OIE Reference Laboratory Reports Activities

Activities in 2020

This report has been submitted : 2021-02-15 11:37:35

Name of disease (or topic) for which you are a designated OIE Reference Laboratory:	Epizootic haematopoietic necrosis
Address of laboratory:	Co-host1: 425 Werombi Road Private Bag 3 Camden NSW 2570 AUSTRALIA AUSTRALIA Co-host2: 5 Portarlington Road Private Bag 24 Geelong, Victoria 3220 AUSTRALIA
Tel.:	+61-2 93 51 16 19; +
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E-mail address:	paul.hick@sydney.edu.au
Website:	www.csiro.au
Name (including Title) of Head of Laboratory (Responsible Official):	Prof Trevor Drew -Director
Name (including Title and Position) of OIE Reference Expert:	Co-host1: Associate Professor Paul Hick, Sydney School of Veterinary Science Co-host2: Dr Nick Moody ACDP Fish Diseases Laboratory Group Leader
Which of the following defines your laboratory? Check all that apply:	Governmental Academic

ToR 1: To use, promote and disseminate diagnostic methods validated according to OIE Standards

1. Did your laboratory perform diagnostic tests for the specified disease/topic for purposes such as disease diagnosis, screening of animals for export, surveillance, etc.? (Not for quality control, proficiency testing or staff training)

Yes

Diagnostic Test	Indicated in OIE Manual (Yes/No)	Total number of test performed last year	
Indirect diagnostic tests		Nationally	Internationally
none	yes	0	0
Direct diagnostic tests		Nationally	Internationally
Ranavirus qPCR	No	35	0
Ranavirus qPCR	No	200	0

ToR 2: To develop reference material in accordance with OIE requirements, and implement and promote the application of OIE Standards.
To store and distribute to national laboratories biological reference products and any other reagents used in the diagnosis and control of the designated pathogens or disease.

2. Did your laboratory produce or supply imported standard reference reagents officially recognised by the OIE?

No

3. Did your laboratory supply standard reference reagents (non OIE-approved) and/or other diagnostic reagents to OIE Member Countries?

Yes

Type of reagent available	Related diagnostic test	Produced/ provide	Amount supplied nationally (ml, mg)	Amount supplied internationally (ml, mg)	No. of recipient OIE Member Countries	Region of recipients
Genomic DNA	molecular assays	provided	0	2 x 50 ul	1	☐Africa ☒Americas ☐Asia and Pacific ☐Europe ☐Middle East
Ethanol-fixed cell culture supernatant	Molecular testing	provided	0	2 x 500 ul	1	☐Africa ☒Americas ☐Asia and Pacific ☐Europe ☐Middle East

4. Did your laboratory produce vaccines?

No

5. Did your laboratory supply vaccines to OIE Member Countries?

No

ToR 3: To develop, standardise and validate, according to OIE Standards, new procedures for diagnosis and control of the designated pathogens or diseases

6. Did your laboratory develop new diagnostic methods validated according to OIE Standards for the designated pathogen or disease?

No

7. Did your laboratory develop new vaccines according to OIE Standards for the designated pathogen or disease?

No

ToR 4: To provide diagnostic testing facilities, and, where appropriate, scientific and technical advice on disease control measures to OIE Member Countries

8. Did your laboratory carry out diagnostic testing for other OIE Member Countries?

No

9. Did your laboratory provide expert advice in technical consultancies on the request of an OIE Member Country?

No

ToR 5: To carry out and/or coordinate scientific and technical studies in collaboration with other laboratories, centres or organisations

10. Did your laboratory participate in international scientific studies in collaboration with OIE Member Countries other than the own?

No

ToR 6: To collect, process, analyse, publish and disseminate epizootiological data relevant to the designated pathogens or diseases

11. Did your Laboratory collect epizootiological data relevant to international disease control?

No

If the answer is no, please provide a brief explanation of the situation:
No. Reduced laboratory testing capability and freight transport nationally and internationally due to COVID-19. Change in University funding model and removal of institutional support for laboratory activities(co-host 1).

12. Did your laboratory disseminate epizootiological data that had been processed and analysed?

No

If the answer is no, please provide a brief explanation of the situation:
No. Reduced laboratory testing capability and freight transport nationally and internationally due to COVID-19. Change in University funding model and removal of institutional support for laboratory activities(co-host 1).

13. What method of dissemination of information is most often used by your laboratory? (Indicate in the appropriate box the number by category)

a) Articles published in peer-reviewed journals: 0

b) International conferences: 0

c) National conferences: 0

d) Other:
(Provide website address or link to appropriate information) 0

ToR 7: To provide scientific and technical training for personnel from OIE Member Countries
To recommend the prescribed and alternative tests or vaccines as OIE Standards

14. Did your laboratory provide scientific and technical training to laboratory personnel from other OIE Member Countries?

No

ToR 8: To maintain a system of quality assurance, biosafety and biosecurity relevant for the pathogen and the disease concerned

15. Does your laboratory have a Quality Management System?

Yes

Quality management system adopted	Certificate scan (PDF, JPG, PNG format)
ISO 17025	18444 20300 70560 nl.pdf
ISO 9001	Co-host 2 accrediation.pdf
ISO 14001	Co-host 2 accrediation.pdf

16. Is your quality management system accredited?

Yes

Test for which your laboratory is accredited	Accreditation body
20.10 Microbiology For companion animals, production animals, production avian species, zoo animals, wildlife, aquatic animals, equine species and avian species	NATA (ILAC affiliated)
20.11 Bacteriology 01 Diagnostic bacteriology - incorporating identification by simple microscopy, cultural methods of detection and identification of organisms 03 Immunological methods of antigen detection	NATA (ILAC affiliated)
20.13 Other Microorganisms 01 Diagnostic microbiology - incorporating identification by simple microscopy, cultural methods of detection and identification of organisms, including Innocuity testing	NATA (ILAC affiliated)
20.14 Virology 01 Diagnostic virology - non-cultural (immunological) methods of detection 02 Diagnostic virology - cultural methods of detection and identification of organisms Including Innocuity testing 05 Quantitative procedures	NATA (ILAC affiliated)
20.15 Prions 01 Histological identification of prion disease lesions 02 Detection of prion protein by immunological methods (including ELISA, Western Blots, immunohistochemistry) 04 Detection of prion protein by bioassay	NATA (ILAC affiliated)
20.25 Serology of Infection For companion animals, production animals, production avian species, zoo animals, wildlife, equine species and avian species 01 Agar gel immunodiffusion tests 02 Complement fixation tests 03 Enzyme linked immunosorbent assays 04 Haemagglutination inhibition 05 Indirect fluorescent antibody tests 06 Microscopic agglutination tests 08 Serum agglutination tests 09 Serum neutralisation tests 10 Latex agglutination tests 99 Other - Testing for rabies and rabies related lyssaviruses on human specimens	NATA (ILAC affiliated)
20.50 Anatomical Pathology For companion animals, production animals, production avian species, laboratory animals, zoo animals, wildlife, aquatic animals, equine species and avian species	NATA (ILAC affiliated)
20.52 Histopathology 01 Processing of fixed specimens for histology 04 Immunohistochemistry 05 Histological interpretation	NATA (ILAC affiliated)
20.53 Electron Microscopy 01 Transmission electron microscopy 02 Scanning electron microscopy 04 Immunohistochemistry electron microscopy	NATA (ILAC affiliated)
20.54 Necropsy	NATA (ILAC affiliated)
20.80 Molecular Diagnostics For companion animals, production animals, production avian species, aquatic animals, equine species and avian species 01 Identification by extraction and amplification 02 Sequencing 03 Genotyping 99 Other - Testing for rabies and rabies related lyssaviruses on human specimens by molecular techniques	NATA (ILAC affiliated)
20.95 Foreign Regulatory Requirements 01 European Union Directives for Animal Health Council Directive 88/407/EEC of 14 June 1988 Council Directive 64/432/EEC of 26 June 1964 Commission Implementing Decision 2011/630/EU of 20 September 2011 Council Directive 89/556/EEC of 25 September 1989 Commission Decision 2006/168/EC of 4 January 2006 Council Directive 91/68/EEC of 28 January 1991 Council Directive 92/65/EEC of 13 July 1992 Commission Decision 2010/472/EU of 26 August 2010 Commission Decision 2004/211/ED of 6 January 2004 Commission Decision 2010/471/EU of 26 August 2010 For the following species for the following diseases using the following methods of testing: Ovine - EHD, c-ELISA, SNT	NATA (ILAC affiliated)
1.12 Weighing devices [In-House Calibration] 01 Precision laboratory balances [In-House Calibration] with least uncertainties of measurement of - 5 in 10^6 or 56 µg (whichever is greater) up to 3 kg	NATA (ILAC affiliated)

1.80 Calibration of temperature measuring equipment [In-House Calibration] 41 Digital temperature indicator systems [In-House Calibration] with least uncertainties of measurement of - 0.5ºC from -20 to 125ºC	NATA (ILAC affiliated)
1.84 Testing of controlled enclosures [In-House Calibration] 02 Incubators [In-House Calibration] with least uncertainties of measurement of - 0.5ºC from 0 to 125ºC by the methods of - AS 2853 03 Autoclaves and sterilising ovens [In-House Calibration] with least uncertainties of measurement of - 0.5ºC from 0 to 125ºC	NATA (ILAC affiliated)
13.69 Controlled environments [In-House Calibration] by the methods of - AS 1807.1, .5, .6, .22, .23 AS/NZS 2243.8 Appendices A and B 01 Clean rooms and workstations [In-House Calibration] .02 Biological safety cabinets [In-House Calibration] .03 Fume cupboards [In-House Calibration]	NATA (ILAC affiliated)
42.01 Human and Veterinary Pathology Services Bacteria – Serology of infection Bacteria – Molecular diagnostics – Identification by Extraction and Amplification Bacteria – Detection and Identification of virus antigen Parasites-Serology of infection Parasites – Molecular diagnostics – Identification by Extraction and Amplification Parasites – Detection and identification of virus antigen Fungi and Yeast – Serology of infection Fungi and Yeast – Molecular diagnostics – Identification by Extraction and Amplification Fungi and Yeast – Detection and identification of virus antigen Molecular Diagnostics – Identification by Extraction and Amplification Blood and Blood products – Serology of infection	NATA (ILAC affiliated)

17. Does your laboratory maintain a “biorisk management system” for the pathogen and the disease concerned?

Yes

(See Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, Chapter 1.1.4)

ToR 9: To organise and participate in scientific meetings on behalf of the OIE

18. Did your laboratory organise scientific meetings on behalf of the OIE?

No

19. Did your laboratory participate in scientific meetings on behalf of the OIE?

Yes

Title of event	Date (mm/yy)	Location	Role (speaker, presenting poster, short communications)	Title of the work presented
2nd meeting of the ad hoc Steering Committee Members of the Regional Collaboration Framework on Aquatic Animal Health	03/2020	Online	Particpant (Moody)	General discussions
3rd meeting of the ad hoc Steering Committee Members of the Regional Collaboration Framework on Aquatic Animal Health	12/2020	Online	Speaker (Moody)	Reference Laboratory report

ToR 10: To establish and maintain a network with other OIE Reference Laboratories designated for the same pathogen or disease and organise regular inter-laboratory proficiency testing to ensure comparability of results

20. Did your laboratory exchange information with other OIE Reference Laboratories designated for the same pathogen or disease?

No

21. Was your laboratory involved in maintaining a network with OIE Reference Laboratories designated for the same pathogen or disease by organising or participating in proficiency tests?

No

22. Did your laboratory collaborate with other OIE Reference Laboratories for the same disease on scientific research projects for the diagnosis or control of the pathogen of interest?

No

ToR 11: To organise inter-laboratory proficiency testing with laboratories other than OIE Reference Laboratories for the same pathogens and diseases to ensure equivalence of results

23. Did your laboratory organise or participate in inter-laboratory proficiency tests with laboratories other than OIE Reference Laboratories for the same disease?

No

Note: See Interlaboratory test comparisons in: Laboratory Proficiency Testing at: http://www.oie.int/en/our-scientific-expertise/reference-laboratories/proficiency-testing see point 1.3

ToR 12: To place expert consultants at the disposal of the OIE

24. Did your laboratory place expert consultants at the disposal of the OIE?

Yes

Kind of consultancy	Location	Subject (facultative)
Dr Nick Moody, ad hoc Group member	email	ad hoc Group on tilapia lake virus
Dr Nick Moody, ad hoc Group member	email	ad hoc Group on the OIE Manual of Diagnostic Tests for Aquatic Animals

25. Additional comments regarding your report:

Due to COVID-19, ACDP has worked on limited operational capacity since March 2020 (for example, adopting roster arrangements for staff site access, reduced site access to ensure physical distancing, no domestic or international travel and visitors unable to attend site for most of the year). This has significantly limited ACDP's capacity to carry out planned research and conduct training and has limited some types of diagnostic submissions to the laboratory.
Changed university funding and priorities has reduced the capacity for co-host 1 to undertake reference laboratory activities.