

OIE Reference Laboratory Reports Activities

Activities in 2019

This report has been submitted : 2020-02-06 04:02:35

Name of disease (or topic) for which you are a designated OIE Reference Laboratory:	Infection with ranavirus
Address of laboratory:	Node 1. 425 Werombi Road Private Bag 3 Camden NSW 2570 AUSTRALIA Node 2. 5 Portarlinton Road Private Bag 24 Geelong, Victoria 3220 AUSTRALIA
Tel.:	+61-2 93 51 16 19; +
Fax:	+61-2 93 51 16 18; +
E-mail address:	paul.hick@sydney.edu.au
Website:	www.csiro.au; https://sydney.edu.au/science/schools/sydney-school-of-veterinary-science.html
Name (including Title) of Head of Laboratory (Responsible Official):	Node 1. Professor Frazer Allan - Head of School Node 2. Prof Trevor Drew -Director
Name (including Title and Position) of OIE Reference Expert:	Node 1. Dr Paul Hick, Senior lecturer veterinary virology, Sydney School of Veterinary Science Node 2. Dr Nick Moody, AAHL Fish Diseases Laboratory Group Leader
Which of the following defines your laboratory? Check all that apply:	Governmental Academic

ToR 1: To use, promote and disseminate diagnostic methods validated according to OIE Standards

1. Did your laboratory perform diagnostic tests for the specified disease/topic for purposes such as disease diagnosis, screening of animals for export, surveillance, etc.? (Not for quality control, proficiency testing or staff training)

Yes

Diagnostic Test	Indicated in OIE Manual (Yes/No)	Total number of test performed last year	
		Nationally	Internationally
Indirect diagnostic tests			
none	n/a	0	0
Direct diagnostic tests			
Ranavirus qPCR	No	132	0

ToR 2: To develop reference material in accordance with OIE requirements, and implement and promote the application of OIE Standards. To store and distribute to national laboratories biological reference products and any other reagents used in the diagnosis and control of the designated pathogens or disease.

2. Did your laboratory produce or supply imported standard reference reagents officially recognised by the OIE?

No

3. Did your laboratory supply standard reference reagents (non OIE-approved) and/or other diagnostic reagents to OIE Member Countries?

No

4. Did your laboratory produce vaccines?

No

5. Did your laboratory supply vaccines to OIE Member Countries?

No

ToR 3: To develop, standardise and validate, according to OIE Standards, new procedures for diagnosis and control of the designated pathogens or diseases

6. Did your laboratory develop new diagnostic methods validated according to OIE Standards for the designated pathogen or disease?

No

7. Did your laboratory develop new vaccines according to OIE Standards for the designated pathogen or disease?

No

ToR 4: To provide diagnostic testing facilities, and, where appropriate, scientific and technical advice on disease control measures to OIE Member Countries

8. Did your laboratory carry out diagnostic testing for other OIE Member Countries?

No

9. Did your laboratory provide expert advice in technical consultancies on the request of an OIE Member Country?

No

ToR 5: To carry out and/or coordinate scientific and technical studies in collaboration with other laboratories, centres or organisations

10. Did your laboratory participate in international scientific studies in collaboration with OIE Member Countries other than the own?

No

ToR 6: To collect, process, analyse, publish and disseminate epizootiological data relevant to the designated pathogens or diseases

11. Did your Laboratory collect epizootiological data relevant to international disease control?

Yes

If the answer is yes, please provide details of the data collected:
Surveillance for freedom from infection

12. Did your laboratory disseminate epizootiological data that had been processed and analysed?

Yes

If the answer is yes, please provide details of the data collected:

Submitted for publication: Whinfield et al., Alien smooth newts (*Lissotriton vulgaris*) in Australia are infected with Chytrid fungus, but test negative for ranaviruses

13. What method of dissemination of information is most often used by your laboratory? (Indicate in the appropriate box the number by category)

a) Articles published in peer-reviewed journals: 1

Laurin, E., Thakur, K., Mohr, P.G., Hick, P., Crane, M.S.J., Gardner, I.A., Moody, N.J.G., Colling, A., Ernst, I., 2019. To pool or not to pool? Guidelines for pooling samples for use in surveillance testing of infectious diseases in aquatic animals. *Journal of Fish Diseases* 42, 1471-1491.

b) International conferences: 1

5th International Symposium on Ranaviruses, June 4th-6th Townsville, Australia

- CURRENT STATUS OF TAXONOMY WITHIN THE FAMILY IRIDOVIRIDAE. ICTV Iridovirus Study Group: VG Chinchar, P Hick, J Huang³, IA Ince, JK Jancovich, R Marschang, Q Qin, K Subramaniam, TB Waltzek⁸, R Whittington², T Williams, QY Zhang.

c) National conferences: 1

Wildlife Disease Association (WDA) Australasian Section Conference, Tasmania 29th September - 4th October 2019.

Jessica Whinfield, Reid Tingley, Alison Tweedie, Jasmin Hufschmid and Paul Hick. Identification of Pathogens Associated with Exotic Smooth Newts (*Lissotriton vulgaris*) in Australia

d) Other:

(Provide website address or link to appropriate information) 0

ToR 7: To provide scientific and technical training for personnel from OIE Member Countries

To recommend the prescribed and alternative tests or vaccines as OIE Standards

14. Did your laboratory provide scientific and technical training to laboratory personnel from other OIE Member Countries?

No

ToR 8: To maintain a system of quality assurance, biosafety and biosecurity relevant for the pathogen and the disease concerned

15. Does your laboratory have a Quality Management System?

Yes

Quality management system adopted	Certificate scan (PDF, JPG, PNG format)
Node 1. ISO 17025(2017)	NATA accreditation 2019.pdf
Node 2. ISO 17025	
Node 2. ISO 9001; ISO 14001; ISO 17043	

16. Is your quality management system accredited?

Yes

Test for which your laboratory is accredited	Accreditation body
Node 1. Ranavirus qPCR	NATA
Node 2. 20.10 Microbiology For companion animals, production animals, production avian species, zoo animals, wildlife, aquatic animals, equine species and avian species	NATA (ILAC affiliated)
Node 2. 20.11 Bacteriology 01 Diagnostic bacteriology - incorporating identification by simple microscopy, cultural methods of detection and identification of organisms 03 Immunological methods of antigen detection	NATA (ILAC affiliated)
Node 2. 20.13 Other Microorganisms 01 Diagnostic microbiology - incorporating identification by simple microscopy, cultural methods of detection and identification of organisms, including Innocuity testing	NATA (ILAC affiliated)
Node 2. 20.14 Virology 01 Diagnostic virology - non-cultural (immunological) methods of detection 02 Diagnostic virology - cultural methods of detection and identification of organisms Including Innocuity testing 05 Quantitative procedures	NATA (ILAC affiliated)
Node 2. 20.15 Prions 01 Histological identification of prion disease lesions 02 Detection of prion protein by immunological methods (including ELISA, Western Blots, immunohistochemistry) 04 Detection of prion protein by bioassay	NATA (ILAC affiliated)
Node 2. 20.25 Serology of Infection For companion animals, production animals, production avian species, zoo animals, wildlife, equine species and avian species 01 Agar gel immunodiffusion tests 02 Complement fixation tests 03 Enzyme linked immunosorbent assays 04 Haemagglutination inhibition 05 Indirect fluorescent antibody tests 06 Microscopic agglutination tests 08 Serum agglutination tests 09 Serum neutralisation tests 10 Latex agglutination tests 99 Other - Testing for rabies and rabies related lyssaviruses on human specimens	NATA (ILAC affiliated)
Node 2. 20.50 Anatomical Pathology For companion animals, production animals, production avian species, laboratory animals, zoo animals, wildlife, aquatic animals, equine species and avian species	NATA (ILAC affiliated)
Node 2. 20.52 Histopathology 01 Processing of fixed specimens for histology 04 Immunohistochemistry 05 Histological interpretation	NATA (ILAC affiliated)
Node 2. 20.53 Electron Microscopy 01 Transmission electron microscopy 02 Scanning electron microscopy 04 Immunohistochemistry electron microscopy	NATA (ILAC affiliated)
Node 2. 20.54 Necropsy	NATA (ILAC affiliated)
Node 2. 20.80 Molecular Diagnostics For companion animals, production animals, production avian species, aquatic animals, equine species and avian species 01 Identification by extraction and amplification 02 Sequencing 03 Genotyping 99 Other - Testing for rabies and rabies related lyssaviruses on human specimens by molecular techniques	NATA (ILAC affiliated)
Node 2. 20.95 Foreign Regulatory Requirements 01 European Union Directives for Animal Health Council Directive 88/407/EEC of 14 June 1988 Council Directive 64/432/EEC of 26 June 1964 Commission Implementing Decision 2011/630/EU of 20 September 2011 Council Directive 89/556/EEC of 25 September 1989 Commission Decision 2006/168/EC of 4 January 2006 Council Directive 91/68/EEC of 28 January 1991 Council Directive 92/65/EEC of 13 July 1992 Commission Decision 2010/472/EU of 26 August 2010 Commission Decision 2004/211/ED of 6 January 2004 Commission Decision 2010/471/EU of 26 August 2010 For the following species for the following diseases using the following methods of testing: Ovine - EHD, c-ELISA, SNT	NATA (ILAC affiliated)

Node 2. 1.12 Weighing devices [In-House Calibration] 01 Precision laboratory balances [In-House Calibration] with least uncertainties of measurement of - 5 in 10 ⁶ or 56 µg (whichever is greater) up to 3 kg	NATA (ILAC affiliated)
Node 2. 1.80 Calibration of temperature measuring equipment [In-House Calibration] 41 Digital temperature indicator systems [In-House Calibration] with least uncertainties of measurement of - 0.5°C from -20 to 125°C	NATA (ILAC affiliated)
Node 2. 1.84 Testing of controlled enclosures [In-House Calibration] 02 Incubators [In-House Calibration] with least uncertainties of measurement of - 0.5°C from 0 to 125°C by the methods of - AS 2853 03 Autoclaves and sterilising ovens [In-House Calibration] with least uncertainties of measurement of - 0.5°C from 0 to 125°C	NATA (ILAC affiliated)
Node 2. 13.69 Controlled environments [In-House Calibration] by the methods of - AS 1807.1, .5, .6, .22, .23 AS/NZS 2243.8 Appendices A and B 01 Clean rooms and workstations [In-House Calibration] .02 Biological safety cabinets [In-House Calibration] .03 Fume cupboards [In-House Calibration]	NATA (ILAC affiliated)
Node 2. 42.01 Human and Veterinary Pathology Services Bacteria – Serology of infection Bacteria – Molecular diagnostics – Identification by Extraction and Amplification Bacteria – Detection and Identification of virus antigen Parasites- Serology of infection Parasites – Molecular diagnostics – Identification by Extraction and Amplification Parasites – Detection and identification of virus antigen Fungi and Yeast – Serology of infection Fungi and Yeast – Molecular diagnostics – Identification by Extraction and Amplification Fungi and Yeast – Detection and identification of virus antigen Molecular Diagnostics – Identification by Extraction and Amplification Blood and Blood products – Serology of infection	NATA (ILAC affiliated)
Node 2. Accreditation No: 13546 (Scope Last Changed 08/12/14)	NATA (ILAC affiliated)

17. Does your laboratory maintain a “biorisk management system” for the pathogen and the disease concerned?

Yes

(See *Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, Chapter 1.1.4*)

ToR 9: To organise and participate in scientific meetings on behalf of the OIE

18. Did your laboratory organise scientific meetings on behalf of the OIE?

No

19. Did your laboratory participate in scientific meetings on behalf of the OIE?

Yes

Title of event	Date (mm/yy)	Location	Role (speaker, presenting poster, short communications)	Title of the work presented
OIE Global Conference on Aquatic Animal Health	2-4 April, 2016	Santiago, Chile	Dr Nick Moody, Attendee	n/a
Regional Side Meeting on aquatic animal health	4 April, 2019	Santiago, Chile	Dr Nick Moody, Speaker	OIE Regional Collaboration Framework - draft plan
Regional Seminar for OIE National Focal Points for Veterinary Laboratories	17-18 June, 2019	Chiang Mai, Thailand	Dr Nick Moody, Invited speaker	Sustainability in aquatic laboratories
13th OIE Seminar			Dr Nick Moody,	
19th WAVLD Symposium			Dr Nick Moody,	

ToR 10: To establish and maintain a network with other OIE Reference Laboratories designated for the same pathogen or disease and organise regular inter-laboratory proficiency testing to ensure comparability of results

20. Did your laboratory exchange information with other OIE Reference Laboratories designated for the same pathogen or disease?

No

21. Was your laboratory involved in maintaining a network with OIE Reference Laboratories designated for the same pathogen or disease by organising or participating in proficiency tests?

No

22. Did your laboratory collaborate with other OIE Reference Laboratories for the same disease on scientific research projects for the diagnosis or control of the pathogen of interest?

No

ToR 11: To organise inter-laboratory proficiency testing with laboratories other than OIE Reference Laboratories for the same pathogens and diseases to ensure equivalence of results

23. Did your laboratory organise or participate in inter-laboratory proficiency tests with laboratories other than OIE Reference Laboratories for the same disease?

No

Note: See Interlaboratory test comparisons in: Laboratory Proficiency Testing at:
<http://www.oie.int/en/our-scientific-expertise/reference-laboratories/proficiency-testing> see point 1.3

ToR 12: To place expert consultants at the disposal of the OIE

24. Did your laboratory place expert consultants at the disposal of the OIE?

Yes

Kind of consultancy	Location	Subject (facultative)
Dr Nick Moody, ad hoc Group member	email	ad hoc Group on tilapia lake virus
Dr Nick Moody, ad hoc Group member	email	ad hoc Group on the OIE Manual of Diagnostic Tests for Aquatic Animals

25. Additional comments regarding your report: