessary. Additional salt should be added regularly as a solution to maintain suitable constant concentrations in the waterbath.

Using waterbaths, birds are stunned in groups and different birds will have different impedances. The voltage should be adjusted so that the total current is the required current per bird as shown in the table hereafter, multiplied by the number of birds in the waterbath at the same time. The following values have been found to be satisfactory when employing a 50 Hertz sinusoidal alternating current.

Birds should receive the current for at least 4 seconds.

<table>
<thead>
<tr>
<th>Species</th>
<th>Current (milliamperes per bird)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Broilers</td>
<td>100</td>
</tr>
<tr>
<td>Layers (spent hens)</td>
<td>100</td>
</tr>
<tr>
<td>Turkeys</td>
<td>150</td>
</tr>
<tr>
<td>Ducks and Geese</td>
<td>130</td>
</tr>
</tbody>
</table>

While a lower current may also be satisfactory, the current shall in any case be such as to ensure that unconsciousness occurs immediately and lasts until the bird has been killed by cardiac arrest or by bleeding. When higher electrical frequencies are used, higher currents may be required.

<table>
<thead>
<tr>
<th>Frequency (Hz)</th>
<th>Chickens</th>
<th>Turkeys</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt; 200 Hz</td>
<td>100 mA</td>
<td>250 mA</td>
</tr>
<tr>
<td>From 200 to 400 Hz</td>
<td>150 mA</td>
<td>400 mA</td>
</tr>
<tr>
<td>From 400 to 1500 Hz</td>
<td>200 mA</td>
<td>400 mA</td>
</tr>
</tbody>
</table>

Every effort shall be made to ensure that no conscious or live birds enter the scalding tank.

In the case of automatic systems, until fail-safe systems of stunning and bleeding have been introduced, a manual back-up system should be in place to ensure that any birds which have missed the waterbath stunner and/or the automatic neck-cutter are immediately stunned and/or killed immediately, and they are dead before entering scald tank.
To lessen the number of birds that have not been effectively stunned reaching neck cutters, steps should be taken to ensure that small birds do not go on the line amongst bigger birds and that these small birds are stunned separately.

4. **Gas stunning (under study)**

   a) **Stunning of pigs by exposure to carbon dioxide (CO$_2$)**

   The concentration of CO$_2$ for stunning should be preferably 90% by volume but in any case no less than 80% by volume. After entering the stunning chamber, the animals should be conveyed to the point of maximum concentration of the gas as rapidly as possible and be kept until they are dead or brought into a state of insensibility which lasts until death occurs due to bleeding. Ideally, pigs should be exposed to this concentration of CO$_2$ for 3 minutes. Sticking should occur as soon as possible after exit from the gas chamber.

   In any case, the concentration of the gas should be such that it minimises as far as possible all stress of the animal prior to loss of consciousness.

   The chamber in which animals are exposed to CO$_2$ and the equipment used for conveying them through it shall be designed, constructed and maintained in such a way as to avoid injury or unnecessary stress to the animals. The animal density within the chamber should be such to avoid stacking animals on top of each others.

   The conveyor and the chamber shall be adequately lit to allow the animals to see their surroundings and, if possible, each other.

   It should be possible to inspect the CO$_2$ chamber whilst it is in use, and to have access to the animals in emergency cases.

   The chamber shall be equipped to continuously measure and display register at the point of stunning the CO$_2$ concentration and the time of exposure, and to give a clearly visible and audible warning if the concentration of CO$_2$ falls below the required level.

   Emergency stunning equipment should be available at the point of exit from the stunning chamber and used on any pigs that do not appear to be dead or completely stunned.

   b) **Inert gas mixtures for stunning pigs**

   Inhalation of high concentrations of carbon dioxide is aversive and can be distressing to animals. Therefore, the use of non-aversive gas mixtures is being developed.

   Such gas mixtures include:

   i) a maximum of 2% by volume of oxygen in argon, nitrogen or other inert gases, or

   ii) a maximum of 30% by volume of carbon dioxide and a maximum of 2% by volume of oxygen in mixtures with carbon dioxide and argon, nitrogen or other inert gases.

   Exposure time to the gas mixtures should be sufficient to ensure that no pigs regain consciousness before death supervenes through bleeding or cardiac arrest is induced.
c) Gas stunning of poultry

The main objective of gas stunning is to avoid the pain and suffering associated with shackling conscious poultry under water bath stunning and killing systems. Therefore, gas stunning should be limited to birds contained in crates or on conveyors only. The gas mixture should be non-aversive to poultry.

Gas stunning of poultry in their transport containers will eliminate the need for live bird handling at the processing plant and all the problems associated with the electrical stunning. Gas stunning of poultry on a conveyor eliminates the problems associated with the electrical water bath stunning.

Live poultry should be conveyed into the gas mixtures either in transport crates or on conveyor belts.

The following gas procedures have been properly documented for chickens and turkeys but do not necessarily apply for other domestic birds. In any case the procedure should be designed as to ensure that all animals are properly stunned without unnecessary suffering.

i) Gas mixtures used for stunning poultry include:

- a minimum of 2 minutes exposure to 40% carbon dioxide, 30% oxygen and 30% nitrogen, followed by a minimum of one minute exposure to 80% carbon dioxide in air; or
- a minimum of 2 minutes exposure to any mixture of argon, nitrogen or other inert gases with atmospheric air and carbon dioxide, provided that the carbon dioxide concentration does not exceed 30% by volume and the residual oxygen concentration does not exceed 2% by volume; or
- a minimum of 2 minutes exposure to argon, nitrogen, other inert gases or any mixture of these gases in atmospheric air with a maximum of 2% residual oxygen by volume; or
- a minimum of 2 minutes exposure to a minimum of 55% carbon dioxide in air.

ii) Requirements for effective use are as follows:

- Compressed gases should be vaporised prior to administration into the chamber and should be at room temperature to prevent any thermal shock. Under no circumstances, should solid gases with freezing temperatures enter the chamber.
- Gas mixtures should be humidified.
- Appropriate gas concentrations of oxygen and carbon dioxide should be monitored and displayed continuously at the level of the birds inside the chamber to ensure that anoxia ensues.

Under no circumstances, should birds exposed to gas mixtures be allowed to regain consciousness. If necessary, the exposure time should be extended.

5. Bleeding

From the point of view of animal welfare, animals which are stunned with a reversible method should be bled without delay. Maximum stun-stick interval depends on the parameters of the stunning method applied, the species concerned and the bleeding method used (full cut or chest stick when possible). As a consequence, depending on those factors, the slaughterhouse operator should set up a maximum stun-stick interval that ensures that no animals recover consciousness during bleeding. In any case the following time limits should be applied.
## Stunning method

<table>
<thead>
<tr>
<th>Stunning method</th>
<th>Maximum delay for bleeding to be started</th>
</tr>
</thead>
<tbody>
<tr>
<td>Electrical methods and non penetrating captive bolt</td>
<td>20 seconds</td>
</tr>
<tr>
<td>CO₂</td>
<td>60 seconds (after leaving the chamber)</td>
</tr>
</tbody>
</table>

All animals should be bled out by incising both carotid arteries, or the vessels from which they arise (e.g. chest stick). However, when the stunning method used cardiac arrest, the incision of all of these vessels is not necessary from the point of view of animal welfare.

It should be possible for staff to observe, inspect and access the animals throughout the bleeding period. Any animal showing signs of recovering consciousness should be re-stunned.

After incision of the blood vessels, no scalding carcass treatment or dressing procedures should be performed on the animals for at least 30 seconds, or in any case until all brain-stem reflexes have ceased.
### Annex XXIII (contd)

**Summary analysis of stunning methods and the associated animal welfare issues**

<table>
<thead>
<tr>
<th>Method</th>
<th>Specific method</th>
<th>AW concerns/ implications</th>
<th>Key AW requirements applicable</th>
<th>Species</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mechanical</td>
<td>Free bullet</td>
<td>Inaccurate targeting and inappropriate ballistics</td>
<td>Operator competence, achieving outright kill with first shot</td>
<td>Cattle, calves, buffalo, deer, horses, pigs (boars and sows)</td>
<td>Personnel safety</td>
</tr>
<tr>
<td>Captive bolt -</td>
<td>Penetrating</td>
<td>Inaccurate targeting, velocity and diameter of bolt</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, buffalo, sheep, goats, deer, horses, pigs, camelids, ratites</td>
<td>(Unsuitable for specimen collection from TSE suspects). A back-up gun should be available in the event of an ineffective shot</td>
</tr>
<tr>
<td>Captive bolt -</td>
<td>Non-penetrating</td>
<td>Inaccurate targeting, velocity of bolt, potentially higher failure rate than penetrating captive bolt</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, sheep, goats, deer, pigs, camelids, ratites</td>
<td>Presently available devices are not recommended for young bulls and animals with thick skull. This method should only be used for cattle and sheep when alternative methods are not available.</td>
</tr>
<tr>
<td>Manual percussive</td>
<td>Blow</td>
<td>Inaccurate targeting; insufficient power; size of instrument</td>
<td>Competent animal handlers; restraint; accuracy. Not recommended for general use</td>
<td>Young and small mammals, ostriches and poultry</td>
<td>Mechanical devices potentially more reliable. Where manual percussive blow is used, unconsciousness should be achieved with single sharp blow delivered to central skull bones</td>
</tr>
<tr>
<td>Electrical</td>
<td>Split application:</td>
<td>Accidental pre-stun electric shocks; electrode positioning; application of a current to the body while animal conscious; inadequate current and voltage</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, sheep and pigs, ratites and poultry</td>
<td>Systems involving repeated application of head-only or head-to-leg with short current durations (&lt;1 second) in the first application should not be used.</td>
</tr>
<tr>
<td></td>
<td>1. across head then head to chest; 2. across head then across chest</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Single application:</td>
<td></td>
<td>Accidental pre-stun electric shocks; inadequate current and voltage; wrong electrode positioning; recovery of consciousness</td>
<td>Competent operation and maintenance of equipment; restraint; accuracy</td>
<td>Cattle, calves, sheep, goats, pigs, ratites, poultry</td>
<td></td>
</tr>
<tr>
<td></td>
<td>1. head only; 2. head to body; 3. head to leg</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Waterbath</td>
<td></td>
<td>Restraint, accidental pre-stun electric shocks; inadequate current and voltage; recovery of consciousness</td>
<td>Competent operation and maintenance of equipment</td>
<td>Poultry only</td>
<td></td>
</tr>
</tbody>
</table>
### Summary analysis of stunning methods and the associated animal welfare issues

<table>
<thead>
<tr>
<th>Method</th>
<th>Specific method</th>
<th>AW concerns/implications</th>
<th>Key AW requirements applicable</th>
<th>Species</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gaseous</td>
<td>CO₂ air/ O₂ mixture; CO₂ inert gas mixture</td>
<td>Aversiveness of high CO₂ concentrations, respiratory distress; inadequate exposure</td>
<td>Concentration; duration of exposure; design, maintenance and operation of equipment; stocking density management</td>
<td>Pigs, poultry</td>
<td></td>
</tr>
<tr>
<td>Inert gases</td>
<td></td>
<td>Recovery of consciousness</td>
<td>Concentration; duration of exposure; design, maintenance and operation of equipment; stocking density management</td>
<td>Pigs, poultry</td>
<td></td>
</tr>
<tr>
<td>Bleeding out by severance of blood vessels in the neck without stunning</td>
<td>Full frontal cutting across the throat</td>
<td>Failure to cut both common carotid arteries; occlusion of cut arteries; pain during and after the cut</td>
<td>High level of operator competency. A very sharp blade or knife, of sufficient length so that the point of the knife remains outside the incision during the cut; the point of the knife should not be used to make the incision. An incision which does not close over the knife during the throat cut.</td>
<td>Cattle, buffalo, horses, camels, sheep, goats, poultry, ratites</td>
<td>No further procedure should be carried out before the bleeding out is completed (i.e. at least 30–60 seconds for mammals). The practice to remove hypothetical blood clots just after the bleeding should be discouraged since this may increase animal suffering.</td>
</tr>
</tbody>
</table>
**Summary analysis of slaughter methods and the associated animal welfare issues**

<table>
<thead>
<tr>
<th>Slaughter methods</th>
<th>Specific method</th>
<th>AW concerns / implications</th>
<th>Key requirements</th>
<th>Species</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bleeding with prior stunning</td>
<td>Full frontal cutting across the throat</td>
<td>Failure to cut both common carotid arteries; occlusion of cut arteries; pain during and after the cut.</td>
<td>A very sharp blade or knife, of sufficient length so that the point of the knife remains outside the incision during the cut, the point of the knife should not be used to make the incision. An incision which does not close over the knife during the throat cut.</td>
<td>Cattle, buffalo, horses, camels, sheep, goats,</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Neck stab followed by forward cut</td>
<td>Neck stab followed by forward cut</td>
<td>Ineffective stunning; failure to cut both common carotid arteries; impaired blood flow; delay in cutting after reversible stunning</td>
<td>Prompt and accurate cutting</td>
<td>Camelids, sheep, goats, poultry, ratites</td>
<td></td>
</tr>
<tr>
<td>Neck stab alone</td>
<td>Neck stab alone</td>
<td>Ineffective stunning; failure to cut both common carotid arteries; impaired blood flow; delay in cutting after reversible stunning</td>
<td>Prompt and accurate cutting</td>
<td>Camelids, sheep, goats, poultry, ratites</td>
<td></td>
</tr>
<tr>
<td>Chest stick into major arteries or hollow-tube knife into heart</td>
<td>Chest stick into major arteries or hollow-tube knife into heart</td>
<td>Ineffective stunning; inadequate size of stick wound; inadequate length of sticking knife; delay in sticking after reversible stunning</td>
<td>Prompt and accurate sticking</td>
<td>Cattle, sheep, goats, pigs</td>
<td></td>
</tr>
<tr>
<td>Neck skin cut followed by severance of vessels in the neck</td>
<td>Neck skin cut followed by severance of vessels in the neck</td>
<td>Ineffective stunning; inadequate size of stick wound; inadequate length of sticking knife; delay in sticking after reversible stunning</td>
<td>Prompt and accurate cutting of vessels</td>
<td>Cattle</td>
<td></td>
</tr>
</tbody>
</table>
### Article 3.7.5.9.

**Summary analysis of slaughter methods and the associated animal welfare issues (contd)**

<table>
<thead>
<tr>
<th>Slaughter methods</th>
<th>Specific method</th>
<th>AW concerns / implications</th>
<th>Key requirements</th>
<th>Species</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bleeding with prior stunning (contd)</td>
<td>Automated mechanical cutting</td>
<td>Ineffective stunning; failure to cut and misplaced cuts. Recovery of consciousness following reversible stunning systems</td>
<td>Design, maintenance and operation of equipment; accuracy of cut; manual back-up</td>
<td>Poultry only</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Manual neck cut on one side</td>
<td>Ineffective stunning; recovery of consciousness following reversible stunning systems</td>
<td>Prior non-reversible stunning</td>
<td>Poultry only</td>
<td>N.B. slow induction of unconsciousness under slaughter without stunning</td>
</tr>
<tr>
<td></td>
<td>Oral cut</td>
<td>Ineffective stunning; recovery of consciousness following reversible stunning systems</td>
<td>Prior non-reversible stunning</td>
<td>Poultry only</td>
<td>N.B. slow induction of unconsciousness in non-stun systems</td>
</tr>
<tr>
<td>Other methods without stunning</td>
<td>Decapitation with a sharp knife</td>
<td>Pain due to loss of consciousness not being immediate</td>
<td></td>
<td>Sheep, goats, poultry</td>
<td>This method is only applicable to Jhatka slaughter</td>
</tr>
<tr>
<td></td>
<td>Manual neck dislocation and decapitation</td>
<td>Pain due to loss of consciousness not being immediate; difficult to achieve in large birds</td>
<td>Neck dislocation should be performed in one stretch to sever the spinal cord</td>
<td>Poultry only</td>
<td>Slaughter by neck dislocation should be performed in one stretch to sever the spinal cord. Acceptable only when slaughtering small numbers of small birds</td>
</tr>
<tr>
<td></td>
<td>Bleeding by evisceration</td>
<td></td>
<td>Induction of cardiac arrest</td>
<td>Quail</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Bleeding by neck cutting</td>
<td></td>
<td></td>
<td>Poultry</td>
<td></td>
</tr>
</tbody>
</table>
Annex XXIII (contd)

Article 3.7.5.10.

Methods, procedures or practices unacceptable on animal welfare grounds

1. The restraining methods which work through immobilisation by injury such as breaking legs, leg tendon cutting, and severing the spinal cord (e.g. using a puntilla or dagger) cause severe pain and stress in animals. Those methods are not acceptable in any species.

2. The use of the electrical stunning method with a single application leg to leg is ineffective and unacceptable in any species.

3. The slaughter method of brain stem severance by piercing through the eye socket or skull bone without prior stunning, is not acceptable in any species.

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- - - - - - - - - - - - - -

 text deleted
APPENDIX 3.7.6.

GUIDELINES FOR THE KILLING OF ANIMALS FOR DISEASE CONTROL PURPOSES

Article 3.7.6.1.

General principles

These guidelines are based on the premise that a decision to kill the animals has been made, and address the need to ensure the welfare of the animals until they are dead.

1. All personnel involved in the humane killing of animals should have the relevant skills and competencies. Competence may be gained through formal training and/or practical experience.

2. As necessary, operational procedures should be adapted to the specific circumstances operating on the premises and should address, apart from animal welfare, aesthetics of the method of euthanasia, cost of the method, operator safety, biosecurity and environmental aspects, aesthetics of the method of euthanasia and cost of the method.

3. Following the decision to kill the animals, killing should be carried out as quickly as possible and normal husbandry should be maintained until the animals are killed.

4. The handling and movement of animals should be minimised and when done, it should be done in accordance with the guidelines described below.

5. Animal restraint should be sufficient to facilitate effective killing, and in accordance with animal welfare and operator safety requirements; when restraint is required, killing should follow with minimal delay.

6. When animals are killed for disease control purposes, methods used should result in immediate death or immediate loss of consciousness lasting until death; when loss of consciousness is not immediate, induction of unconsciousness should be non-aversive and should not cause anxiety, pain, distress or suffering in the animals.

7. For animal welfare considerations, young animals should be killed before older animals; for biosecurity considerations, infected animals should be killed first, followed by in-contact animals, and then the remaining animals.

8. There should be continuous monitoring of the procedures by the Competent Authorities to ensure they are consistently effective with regard to animal welfare, operator safety and biosecurity.

9. When the operational procedures are concluded, there should be a written report describing the practices adopted and their effect on animal welfare, operator safety and biosecurity.

10. These general principles should also apply when animals need to be killed for other purposes such as after natural disasters or for culling animal populations.

Article 3.7.6.2.

Organisational structure

Disease control contingency plans should be in place at a national level and should contain details of management structure, disease control strategies and operational procedures; animal welfare considerations should be addressed within these disease control contingency plans. The plans should also include a strategy to ensure that an adequate number of personnel competent in the humane killing of animals is available. Local level plans should be based on national plans and be informed by local knowledge.
Disease control contingency plans should address the animal welfare issues that may result from animal movement controls.

The operational activities should be led by an official veterinarian who has the authority to appoint the personnel in the specialist teams and ensure that they adhere to the required animal welfare and biosecurity standards. When appointing the personnel, he/she should ensure that the personnel involved have the required competencies.

The official veterinarian should be responsible for all activities across one or more affected premises and should be supported by coordinators for planning (including communications), operations and logistics to facilitate efficient operations.

The official veterinarian should provide overall guidance to personnel and logistic support for operations on all affected premises to ensure consistency in adherence to the OIE animal welfare and animal health guidelines.

A specialist team, led by a team leader answerable to the official veterinarian, should be deployed to work on each affected premises. The team should consist of personnel with the competencies to conduct all required operations; in some situations, personnel may be required to fulfil more than one function. Each team should contain a veterinarian or have access to veterinary advice at all times.

In considering the animal welfare issues associated with the killing of animals, the key personnel, their responsibilities and competencies required are described in Article 3.7.6.3.

Article 3.7.6.3.

Responsibilities and competencies of the specialist team

1. Team leader
   a) Responsibilities:
      i) plan overall operations on an affected premises;
      ii) determine and address requirements for animal welfare, operator safety and biosecurity;
      iii) organise, brief and manage team of people to facilitate humane killing of the relevant animals on the premises in accordance with national regulations and these guidelines;
      iv) determine logistics required;
      v) monitor operations to ensure animal welfare, operator safety and biosecurity requirements are met;
      vi) report upwards on progress and problems;
      vii) provide a written report at the conclusion of the killing, describing the practices adopted and their effect on the animal welfare, operator safety and biosecurity outcomes.
   b) Competencies
      i) appreciation of normal animal husbandry practices;
      ii) appreciation of animal welfare and the underpinning behavioural, anatomical and physiological processes involved in the killing process;
      iii) skills to manage all activities on premises and deliver outcomes on time;
      iv) awareness of psychological effects on farmers, team members and general public;
      v) effective communication skills;
      vi) appreciation of the environmental impacts caused by their operation.
2. **Veterinarian**
   
a) **Responsibilities**
   
i) determine and supervise the implementation of the most appropriate killing method to ensure that animals are killed without avoidable pain and distress;
   
ii) determine and implement the additional requirements for animal welfare, including the order of killing;
   
iii) ensure that confirmation of animals deaths is carried out by competent persons at appropriate times after the killing procedure;
   
iv) minimise the risk of disease spread within and from the premises through the supervision of biosecurity procedures;
   
v) continuously monitor animal welfare and biosecurity procedures;
   
vi) in cooperation with the leader, prepare a written report at the conclusion of the killing, describing the practices adopted and their effect on animal welfare.
   
b) **Competencies**
   
i) ability to assess animal welfare, especially the effectiveness of stunning and killing, and to correct any deficiencies;
   
ii) ability to assess biosecurity risks.

3. **Animal handlers**
   
a) **Responsibilities**
   
i) review on-site facilities in terms of their appropriateness;
   
ii) design and construct temporary animal handling facilities, when required;
   
iii) move and restrain animals;
   
iv) continuously monitor animal welfare and biosecurity procedures.
   
b) **Competencies**
   
i) animal handling in emergency situations and in close confinement is required;
   
ii) an appreciation of biosecurity and containment principles.

4. **Animal killing personnel**
   
a) **Responsibilities**
   
Humane killing of the animals through effective stunning and killing should be ensured.
   
b) **Competencies**
   
i) when required by regulations, licensed to use necessary equipment;
   
ii) competent to use and maintain relevant equipment;
   
iii) competent to use techniques for the species involved;
   
iv) competent to assess effective stunning and killing.

5. **Carcass disposal personnel**
   
a) **Responsibilities**
   
An efficient carcass disposal (to ensure killing operations are not hindered) should be ensured.
b) Competencies

The personnel should be competent to use and maintain available equipment and apply techniques for the species involved.

6. Farmer/owner/manager

a) Responsibilities

i) assist when requested.

b) Competencies

i) specific knowledge of his/her animals and their environment.

Article 3.7.6.4.

Considerations in planning the humane killing of animals

Many activities will need to be conducted on affected premises, including the humane killing of animals. The team leader should develop a plan for humanely killing animals on the premises which should include consideration of:

1. minimising handling and movement of animals;
2. killing the animals on the affected premises; however, there may be circumstances where the animals may need to be moved to another location for killing; when the killing is conducted at an abattoir, the guidelines in Appendix 3.7.5. on slaughter of animals should be followed;
3. the species, number, age and size of animals to be killed, and the order of killing them;
4. methods of killing the animals, and their cost;
5. housing, husbandry, location of the animals, as well as accessibility of the farm;
6. the availability and effectiveness of equipment needed for killing of the animals, as well as the time necessary to kill the required number of animals using such methods;
7. the facilities available on the premises that will assist with the killing including any additional facilities that may need to be brought on and then removed from the premises;
8. biosecurity and environmental issues;
9. the health and safety of personnel conducting the killing;
10. any legal issues that may be involved, for example where restricted veterinary drugs or poisons may be used, or where the process may impact on the environment;
11. the presence of other nearby premises holding animals;
12. possibilities for removal, disposal and destruction of carcasses.

The plan should minimise the negative welfare impacts of the killing by taking into account the different phases of the procedures to be applied for killing (choice of the killing sites, killing methods, etc.) and the measures restricting the movements of the animals.

Competences and skills of the personnel handling and killing animals.

In designing a killing plan, it is essential that the method chosen be consistently reliable to ensure that all animals are humanely and quickly killed.
Article 3.7.6.5.

<table>
<thead>
<tr>
<th>Species</th>
<th>Age range</th>
<th>Procedure</th>
<th>Restraint necessary</th>
<th>Animal welfare concerns with inappropriate application</th>
<th>Article reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cattle</td>
<td>all</td>
<td>free bullet</td>
<td>no</td>
<td>non-lethal wounding</td>
<td>3.7.6.6.</td>
</tr>
<tr>
<td></td>
<td>all except neonates</td>
<td>captive bolt - penetrating, followed by pithing or bleeding</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>3.7.6.7.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>captive bolt - non-penetrating, followed by bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before killing</td>
<td>3.7.6.8.</td>
</tr>
<tr>
<td></td>
<td>calves only</td>
<td>electrical, two stage application</td>
<td>yes</td>
<td>pain associated with cardiac arrest after ineffective stunning</td>
<td>3.7.6.10.</td>
</tr>
<tr>
<td></td>
<td>calves only</td>
<td>electrical, single application (method 1)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>3.7.6.11.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>injection with barbiturates and other drugs</td>
<td>yes</td>
<td>non-lethal dose, pain associated with injection site</td>
<td>3.7.6.15.</td>
</tr>
<tr>
<td>Sheep and goats</td>
<td>all</td>
<td>free bullet</td>
<td>no</td>
<td>non-lethal wounding</td>
<td>3.7.6.6.</td>
</tr>
<tr>
<td></td>
<td>all except neonates</td>
<td>captive bolt - penetrating, followed by pithing or bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before death</td>
<td>3.7.6.7.</td>
</tr>
<tr>
<td></td>
<td>all except neonates</td>
<td>captive bolt - non-penetrating, followed by bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before death</td>
<td>3.7.6.8.</td>
</tr>
<tr>
<td></td>
<td>neonates</td>
<td>captive bolt - non-penetrating</td>
<td>yes</td>
<td>non-lethal wounding</td>
<td>3.7.6.8.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>electrical, two stage application</td>
<td>yes</td>
<td>pain associated with cardiac arrest after ineffective stunning</td>
<td>3.7.6.10.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>electrical, single application (Method 1)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>3.7.6.11.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>CO₂ / air mixture</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>3.7.6.12.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>nitrogen and/or inert gases mixed with CO₂</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>3.7.6.13.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>nitrogen and/or inert gases</td>
<td>yes</td>
<td>slow induction of unconsciousness</td>
<td>3.7.6.14.</td>
</tr>
</tbody>
</table>
### Table summarising killing methods described in Articles 3.7.6.6.-3.7.6.17. (Contd)

<table>
<thead>
<tr>
<th>Species</th>
<th>Age range</th>
<th>Procedure</th>
<th>Restraint Necessary</th>
<th>Animal welfare concerns with inappropriate application</th>
<th>Article reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sheep and goats (cont)</td>
<td>all</td>
<td>injection of barbiturates and other drugs</td>
<td>yes</td>
<td>non-lethal dose, pain associated with injection site</td>
<td>3.7.6.15.</td>
</tr>
<tr>
<td>Pigs</td>
<td>all, except neonates</td>
<td>free bullet</td>
<td>no</td>
<td>Non-lethal wounding</td>
<td>3.7.6.6.</td>
</tr>
<tr>
<td></td>
<td>all except neonates</td>
<td>captive bolt - penetrating, followed by pithing or bleeding</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before death</td>
<td>3.7.6.7.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>captive bolt - non-penetrating</td>
<td>yes</td>
<td>Non-lethal wounding</td>
<td>3.7.6.8.</td>
</tr>
<tr>
<td></td>
<td>all §</td>
<td>electrical, two stage application</td>
<td>yes</td>
<td>pain associated with cardiac arrest after ineffective stunning</td>
<td>3.7.6.10.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>electrical, single application (Method 1)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>3.7.6.11.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>CO₂/ air mixture</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>3.7.6.12.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>nitrogen and/ or inert gas mixed with CO₂</td>
<td>yes</td>
<td>slow induction of unconsciousness, aversiveness of induction</td>
<td>3.7.6.13.</td>
</tr>
<tr>
<td></td>
<td>neonates only</td>
<td>nitrogen and/ or inert gases</td>
<td>yes</td>
<td>slow induction of unconsciousness,</td>
<td>3.7.6.14.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>injection with barbiturates and other drugs</td>
<td>yes</td>
<td>non-lethal dose, pain associated with injection site</td>
<td>3.7.6.15.</td>
</tr>
<tr>
<td>Poultry</td>
<td>adults only</td>
<td>captive bolt - non-penetrating</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>3.7.6.8.</td>
</tr>
<tr>
<td></td>
<td>day-olds and eggs only</td>
<td>Maceration</td>
<td>no</td>
<td>non-lethal wounding, non- immediacy,</td>
<td>3.7.6.9.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>electrical single application (Method 2)</td>
<td>yes</td>
<td>ineffective stunning</td>
<td>3.7.6.11.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>electrical single application, followed by killing (Method 3)</td>
<td>yes</td>
<td>ineffective stunning, regaining of consciousness before death</td>
<td>3.7.6.11.</td>
</tr>
</tbody>
</table>
### Table summarising killing methods described in Articles 3.7.6.6.-3.7.6.17. (Contd)

<table>
<thead>
<tr>
<th>Species</th>
<th>Age range</th>
<th>Procedure</th>
<th>Restraint Necessary</th>
<th>Animal welfare concerns with inappropriate application</th>
<th>Article reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Poultry (cont)</td>
<td>all</td>
<td>CO₂ / air mixture Method 1 Method 2</td>
<td>yes/no</td>
<td>slow induction of unconsciousness, averseness of induction</td>
<td>3.7.6.12.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>nitrogen and/ or inert gas mixed with CO₂</td>
<td>yes</td>
<td>slow induction of unconsciousness, averseness of induction</td>
<td>3.7.6.13.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>nitrogen and/ or inert gases</td>
<td>yes</td>
<td>slow induction of unconsciousness</td>
<td>3.7.6.14.</td>
</tr>
<tr>
<td></td>
<td>all</td>
<td>injection of barbiturates and other drugs</td>
<td>yes</td>
<td>Non-lethal dose, pain associated with injection site</td>
<td>3.7.6.15.</td>
</tr>
<tr>
<td></td>
<td>adults only</td>
<td>addition of anaesthetics to feed or water, followed by an appropriate killing method</td>
<td>no</td>
<td>ineffective or slow induction of unconsciousness</td>
<td>3.7.6.16.</td>
</tr>
</tbody>
</table>

- The methods are described in the order of mechanical, electrical and gaseous, not in an order of desirability from an animal welfare viewpoint.

§ The only preclusion against the use of this method for neonates is the design of the stunning tongs that may not facilitate their application across such a small-sized head/body.

**Free bullet**

1. **Introduction**
   a) A free bullet is a projectile fired from a shotgun, rifle, handgun or purpose-made humane killer.
   b) The most commonly used firearms for close range use are:
      i) humane killers (specially manufactured/ adapted single-shot weapons);
      ii) shotguns (12, 16, 20, 28 bore and .410);
      iii) rifles (.22 rimfire);
      iv) handguns (various calibres from .32 to .45).
   c) The most commonly used firearms for long range use are rifles (.22, .243, .270 and .308).
   d) A free bullet used from long range should be aimed to penetrate the skull or soft tissue at the top of the neck of the animal (high neck shot), to cause irreversible concussion and death and should only be used by properly trained and competent marksmen.

2. **Requirements for effective use**
   a) The marksman should take account of human safety in the area in which he/ she is operating. Appropriate vision and hearing protective devices should be worn by all personnel involved.
b) The marksman should ensure that the animal is not moving and in the correct position to enable accurate targeting and the range should be as short as possible (5 – 50 cm for a shotgun) but the barrel should not be in contact with the head of the animal.

c) The correct cartridge, calibre and type of bullet for the different species age and size should be used. Ideally the ammunition should expand upon impact and dissipate its energy within the cranium.

d) Shot animals should be checked to ensure the absence of brain stem reflexes.

**Figure 1.** The optimum shooting position for cattle is at the intersection of two imaginary lines drawn from the rear of the eyes to the opposite horn buds.

![Figure 1](https://www.hsa.org.uk)

Figure Source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire, AL4 8AN, United Kingdom (www.hsa.org.uk).

**Figure 2** The optimum position for hornless sheep and goats is on the midline, with the shot aiming at the angle of the jaw.

![Figure 2](https://www.hsa.org.uk)

Figure Source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire, AL4 8AN, United Kingdom (www.hsa.org.uk).
Figure 3. The optimum shooting position for heavily horned sheep and horned goats is behind the poll aiming towards the angle of the jaw.

Figure Source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire, AL4 8AN, United Kingdom (www.hsa.org.uk).

Figure 4. The optimum shooting position for pigs is just above eye level, with the shot directed down the line of the spinal cord.

Figure Source: Humane Slaughter Association (2005) Guidance Notes No. 3: Humane Killing of Livestock Using Firearms. Published by the Humane Slaughter Association, The Old School, Brewhouse Hill, Wheathampstead, Hertfordshire, AL4 8AN, United Kingdom (www.hsa.org.uk).

3. Advantages
   a) Used properly, a free bullet provides a quick and effective method for killing.
   b) It requires minimal or no restraint and can be used to kill from a distance by properly trained and competent marksmen.
   c) It is suitable for killing agitated animals in open spaces.

4. Disadvantages
   a) The method is potentially dangerous to humans and other animals in the area.
b) It has the potential for non-lethal wounding.

c) Destruction of brain tissue may preclude diagnosis of some diseases.

d) Leakage of bodily fluids may present a biosecurity risk.

e) Legal requirements may preclude or restrict use.

f) There is a limited availability of competent personnel.

5. Conclusions

The method is suitable for cattle, sheep, goats and pigs, including large animals in open spaces.

Annex XXIII (contd)

Penetrating captive bolt

1. Introduction

A penetrating captive bolt is fired from a gun powered by either compressed air or a blank cartridge. There is no free projectile.

The captive bolt should be aimed on the skull in a position to penetrate the cortex and mid-brain of the animal. The impact of the bolt on the skull produces unconsciousness. Physical damage to the brain caused by penetration of the bolt may result in death, however pithing or bleeding should be performed as soon as possible after the shot to ensure the death of the animal.

2. Requirements for effective use

a) For cartridge powered and compressed air guns, the bolt velocity and the length of the bolt should be appropriate to the species and type of animal, in accordance with the recommendations of the manufacturer.

b) Captive bolt guns should be frequently cleaned and maintained in good working condition.

c) More than one gun may be necessary to avoid overheating and a back-up gun should be available in the event of an ineffective shot.

d) Animals should be restrained; at a minimum they should be penned for cartridge powered guns and in a race for compressed air guns.

e) The operator should ensure that the head of the animal is accessible.

f) The operator should fire the captive bolt at right angles to the skull in the optimal position (see figures 1, 3 & 4. The optimum shooting position for hornless sheep is on the highest point of the head, on the midline and aim towards the angle of the jaw).

h) To ensure the death of the animal, pithing or bleeding should be performed as soon as possible after stunning.

h) Animals should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

3. Advantages

a) Mobility of cartridge powered equipment reduces the need to move animals.

b) The method induces an immediate onset of a sustained period of unconsciousness.
4. Disadvantages

a) Poor gun maintenance and misfiring, and inaccurate gun positioning and orientation may result in poor animal welfare.

b) Post stun convulsions may make pithing difficult and hazardous.

c) The method is difficult to apply in agitated animals.

d) Repeated use of a cartridge powered gun may result in over-heating.

e) Leakage of bodily fluids may present a biosecurity risk.

f) Destruction of brain tissue may preclude diagnosis of some diseases.

5. Conclusions

The method is suitable for cattle, sheep, goats and pigs (except neonates), when followed by pithing or bleeding.

Captive bolt - non-penetrating

1. Introduction

A non-penetrating captive bolt is fired from a gun powered by either compressed air or a blank cartridge. There is no free projectile.

The gun should be placed on the front of the skull to deliver a percussive blow which produces unconsciousness in cattle (adults only), sheep, goats and pigs, and death in poultry and neonate sheep, goats and pigs up to a maximum live weight of 10 kg. Bleeding should be performed as soon as possible after the blow to ensure the death of the animal.

2. Requirements for effective use

a) For cartridge powered and compressed air guns, the bolt velocity should be appropriate to the species and type of animal, in accordance with the recommendations of the manufacturer.

b) Captive bolt guns should be frequently cleaned and maintained in good working condition.

c) More than one gun may be necessary to avoid overheating and a back-up gun should be available in the event of an ineffective shot.

d) Animals should be restrained; at a minimum mammals should be penned for cartridge powered guns and in a race for compressed air guns; birds should be restrained in cones, shackles, crushes or by hand.

e) The operator should ensure that the head of the animal is accessible.

f) The operator should fire the captive bolt at right angles to the skull in the optimal position (figures 1-4).
g) To ensure death in non-neonate mammals, bleeding should be performed as soon as possible after stunning.

h) Animals should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

3. Advantages

a) The method induces an immediate onset of unconsciousness, and death in birds and neonate mammals.

b) Mobility of equipment reduces the need to move animals.

4. Disadvantages

a) As consciousness can be regained quickly in non-neonate mammals, they should be bled as soon as possible after stunning.

b) Laying hens in cages have to be removed from their cages and most birds have to be restrained.

c) Poor gun maintenance and misfiring, and inaccurate gun positioning and orientation may result in poor animal welfare.

d) Post stun convulsions may make bleeding difficult and hazardous.

e) Difficult to apply in agitated animals; such animals may be sedated in advance of the killing procedure.

f) Repeated use of a cartridge powered gun may result in overheating.

g) Bleeding may present a biosecurity risk.

5. Conclusions

a) The method is suitable for poultry, and neonate sheep, goats and pigs up to a maximum weight of 10 kg.

Article 3.7.6.9.

Maceration

1. Introduction

Maceration, utilising a mechanical apparatus with rotating blades or projections, causes immediate fragmentation and death in day-old poultry and embryonated eggs.

2. Requirements

a) Maceration requires specialised equipment which should be kept in excellent working order.
b) The rate of introducing the birds should not allow the equipment to jam, birds to rebound from the blades or the birds to suffocate before they are macerated.

3. **Advantages**
   a) Procedure results in immediate death.
   b) Large numbers can be killed quickly.

4. **Disadvantages**
   a) Specialised equipment is required.
   b) Macerated tissues may present a biosecurity or human health risks.
   c) The cleaning of the equipment can be a source of contamination.

5. **Conclusion**
   The method is suitable for killing day-old poultry and embryonated eggs.

**Electrical - two-stage application**

1. **Introduction**
   A two stage application of electric current comprises firstly an application of current to the head by scissor-type tongs, immediately followed by an application of the tongs across the chest in a position that spans the heart.

   The application of sufficient electric current to the head will induce ‘tonic/clonic’ epilepsy and unconsciousness. Once the animal is unconscious, the second stage will induce ventricular fibrillation (cardiac arrest) resulting in death. The second stage (the application of low frequency current across the chest) should only be applied to unconscious animals to prevent unacceptable levels of pain.

2. **Requirements for effective use**
   a) The stunner control device should generate a low frequency (AC sine wave 50 Hz) current with a minimum voltage and current as set out in the following table:

<table>
<thead>
<tr>
<th>Animal</th>
<th>Minimum voltage (V)</th>
<th>Minimum current (A)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cattle</td>
<td>220</td>
<td>1.5</td>
</tr>
<tr>
<td>Sheep</td>
<td>220</td>
<td>1.0</td>
</tr>
<tr>
<td>Pigs &gt; 6 weeks</td>
<td>220</td>
<td>1.3</td>
</tr>
<tr>
<td>Pigs &lt; 6 weeks</td>
<td>125</td>
<td>0.5</td>
</tr>
</tbody>
</table>

   b) Appropriate protective clothing (including rubber gloves and boots) should be worn.
c) Animals should be restrained, at a minimum free-standing in a pen, close to an electrical supply.

d) Two team members are required, the first to apply the electrodes and the second to manipulate the position of the animal to allow the second application to be made.

e) A stunning current should be applied via scissor-type stunning tongs in a position that spans the brain for a minimum of \(3-10\) seconds; immediately following the application to the head, the electrodes should be transferred to a position that spans the heart and the electrodes applied for a minimum of 3 seconds.

f) Electrodes should be cleaned regularly and after use, to enable optimum electrical contact to be maintained.

g) Animals should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

h) Electrodes should be applied firmly for the intended duration of time and pressure not released until the stun is complete.

3. Advantages

a) The application of the second stage minimises post-stun convulsions and therefore the method is particularly effective with pigs.

b) Non-invasive technique minimises biosecurity risk.

4. Disadvantages

a) The method requires a reliable supply of electricity.

b) The electrodes must be applied and maintained in the correct positions to produce an effective stun and kill.

c) Most stunner control devices utilise low voltage impedance sensing as an electronic switch prior to the application of high voltages; in unshorn sheep, contact impedance may be too high to switch on the required high voltage (especially during stage two).

d) The procedure may be physically demanding, leading to operator fatigue and poor electrode placement.

5. Conclusion

The method is suitable for calves, sheep and goats, and especially for pigs (over one week of age).

Article 3.7.6.11.

**Electrical - single application**

1. **Method 1**

Method 1 comprises the single application of sufficient electrical current to the head and back, to simultaneously stun the animal and fibrillate the heart. Provided sufficient current is applied in a position that spans both the brain and heart, the animal will not recover consciousness.
a) Requirements for effective use

i) The stunner control device should generate a low frequency (30 – 60 Hz) current with a minimum voltage of 250 volts true RMS under load.

ii) Appropriate protective clothing (including rubber gloves and boots) should be worn.

iii) Animals should be individually and mechanically restrained close to an electrical supply as the maintenance of physical contact between the stunning electrodes and the animal is necessary for effective use.

iv) The rear electrode should be applied to the back, above or behind the heart, and then the front electrode in a position that is forward of the eyes, with current applied for a minimum of \(3 \times 10^2\) seconds.

v) Electrodes should be cleaned regularly between animals and after use, to enable optimum electrical contact to be maintained.

vi) Water or saline may be necessary to improve electrical contact with sheep.

vii) An effective stun and kill should be verified by the absence of brain stem reflexes.

b) Advantages

i) Method 1 stuns and kills simultaneously.

ii) It minimises post-stun convulsions and therefore is particularly effective with pigs.

iii) A single team member only is required for the application.

iv) Non-invasive technique minimises biosecurity risk.

c) Disadvantages

i) Method 1 requires individual mechanical animal restraint.

ii) The electrodes must be applied and maintained in the correct positions to produce an effective stun and kill.

iii) Method 1 requires a reliable supply of electricity.

d) Conclusion

Method 1 is suitable for calves, sheep, goats, and pigs (over 1 week of age).

2. Method 2

Method 2 stuns and kills by drawing inverted and shackled poultry through an electrified waterbath stunner. Electrical contact is made between the ‘live’ water and earthed shackle and, when sufficient current is applied, poultry will be simultaneously stunned and killed.

a) Requirements for effective use

i) A mobile waterbath stunner and a short loop of processing line are required.

ii) A low frequency (50-60 Hz) current applied for a minimum of 3 seconds is necessary to stun and kill the birds.

iii) Poultry need to be manually removed from their cage, house or yard, inverted and shackled onto a line which conveys them through a waterbath stunner with their heads fully immersed.
Annex XXIII (contd)

iv) The required minimum currents to stun and kill dry birds are:
   - Quail - 100 mA/bird
   - Chickens - 160 mA/bird
   - Ducks & Geese - 200 mA/bird
   - Turkeys - 250 mA/bird.
   A higher current is required for wet birds.

v) An effective stun and kill should be verified by the absence of brain stem reflexes.

b) Advantages
   i) Method 2 stuns and kills simultaneously.
   ii) It is capable of processing large numbers of birds reliably and effectively.
   iii) This non-invasive technique minimises biosecurity risk.

c) Disadvantages
   i) Method 2 requires a reliable supply of electricity.
   ii) Handling, inversion and shackling of birds are required.

d) Conclusion
   Method 2 is suitable for large numbers of poultry.

3. Method 3

Method 3 comprises the single application of sufficient electrical current to the head of poultry in a position that spans the brain, causing unconsciousness; this is followed by a killing method (Article 3.7.6.17.).

a) Requirements for effective use
   i) The stunner control device should generate sufficient current (more than 600 mA/duck, more than 300 mA/bird) to stun.
   ii) Appropriate protective clothing (including rubber gloves and boots) should be worn.
   iii) Birds should be restrained, at a minimum manually, close to an electrical supply.
   iv) A stunning current should be applied in a position that spans the brain for a minimum of 3-7 seconds; immediately following this application, the birds should be killed (Article 3.7.6.17.).
   v) Electrodes should be cleaned regularly and after use, to enable optimum electrical contact to be maintained.
   vi) Birds should be monitored continuously after stunning until death to ensure the absence of brain stem reflexes.

b) Advantages
   Non-invasive technique (when combined with cervical dislocation) minimises biosecurity risk.
c) Disadvantages

i) Method 3 requires a reliable supply of electricity and is not suitable for large-scale operations.

ii) The electrodes must be applied and maintained in the correct position to produce an effective stun.

iii) Birds must be individually restrained.

iv) It must be followed by a killing method.

d) Conclusion

Method 3 is suitable for small numbers of poultry.

Article 3.7.6.12.
(under study)

CO₂ / air mixture

1. Introduction

Controlled atmosphere killing is performed by exposing animals to a predetermined gas mixture, either by placing them in a gas-filled container or apparatus (Method 1) or by the gas being introduced into a poultry house (Method 2). Method 2 should be used whenever possible, as it eliminates welfare issues resulting from the need to manually remove live birds.

Inhalation of carbon dioxide (CO₂) induces respiratory and metabolic acidosis and hence reduces the pH of cerebrospinal fluid (CSF) and neurones thereby causing unconsciousness and, after prolonged exposure, death.

2. Method 1

The animals are placed in a gas-filled container or apparatus.

a) Requirements for effective use in a container or apparatus

i) Containers or apparatus should allow the required gas concentration to be maintained and accurately measured.

ii) When animals are exposed to the gas individually or in small groups in a container or apparatus, the equipment used should be designed, constructed, and maintained in such a way as to avoid injury to the animals and allow them to be observed.

iii) Animals can also be introduced to low concentrations (as low concentrations are not aversive) and the concentration could be increased afterwards and the animals then held in the higher concentration until death is confirmed.

iv) Team members should ensure that there is sufficient time allowed for each batch of animals to die before subsequent ones are introduced into the container or apparatus.

iv) Containers or apparatus should not be overcrowded and measures are needed to avoid animals suffocating by climbing on top of each other.
b) Advantages
   i) CO₂ is readily available.
   ii) Application methods are simple.

c) Disadvantages
   i) The need for properly designed container or apparatus.
   ii) The aversive nature of high CO₂ concentrations.
   iii) No immediate loss of consciousness.
   iv) The risk of suffocation due to overcrowding.
   v) Difficulty in verifying death while the animals are in the container or apparatus.

d) Conclusion
   Method 1 is suitable for use in poultry and neonatal sheep, goats and pigs.

3. Method 2

   The gas is introduced into a poultry house.

   a) Requirements for effective use in a poultry house
      i) Prior to introduction of the CO₂ the poultry house should be appropriately sealed to allow
         control over the gas concentration.
      ii) The house should be gradually filled with CO₂ so that all birds are exposed to a
         concentration of >40% until they are dead; a vaporiser may be required to prevent
         freezing.
      iii) Devices should be used to accurately measure the gas concentration at the maximum
         height accommodation of birds.

   b) Advantages
      i) Applying gas to birds in situ eliminates the need to manually remove live birds.
      ii) CO₂ is readily available.
      iii) Gradual raising of CO₂ concentration minimises the aversiveness of the induction of
           unconsciousness.

c) Disadvantages
   i) It is difficult to determine volume of gas required to achieve adequate concentrations of
      CO₂ in some poultry houses.
   ii) It is difficult to verify death while the birds are in the poultry house.

d) Conclusion
   Method 2 is suitable for use in poultry in closed-environment sheds.
Nitrogen and/or inert gas mixed with CO₂

1. Introduction

CO₂ may be mixed in various proportions with nitrogen or an inert gas (e.g. argon), and the inhalation of such mixtures leads to hypercapnic-hypoxia and death when the oxygen concentration by volume is = 2%. This method involves the introduction of animals into a container or apparatus containing the gases. Such mixtures do not induce immediate loss of consciousness, therefore the aversiveness of various gas mixtures containing high concentrations of CO₂ and the respiratory distress occurring during the induction phase, are important animal welfare considerations.

Pigs and poultry appear not to find low concentrations of CO₂ strongly aversive, and a mixture of nitrogen or argon with =30% CO₂ by volume and =2% O₂ by volume can be used for killing poultry and neonatal sheep, goats and pigs.

2. Requirements for effective use

a) Containers or apparatus should allow the required gas concentrations to be maintained, and the O₂ and CO₂ concentrations accurately measured during the killing procedure.

b) When animals are exposed to the gases individually or in small groups in a container or apparatus, the equipment used should be designed, constructed, and maintained in such a way as to avoid injury to the animals and allow them to be observed.

c) Animals should be introduced into the container or apparatus after it has been filled with the required gas concentrations (with =2% O₂), and held in this atmosphere until death is confirmed.

d) Team members should ensure that there is sufficient time allowed for each batch of animals to die before subsequent ones are introduced into the container or apparatus.

e) Containers or apparatus should not be overcrowded and measures are needed to avoid animals suffocating by climbing on top of each other.

3. Advantages

Low concentrations of CO₂ cause little aversiveness and, in combination with nitrogen or an inert gas, produces a fast induction of unconsciousness.

4. Disadvantages

a) A properly designed container or apparatus is needed.

b) It is difficult to verify death while the animals are in the container or apparatus.

c) There is no immediate loss of consciousness.

d) Exposure times required to kill are considerable.

5. Conclusion

The method is suitable for poultry and neonatal sheep, goats and pigs.
Nitrogen and/or inert gases

1. **Introduction**

This method involves the introduction of animals into a container or apparatus containing nitrogen or an inert gas such as argon. The controlled atmosphere produced leads to unconsciousness and death from hypoxia.

Research has shown that hypoxia is not aversive to pigs and poultry, and it does not induce any signs of respiratory distress prior to loss of consciousness.

2. **Requirements for effective use**

   a) Containers or apparatus should allow the required gas concentrations to be maintained, and the \( O_2 \) concentration accurately measured.

   b) When animals are exposed to the gases individually or in small groups in a container or apparatus, the equipment used should be designed, constructed, and maintained in such a way as to avoid injury to the animals and allow them to be observed.

   c) Animals should be introduced into the container or apparatus after it has been filled with the required gas concentrations (with \( \approx 2\% \ O_2 \)), and held in this atmosphere until death is confirmed.

   d) Team members should ensure that there is sufficient time allowed for each batch of animals to die before subsequent ones are introduced into the container or apparatus.

   e) Containers or apparatus should not be overcrowded and measures are needed to avoid animals suffocating by climbing on top of each other.

3. **Advantages**

   Animals are unable to detect nitrogen or inert gases, and the induction of hypoxia by this method is not aversive to animals.

4. **Disadvantages**

   a) A properly designed container or apparatus is needed.

   b) It is difficult to verify death while the animals are in the container or apparatus.

   c) There is no immediate loss of consciousness.

   d) Exposure times required to kill are considerable.

5. **Conclusion**

   The method is suitable for poultry and neonatal sheep, goats and pigs.

Lethal injection

1. **Introduction**

   A lethal injection using high doses of anaesthetic and sedative drugs causes CNS depression, unconsciousness and death. In practice, barbiturates in combination with other drugs are commonly used.
2. **Requirements for effective use**
   a) Doses and routes of administration that cause rapid loss of consciousness followed by death should be used.
   b) Prior sedation may be necessary for some animals.
   c) Intravenous administration is preferred, but intraperitoneal or intramuscular administration may be appropriate, especially if the agent is non-irritating.
   d) Animals should be restrained to allow effective administration.
   e) Animals should be monitored to ensure the absence of brain stem reflexes.

3. **Advantages**
   a) The method can be used in all species.
   b) Death can be induced smoothly.

4. **Disadvantages**
   a) Restraint and/or sedation may be necessary prior to injection.
   b) Some combinations of drug type and route of administration may be painful, and should only be used in unconscious animals.
   c) Legal requirements and skill/training required may restrict use to veterinarians.
   d) Contaminated carcasses may present a risk to other wild or domestic animals.

5. **Conclusion**
   The method is suitable for killing small numbers of cattle, sheep, goats, pigs and poultry.

   Article 3.7.6.16.

**Addition of anaesthetics to feed or water**

1. **Introduction**
   An anaesthetic agent which can be mixed with poultry feed or water may be used to kill poultry in houses. Poultry which are only anaesthetised need to be killed by another method such as cervical dislocation.

2. **Requirements for effective use**
   a) Sufficient quantities of anaesthetic need to be ingested rapidly for effective response.
   b) Intake of sufficient quantities is facilitated if the birds are fasted or water is withheld.
   c) Must be followed by killing (see Article 3.7.6.17.) if birds are anaesthetised only.
Annex XXIII (contd)

3. **Advantages**
   a) Handling is not required until birds are anaesthetised.
   b) There may be biosecurity advantages in the case of large numbers of diseased birds.

4. **Disadvantages**
   a) Non-target animals may accidentally access the medicated feed or water when provided in an open environment.
   b) Dose taken is unable to be regulated and variable results may be obtained.
   c) Animals may reject adulterated feed or water due to illness or adverse flavour.
   d) The method may need to be followed by killing.
   e) Care is essential in the preparation and provision of treated feed or water, and in the disposal of uneaten treated feed/water and contaminated carcasses.

5. **Conclusion**
   The method is suitable for killing large numbers of poultry in houses.

Article 3.7.6.17.

**Cervical dislocation and decapitation**

1. **Cervical dislocation (manual and mechanical)**
   a) **Introduction**

   Unconscious poultry may be killed by either manual cervical dislocation (stretching) or mechanical neck crushing with a pair of pliers. Both methods result in death from cerebral anoxia due to cessation of breathing and/or blood supply to the brain.

   However, conscious birds of less than 3 kilograms in case of small numbers of birds where other methods are not available or impracticable, may be killed using cervical dislocation in a way that the blood vessels of the neck are severed and death is instantaneous.

   b) **Requirements for effective use**

   i) Killing should be performed either by manually or mechanically stretching the neck to sever the spinal cord or by using mechanical pliers to crush the cervical vertebrae with consequent major damage to the spinal cord.

   ii) Consistent results require strength and skill so team members should be rested regularly to ensure consistently reliable results.

   iii) Birds should be monitored continuously until death to ensure the absence of brain stem reflexes.

   c) **Advantages**

   i) It is a non-invasive killing method.

   ii) It can be performed manually on small birds.
d) Disadvantages
   i) Operator fatigue.
   ii) The method is more difficult in larger birds. Its use should be avoided in any case for birds over 3 kg of live weight.
   iii) Requires trained personnel to perform humanely.

2. Decapitation
   a) Introduction
      Decapitation results in death by cerebral ischaemia using a guillotine or knife.
   b) Requirements for effective use
      The required equipment should be kept in good working order.
   c) Advantages
      The technique is effective and does not require monitoring.
   d) Disadvantages
      The working area is contaminated with body fluids, which increases biosecurity risks.

Article 3.7.6.18.

Pithing and bleeding

1. Pithing
   a) Introduction
      Pithing is a method of killing animals which have been stunned by a penetrating captive bolt, without immediate death. Pithing results in the physical destruction of the brain and upper regions of the spinal cord, through the insertion of a rod or cane through the bolt hole.
   b) Requirements for effective use
      i) Pithing cane or rod is required.
      ii) An access to the head of the animal and to the brain through the skull is required.
      iii) Animals should be monitored continuously until death to ensure the absence of brain stem reflexes.
   c) Advantages
      The technique is effective in producing immediate death.
   d) Disadvantages
      i) A delayed and/or ineffective pithing due to convulsions may occur.
      ii) The working area is contaminated with body fluids, which increases biosecurity risks.
2. **Bleeding**

   a) **Introduction**

   Bleeding is a method of killing animals through the severance of the major blood vessels in the neck or chest that results in a rapid fall in blood pressure, leading to cerebral ischaemia and death.

   **Bleeding out should be completed and any incision made should ensure the complete severance off both carotid arteries, or the vessels from which they arise (e.g. chest stick).**

   b) **Requirements for effective use**

   i) A sharp knife is required.

   ii) An access to the neck or chest of the animal is required.

   iii) Animals should be monitored continuously until death to ensure the absence of brain stem reflexes.

   c) **Advantages**

   The technique is effective in producing death after an effective stunning method which does not permit pithing.

   d) **Disadvantages**

   a) A delayed and/or ineffective bleeding due to convulsions may occur.

   b) The working area is contaminated with body fluids, which increases biosecurity risks.
Annex XXIII (contd)

DRAFT GUIDELINES ON DOG POPULATION CONTROL

Preamble: Stray and feral dogs pose serious human health, socio-economic, political and animal welfare problems in many countries of the world. Many of these are developing countries and others fall in the least developed category. Whilst acknowledging human health is a priority including the prevention of zoonotic diseases notably rabies, the OIE recognises the importance of controlling dog populations without causing unnecessary or avoidable animal suffering. Veterinary Services should play a lead role in preventing zoonotic diseases and ensuring animal welfare and should be involved in dog population control.

Guiding principles

The following guidelines are based on those laid down in Section 3.7. of the Terrestrial Animal Health Code. Some additional principles are relevant to these guidelines:

1. The promotion of responsible dog ownership can significantly reduce the numbers of stray dogs and the incidence of zoonotic diseases

2. Because dog ecology is linked with human activities, management of dog populations has to be accompanied by changes in human behaviour to be effective.

Article 1

Definitions

a) Stray Dog: any dog not under direct control or not prevented from roaming

Types of stray dog

a) free roaming owned dog not under direct control or restriction at a particular time;

b) free roaming dog with no owner;

c) feral dog: domestic dog that has reverted to the wild state and is no longer directly dependent upon humans for successful reproduction.

b) Owned Dog: dog with a person that claims responsibility is responsible for this animal.

c) Person: This can include more than one individual, and could comprise family/household members or an organisation.

d) Responsible Ownership: The situation whereby a person (as defined above) accepts and commits to perform various duties focused on the satisfaction of the psychological, environmental and physical needs of a dog (or other pet) and to the prevention of risks (aggression, disease transmission or causing injuries) that the pet dog may cause to the community, other animals or the environment.

e) Euthanasia: the act of inducing death in a humane manner.

f) Competent Authority: means the Veterinary Services, or other Authority of a Member Country, having the responsibility and competence and for ensuring or supervising the implementation of animal health measures or other standards in the Terrestrial Code.
Annex XXIII (contd)

g) Dog population control programme: A programme with the objective of reducing the number of stray dogs. A programme with the aim of reducing a dog population to a particular level and/or maintaining it at that level and/or managing it in order to meet a predetermined objective (see Article 2).

h) Carrying capacity: is the upper limit of the dog population density that could be supported by the habitat based on the availability of resources (food, water, shelter), and human acceptance.

Article 2

Dog population control programme optional objectives

The objectives of a programme to control the dog population may include the following:

1. Improve health and welfare of owned and stray dog population
2. Reduce numbers of stray dogs
3. Create assist in the creation and maintenance of a rabies immune dog population
4. Promote responsible ownership;
5. Reduce the risk of zoonotic diseases other than rabies;
6. Manage other risks to human health e.g. parasites;
7. Prevent harm to the environment
8. Prevent illegal trade and trafficking.

Article 3

Responsibilities and competencies

1. Veterinary Authority Administration

The Veterinary Authority Administration is responsible for the implementation of animal health legislation and for controlling outbreaks of notifiable animal diseases such as foot and mouth disease and avian influenza. Control of endemic zoonotic diseases such as rabies and parasitic infections (e.g. Echinococcus spp.) would require technical advice from the Veterinary Authority Administration, as animal health and some aspects of public health are within this Authority's competence but organising and/or supervising dog control schemes is frequently the responsibility of government agencies other than the Veterinary Authority Administration.

In many countries the Veterinary Authority Administration is in the Ministry of Agriculture.

2. Other government agencies

The responsibilities of other government agencies will depend on the disease risk being managed and the objective/nature of the dog population control measures employed.
Annex XXIII (contd)

The Ministry or other Agency responsible for Public Health would normally play a leadership role and may have legislative authority in dealing with zoonotic diseases. Control of stray dogs, in regard to other human health risks (e.g. stray dogs on roads; dog attacks within communities) may fall within the responsibility of the Public Health Agency but is more likely to be the responsibility of police or other agencies for public safety/security operating at the State/ Provincial or municipal level.

Environment Protection Agencies (normally within a National or State/Provincial Ministry for the Environment) may take responsibility for the controlling problems associated with stray dogs when they present a hazard to the environment (e.g. control of feral dogs in national parks; prevention of dog attacks on wildlife or transmission of diseases to wildlife) or where a lack of environmental controls is giving rise to stray dog populations that threaten human health or access to amenities. For example, Environmental Protection agencies may regulate and enforce measures to prevent dogs (and other wild animals) from accessing waste or human sewage.

3. Private sector veterinarians

The private sector veterinarian is responsible for providing advice to pet owners consulting the veterinarian for advice or treatment of a dog. The private sector veterinarian can play an important role in disease surveillance because he/she might be the first to see a dog suffering from a notifiable disease such as rabies. It is necessary that the private sector veterinarian follow the procedure established by the Veterinary Authority for responding to and reporting a suspected rabies case or a dog that is suffering from any other notifiable disease. Private sector veterinarians also play an important role (often in liaison with the police) in dealing with cases of neglect that can lead to problems with stray and mismanaged dogs.

The private veterinarian has competence and will normally be involved in pet dog health programmes and population control measures, including health testing and vaccination, kennelling during the absence of the owner, sterilisation and euthanasia. Two-way communication between the private sector veterinarian and Veterinary Authority, often via the medium of a veterinary professional organisation, is very important and the Veterinary Authority is responsible to set up appropriate mechanisms for this action.

4. Non Governmental Organisations (NGOs)

NGOs are potentially important partners of the Veterinary Services in contributing to public awareness and understanding and helping to obtain resources to contribute in a practical way to the design and successful implementation of dog control programmes. NGOs can supply local knowledge on dog populations and features of ownership, as well as expertise in handling and kennelling dogs and the implementation of large scale vaccination and sterilisation programmes. NGOs can also contribute, together with veterinarians and the authorities in educating the public in responsible dog ownership. NGOs can help to obtain funding for control programmes, particularly in countries where governments may depend on support from NGOs for programs carried out to assist poor communities.

5. Local Government Authorities

Local Government Authorities are responsible for many services and programmes that relate to health, safety and public good within their jurisdiction. In many countries the legislative framework gives authority to local government agencies in regard to aspects of public health, environmental health/ hygiene and inspection/ compliance activities.
Annex XXIII (contd)

In many countries local government agencies are responsible for the control of stray dogs (e.g. dog catching and shelters) and the alleviation of the problems stray dogs cause. This would normally be done with advice from a higher level (national or state/provincial) authority with specialised expertise in regard to public health and animal health. Collaboration with the private sector veterinarians (e.g. in programs to sterilise and vaccinate stray dogs) is a common feature of dog control programs. Regardless of the legislative basis, it is essential to have the co-operation of local government authorities in the control of stray dogs.

6. Dog owners

When a person takes on the ownership of dog there should be an immediate acceptance of responsibility for that dog, and for any offspring it may produce, for the duration of its life or until a subsequent owner is found. The owner must ensure the dog is protected, as far as possible, from infectious diseases (e.g. through vaccination and parasite control) and from unwanted reproduction (e.g. through surgical sterilisation). Owners should ensure that the dog's ownership is clearly identified (preferably with permanent identification such as a tattoo or microchip) and, where required by legislation, registered on a centralised database. All reasonable steps should be taken to ensure that the dog does not roam out of control in a manner that would pose a problem to the community and/or the environment.

Article 4

Considerations in planning a dog population control programme measures

In the development of a dog population control programme it is recommended that the authorities establish an advisory group, which should include appropriate veterinarians, experts in dog ecology, dog ownership and zoonotic diseases, and representatives of relevant stakeholders (local authorities, human health services/authorities, environmental control services/authorities and the public). The main purpose of this advisory group would be to analyse the problem, identify the causes and propose the most effective approaches to use in the short and long term.

Important considerations are as follows:

1. Identifying the sources of stray dogs
   a) Owned animals that roam freely
   b) Animals that have been abandoned by their owner, including animals resulting from:
      i) uncontrolled breeding of owned dogs;
      ii) unowned dogs that reproduce successfully.

2. Estimating the existing number, distribution and ecology (To be completed)

Practical tools that are available include using available practical tools such as registers of dogs, population estimates, surveys of dogs, owners, dog shelters and associated veterinarians etc. The important factors relevant to the dog carrying capacity of the environment include food, shelter, water and human behaviour.

A methodology, including generalised dog identification and centralised registration, must be established in order to make an estimate of the total dog population.

An overview of appropriate methodologies may be found in Annex I.
The same methodology must be used at appropriate intervals to assess population trends. Find references if possible:

- Identify the important factors relevant to the dog carrying capacity of the environment. These generally include food, shelter, water, and human behaviour.
- Add examples of good methodology if possible.

3. Legislation

Legislation that would help authorities establish successful dog control programmes should include the following key elements:

a) registration and identification of dogs and licensing of dog breeders/owners;
b) rabies vaccination;
c) veterinary procedures (e.g. surgical procedures);
d) control of dog movement (restrictions within the country);
e) control of dog movement (international movement);
f) control of dangerous dogs;
g) regulations on the breeding and sale of dogs (commercial dog production);
h) environmental controls (e.g. abattoirs, rubbish dumps, dead stock facilities);
i) dog shelters;
j) animal welfare, including humane capture and killing methods.

4. Resources available to authorities

a) Human resources
b) Financial resources
c) Technical tools
d) Infrastructure
e) Cooperative activities
f) Public-private-NGO partnerships
g) Central-state or province-local partnerships

**Article 5**

**Control measures**

The following control measures should be implemented according to the situation in national context and local circumstances of Member Countries. They may be used in combination, or singly. Killing of dogs, used alone, is not an effective control measure. If used, it should be combined with other measures to achieve effective long term control. It is also important that authorities gain an understanding of people's attitudes towards dog ownership so that they can develop a cooperative approach to the control of dog populations.

1. **Education and promotion legislation of responsible ownership (To be completed)**

The health and welfare of domestic dogs may be improved through the promotion of responsible human ownership. Minimizing stray dogs population, in combination with educating humans, particularly children about specific behaviours, can reduce dog bite injury and prevent some major zoonotic diseases.
Responsible dog ownership includes the control of reproduction of dogs under direct human supervision such that offspring of owned dogs are not abandoned.

The owned dog population is a primary source of stray dogs, through the abandonment of unwanted dogs and their offspring, and through allowing owned dogs to roam unrestricted, contributing to the stray population. Encouraging dog owners to be more responsible will reduce the number of dogs allowed to roam, improve the health and welfare of dogs, and minimise the risk that dogs pose to the community. The promotion of responsible dog ownership though legislation and education is a necessary part of a dog population management programme. Collaboration with responsible animal welfare NGO’s and private veterinarians will assist Veterinary Authorities in establishing and maintaining programmes.

Education on responsible dog ownership (for the currently owned dog and any offspring it produces) should address the following elements:

a) the importance of proper care to ensure the welfare of the dog and any offspring; this may include preparing the dog to cope with its environment through attention to socialisation and training;

b) registration and identification of dogs (see Article 5.2.);

c) prevention of zoonotic diseases, eg through regular vaccination in rabies endemic areas;

d) preventing negative impacts of dogs on the community, via pollution (eg faeces and noise), risks to human health through biting or traffic accidents and risks to wildlife, livestock and other companion animal species;

e) control of dog reproduction

In order to achieve a shift towards responsible ownership, a combination of legislation, public awareness, education, and promotion of these elements will be required. It may also be necessary to improve access to resources supporting responsible ownership, such as veterinary care, identification and registration services and measures for control of zoonotic diseases.

2. Registration and identification of dogs (licensing)

A core component of dog population management by the Competent Authorities is the registration and identification of owned dogs. This and may include granting licences to owners. Registration and identification may be emphasized as part of responsible dog ownership and are often linked to animal health programs, for example, mandatory rabies vaccination.

Registration and identification of animals in a centralised database can be used to support the enforcement of legislation, the reuniting of lost animals with owners and may be used as a tool to encourage control of dog reproduction control of owned dogs through financial incentives reduced fee schedule to register neutered sterlise dogs.

3. Reproductive control

Controlling reproduction in dogs prevents the birth of unwanted litters of puppies and can help address the balance between demand for dogs and the size of the population. It is advisable to focus efforts to control reproduction on those individuals or groups in the dog population identified as the most productive and the most likely to be the sources of unwanted and stray dogs, as this will to ensure best use of resources. Methods of controlling reproduction will require direct veterinary input to individual animals; involvement of both private and public veterinary sectors may be required to meet demand. The control of reproduction is essentially the responsibility of owners and can be incorporated into education on responsible ownership (section 5 a.). Methods for controlling reproduction in dogs include:
a) surgical sterilisation;

b) chemical sterilisation;

c) chemical contraception;

d) separation of female dogs during oestrus from entire unsterilised males.

Surgical sterilisation should be carried out in a humane manner and include appropriate use of pain relief.

Any chemicals or drugs used in controlling reproduction should be shown to have appropriate safety, quality and efficacy for the function required and used according to the manufacturer's and Competent Authority's regulations. In the case of chemical sterilants and contraceptives, this may require further research and field trials may need to be completed before use.

4. Removal and handling

The Competent Authority should collect dogs that are not under direct supervision and verify their ownership. Capture, transport, and holding of the animals should be done humanely. The Competent Authority should develop and implement appropriate legislation and training to regulate these activities. Capture should be achieved with the minimum force required and equipment should be used that supports humane handling. Snares and uncovered wire loops should not be used for capture.

5. Management of dogs removed from communities

Competent authorities have the responsibility to develop minimum standards for the housing (physical facilities) and care of these dogs. There should be a provision for holding the dogs for a reasonable period of time to allow for reunion with the owner and, as appropriate, for rabies observation. A period of 7-10 days is often used for this purpose.

a) Minimum standards for housing should include the following provisions:

   i) site selection: Access to drainage, water and electricity are essential and environmental factors such as noise and pollution should be taken into account;

   ii) kennel size, design and occupancy taking exercise into account;

   iii) disease control measures including isolation facilities;

b) Management should address:

   i) adequate fresh water and nutritious food;

   ii) regular hygiene and cleaning;

   iii) routine inspection of the dogs;

   iv) monitoring of health and provision of required veterinary treatments;

   v) policies and procedures for rehoming, sterilisation and euthanasia;
Annex XXIII (contd)

vi record keeping and reporting to authorities.

Dogs that are removed from a community may be reunited with the owner or offered to new owners for adoption (rehoming). This provides an opportunity to promote responsible ownership and good animal health care (including rabies vaccination), including animal health care through vaccination against common diseases of dogs, control of ecto- and endo-parasites, and vaccination against major zoonotic diseases such as rabies. Incentives for dog reproduction control may be provided through the provision of neutering services at a reduced rate or the release for adoption of only neutered animals. Sterilisation of dogs prior to adoption should be considered. The suitability of new owners to adopt dogs should be assessed and owners matched with available animals. The effectiveness of this strategy, i.e., offering dogs to new owners, rehoming may be limited due to the suitability and number of dogs.

Dogs that are removed from a community may in some cases be provided health care (including rabies vaccination), sterilised, and released to their local community at or near the place of capture, who agree to take responsibility for the health, welfare and management of the animal. The beneficial effect of this practice for dog welfare and population management is unknown. With regard to disease control, such as for rabies and possibly others, some beneficial effect may be realized. This may be short or long term. This method is more likely to be accepted in the situation where the presence of stray dogs is considered to be inevitable and is well tolerated by the local community.

This method is not applicable in all situations and may be illegal in countries where legislation prohibits the abandonment of dogs. Problems caused by dogs, such as noise, faecal pollution and traffic accidents, would not be alleviated as dogs are returned to the local community and their movements are not restricted. If the local community has owned dogs, consideration should be given to the potential encouragement of abandonment of unwanted. In the situation where many dogs are owned, a population control programme that focuses on neutering and responsible ownership may be more appropriate.

It is recommended that before adopting this approach, a cost-benefit analysis is conducted. Factors such as the monetary costs, impact on culture of ownership and public safety should be assessed as well as the benefits for disease control and animal welfare as well as any societal benefits.

c) If this method is adopted, the following factors should be addressed:

i) Raising awareness of the programme within the local community to ensure understanding and support.

ii) Use of humane methods for catching, transporting and holding dogs.

iii) Correct surgical technique, anaesthesia and analgesia, followed by post-operative care.

iv) Disease control may include blanket vaccination (e.g. rabies) and treatments and testing for disease (e.g. leishmaniasis) followed, as appropriate by treatment or euthanasia of the dog.

v) Behavioural observation may be used to assess if dogs are suitable for release. If not suitable for release or re-homing euthanasia should be considered.

vi) Permanent marking (e.g. tattoo) to indicate that the animal has been sterilised. Individual identification allows for tracking of vaccination status and treatment history. A visible identification (e.g. collar) may also be used to prevent unnecessary recapture. Identification can also be taken to indicate a level of “ownership” by the organisation/authority responsible for carrying out this intervention.
The dog should be returned to a place that is as near as possible to the place of capture.

The welfare of dogs after release should be monitored and action taken if required.

Dogs that are removed from a community may, in some cases, be too numerous or may be unsuitable to place responsible ownership. If elimination of the excess animals is the only option, killing should be under regulation by a for any rehoming scheme. If euthanasia of these unwanted animals is the only option, the procedure should be conducted in accordance with the regulations of the Competent Authority and conducted humanely (see Article 4 k).

A number of selected animals could be released if "environmentally compatible", meaning that, once again, the feasibility of this strategy is very much related to the local people attitude/resources availability:

- Risk-benefit evaluation of Catch-Neuter-Release & Monitoring (CNR&M) in terms of public safety and AW.
- Proper behavioural evaluation of dogs when removed for problems related to public nuisance
- Monitoring needed to evaluate individual health and welfare
- Sufficient level of public tolerance, food and assistance provided by responsible people/community
- Permanent identification (i.e. surgical sterilization, rabies vaccination, echinococcosis treatment, Leishmaniasis negative test). These actions clearly reconnect the animal to an "owner", both intended as public (local municipality, regional government) or private
- Possibly clearly visible at distance (i.e. painted collars)

**Advantages:** Possible strategy when scarce resources are in place, if adopted in very specific situation it may also promote the societal value of animals and the benefits of a positive human-animal relationship (Rome's cat colony, "community" dogs)

**Disadvantages:** Ineffective over a long term since not promoting responsible ownership concept, possible AW concerns due to persistent intolerance by the community, possible risk to human safety and damage of the private property due to improper selection of animals.

Preferably to be used as a "spot" solution in specific situations and only in addition to other measures (humane education, door-to-door reuniting programs, adoption programs), possibly not to be used as the sole method of stray dog population control as a long term strategy.

6. Environmental controls

Steps should be taken to reduce the carrying capacity, such as excluding dogs from sources of food (e.g. rubbish dumps and abattoirs, and installing animal-proof rubbish containers).

This should be linked to a reduction in the animal population by other methods, to avoid animal welfare problems.

7. Control of dog movement – international (export/import)

Chapter 2.2.5 of the Terrestrial Animal Health Code provides recommendations on the international movement of dogs between rabies free countries and countries considered to be infected with rabies.
8. Control of dog movements – within country (e.g. leash laws, roaming restrictions)

Measures for the control of dog movement in a country are generally invoked for two following reasons:

a) for rabies control when the disease is present in a country
b) for public safety reasons
c) for the safety of “owned dogs” in an area or locality when a stray dog control programme is in place
d) to protect wildlife and livestock.

In both cases, it is essential that dogs are registered and permanently identified to control or confine these dogs, reunite them if collected and to keep the relevant sanitary information recorded.

It is necessary to have empowering legislation to give the necessary power is necessary and a national or local infrastructure comprising of organization, administration, staff and resources is essential to encourage the finders of a stray dog to report to the Competent Authority.

The following 3 grades of movement control can be applied:

- Absolute control (confinement, leash-end muzzle), feasible during a limited periods such as for an emergency
- Partial control (obedience if not on leash during daylight, confinement between the relevant information times of 5pm and 8 am)
- Control during specific times (rabies vaccination campaign, stray dog roundup)

9. Regulation of commercial Animal dog dealers

While the majority of animal breeders and dealers are committed to raising and selling physically and psychologically healthy pets, regulation is necessary to ensure that all of these operations provide adequate care.

The law should require the humane care and treatment of certain animals sold as pets in retail stores as well as at the wholesale level, transported in commerce, and used in research or exhibits.

Individuals using or working with such animals should be licensed and they must comply with regulations and standards.

- Standards of Care and Recordkeeping

Businesses in the commercial pet trade must maintain minimum standards for veterinary care and animal management. The requirements should cover housing, handling, sanitation, food, water, and protection against extremes of weather and temperature.

To prevent lost or stolen animals from entering trade channels, breeders and dealers are required to keep records that identify the source and disposition of all regulated animals that come into their possession.

- Shipping and Handling

Specific regulations and standards are needed to regulate the transport of animals by commercial carriers. These rules help ensure that licensed dealers, contract carriers, and intermediate handlers treat regulated animals humanely. Transported animals must meet established minimum age and health certification requirements.
Regulation is needed to ensure that dog breeders and dealers are identified by the Competent Authority and are committed to raising and selling physically and psychologically healthy animals, as unhealthy animals may be more likely to be abandoned to become part of the stray population. Regulations should include specific requirements for accommodation, provision of suitable food, drink and bedding, adequate exercise, veterinary care and disease control. Breeders and dealers establishments should be inspected at regular intervals, including veterinary inspections. Advice on proper animal care should be given to all new owners of dogs.

10. Reduction in dog bite incidence

Propensity to bite is influenced by heredity, early experience, socialisation & training, health and human behaviour towards the dog. Breed or type specific bans are difficult and costly to enforce, provide a false sense of security to the community and, where enacted, no data currently supports them as effective in reducing incidence of dog bites, therefore, they are not recommended. Specific behaviours or incidences can be used as criteria to facilitate identification of a dog as ‘dangerous’ and appropriate measures taken to control the animal by the competent authority. For example, a dog that has been reported to have bitten someone or something (livestock or pets) may be required by law to be confined on the owner’s property and kept on a lead (and if necessary muzzled) when in public. Note that confinement by tethering should be avoided as this can increase the likelihood of aggressive behaviour.

The most effective means of reducing prevalence of dog bites are education and placing responsibility on the owner, not the animal. Dog owners should be educated trained in principles of responsible pet ownership as described in Article 5.a. Legal mechanisms that enable the competent authorities to impose penalties or otherwise deal with irresponsible owners are necessary. Mandatory registration and identification schemes will facilitate the effective application of such mechanisms. Young children are the group at highest most at-risk group for dog bites. Education programmes focussed on appropriate dog-directed behaviour have been demonstrated to be effective in reducing dog bite prevalence and these programmes should be encouraged.

11. Euthanasia

When euthanasia is practised, the procedures used should comply with general principles the presented laid down in the Terrestrial Animal Health Code – 2006 (Article 3.7.6.1) should be followed, with the emphasis on using the most practical, rapid and humane methods and ensuring operator safety.

For practical reasons, different procedures may be used in rural and urban areas.

For reasons of convenience, different procedures could be used in rural and in urban areas. Dogs should only be euthanized after holding for a period of time to allow the owner to locate his/her dog.

Table 1 shows a list of methods for the euthanasia of dogs.

They fall into two major categories based on whether it is necessary to handle or restrain the dog or not in order to euthanize it.

Where capture or restraint procedures give rise to a risk or potential risk of human exposure to rabies, procedures that do not require restraint of dogs are preferable.

The methods are not described in any particular order.
### Annex XXIII (contd)

<table>
<thead>
<tr>
<th>Procedure</th>
<th>Capture</th>
<th>Restraint</th>
<th>Advantages/Disadvantages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Electrocutation</td>
<td>Yes</td>
<td>No</td>
<td>Affordable equipment: 220V mains current; gloves + boots.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Instant death.</td>
</tr>
<tr>
<td>Carbon monoxide (CO)</td>
<td>Yes</td>
<td>No</td>
<td>Needs appropriate premises; puts personnel at risk.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Slow death.</td>
</tr>
<tr>
<td>CO2</td>
<td>Yes</td>
<td>No</td>
<td>As CO2 is heavier than air, the dogs can lift their heads over the CO2 layer and death is slow.</td>
</tr>
<tr>
<td>Barbiturates</td>
<td></td>
<td></td>
<td>Requires an appropriate dose and pre-anaesthetic.</td>
</tr>
<tr>
<td>Intravenous</td>
<td>Yes</td>
<td>Yes</td>
<td>Administered under veterinary supervision and requires trained personnel.</td>
</tr>
<tr>
<td>Intracardial</td>
<td>Yes</td>
<td>Yes</td>
<td>Slow death.</td>
</tr>
<tr>
<td>Intraperitoneal</td>
<td>Yes</td>
<td>Yes</td>
<td></td>
</tr>
<tr>
<td>T61 - Tanax</td>
<td></td>
<td></td>
<td>Dangerous for personnel in the event of accidental injection.</td>
</tr>
<tr>
<td>Intravenous</td>
<td>Yes</td>
<td>Yes</td>
<td>Slow death.</td>
</tr>
<tr>
<td>Intracardial</td>
<td>Yes</td>
<td>Yes</td>
<td></td>
</tr>
<tr>
<td>Intrapulmonary</td>
<td>Yes</td>
<td>Yes</td>
<td></td>
</tr>
<tr>
<td>Free bullet used from long range</td>
<td>No</td>
<td>No</td>
<td>Fast death.</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Risk of accident (same as for hunting).</td>
</tr>
</tbody>
</table>
**Table 1: List of methods for the euthanasia of dogs**

| Euthanasia method | Specific method                | Animal welfare concerns/ implications | Key animal welfare requirements                                                                 | Considerations relating to operator security                                                                 | Advantages                                                                                       | Disadvantages                                                                                   |
|-------------------|--------------------------------|--------------------------------------|------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------|
| Chemical          | Barbiturates                   | Correct restraint is needed.          | Recommend to use IV injection. When using IP injection, the solution may be diluted or local anaesthetic agent used in conjunction. IC should only be performed on unconscious animal and by skilled operator. | Correct restraint is needed. Administered under veterinary supervision and requires trained personnel. | Speed of action generally depends on the dose, concentration, route and rate of injection. Barbiturates induce euthanasia smoothly, with minimal discomfort to the animal. Barbiturates are less expensive than many other euthanasia agents. | Mild aesthetic objection as terminal gasps may occur in unconscious animals. These drugs persist in the carcass and may cause sedation or death in animals that consume the cadaver. |
| -via injection    | Embutramide + Mebezonium + Tetracaine | Muscle paralysis may occur before lost of consciousness if injection given rapidly | Use slow IV injection with sedation to permit slow rate of injection. | Correct restraint is needed. To be administered under veterinary supervision and by trained personnel. | Quite low cost. | Unavailable/unlicensed in some countries |
|                   | Anaesthetic agent overdose (thiopentone or propofenol) | Underdosing may lead to recovery | IV injection of a sufficient dose | Correct restraint is needed. To be administered under veterinary supervision and by trained personnel. | Generally quick action and minimal discomfort to animal. | Large volume required (cost implications) |
|                   | Potassium chloride (KCl)       | K⁺ is cardiotoxic and very painful if used without anaesthetic agent. | Only use on anaesthetised animals, IV injection | Requires trained personnel. | Readily available without veterinary control. | Prior need for anaesthetic (cost and availability implications) |
Table 1: List of methods for the euthanasia of dogs

<table>
<thead>
<tr>
<th>Euthanasia method</th>
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<th>Advantages</th>
<th>Disadvantages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Free bullet</td>
<td>Can be inhumane if shot is inaccurate and dog is only wounded; dog may also escape.</td>
<td>Skilled operator essential.</td>
<td>Risk of injury of operator.</td>
<td>Not necessary to handle or capture dog.</td>
<td>Brain tissue may be unavailable for rabies diagnosis. Risk of injury to bystanders. Legal constraints on use of firearms.</td>
<td></td>
</tr>
<tr>
<td>Mechanical</td>
<td>Penetrating captive bolt</td>
<td>Can be inhumane if shot is inaccurate and dog is only wounded.</td>
<td>Skilled operator essential.</td>
<td>Animal must be restrained. Skilled operator essential.</td>
<td>No risk to operator (cf free bullet)</td>
<td>Brain tissue may be unavailable for rabies diagnosis. Legal constraints on use of firearms. May raise aesthetic objections.</td>
</tr>
<tr>
<td>Exsanguination</td>
<td>Onset of hypovolaemia may cause dog to become anxious.</td>
<td>Only use on unconscious animal</td>
<td>Danger to operator through use of sharp instrument.</td>
<td>Material requirements minimal.</td>
<td>Must be done on unconscious animal. Aesthetically objectionable</td>
<td></td>
</tr>
</tbody>
</table>
### Table 1: List of methods for the euthanasia of dogs (cont)

<table>
<thead>
<tr>
<th>Euthanasia method</th>
<th>Specific method</th>
<th>Animal welfare concerns/ implications</th>
<th>Key animal welfare requirements</th>
<th>Considerations relating to operator security</th>
<th>Advantages</th>
<th>Disadvantages</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Gaseous</strong></td>
<td>Carbon monoxide (CO)</td>
<td>Gas is aversive. Inadequate concentration of CO is not lethal and can cause suffering. Signs of distress (convulsions, vocalization and agitation) may occur.</td>
<td>Compressed CO in cylinders must be used to achieve and maintain adequate concentration, which must be monitored. Note: fumes from gasoline engines are irritant and this source of CO is not recommended.</td>
<td>Very hazardous for operator - gas is odourless and causes both high and chronic toxicity.</td>
<td>Dog dies quite rapidly if concentration of 4 to 6% used. No odour (therefore no aversive effect). Gas is not flammable or explosive except at concentration greater than 10%.</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Carbon dioxide (CO&lt;sub&gt;2&lt;/sub&gt;)</td>
<td>Gas is highly aversive. Inadequate concentration of CO&lt;sub&gt;2&lt;/sub&gt; is not lethal and can cause suffering. CO&lt;sub&gt;2&lt;/sub&gt; is heavier than air, so when incomplete filling of the chamber occurs, dogs may raise their head and avoid exposure. Few studies on adequate concentration and animal welfare.</td>
<td>Compressed CO&lt;sub&gt;2&lt;/sub&gt; gas chamber is the only recommended method because the concentration can be monitored and regulated.</td>
<td>Minimal hazard to operator when properly designed equipment used.</td>
<td>Gas is not flammable or explosive and causes quite rapid anaesthesia when correct concentrations used. Low cost. Readily available as compressed gas</td>
<td>Anaesthesia can be quite rapid but death may take some time.</td>
</tr>
<tr>
<td></td>
<td>Inert gas (nitrogen, N&lt;sub&gt;2&lt;/sub&gt;, argon, Ar)</td>
<td>Loss of consciousness is preceded by hypoxemia and ventilatory stimulation, which may be distressing to the dog. Re-establishing a low concentration of O&lt;sub&gt;2&lt;/sub&gt; (ie greater than or equal to 6%) in the chamber before death will allow immediate recovery.</td>
<td>Concentration above 98% must be achieved rapidly and maintained. Properly designed equipment must be used</td>
<td>Minimal hazard to operator when properly designed equipment used.</td>
<td>Gas is not flammable or explosive and is odourless. Readily available as compressed gas</td>
<td>High cost. Little data on animal welfare implications in dogs.</td>
</tr>
</tbody>
</table>

*Annex XXIII (cont)*
Annex XXIII (contd)

### Table 1: List of methods for the euthanasia of dogs (cont)

<table>
<thead>
<tr>
<th>Euthanasia method</th>
<th>Specific method</th>
<th>Animal welfare concerns/implications</th>
<th>Key animal welfare requirements</th>
<th>Considerations relating to operator security</th>
<th>Advantages</th>
<th>Disadvantages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gaseous</td>
<td>Anaesthetic gas overdose (halothane or enflurane)</td>
<td>Animal may struggle and become anxious during induction. Vapours may be irritating and can induce excitement.</td>
<td>Supplementation with air or O₂ required to avoid hypoxemia during induction phase.</td>
<td>Some gases may be hazardous, especially for pregnant women. General recommendation: Avoid human exposure to greater than or equal to 2ppm to avoid narcosis.</td>
<td>Gas is not flammable or explosive. Valuable for use with small animals.</td>
<td>High cost. Anaesthetic and euthanasia properties of the gas used must be known. Isoflurane has a pungent odour. Methoxyflurane's action is slow and dog may become agitated.</td>
</tr>
<tr>
<td>Electrical</td>
<td>Electrocution</td>
<td>Cardiac fibrillation occurs before onset of unconsciousness, causing severe pain if dog is conscious. Pain can also be caused by violent extension of the limbs, head and neck. Method may not be effective if insufficient current applied.</td>
<td>Dogs must be unconscious before being electrocuted. This can be accomplished by electrical stunning (current through the brain to produce an instantaneous stun) or anaesthesia. Electrodes should span the brain in order that the current passed through the brain.</td>
<td>May be hazardous for operator, who should use protective equipment (boots and gloves).</td>
<td>Low cost.</td>
<td>Inhumane if performed on conscious dog. May raise aesthetic objections.</td>
</tr>
</tbody>
</table>

**KEY to abbreviations used in Table 1:**
- IV: intravenous
- IP: Intraperitoneal
- IC: Intracardiac
To be developed for each method

1. Introduction
2. Requirements for effective use
3. Advantages
4. Disadvantages
5. Conclusions

a) Summary assessment of Comments on methods for the euthanasia of dogs:

i) Restraint

When a dog needs to be restrained for any procedure, including euthanasia, this should always be done with full regard for operator security and animal welfare. In order to use some euthanasia methods must be used in association with sedation or anaesthesia in order to be considered humane ways, may be required.

ii) Special equipment

When special equipment is needed to perform euthanasia (e.g. gas chamber) the system should be properly designed for the purpose and regularly maintained in order to achieve operator security and animal welfare.

iii) The following methods, procedures and practices are unacceptable on animal welfare grounds:

- Chemical methods:
  - Embutramide + Mebezonium + Tetracaine without sedation or by other than IV injection
  - Chloral hydrate
  - Nitrous oxide: may be used with other inhalants to speed the onset of anaesthesia but alone it does not induce anaesthesia in dogs
  - Ether
  - Chloroform
  - Cyanide
  - Strychnine
  - Neuromuscular blocking agents (nicotine, magnesium sulphate, potassium chloride, all curariform agents): when used alone, respiratory arrest occurs before loss of consciousness, so the dog may perceive pain
  - Formalin
  - Household products and solvents

- Mechanical methods:
  - Air embolism on conscious animal
Annex XXIII (contd)

- Burning
- Exsanguination of conscious animal
- Decompression: expansion of gas trapped in body cavities may be very painful
- Drowning
- Hypothermia, rapid freezing
- Stunning: stunning is not a euthanasia method, it should always be followed by a method which ensures death.
- Kill-trapping
- Electrocution of conscious animal.

Because neonatal animals are resistant to hypoxia, methods that depend upon achieving a hypoxic state (eg CO₂, CO, N₂, Ar) should not be used. These methods should not be used in animals aged less than 4 months, except to produce loss of consciousness and should be followed by another method to cause death. Cervical dislocation and concussion may be used in neonatal dogs. Operators must be well trained in the use of physical techniques to ensure that they are correctly and humanely carried out. The dog must be exsanguinated immediately after concussion or cervical dislocation.

iv) Confirmation of death

For all methods of euthanasia used, death must be confirmed before animals are disposed of or left unattended. If an animal is not dead, another method of euthanasia must be performed.

v) Carcass disposal

Carcasses should be disposed of in a manner that complies with legislation. Attention must be paid to the risk of residues occurring in the carcase. Incineration is generally the safest way of carcass disposal.

**Article 6**

**Monitoring and evaluation of dog population control programmes**

Monitoring and evaluation allows for comparison of important indicators against the baselines measured during initial assessment (Article 4). The three main reasons for carrying out monitoring and evaluation are:

1. To help improve performance, by highlighting both problems and successful elements of interventions.
2. For accountability, to demonstrate that the programme is achieving its aims.
3. Assuming methods are standardised, to compare the success of strategies used in different locations and situations.

Monitoring is a continuous process that aims to check the programme progress against targets and allows for regular adjustments. Evaluation is a periodic assessment, usually carried out at particular milestones to check the programme is having the desired and stated impact. These procedures involve the measurement of ‘indicators’ that are chosen because they reflect important components of the programme at different stages. Selection of suitable indicators requires clear planning of what the programme is aiming to achieve, the best selection of indicators will be one that reflects the interest of all relevant stakeholders. Standardised methodology will facilitate comparison of data from subsequent evaluations and performance between different projects. Indicators can be direct measurements of an area targeted to change (e.g. population of free roaming dogs on public property) or indirect measures that reflect change in a targeted area (e.g. number of reported dog bites as a reflection of rabies prevalence).
4. Elements that should generally be monitored and evaluated most programmes will need to monitor and evaluate include:
   a) Dog population size, separated by sub-populations according to ownership and restriction of movement (i.e. roaming unrestricted or restricted by an owner);
   b) Dog welfare in the target population (e.g. body condition score, skin conditions and injuries or lameness) and as a result of the programme (if interventions involve direct handling of dogs, the welfare of the dogs as result of this handling should be monitored);
   c) Prevalence of zoonotic diseases, such as rabies; prevalence in both the animal and human population can be measured;
   d) Responsible animal ownership, including measures of attitudes and understanding of responsible ownership and evidence that this is translating into actual responsible behaviour.

5. There are many sources of information for measuring indicators can be widespread, including:
   a) Feedback from the local community (e.g. through the use of structured questionnaires or ‘open format’ consultation processes)
   b) Records and opinions obtained from relevant professionals (e.g. veterinarians, medical doctors, law enforcement agencies, educators)
   c) Animal based measurements (e.g. direct observation surveys of population size and welfare status)

The output of activities against budget should be carefully recorded in order to evaluate balance the effort (or cost) against the outcomes and impact (or benefit) that are reflected in the results of monitoring and evaluation results.

Article 7

Research needs

To be completed

Article 8

International cooperation

To be completed
Annex XXIII (contd)

Annex I:

An overview of appropriate methodologies for estimating the size of dog populations.

Population estimates are necessary for making realistic plans for dog population management and zoonosis control, and for monitoring the success of such interventions. However, for designing effective management plans, data on population sizes alone are insufficient. Additional information is required, such as degrees of supervision of owned dogs, the origin of ownerless dogs, accessibility, etc.

The term “owned” may be restricted to a dog that is registered with licensing authorities, or it may be expanded to unregistered animals that are somewhat supervised and receive shelter and some form of care in individual households. Owned dogs may be well supervised and restrained at all times, or they may be left without control for various time periods and activities. Dogs without owners that claim responsibility may still be accepted or tolerated in the neighbourhood, and individuals may provide food and protection. Such animals are sometimes called “community owned dogs” or “neighbourhood dogs”. For an observer it is frequently impossible to decide if a free roaming dog belongs to someone or not.

The choice of methods for assessing the size of a dog population depends on the ratio of owned versus ownerless dogs, which may not always easy to judge. For populations with a large proportion of owned dogs it may be sufficient to consult dog registration records or to conduct household surveys. These surveys should establish the number of owned dogs and the dog to human ratio in the area. In addition, questions on dog reproduction and demographics, care provided, zoonosis prevention, dog bite incidence, etc. may be asked. Sample questionnaires can be found in the “Guidelines for Dog Population Management” (WHO/WSPA 1990). Standard polling principles must be applied.

If the proportion of ownerless dogs is high or difficult to assess, then one must resort to more experimental approaches. Methods borrowed from wildlife biology can be applied. These methods are described WHO/WSPA’s “Guidelines for Dog Population Management” (1990), and in more detail in numerous professional publications and handbooks, such as Bookhout (1994) and Sutherland (2006). Being generally diurnal and tolerant to human proximity, dogs lend themselves to direct observation and the application of mark-recapture techniques. Nevertheless, a number of caveats and limitations have to be taken into account. The methods are relatively labour intensive, they require some understanding of statistics and population biology, and most importantly, they are difficult to apply to very large areas. One must take into account that dog distribution is non-random, that their populations are not static, and that individual dogs are fairly mobile.

Counting of dogs visible in a defined area is the simplest approach to getting information on population size. One has to take into account that the visibility of dogs depends on the physical environment, but also on dog and human activity patterns. The visibility of animals changes with the time of the day and with seasons as a function of food availability, shelter (shade), disturbance, etc. Repeated standardized counting of dogs visible within defined geographical localities (e.g. wards) and specific times will provide indications of population trends. Direct counting is most reliable if it is applied to small and relatively confined dog populations, e.g. in villages, where it might be possible to recognize individual dogs based on their physical appearance.

Methods using mark-recapture procedures are often considered more reliable. However, they also produce trustworthy results only when a number of preconditions are met. Mortality, emigration and recruitment into the population must be minimal during the census period. One may be able to incorporate corrective factors into the calculations.
Annex XXIII (contd)

It is therefore important that the recommended census procedures are applied at times of low dispersal and that one selects study plots of shape and size that minimize the effect of dog movements in and out of the observation area. Census surveys should be completed within a few days to a maximum of two weeks in order to reduce demographic changes. In addition, all individuals in the population must have an equal chance of being counted. This is a highly improbable condition for dogs, whose visibility depends on ownership status and degrees of supervision. It is therefore recommended that the investigator determines what fraction of the total population he/she might cover with an observational method and how much this part overlaps with the owned dog segment that he/she assesses with household surveys.

There are essentially two ways to obtain a population estimate if it is possible, in a defined area and within a few days, to tag a large number of dogs with a visible mark, e.g. a distinctive collar or a paint smudge. The first method requires that the capture (marking) effort remains reasonably constant for the whole length of the study. By plotting the daily number of dogs marked against the accumulated total of marked dogs for each day one can extrapolate the value representing the total number of dogs in the area. More commonly used in wildlife studies are mark recapture methods (Peterson-Jackson, Lincoln indices). Dogs are marked (tagged) and released back into the population. The population is subsequently sampled by direct observation. The number of marked and unmarked dogs is recorded. One multiplies the number of dogs that were initially marked and released by the number of subsequently observed dogs divided by the number of dogs seen as marked during the re-observation to obtain a total population estimate. Examples for the two methods are given in WHO/WSPA’s “Guidelines for Dog Population Management” (1990).

Since the dog populations of entire countries, states, provinces or even cities are much too large for complete assessment, it is necessary to apply the methods summarized above to sample areas. These should be selected (using common sense) so that results can be extrapolated to larger areas.


CHAPTER 2.9.X.

AETHINA TUMIDA (SMALL HIVE BEETLE)
INFESTATION OF HONEY BEES

Article 2.9.X.1.

For the purposes of this chapter, small hive beetle (SHB) is an infestation of bee colonies by the beetle Aethina tumida, which is an ectoparasite affecting populations of the honey bee Apis mellifera L. It can also parasitise bumble bee Bombus terrestris colonies under experimental conditions, and although infestation has not been demonstrated in wild populations, Bombus spp. must also be considered to be susceptible to infestation.

The adult beetle is attracted to bee colonies to reproduce, although it can survive and reproduce independently in other natural environments, using other food supplies as its nutritional source, including certain types of fruit. Hence once it is established within a localised environment, it is extremely difficult to eradicate.

The life cycle of Aethina tumida begins with the adult beetle laying eggs within infested hives. These are usually laid in irregular masses in hive crevices or brood combs. After 2-6 days, the eggs hatch and the emerging larvae begin to feed vociferously on brood comb, bee eggs, pollen and honey within the hive. The SHB has a high reproductive potential. Each female can produce about 1,000 eggs in its four to six months of life. At maturation (approximately 10-29 days after hatching), the larvae exit the hive and burrow into soil around the hive entrance. Adult beetle emerge after an average of 3-4 weeks, although pupation can take between 8 and 60 days depending on temperature and moisture levels (usually takes 3 to 4 weeks).

The life span of an adult beetle depends on environmental conditions such as temperature and humidity but, in practice, adult beetles can live for at least 6 months and, in favourable reproductive conditions, the female is capable of laying new egg batches every 5-12 weeks. The beetle is able to survive at least two weeks without food and 50 days on brood combs.

Early signs of infestation may go unnoticed, but the growth in the beetle population is rapid, leading to high mortality in the hive. Because Aethina tumida can be found and can thrive within the natural environment, and can fly up to a distance of 6-13km from its nest site, it is capable of dispersing rapidly and directly colonising hives. This includes following or accompanying swarms. It also does not require direct contact between adult bees to spread infestation. However, the movement of adult bees, honeycomb and other apiculture products may all cause infestations to spread to previously unaffected colonies.

[Standards for diagnostic tests are described in the Terrestrial Manual.]

Article 2.9.X.2.

The Aethina tumida status of a country or zone can only be determined after considering the following criteria:

1. a risk assessment has been conducted, identifying all potential factors for Aethina tumida occurrence and their historic perspective, including disease/pest incidence data from permanent official sanitary surveillance of apiaries programme;
Annex XXIV (contd)

2. Aethina tumida infestation should be notifiable in the whole country, and all clinical signs suggestive of Aethina tumida infestation should be subjected to field and laboratory investigations;

3. on-going awareness and training programmes should be in place to encourage reporting of all cases suggestive of Aethina tumida infestation;

4. the Veterinary Authority or other competent authority with responsibility for the health of honey bees should have current knowledge of, and authority over, all domesticated apiaries in the country.

Article 2.9.X.3.

Country or zone free from Aethina tumida

1. Historically free status

A country or zone may be considered free from the disease after conducting a risk assessment as referred to in Article 2.9.X.2. but without formally applying a specific surveillance programme if the country or zone complies with the provisions of Article 3.8.1.2.

2. Free status as a result of an eradication programme

A country or zone which does not meet the conditions of point 1 above may be considered free from Aethina tumida infestation after conducting a risk assessment as referred to in Article 2.9.X.2. and when:

a) the Veterinary Authority or other competent authority with responsibility for the health of honey bees has current knowledge of, and authority over, all domesticated apiaries existing in the country or zone;

b) Aethina tumida infestation is notifiable in the whole country or zone, and any clinical cases suggestive of Aethina tumida infestation are subjected to field and laboratory investigations; a contingency plan is in place describing controls and inspection activities;

c) for the 5 years following the last reported case of Aethina tumida infestation, an annual survey supervised by the Veterinary Authority, with negative results, have been carried out on a representative sample of apiaries in the country or zone to provide a confidence level of at least 95% of detecting Aethina tumida infestation if at least 1% of the apiaries were infested at a within-apiary prevalence rate of at least 5% of the hives; such surveys may be targeted towards areas with a higher likelihood of infestation;

d) to maintain free status, an annual survey supervised by the Veterinary Authority, with negative results, is carried out on a representative sample of apiaries in the country or zone to indicate that there has been no new cases; such surveys may be targeted towards areas with a higher likelihood of infestation;

e) all equipment associated with previously infested apiaries has been destroyed, or cleaned and sterilised to ensure the destruction of Aethina tumida spp., in conformity with one of the procedures referred to in Appendix X.X.X. (under study);

f) the soil and undergrowth in the immediate vicinity of all infested apiaries has been treated with a soil drench or similar suitable treatment that is efficacious in destroying incubating Aethina tumida larvae and pupae;
g) the importation of the commodities listed in this Chapter into the country or zone is carried out, in conformity with the recommendations of this Chapter.

Article 2.9.X.4.
Regardless of the status of the exporting country with regard to Aethina tumida infestation, Veterinary Authorities should authorise without restriction the import or transit through their territory of the following commodities:
1. honey bee semen and honey bee venom;
2. extracted honey, refined or rendered beeswax, propolis and royal jelly.

Article 2.9.X.5.
Veterinary Authorities of importing countries should require:

for individual consignments containing a single live queen honey bee, accompanied by a small number of associated attendants (a maximum of 20 attendants per queen)

the presentation of an international veterinary certificate attesting that the bees come from a country or zone officially free from Aethina tumida infestation

OR

the presentation of an international veterinary certificate including an attestation from the competent authority of the exporting third country stating that:
1. the bees come from hives or colonies which were inspected immediately prior to dispatch and show no clinical signs or suspicion of the presence of Aethina tumida or its eggs, larvae or pupae;
2. come from an area of at least 100 km radius where no apiary has been subject to any restrictions associated with the occurrence of Aethina tumida for the previous 6 months; and
3. the bees and accompanying packaging presented for export have been thoroughly and individually inspected and do not contain Aethina tumida or its eggs, larvae or pupae.

Article 2.9.X.6.
Veterinary Authorities of importing countries should require:

for live worker bees, drone bees or bee colonies with or without associated brood combs

the presentation of an international veterinary certificate attesting that the bees come from a country or zone officially free from Aethina tumida infestation, and the bees and accompanying packaging presented for export have been inspected and do not contain Aethina tumida or its eggs, larvae or pupae.

Article 2.9.X.7.
Veterinary Authorities of importing countries should require:

for eggs, larvae and pupae of honey bees

the presentation of an international veterinary certificate attesting that the products:
Annex XXIV (contd)

1. were sourced from a free country or zone (under study); 
   OR
2. have been isolated from queens in a quarantine station; and
3. are from hives or come from hives or colonies which were inspected immediately prior to entry into
   the quarantine station and show no clinical signs or suspicion of the presence of Aethina tumida or its
   eggs or larvae or pupae then and during the quarantine period.

   Article 2.9.X.8.

Veterinary Authorities of importing countries should require:

for used equipment associated with beekeeping

the presentation of an international veterinary certificate attesting that:

1. the equipment:
   EITHER
   a) comes from a country or zone free from Aethina tumida infestation; and
   b) contains no live honey bees or bee brood;
   OR
   c) contains no live honey bees or bee brood;
   d) has been thoroughly cleaned, and treated to ensure the destruction of Aethina tumida spp., in
      conformity with one of the procedures referred to in Appendix XXX (under study); and
2. all precautions have been taken to prevent infestation/contamination.

   Article 2.9.X.9.

Veterinary Authorities of importing countries should require:

for honey-bee collected pollen and beeswax (in the form of honeycomb)

the presentation of an international veterinary certificate attesting that:

1. the products:
   EITHER
   a) comes from a country or zone free from Aethina tumida infestation; and
   b) contains no live honey bees or bee brood;
   OR
   c) contains no live honey bees or bee brood;
   d) has been thoroughly cleaned, and treated to ensure the destruction of Aethina tumida spp., in
      conformity with one of the procedures referred to in Appendix XXX (under study); and
2. all precautions have been taken to prevent infestation/contamination.
Article 2.9.X.10.

Veterinary Authorities of importing countries should require:

for comb honey

the presentation of an international veterinary certificate attesting that the products:

1. comes from a country or zone free from Aethina tumida infestation; and
2. contains no live honey bees or bee brood.